CTS Immune Cell SR for Serum Free Culture and Expansion of Human T Cells



Angel Varela-Rohena¹, Karoline W. Schjetne², Andrew Medvec³, James L. Riley³, Sandy Kuligowski¹, Grethe Økern², Tanja Aarvak² Thermo Fisher Scientific, ¹Grand Island, NY and ²Oslo, Norway; ³University of Pennsylvania, Philadelphia, PA

INTRODUCTION

The manufacture of a majority of clinical T cell products for immunotherapy applications requires *in vitro* T cell culture and expansion. Commercialization of T cell manufacturing processes requires reagents that meet regulatory guidelines and ultimately help reduce manufacturing cost of goods. A key component in many T cell culture protocols, in addition to cell culture media and growth factors, is human serum. Human serum is expensive and requires extensive testing prior to use for manufacturing of a cGMP-compliant T cell product. To this end, we have tested a xeno-free serum replacement, CTS™ Immune Cell SR. CTS Immune Cell SR contains only defined components and can be used in combination with several different cell culture media to support *in vitro* culture and expansion of T cells.

EXPANSION OF OKT3-ACTIVATED T CELLS

METHODS

Polyclonal T cells were negatively isolated from fresh PBMC, activated *in vitro* with OKT3 mAb, irradiated pooled feeder cells and high dose IL-2 and expanded for two weeks.

Cell culture media tested were CTS™ OpTmizer™ T Cell Expansion SFM, Lonza X-VIVO™ 15 and CTS AIM-V Media.

Day 0 Isolate CD3+ T cells and activate	Day 6 Split and feed	Day9 Splitand feed	Day 11 Split and feed	Day 13 Analyse
1x10 ⁶ CD3 isolated polyclonal T cells 1x10 ⁷ irradiated pooled feeder cells 30 ngiml OKT3 30000 UI-2/mL in 5 ml static cultures (6-well plate)				Count cells Flow analys

RESULTS

Total Fold Expansion	Fold Expansion of CD4 T Cells	Fold Expansion of CD8 T Cells			
bilitorias bilingenia di polarizza standarda 33 tiple dansi ang polarizza di naga tiple dansi 32 tiple dansi ang polarizza di naga tiple dansi di naga tiple dansi ang polarizza di naga tiple dansi d		Adder tabilit expense of CC 7 mb			

T cells expanded in cultures supplemented with pooled human serum or CTS Immune Cell SR show similar growth kinetics and total fold expansion after 2 weeks in culture. Both CD4+ and CD8+ T cell subsets are expanded.

EXPANSION OF CTS™ DYNABEADS™ CD3/CD28 ISOLATED AND ACTIVATED T CELLS

METHODS

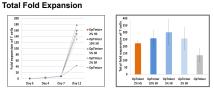
Polyclonal T cells from fresh PBMC were isolated and activated with Dynabeads CD3/CD28 CTS and expanded for two weeks.

Cell culture media tested were CTS OpTmizer T Cell Expansion SFM and Lonza X-VIVO™ 15 (not shown).

Cell culture media were supplemented with either pooled AB human serum or 10% CTS Immune Cell SR.

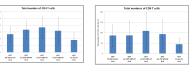


RESULTS



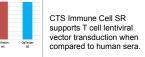
T cells expanded in CTS OpTmizer supplemented with human serum or CTS Immune Cell SR show similar growth kinetics and total fold expansion after two weeks in culture. Left panel: Growth kinetics from 1 representative donor. Right panel: Fold expansion of total T cells at the end of culture (day 12). Bars represent averages of 4 normal donors.

CTS Immune Cell SR supports expansion of CD4 and CD8 T cells



Lentiviral transduction

NT N= 2



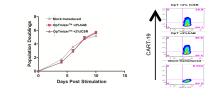
CTS™ IMMUNE CELL SR EXPANDED CART-19 T CELLS ARE EFFECTIVE *IN VIVO*

METHODS

Human CD4+ and CD8+ T cells were procured from the Human Immunology Core at the University of Pennsylvania and stimulated with Dynabeads CD3/CD28 CTS in CTS OpTmizer SFM supplemented with 2% pooled human AB serum or 2% CTS Immune Cell SR and transduced with CART-19 lentiviral vectors. 6 —10 week ddNOD-SCID-Yc^-/- (NSG) mice were bred in-house under an approved IACUC protocol and maintained in pathogenfree conditions. Animals were injected I.V. via tail vein with 10^o6 Nalm-6 leukemia followed by 10^o6 T cells injected 7 days after. Animals were monitored for signs of high tumor burden by luciferase bioluminescence.

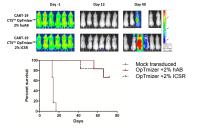
RESULTS

Comparable expansion and transduction of CART-19 cells in human AB serum and CTS Immune Cell SR



CART-19 cells grown with CTS Immune Cell SR supplementation expanded to comparable yields (left panel) and transduction efficiencies (right panel) when compared to cells grown in human serum. Mock transduced cells shown as negative control.

In vivo efficacy of CART-19 cells expanded in CTS Immune Cell SR



CART-19 cells grown in CTS OpTmizer with CTS Immune Cell SR supplementation showed similar potency and efficacy as control CART-19 grown in CTS OpTmizer with human serum as measured by tumor burden bioluminescence (top) and animal survival (bottom panel).

CONCLUSIONS

- CTS Immune Cell SR supports expansion of Dynabeads CD3/CD28 CTS-activated polyclonal T cells and REM – expanded T cells when supplemented into commonly used T cell culture media.
- CTS Immune Cell SR supports lentiviral transduction and expansion of gene-modified T cells, and maintains favourable immune function and *in vivo* efficacy of CAR-T cells.

REFERENCES

 Ex vivo expansion of human T cells for adoptive immunotherapy using the novel Xeno-free CTS Immune Cell Serum Replacement. Smith C et al. *Clin Transl Immunology* 2015 Jan 15; 4

TRADEMARKS/LICENSING

©2015 Thermo Fisher Scientific. All rights reserved. All trademarks are the property of Thermo Fisher Scientific and its subsidiaries unless otherwise specified.

