## **thermo**scientific

STUDY REPORT

# Thermo Scientific SureTect Salmonella species PCR Assay Workflow NF VALIDATION; ISO 16140-2:2016

Jessica Williams, Thermo Fisher Scientific, Wade Road, Basingstoke, Hampshire, RG24 8PW, UK

#### **Summary**

The Thermo Scientific™ SureTect™ Salmonella species PCR Assay workflow (PT0100A) (alternative method) has been certified by NF VALIDATION (UNI 03/07-11/13) for the detection of *Salmonella* species from meat, dairy, seafood, vegetables, raw beef meats with and without aromatics, production environment samples and powdered infant formula (PIF) 10 g and 375 g samples. The following report gives a summary of the validation studies performed to gain NF VALIDATION.

#### Methodology

Study	Reference method
Initial validation PCR analysis was conducted using the Thermo Scientific™ SureTect™ PikoReal™ Real-Time PCR Instrument	ISO 6579:2002 'Microbiology of the food chain— Horizontal method for the detection of Salmonella species.'
Extension study  To incorporate the Applied Biosystems™ 7500 Fast Food Safety Real-Time PCR System (7500 Fast Real-Time PCR Instrument and Applied Biosystems™ RapidFinder™ Express Software (version 2.0 or higher))	ISO/TS 22964:2006 'Microbiology of the food chain— Horizontal method for the detection of Salmonella species.'
<ul> <li>Extension study</li> <li>To incorporate the following;</li> <li>Applied Biosystems™ QuantStudio™ 5 Food Safety Real-Time PCR System (Applied Biosystems QuantStudio 5 Real-Time PCR Instrument and RapidFinder™ Analysis Software version 1.0)</li> <li>Powdered milk products (375 g)</li> </ul>	ISO 6579-1:2017 'Microbiology of the food chain— Horizontal method for the detection, enumeration and serotyping of Salmonella – Part 1: Detection of Salmonella species.'



The initial certification study and all subsequent extension studies were conducted by ADRIA Développement, Quimper, France.

The protocols for the alternative method and the reference method are summarized in Appendix 1 and 2, respectively.

#### **Inclusivity & exclusivity study**

Fifty-five inclusivity isolates were cultured in Thermo Scientific™ Brain Heart Infusion (BHI) Broth (incubated for 24 hours at 37±1°C). The BHI culture was inoculated into Thermo Scientific™ Buffered Peptone Water (BPW) (ISO) at a level of 10 CFU/225 mL (incubated for 8 hours at 41.5±°C).

Thirty exclusivity isolates were cultured in BHI Broth (incubated for 24 hours at 37±1°C). The BHI culture was inoculated into BPW (ISO) to obtain approximately 10<sup>5</sup> CFU/mL (incubated for 24 hours at 37±1°C).

Following incubation, the enrichment samples were analyzed following the alternative method protocol (Appendix 1).

#### Inclusivity & exclusivity results

The alternative method successfully identified all 55 inclusivity isolates and correctly gave a negative result for all 30 exclusivity isolates. The results show the alternative method is sensitive and specific with equivalent performance to the ISO reference method.

#### **Inter-laboratory study**

An inter-laboratory study was performed as part of the initial validation. A raw ground beef matrix was prepared and spiked with *Salmonella* Typhimurium and sent to all participating laboratories. Samples were analyzed following both the alternative method and the ISO reference method (Appendix 1 and 2). Of all the samples tested, one third were unspiked, one third were spiked with a low level inoculum (4.1 CFU/25 g) and the remaining samples were spiked with a high level inoculum (19.2 CFU/25 g).

#### Inter-laboratory study results

Table 1. Inter-laboratory study results summary

Relative accuracy	99.6%
Relative sensitivity	100.0%
Relative specificity	100.0%

The results displayed in Table 1 demonstrate that the alternative method is a reliable method for the detection of *Salmonella* species.

#### **Method comparison study**

A total of 456 samples including meat products, milk and dairy products, seafood and fishery products, vegetables, raw beef meats, environmental samples and PIF (10 g) categories were analyzed using the alternative method with the Applied Biosystems 7500 Fast Food Safety System and the Applied Biosystems QuantStudio 5 Food Safety System.

The current extension study aimed to extend the claims of the NF VALIDATION to include milk powders (375 g), and to incorporate the use of the Applied Biosystems QuantStudio 5 Food Safety System.

For the milk and dairy category and the raw beef meats category, two different enrichment protocols were analyzed as part of the method comparison study on the Applied Biosystems 7500 Fast Food Safety System and the Applied Biosystems QuantStudio 5 Food Safety System. Table 3 summarizes the categories and enrichment protocols.

**Table 3: Enrichment protocol summary** 

egory or type	Enrichment broth	Incubation
Meat products	BPW (ISO) + 12 mg/L Novobiocin	20-24 h at 37±1°C
	BPW (ISO) + 12 mg/L Novobiocin	20-24 h at 37±1°C
Dairy products	Thermo Scientific™ ONE Broth-Salmonella (OBS) with 12 mg/L Novobiocin	20-24 h at 37±1°C
Infant formula	BPW (ISO)	16-20 h at 37±1°C
Vegetables	BPW (ISO) with 12 mg/L Novobiocin	20-24 h at 37±1°C
Seafood	BPW (ISO) with 12 mg/L Novobiocin	20-24 h at 37±1°C
Raw beef meats with and	Pre-warmed BPW (ISO)	9 h at 41.5±1°C
without aromatics	Pre-warmed BPW (ISO)	24 h at 41.5±1°C
Environmental samples	BPW (ISO)	20-24 h at 37±1°C
Powdered milk products	BPW (ISO) with 6 mg/L Vancomycin	18-22 h at 37±1°C
	Meat products  Dairy products  Infant formula  Vegetables  Seafood  Raw beef meats with and without aromatics  Environmental samples	Meat productsBPW (ISO) + 12 mg/L NovobiocinDairy productsThermo Scientific™ ONE Broth-Salmonella (OBS) with 12 mg/L NovobiocinInfant formulaBPW (ISO)VegetablesBPW (ISO) with 12 mg/L NovobiocinSeafoodBPW (ISO) with 12 mg/L NovobiocinRaw beef meats with and without aromaticsPre-warmed BPW (ISO)Environmental samplesBPW (ISO)

PA = Positive agreement PD = Positive deviation PP = Positive deviation positive presumptive non-confirmed samples NA = Negative agreement ND = Negative deviation PPND = Predictive positive negative deviation PPNA = Predictive positive negative agreement

The results for the method comparison study using the Applied Biosystems 7500 Fast and the Applied Biosystems QuantStudio 5 Food Safety Systems are detailed in Tables 4 and 5, respectively.

### Method comparison study results

Table 4: Method comparison study results using the Applied Biosystems 7500 Fast Food Safety System

Cate	gory	PA	NA	PD	ND	PPND	PPNA	Total
1	Meat products	26	35	7	5	0	0	73
	Milk and dairy products (BPW + Novobiocin)	21	32	4	4	1	0	62
2	Milk and dairy products ((One-Broth Salmonella) (OBS) + Novobiocin)	21	31	5	5	0	0	62
3	Infant formula	33	30	0	1	0	0	64
4	Vegetables	22	33	5	3	1	0	64
5	Seafood products	22	33	3	4	1	0	63
6	Raw beef meats 9 h	20	31	5	5	0	0	61
6	Raw beef meats 24 h	21	31	5	4	0	0	61
7	Environmental samples	27	39	0	3	0	0	69
8	Powdered milk products (375 g)	21	30	6	6	0	0	63
	tegories + Dairy enriched with BPW + Raw Beef ated for 9 hours	192	263	30	31	3	0	519
	All categories + Dairy enriched with OBS + Raw Beef incubated for 9 hours		263	31	32	2	0	519
	All products + Dairy enriched with BPW + Raw Beef incubated for 24 hours		263	30	30	3	0	519
	oducts + Dairy enriched with OBS + Raw Beef ated for 24 hours	193	262	31	31	2	0	519

Table 5: Method comparison study results using the Applied Biosystems QuantStudio 5 **Food Safety System** 

Cate	gory	PA	NA	PD	ND	PPND	PPNA	Total
1	Meat products	26	35	7	5	0	0	73
0	Milk and dairy products (BPW + Novobiocin)	21	32	4	4	1	0	62
2	Milk and dairy products (OBS + Novobiocin)	21	29	6	3	2	1	62
3	Infant formula	33	29	0	1	0	1	64
4	Vegetables	22	31	6	3	1	1	64
5	Seafood products	22	32	4	4	1	0	63
6	Raw beef meats 9 h	21	32	4	3	1	0	61
6	Raw beef meats 24 h	21	31	5	4	0	0	61
7	Environmental samples	28	39	0	2	0	0	69
8	Powdered milk products (375 g)	20	29	6	7	0	1	63
	tegories + Dairy enriched with BPW + Raw Beef ated for 9 hours	193	259	31	29	4	3	519
	All categories + Dairy enriched with OBS + Raw Beef incubated for 9 hours		256	33	28	5	4	519
	All products + Dairy enriched with BPW + Raw Beef incubated for 24 hours		258	32	30	3	3	519
	oducts + Dairy enriched with OBS + Raw Beef ated for 24 hours	193	255	34	29	4	4	519

PA = Positive agreement PD = Positive deviation PP = Positive deviation positive presumptive non-confirmed samples NA = Negative agreement 

During the current extension study, six negative deviations were observed for the Applied Biosystems 7500 Fast Food Safety System and seven negative deviations were observed for the Applied Biosystems QuantStudio 5 Food Safety System.

Among the samples in negative deviation, the presence of Salmonella was confirmed for three samples. For one sample, one of the three PCR replicates tested gave a negative result on both Food Safety Systems. For the remaining two samples, three PCR replicates were analyzed and all gave negative results. The contamination in the enrichment broths for these samples was likely below the limit of detection of the alternative method.

During the current extension study, six positive deviations were recorded when using both Food Safety Systems. The positive deviations were detected as positive using the alternative method but failed to be detected with the reference method. All six positive deviations were confirmed as true positives.

The results from all validation studies has been combined and is presented in Table 6.

Table 6: Sensitivity, relative trueness and false positive ratio summary

	Applied Biosystems 7500 Fast				Applied Biosystems QuantStudio 5			
	Α	В	С	D	A	В	С	D
Sensitivity of the alternative method	88.7%	86.8%	87.1%	87.2%	87.2%	87.3%	87.2%	87.3%
Sensitivity of the reference method	88.3%	87.9%	88.3%	87.9%	87.9%	87.3%	87.6%	86.9%
Relative trueness	87.7%	87.5%	87.9%	87.7%	87.7%	87.3%	87.5%	87.1%
False positive ratio	1.1%	0.8%	1.1%	0.8%	2.7%	3.5%	2.3%	3.1%

 $A = All \ categories + Dairy \ enriched \ with \ BPW + Raw \ Beef \ incubated \ for \ 9 \ hours \\ B = All \ categories + Dairy \ enriched \ with \ OBS + Raw \ Beef \ incubated \ for \ 9 \ hours$ 

The method comparison study results demonstrate that the alternative method was shown to be a reliable alternative to the ISO reference method for the detection of *Salmonella* spp. from the meat products, milk and dairy products, seafood and fishery product, vegetables, raw beef meats, environmental samples, powdered infant formula and powdered milk products categories.

### Relative level of detection study

For the relative level of detection (RLOD) studies, eight *Salmonella* species isolates were spiked into eight matrices (see Table 7 for matrix/strain pairs) and analyzed using the Applied Biosystems 7500 Fast and the Applied Biosystems QuantStudio 5 Food Safety Systems.

Table 7: Defined matrix/strain pairs for the RLOD determination.

Ca	tegory	Matrix	Inoculated strain
1	Meat products	Raw chicken meat	Salmonella Bredeney 975
2	Milk and dairy products	Raw milk	Salmonella Ohio Ad1482
3	Powdered infant formula (10 g)	Powdered infant formula with probiotics	Salmonella Anatum Ad298
4	Vegetables	Frozen spinach	Salmonella Virchow Ad1721
5	Seafood	Fish terrine	Salmonella Derby Ad1093
6	Raw beef meats	Ground beef	Salmonella Typhimurium A00C060
7	Production environmental samples	Process water	Salmonella Livingstone A00L058
8	Milk powder, powdered infant formula and infant cereals with and without probiotics including ingredients (375 g) <sup>a</sup>	Powdered infant formula with probiotics	Salmonella Mbandaka Ad1810

<sup>&</sup>lt;sup>a</sup> Categories analyzed in current extension study

 $C = All \ products + Dairy \ enriched \ with \ BPW + Raw \ Beef \ incubated \ for 24 \ hours$ 

D = All products + Dairy enriched with OBS + Raw Beef incubated for 24 hours

The samples were analyzed using the reference method detailed in ISO 6579-1:2017 prior to inoculation in order to verify the absence of *Salmonella* spp. After inoculation, samples were tested using the ISO reference method and the alternative method.

Table 8: Relative level of detection results

Matrix	Strain	QuantStudio 5 RLOD	7500 Fast RLOD	Acceptability level (≤)
Raw chicken meat	Salmonella Bredeney 975	1.629	1.629	2.5
Raw milk (BPW + Novobiocin)	Salmonella Ohio Ad1482	0.761	1.000	2.5
Raw milk (OBS + Novobiocin)	Salmonella Ohio Ad1482	1.336	1.336	2.5
Powdered infant formula with probiotics (10 g) <sup>a</sup>	Salmonella Anatum Ad298		1.000	1.5
Frozen spinach	Salmonella Virchow Ad1721	1.000	1.000	2.5
Fish terrine	Salmonella Derby Ad1093	1.000	1.000	2.5
Ground beef (9 h and 24 h)	Salmonella Typhimurium A00C060	0.554	0.554	2.5
Process water	Salmonella Livingstone A00L058	1.000	1.170	2.5
Powdered infant formula with probiotics (375 g) <sup>b</sup>	Salmonella Mbandaka Ad1810	0.195	0.195	2.5
Combined RLOD		0.822	0.852	/

<sup>&</sup>lt;sup>a</sup> Paired study design

As shown in Table 8, the alternative method gave a RLOD within the acceptability limit of  $\leq$ 1.5 for the paired study and  $\leq$ 2.5 for the unpaired study designs, when used with the Applied Biosystems QuantStudio 5 Food Safety System and the Applied Biosystems 7500 Fast Food Safety System.

#### Conclusion

The NF VALIDATION studies demonstrate that the SureTect Salmonella species PCR Assay workflow gives equivalent or improved performance compared to the ISO reference method detailed in ISO 6579-1:2017 for the detection of *Salmonella* spp. from a broad range of foods and environmental samples when using the Applied Biosystems 7500 Fast or the Applied Biosystems QuantStudio 5 Food Safety Systems. The NF VALIDATION certificate and a summary of the validation report for this study are available from www.thermofisher.com/foodsafety.

#### www.thermofisher.com

© 2019 Thermo Fisher Scientific Inc. All rights reserved. All trademarks are the property of Thermo Fisher Scientific and its subsidiaries unless otherwise specified. NF VALIDATION is a trademark of Association Française de Normalisation (AFNOR). This information is presented as an example of the capabilities of Thermo Fisher Scientific Inc. products. It is not intended to encourage use of these products in any manners that might infringe the intellectual property rights of others.



<sup>&</sup>lt;sup>b</sup> Categories analyzed in current extension study

## Appendix 1: Workflow for alternative method—SureTect Salmonella Species PCR Assay for use on the QuantStudio 5 Food Safety System

	Day 0								
Environmental samples  • 25 g + 225 mL BPW  • 1 swab + 10 mL BPW  • 1 sponge + 100 mL BPW  • 1 wipe + 225 mL BPW  20–24 h at 37±1°C	Meat products, seafood and vegetables  25 g + 225 mL  BPW + 12 mg/L  Novobiocin  20–24 h at 37±1°C	Raw beef meats 25 g + 225 mL pre-warmed BPW 9-24 h at 41.5±1°C	Dairy products  • 25 g + 225 mL  BPW + 12 mg/L  Novobiocin  20–24 h at  37±1°C  • 25 g + 225 mL  OBS + 12 mg/L  Novobiocin  20–24 h at 37±1°C	Powdered infant formula (10 g) 25 g + 225 mL BPW  16–20 h at 37±1°C	Powdered milk products  375 g + 3375 mL  BPW + 6 mg/L  vancomycin  18–22 h at 37±1°C				

#### Day 1

Add 10 µL of SureTect Proteinase K to each required SureTect Lysis Tube (supplied pre-filled with Lysis Reagent 1).

Add 10  $\mu$ L enriched sample to the SureTect Lysis Tube. For the 7500 Fast; add 10  $\mu$ L sterile nuclease-free water to a SureTect Lysis Tube as a negative control.

Incubate SureTect Lysis Tubes in the Applied Biosystems™ SimpliAmp™ Thermal Cycler at 37±1°C for 10 minutes followed by 95±1°C for 5 minutes, and 2 minutes at 10±1°C.

Transfer 20 µL of lysate to SureTect PCR Tubes.

Report negative results

Load SureTect PCR Tubes to the Applied Biosystems 7500 Fast or the Applied Biosystems QuantStudio 5 PCR Instrument. Start PCR and review results at end of run.

## Day 2

Confirm PCR positive results by plating 10  $\mu$ L of enrichment broth onto Thermo Scientific<sup>M</sup> Brilliance<sup>M</sup> Salmonella Agar. For samples with high background microflora first subculture in RVS broth (incubate for 24 h±3 h at 41.5°C±1°C).

Confirm presumptive-positive colonies with the Oxoid™ Salmonella Latex Kit, MicroBact™ GNB24E or confirmatory tests of the ISO method.

## Appendix 2: Protocol for the reference method: ISO 6579-1

## Day 0

25 g + 225 mL BPW 1 swab + 10 mL BPW 1 sponge + 100 mL BPW 1 wipe + 225 mL BPW

Incubate for 18±2 h at 34-38°C

## Day 1

Add 100  $\mu$ L of primary enrichment to 10 mL of Rappaport-Vassiliadis soya peptone broth (RVS) or Modified Semi-solid Rappaport Vassiliadis (MRSV) agar.

Incubate for 24±3 h at 41.5±1°C

Add 100  $\mu$ L of primary enrichment broth to 10 mL of Muller-Kauffman Tetrathionate Broth supplemented with Novobiocin.

Incubate for 24±3 h at 37±1°C

## Day 2

Plate 10 µL of secondary enrichment broth onto XLD agar and a second chromogenic agar.

Incubate for 24±3 hours at 37±1°C

## Day 3

Confirm up to five presumptive-positive colonies by biochemical identification and serological tests.