

Key products for cardiovascular primary cell culture

- Primary cells give you greater physiological relevance
- Complete cell culture systems designed and optimized to work together
- Backed by expert technical support

Invitrogen's Cascade Biologics® primary cells have been developed to work together for optimal performance. When your cell culture research demands relevance, reliability, and robustness, choose Cascade Biologics® primary cells to meet your most critical needs.

Table 1—Cascade Biologics® products for cardiovascular primary cell culture.*

Primary human cells (cryopreserved)	Aortic endothelial cells (C-006-5C)	Aortic smooth muscle cells (C-007-5C)	Dermal microvascular endothelial cells • Neonatal (C-010-5C) • Adult (C-011-5C)
	Pulmonary artery endothelial cells (C-008-5C)	Pulmonary artery smooth muscle cells (C-009-5C)	
	Umbilical vein endothelial cells • Single-donor (C-003-5C) • Pooled (C-015-5C, C-015-10C)	Coronary smooth muscle cells (C-017-5C)	
Basal media and growth supplements	Medium 200 (500 ml) • Standard (M-200-500) • Phenol red-free (M-200PRF-500)	Medium 231 (500 ml) (M-231-500)	Medium 131 with attachment factor (500 ml) (M-131-500)
	Low-serum growth supplement (LSGS) • Single-addition (S-003-10) • Kit (S-003-K)	Smooth muscle growth supplement (SMGS) (S-007-25)	Microvascular growth supplement (MVGS) (S-005-25)
		Smooth muscle differentiation supplement (SMDS) (S-008-5)	Attachment factor (100 ml) (S-006-100)

Subculture and other reagents

- Gentamicin/amphotericin 10-pack (R-015-10)
- Synth-a-Freeze® cryopreservation medium (R-005-50)
- Trypsin/EDTA (R-001-100)
- Trypsin neutralizer (R-002-100)

* The cells listed in Table 1 are also available as proliferating cultures (catalog numbers for proliferating cultures take the form C-xxx-25P). All cells have tested negative for HIV-1, hepatitis B, hepatitis C, mycoplasmas, bacteria, yeast, and other fungi and are highly characterized (Table 2).

Table 2—Characteristics of Cascade Biologics® cells for cardiovascular primary cell culture.

Cell type	α-actin	vWf	CD31	CD36	dil-Ac-LDL	Population doublings	Viability (upon thawing)
Large vessel endothelial cells (aortic, pulmonary artery, and umbilical vein cells)	–	+	+	N/A	+	>16	>70%
Smooth muscle cells	+	–	N/A	N/A	N/A		
Dermal microvascular cells	–	+	+	+	+		

Visit www.invitrogen.com/primarycells to see the entire range of Cascade Biologics® primary cells and optimized media from Invitrogen Cell Culture.

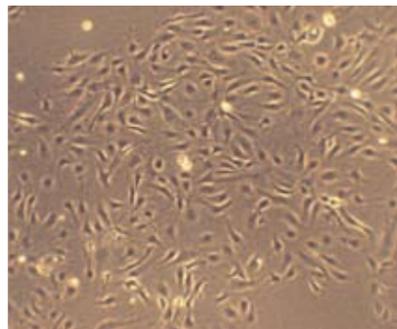


Figure 1—Phase-contrast image of human umbilical vein endothelial cells.

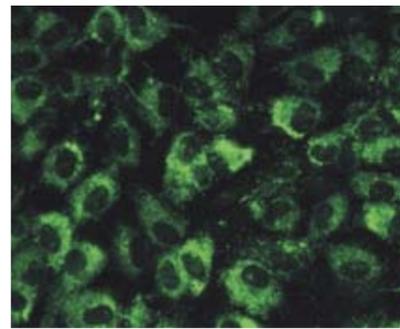


Figure 2—Human umbilical vein endothelial cells, anti-vWf immunofluorescence.

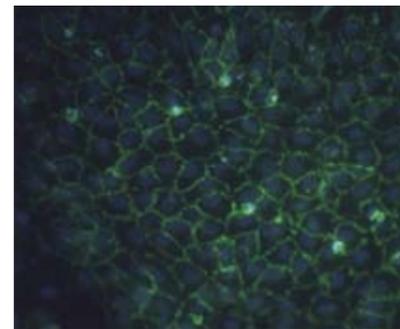
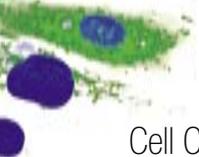


Figure 3—Microvascular endothelial cells were stained using an anti-CD31 primary antibody in conjunction with a fluorescein-labeled secondary antibody. Nuclei were counterstained with DAPI.

Selected references

Cardiovascular cell research references that cite the use of Cascade Biologics® products:

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(Human umbilical vein endothelial cells; medium 200; low-serum growth supplement)
- Barresi, R. et al. (2000) Expression of γ -sarcoglycan in smooth muscle and its interaction with the smooth muscle sarcoglycan-sarcospan complex. *J Biol Chem* 275(49): 38554–38560.
(Human coronary artery smooth muscle cells; medium 231; smooth muscle cell growth supplement; smooth muscle cell differentiation supplement)
- Bhaskar, V. et al. (2003) E-selectin up-regulation allows for targeted drug delivery in prostate cancer. *Cancer Res* 63(19): 6387–6394.
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- Bulin, C. et al. (2005) Differential effects of vasodilatory prostaglandins on focal adhesions, cytoskeletal architecture, and migration in human aortic smooth muscle cells. *Arterioscler Thromb Vasc Biol* 25(1): 84–89.
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- Chou, H.-H. et al. (2005) Porphyromonas gingivalis fimbria-dependent activation of inflammatory genes in human aortic endothelial cells. *Infect Immun* 73(9): 5367–5378.
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(Human coronary artery smooth muscle cells; medium 231; smooth muscle cell growth supplement)
- Fujita, H. et al. (2005) Local activation of Rap1 contributes to directional vascular endothelial cell migration accompanied by extension of microtubules on which RAP1, a Rap1-associated molecule, localizes. *J Biol Chem* 280(6): 5022–5031.
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- Leung, W.C.Y. et al. (2004) Apolipoprotein D and platelet-derived growth factor-BB synergism mediates vascular smooth muscle cell migration. *Circ Res* 95(2): 179–186.
(Human pulmonary artery smooth muscle cells)
- Li, S. et al. (2003) Genomic analysis of smooth muscle cells in three-dimensional collagen matrix. *FASEB J* 17(1): 97–99.
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- Panyam, J. et al. (2002) Rapid endo-lysosomal escape of poly(DL-lactide-co-glycolide) nanoparticles: implications for drug and gene delivery. *FASEB J* 16(10): 1217–1226.
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- Romagnani, P. et al. (2001) Cell cycle-dependent expression of CXC chemokine receptor 3 by endothelial cells mediates angiostatic activity. *J Clin Invest* 107: 53–63.
(Human microvascular endothelial cells; medium 131; microvascular growth supplement)
- Sun, Q. et al. (2006) Defining the mammalian CArGome. *Genome Res* 16(2): 197–207.
(Human coronary artery smooth muscle cells; medium 231; smooth muscle cell growth supplement; medium 200; low-serum growth supplement)



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