Evaluation and Comparison of Liquid Biopsy Reference Materials from Commercial Sources using Oncomine™ Pan-Cancer Cell-free Assay

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Over the last few years, analysis of liquid biopsies has grown rapidly to interrogate cell-free nucleic acid and characterize tumor mutation profiles from plasma. To date, a number of liquid biopsy research assays utilizing next generation sequencing technologies have been made commercially available to detect various types of variants at low allele frequency with minimal sample input. To ensure each assay is performed consistently and adopted successfully by researchers, reference materials are essential in assay validation and can be used as standards to compare technologies and standardize protocols across different laboratories.

MATERIALS AND METHODS

Reference material

Multiplex cfDNA reference standard set: this standard set (Horizon Discovery, Cat# HD780) contains 4 contrived DNA samples which mimic cfDNA type of material. Each sample covers 8 engineered single nucleotide variants (SNVs/SNPs) with allele frequency at 5%, 1%, 0.1%, and 0% (100% wild type), respectively.

Structural Multiplex cfDNA Reference Standard: this standard (Horizon Discovery, Cat# HD786) contains RET and ROS1 fusion variants, MYC and MET amplifications.

Seraseq® cfDNA completeTM mutation mix: four ctDNA mutation mix samples were provided by Seracare including wild type (Cat# 0710-0533), 1% (Cat# 0710-0530), 0.5% (Cat# 0710-0531), and 0.1% (Cat# 0710-0532). Each sample contains 25 unique multiplexed variants in 16 genes, covering 12 SNVs, 7 INDELs, 3 CNVs, and 3 SVs at different allele frequencies.

Seraseg® fusion RNA mix: this contrived fusion standard (Cat# 0710-0431) contains 16 clinically relevant RNA fusions mixed in total RNA from GM24385 cell line as background. In this study, it was further diluted to 10%, 5%, and 2% using cfTNA from healthy donor as background to generate a serial of standards with different copy numbers of fusion variants for limit of detection (LOD) evaluation.

Library preparation and sequencing:

Targeted libraries were prepared using Oncomine[™] Pan-Cancer Cellfree assay reagents (Thermo Fisher Scientific, Cat# A37664).

For Horizon multiplex cfDNA reference standard, 25 ng was used as input in library preparation. In addition, input amount for 0.1% standard was increased to 50 ng to mimic 6000 copies of DNA allowing for improved sensitivity at lower LOD.

For Seraseq® cfDNA completeTM mutation mix, 25 ng was used as input in library preparation. For the serial dilutions from Seraseq® fusion RNA mix in cfTNA background, 20 ng was used as input in library preparation.

The Ion 540[™] Kit-Chef was used for template preparation on Ion Chef[™] (Thermo Fisher Scientific, Cat# 4484177) and followed by sequencing on Ion GeneStudio S5 XL system (Thermo Fisher Scientific, Cat# A27214) using the Ion 540[™] Chip (Thermo Fisher Scientific, Cat# A27766).

Data analysis:

Data analysis was performed using Torrent Suite™ Software v5.6 and Ion Reporter™ v5.6 for simultaneous SNV/Indel, CNV, and fusion variant calls

Oncomine[™] Pan-Cancer Cell-free Assay Content

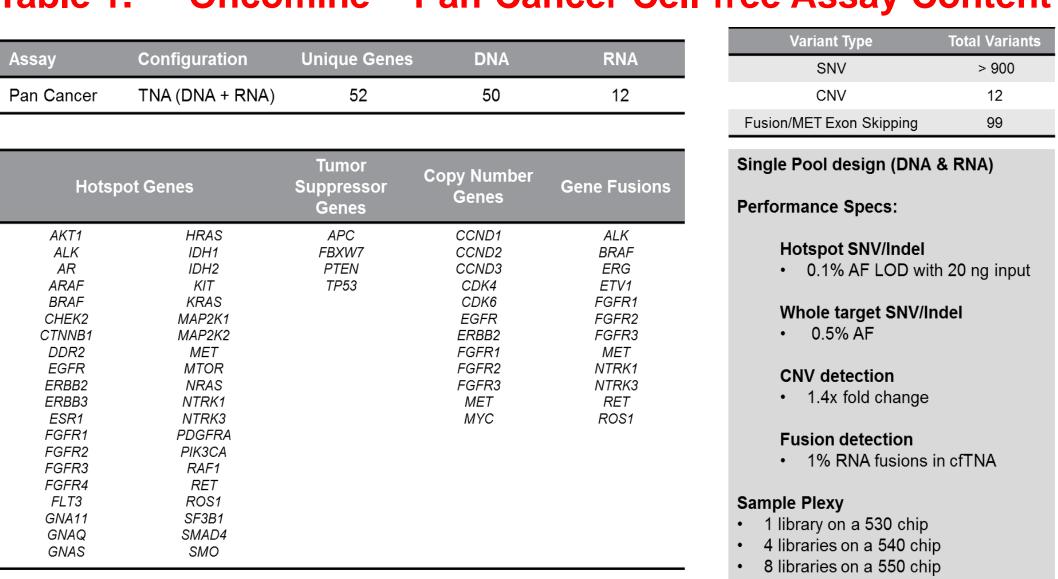
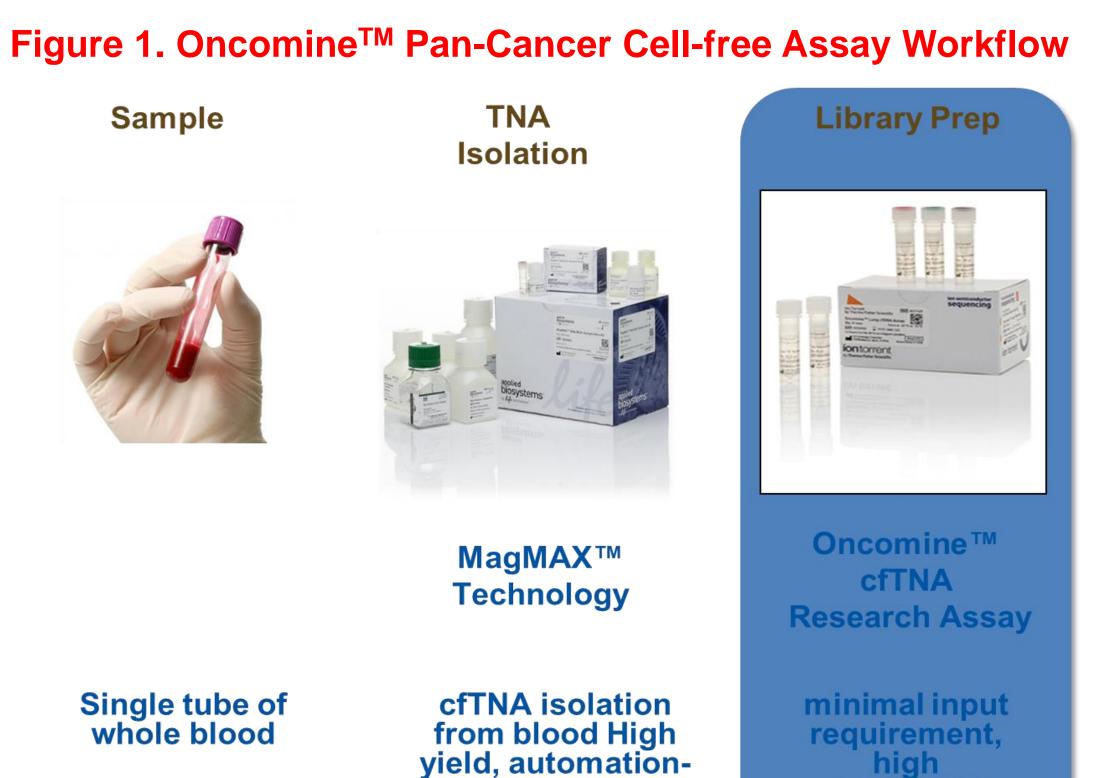


Table 1. Oncomine[™] Pan-Cancer cell-free assay provides broad coverage across 52 unique genes that detects DNA and RNA variants across multiple cancer types. The workflow also offers multiplexing flexibility utilizing different chip configurations available to GeneStudio S5 systems to accommodate the needs for desired throughput...

RESULTS

Blood Sample



ready.

Ion Torrent™ Chef™/S5™ Ion S5™ Fast accurate sequencing, flexible throughput multiplexing

Templating &

Sequencing

Lab-created **Analysis** Report **Torrent Suite™ Oncomine**[™] Ion Reporter™ Knowledgebase **Detection of Annotation and** variants at reporting with compendium of onco-genomic

Custom Report

Figure 1. The OncomineTM Pan-Cancer cell-free assay enables a fast 3-day workflow that starts with cell-free nucleic acid purification from a single tube of blood and library preparation to interrogate both DNA and RNA in a single reaction. The amplified library will undergo high-throughput semiconductor sequencing, with results analyzed and reported through an integrated bioinformatics solution.

Table 2. Result Summary for Horizon Multiplex cfDNA **Reference Standards**

Sample			SNV / Indel			
	Gene	AA Chg	AF % (R1)	AF % (R2)	AF % (R3)	AF % (R4)
	CTNNB1	p.S33Y	24.0	24.3	26.7	24.2
Horizon	PIK3CA	p.H1047R	24.9	24.0	26.5	24.7
cfDNA RS WT		p.G719S	20.6	23.5	22.9	22.2
	BRAF	p.V600E	29.9	29.3	30.7	33.5
	MAP2K1	p.Q56P	22.4	23.7	23.7	22.4
Sample			SNV / Indel			
-	Gene	AA Chg	AF % (R1)	AF % (R2)	AF % (R3)	AF % (R4
	NRAS	p.Q61K	7.37	6.86	6.46	6.22
	NRAS	p.A59T	7.20	7.22	6.34	6.96
Horizon	PIK3CA	p.E545K	6.94	6.55	6.33	5.96
cfDNA RS 5%	EGFR	p.Q746 A750del	5.48	6.98	6.70	5.80
(25ng)	EGFR	p.M766 A767insASV	3.60	3.78	3.37	3.12
	EGFR	p.T790M	3.72	3.53	3.01	3.98
	EGFR	p.L858R	6.05	4.74	4.20	5.09
	KRAS	p.G12D	6.33	5.28	6.22	6.27
Sample		•	SNV / Indel			
-	Gene	AA Chg	AF % (R1)	AF % (R2)	AF % (R3)	AF % (R4
	NRAS	p.Q61K	1.12	1.47	1.15	1.46
	NRAS	p.A59T	1.46	0.78	1.77	1.19
Horizon	PIK3CA	p.E545K	1.10	1.13	1.52	1.12
cfDNA RS EGFR		p.Q746 A750del	1.12	1.53	1.45	1.71
1.0% (25ng)	EGFR	p.M766 A767insASV	0.74	0.62	0.69	0.68
	EGFR	p.T790M	0.81	0.80	0.69	0.58
	EGFR	p.L858R	0.57	0.86	0.80	0.94
	KRAS	p.G12D	0.90	1.43	1.24	1.49
Sample			SNV / Indel			
	Gene	AA Chg	AF % (R1)	AF % (R2)	AF % (R3)	AF % (R4
	NRAS	p.Q61K	ND	0.21	0.17	
	NRAS	p.A59T	ND	0.13	0.17	0.13
Horizon PIK3CA p		p.E545K	ND	0.11	0.19	0.15
		p.Q746_A750del	0.13	0.08	ND	ND
(25ng)	EGFR	p.M766_A767insASV	0.11	ND	0.08	0.21
	EGFR	p.T790M	0.20	0.16	0.11	0.15
	EGFR	p.L858R	ND	ND	ND	ND
	KRAS	p.G12D	ND	ND	0.09	ND
Sample		SNV / Ind	el			
	Gene	AA Chg	AF % (R1)	AF % (R2)]	
	NRAS	p.Q61K	ND	0.098		
	NRAS	p.A59T	0.096	0.117		
Horizon	PIK3CA	p.E545K	0.079	0.212		
cfDNA RS	EGFR	p.Q746_A750del	0.101	0.120		
0.1% (50ng)	EGFR	p.M766_A767insASV	0.073	ND		
	EGFR	p.T790M	0.193	0.117		
	EGFR	p.L858R	0.136	0.117		
	KRAS	p.G12D	ND	ND	ĺ	

Table 2. All 13 variants were successfully detected in Horizon Multiplex I cfDNA RS standards above 1% AF (25 ng input) using Pan-Cancer assay, including 8 SNV/Indels at several important gene loci and 5 additional SNVs from parental cell lines as shown in RS wild-type. For 0.1% RS, high DNA input at 50 ng that is equivalent 6000 copies of DNA molecules significantly improved variant detection sensitivity from ~73% to ~85% comparing to 25 ng input. No FPs were observed in any samples.

Table 3. Result Summary for Seraseq® cfDNA Complete™ **Mutation Mix**

Sample			SNV / Indel		
Seraseq ctDNA complete	Gene	AA Chg	R1 (%AF)	R2 (%AF)	R3 (%AF
mix WT		FP	1	2	2
	Gene	AA Chg	R1 (%AF)	R2 (%AF)	R3 (%AF
	NRAS	p.Q61R	1.09	1.33	0.92
	ALK	p.G1202R	1.14	1.20	1.34
	ALK	p.F1174L	0.74	1.26	1.05
	PIK3CA	p.H1047R	1.18	1.50	1.13
	KIT	p.D816V	1.45	0.88	1.26
	EGFR	p.E746_A750del	1.31	1.48	0.95
	EGFR	p.T790M	1.29	1.43	0.91
	EGFR	p.L858R	1.02	1.29	0.77
Seraseq ctDNA	BRAF	p.V600E	0.70	0.47	1.08
complete mix 1.0%	KRAS	p.Q61H	1.02	1.04	1.04
	KRAS	p.G12D	0.93	0.81	1.06
	KRAS	p.G12C	1.00	1.07	1.03
	EGFR	p.L747_P753>S	1.77	1.24	1.68
	EGFR	· —	1.16	1.03	1.11
		p.S752_I759del			
	ERBB2	p.E770_A771insAYVM	0.94	1.00	0.87
	AKT1	p.E17K	0.75	0.76	0.69
	TP53	p.G245D	0.37	0.66	0.44
		FP	1	2	2
	Gene	AA Chg	R1 (%AF)	R2 (%AF)	R3 (%AF
	NRAS	p.Q61R	0.70	0.68	0.54
	ALK	p.G1202R	0.50	0.79	0.50
	ALK	p.F1174L	0.50	0.60	0.46
	PIK3CA	p.H1047R	0.51	0.58	0.52
	KIT	p.D816V	0.88	0.82	0.91
	EGFR	p.E746_A750del	0.56	0.59	0.68
	EGFR	p.T790M	0.20	0.42	0.45
CaracasalDNA	EGFR	p.L858R	0.55	0.64	0.72
Seraseq ctDNA	BRAF	p.V600E	0.28	0.64	0.55
complete mix 0.5%	KRAS	p.Q61H	0.63	0.55	0.77
	KRAS	p.G12D	0.36	0.59	0.50
	KRAS	p.G12C	0.72	0.59	0.61
	EGFR	p.L747_P753>S	0.88	0.59	0.74
	EGFR	p.S752_I759del	0.88	0.59	0.59
	ERBB2	p.E770_A771insAYVM	0.18	0.28	0.20
	AKT1	p.E17K	0.53	0.65	0.52
		·			0.32
	TP53	p.G245D FP	0.41	0.39	2
	Cono		1	1	
	Gene	AA Chg	R1 (%AF)	R2 (%AF)	R3 (%AF
	NRAS	p.Q61R	0.24	0.13	0.21
	ALK	p.G1202R	0.10	0.15	0.12
	ALK	p.F1174L	0.09	ND	ND
	PIK3CA	p.H1047R	ND	ND	0.24
	KIT	p.D816V	0.09	0.09	ND
	EGFR	p.E746_A750del	ND	0.31	0.12
	EGFR	p.T790M	ND	0.15	ND
Seraseq ctDNA	EGFR	p.L858R	0.21	0.13	ND
complete mix 0.1%	BRAF	p.V600E	0.21	ND	ND
complete mix 0.1/0	KRAS	p.Q61H	0.43	ND	ND
	KRAS	p.G12D	ND	ND	0.14
	KRAS	p.G12C	ND	0.08	0.20
	EGFR	p.L747_P753>S	0.09	0.15	ND
	EGFR	p.S752 I759del	0.09	0.15	ND
	ERBB2	p.E770_A771insAYVM	0.13	ND	ND
	AKT1	p.E17K	0.15	ND	0.14
	TP53	p.E17K p.G245D	0.65	0.29	0.14
	11 73	P.ULHJU	0.03	ひ.とフ	U.41

Table 3. All 16 variants were successfully detected in Seraseq[™] cfDNA Complete Mutation Mix with higher allele frequency ≥ 0.5%. As observed previously (data not shown) additional FP calls were made in these libraries.

Table 4. Result Summary for Seraseq® Fusion RNA Dilutions

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Fusion Variants	Copies (20%)*	Molecule Count∆				
Fusion Variants	ddPCR_SeraCare	20%	10%	5%	2%	
TPM3-NTRK1	95	77	66	22	11	
LMNA-NTRK1	97	123	30	16	15	
SLC45A3-BRAF	132	110	35	6	2	
EML4-ALK	184	78	36	15	19	
FGFR3-BAIAP2L1	259	38	16	16	3	
FGFR3-TACC3	147	35	20	14	3	
SLC34A2-ROS1	101	24	4	3	ND	
CD74-ROS1	153	40	18	4	8	
MET Exon14 Skipping	166	68	45	15	9	
KIF5B-RET	83	76	45	15	5	
NCOA4-RET	144	127	30	13	14	
ETV6-NTRK3	128	87	51	15	8	
TMPRSS2-ERG	83	8	ND	ND	ND	
PAX8-PPARG1	259					
EGFR-SEPT14	133	Not covered				
EGFR variant III	124					

* Estimated copy number in 20% dilution based on ddPCR measurement from SeraCare

△ Measured copy number using Pan-Cancer assay

Table 4. All 13 fusion variants covered by Pan-Cancer assay were detected in Seraseq fusion standards diluted down to 5% in cfTNA background. Based on the molecule counts, LODs for each fusion target were able to achieve at 2% standard for Pan-Cancer assay.

Table 5. Result Summary for Horizon Structural Multiplex cfDNA Reference Standard

Sample	SNV / Indel			CNV		Fusion		
	Gene	AA Chg	AF %	Gene	CNV	Variant Exon	Molecule	
			(R1)		Ratio		Counts	
	CTNNB1	p.S33Y	5.4	FGFR3*	1.7	SLC34A2-ROS1.S4R32	50	
	PIK3CA	p.E545K	5.8	CDK6*	1.4	SLC34A2-ROS1.S4R34	6	
	PIK3CA	p.H1047R	15.7	MET	1.8	CCDC6-RET.C1R12	26	
Structural	EGFR	p.G719S	5.8	MYC	3.1			
Multiplex	EGFR	p.Q746_A750del	5.7					
cfDNA	EGFR	p.M766_A767insASV	4.9					
Reference	BRAF	p.V600E	16.4					
Standard	KRAS	p.G13D	4.7					
	AKT1	p.E17K	5.0					
	MAP2K1	p.Q56P	4.7					
	TP53	p.S241F	6.9					
	TP53	p.R175C	0.2					
	GNA11	p.Q209L	5.8					

*FGFR3 and CDK6 gene amplification false positives due to baseline not compatible with contrived samples using genomic DNA material.

Table 5. All the expected variants in Horizon structural multiplex cfDNA standard were detected including 13 SNC/Indel, ROS1 and RET RNA fusion, MET and MYC gene amplifications.

CONCLUSIONS

- This study established overall performance of commercial cfDNA control materials using Oncomine™ Pan-Cancer cell-free assay and provided reference information for laboratories when selecting appropriate standards during assay evaluation and validation.
- Horizon Multiplex I cfDNA reference standard set contains 8 SNV/Indels at several important gene loci and 5 additional SNVs from parental cell lines using Pan-Cancer assay. All 13 variants were successfully detected from standards > 1%; for 0.1% standard, majority of variants were detected consistently at 50 ng DNA input with sensitivity of 85% and specificity of 100%.
- In SeraseqTM cfDNA Complete Mutation Mix, Pan-Cancer assay covered 16 out of 19 claimed SNV/Indels. In high AF samples above 0.5%, all 16 SNVs were successfully detected using Pan-Cancer assay. In 0.1% standard, quite a few variants were missed with sensitivity < 60%. In addition. More false positives were reported in this set of control samples.
- Similarly, all variant types in Horizon Structural Multiplex standard were successfully detected including SNV/Indel, CNV and fusion with 100% sensitivity. In addition, gene amplification of FGFR3 and CDK6 was detected possibly due to the CNV baseline in Pan-Cancer assay which were constructed mainly using cell free type of material and not including this type of contrived sample using genomic DNA material.

TRADEMARKS/LICENSING

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