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A High Throughput System for Profiling Respiratory Tract Microbiota

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INTRODUCTION

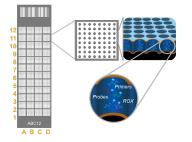
As one of the leading causes of death globally, respiratory infections could be caused by singleor multiple types of viral, bacterial or fungal pathogens that present in the upper and lower respiratory tract. Panel-basedtesting using molecular methods to identify multiple pathogens simultaneous ly can contribute to better understanding of respiratory infections.

MATERIALS AND METHODS

To develop a comprehensive and flexible research panel, we chose TaqMan® OpenArray™platform to identify common respiratory tractorganisms via real-time PCR technology. Target organism sequences were acquired from IMG and NCB database. Divergent gene targets were chosen to design research assays with highs pecificity (ANI>96%) and wide strain coverage. For each target, multiple assays were designed in silico and then assay performance was evaluated using various controlsincluding synthetic and natural genomic DNA and RNA, as well as human respiratory specimen. As say specificity was evaluated with genomic RNA and DNA of standard reference viral and bacterial organisms from American Type Culture Collection (ATCC), To achieve higher sensitivity, optimized components and thermal cyclingcondition for PCR pre-amplification was determined by extensive Design of Experiments (DOE) studies.

RESULTS

Figure 1. OpenArray[™] system for high throughput RTM profiling.



OpenArray™echnologyutilizes a microscope silde-sized plate with 3072 through-holes. Eachplate contains 48 subarrays with 64 through-holes. Assays for different RTM targets are loaded in individual through-holes. 1-4 subarrays can form a RTM parel depending on customerspecificationof the number of targets and replicates. 12 – 48 samples may beprocessed simultaneously on one OpenArray plate for RTM studies.

Figure 2. RTM study workflow

(SP) at serial dilutions.



DNA and RNA from various types of samples of respiratory tractmay be extracted with KingFisher™Flex purification system, followed by reverse transcription and pre-amplification. The reaction is thendiluted and loaded into Open Array™plates with AccuFill™system. Real time PCR of RTM assays performed in QuantStudio™12K Flex instrument and results of positiveor negative are analyzed with QuantStudio™12 Flex software.

Table 1. List of targeted organisms of Thermo Fisher Scientific OpenArray[™] Respiratory Tract Microbiota (RTM) research panel

		_		sm Namo		
			Adenovirus pool 3_1	Influenza A H3		
	Viral	DNA	Adenovirus pool 3_2	Influenza A pan assay		Viral
			Bocavirus	Influenza B		
			HHV3 (VZV)	Measles virus		
			HHV4 (EBV)	MERS	RNA	
			HHV5 (CMV)	Mumps virus		
			HHV6	RSVA specific		
		RNA	Human Coronavirus 229E	RSVB specific		
			Human Coronavirus HKU1	SARS		
			Human Coronavirus NL63	Pneumocystis iirovecii	DNA	Fungal
			Human Coronavirus OC43	Bordetella		Bacterial
			Human enterovirus D68 strain	Bordetella holmesii		
			Human enterovirus pan assav	Bordetella pertussis		
			Human Metapneumovirus	Chlamydophila pneumoniae		
			Human Parainfluenza virus 1	Coxiella burnetii		
			Human Parainfluenza virus 2	Haemophilus influenzae		
			Human Parainfluenza virus 3	Klebsiella pneumoniae	UNA	
			Human Parainfluenza virus 4	Legionella pneumophila		
			Human Parechovirus v2	Moraxela catarrhalis		
			Human Rhinovirus A	Mycoplasma pneumoniae		
			Human Rhinovirus B/C	Staphylococcus aureus		
			Influenza A H1 seasonal	Streptococcus pneumoniae		

Figure 3. gPCR results of Thermo Fisher Scientific Respiratory Tract

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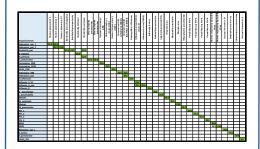
Plots show Crtvalues of selected assays at different input (copies/µL) of synthetic

targets. The resulted Ct values show linear response to the copy number of targets,

demonstrating high PCR efficiency, which confirms in silico design prediction.

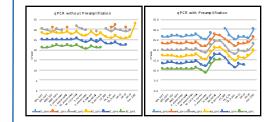
Microbiota (RTM) panel assays tested with synthetic assay targets

Table 2. Specificity of OpenArray[™] Respiratory Tract Microbiota (RTM) Panel



When tested with genomic RNA and DNA of standard reference viral and bacterial organisms from American TypeCultureCollection (ATCC), the assays in RTM panel displayed high specificity. All organisms are deteded by relevant assays.

Figure 4. Pre-amplification enhances assay sensitivity



Genomic materials from ATCC were pooled and titrated at serial dilutions for the tests. The left panel shows Ct values of pools when applied for real-time PCR teston OpenArray "Micredy after reverse transcription. The right panel shows Ct values of corresponding pools underwert a preamplification process at which 14 cycles of polymerase chain reaction was carried outfollowing reverse transcription.

Figure 5. Linearity range of RTM assays

Adv_1ot2	AdV_2of2	R.holmesii	Bordetella
	38.3	314	80
•	21	21.1	800
		24	31.0
-		25.8	800
Y = 3.1654 + 36.073	11.1 Y- 1.14(20) St.211.	26.8 V + 2.8961	11.0 F 10.000
8 ⁷ - 2.5954			300
	5.0	5.8	5.0
1 2 3 4 5	10		60
1 7 5 7 5		0 1 2 3 1 3	
HHV3	HRV5	R pertussis	Gay 2296
	300	160	NA .
	80	80	100
~	800	no	
		200	200 10.000000000000000000000000000000000
Y-3.205 26.802	11.0	11.0 11.0 11.0 11.0 10 10 10 10 10 10 10 10 10 10 10 10 10	100 P-11940
N*+3,5996		87+03880	100
	5.0	5.0	34
	60	6.6	44
3 2 3 4 5	0 1 3 3 A 5	1 1 2 3 4 5	- 1 - 2
			Spreumoniee
No.A.HL	Plu_A_pen	EV_068	820 250 250 250
-	35.0	85.0 35.0 35.0 35.0 15.0	800 250 300 350
1. 1000-100	800 150 300 150 150 150 150	81.0 91.0	80.0 25.0 25.0 15.0
7- 5.0000-1849 P+1.064	800 300 300 300 300 300 300 300	810 300 300 300 300 150 150 150 150 150 150 150 1	800 250 350 150 150 900 8 × 3.3766 - 2.441 300 8 × 0.5690
L. Martin B.D.	840 350 350 350 350 350 350 97 * 33940* 34792 350 97 * 33940* 34792	81.0 31.0	800 350 350 350 350 50 8° × 0.5990
1 2 3 4	10 10 10 10 10 10 10 10 10 10	Bac	920 350 350 920 920 920 920 920 920 920 92
PARKA	5 A C C C C C C C C C C C C C C C C C C	100 100 100 100 100 100 100 100	820 230 320 320 320 320 9 1 2 3 4 HWV2
1 2 3 4	320 320 320 320 320 320 320 320	110 110 110 110 110 110 110 110	820 310 310 310 310 310 310 310 31
1 2 3 4	320	100 100 100 100 100 100 100 100	320 320
r - 5.0507 - 79.05 F - 2.064 1 2 3 4 E. preumotike	820 320 320 320 320 320 520 520 520 520 520 520 520 5	100 100 100 100 100 100 100 100	320 330 330 330 300 4 - 33786 - 24 4 8 - 3 - 2 - 3 - 4 100 20 10 - 1 - 2 - 3 - 4
r = 1.0007 - 7.000 F = 2.004 L = 2. 8 = 4 E. preumotike	320	100 100 100 100 100 100 100 100	33.0
r - 5.0507 - 79.05 F - 2.064 1 2 3 4 E. preumotike	100 100 100 100 100 100 100 100	100 100 100 100 100 100 100 100	330
r - 5.0111 - 11.00 r - 5.0111 - 11.00 r - 5.010 L 2 2 4 C preumotile r - 3.1024 - 75.10	320	100 100 100 100 100 100 100 100	33.0
r - 4.0111 - 11.00 r - 4.0111 - 11.00 r - 4.0111 - 11.00 E preumatile r - 3.0221 - 75.10	100 100 100 100 100 100 100 100	100 100 100 100 100 100 100 100	320

Charts show Ctvalues (Yaxes) corresponding to different amount of initial input copies (X-axes, at power of 10 copies/ μ L) of ATCC viral and bacterial controls.

CONCLUSIONS

Anovel researchapplication hasbeen developed for Respiratory TractMicrobio (RTM) profiling Assays in the pand demonstrate desirable performance in terms of sensitivity, specificity and linearity range. The application enables both customizable and highthroughputpanels for respiratory infection research and provides a cost-effective both or researchersts understand pathogenicity in respiratory tractifications.

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