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Axiom™ Propel XPRES 384HT Workflow

USER GUIDE

for use with:
Axiom™ 384HT Array Plates
Axiom™ Propel XPRES Reagent Kit, 2x384HT
Multidrop™ Combi Reagent Dispenser

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Products:
Axiom™ 384HT Array Plates
Axiom™ myDesign™ Array Plates

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Revision	Date	Description
C.0	30 March 2021	• Added the Thermo Scientific™ Digital Microplate Shaker as a shaker option. • Incorporated new plate sealing parameters for 96-deepwell plates (ABgene™ AB0932) on the ALPS™ 3000 Automated Microplate Heat Sealer. • Extended the storage time for GeneTitan™ master mixes to provide workflow timing flexibility. • Added the recommendation to use GeneTitan™ Barcoded Stain Trays to the general guidelines section. • Added instruction to the Stage 7 prerun checklist to ensure that each labeled stain tray is placed next to the correct Multidrop™ Combi dispensing the corresponding reagent. • Corrected the amounts of reduced EDTA TE buffer to use to dilute human and animal DNA samples.
B.0	12 August 2020	Correcting instructions for thawing the frozen Amplification Plates.
A.0	28 July 2020	New publication.

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About the Axiom™ Propel XPRES 384HT Workflow

The Axiom™ Propel XPRES 384HT Workflow is a new workflow for ultra high-throughput microarray genotyping. The workflow includes:

- DNA target preparation using Multidrop™ Combi Reagent Dispenser stations setup for: DNA amplification, fragmentation, purification, and resuspension of the pelleted DNA in hybridization cocktail.
- Array hybridization in an off-line oven.
- Automated array plate processing (ligation, stain, wash, and imaging) in the Applied Biosystems™ GeneTitan™ Multi-Channel (MC) Instrument.
- Processing of CEL files generated by the GeneTitan™ MC Instrument, using the Axiom™ Genotyping Algorithm version 1 (Axiom GT1), available through Applied Biosystems™ Array Power Tools or Axiom™ Analysis Suite v5.0 or later.

The Axiom™ Propel XPRES Reagent Kit, 2x384HT provides all necessary large-filled reagents for target preparation and GeneTitan™ reagents in volumes that are optimized for processing the modular workflow.

IMPORTANT! The Applied Biosystems™ Axiom™ Propel XPRES Reagent Kit, 2x384HT is for single use only. This large fill reagent kit is configured to include prime volumes required for use with the Multidrop™ Combi. Discard all excess reagents after use.

About the Axiom™ Genotyping Solution

The Axiom™ Propel XPRES 384HT Workflow is part of the Axiom™ Genotyping Solution. The Axiom™ Genotyping Solution is a genotyping microarray platform that includes novel assay biochemistry, array configuration and processing, and automated target preparation on various array plate formats. It offers the capability to genotype approximately 65,000 variants (of single nucleotide polymorphisms (SNPs) and insertion/deletion polymorphisms (indels) from diploid species or 43,000 variants from polyploid species, with a processing throughput of greater than 3,000 samples per week.

High-throughput genotyping through microarray technology has applications in human disease research and basic and applied agriculture research.

- For human disease research applications, Thermo Fisher Scientific conducted an empirical screen of genomic content from dbSNP (**ncbi.nlm.nih.gov/projects/SNP/**). The screen included markers from HapMap and the 1,000 Genomes Project and other sources, using HapMap phase 3 samples and/or the original 270 HapMap samples. All this information has gone into creating a proprietary database of verified markers that can be interrogated using the Axiom™ Propel XPRES 384HT Assay.
- For agriculture applications, the Axiom™ Genotyping Solution can genotype samples using DNA extracted from leaves and seeds, playing an important role in genotype-trait association studies and marker-assisted selection in both plant and animal breeding programs.
- For molecular breeding programs, where turn-around time, accuracy, and ease-of-use are all important, the Axiom™ Genotyping Solution is ideal for high-throughput screening.

The Axiom™ 96-array layout and the Axiom™ 384HT-array layout retain full compatibility with the existing Axiom™ instrumentation platform and downstream data analysis.

What's new

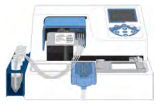
The new Axiom™ Propel XPRES 384HT Workflow includes enhancements to the Axiom™ chemistry which enable maximum assay workflow efficiency and flexibility:

- Whole genome amplification (WGA) incubation time: An option for an accelerated 4-hour WGA or up to 24 hours to maximize steady-state production schedules.
- DNA precipitation incubation time: An option for an accelerated 3-hour precipitation incubation time up to the standard overnight precipitation.
- GeneTitan™ MC Instrument: A reduced overall fluidics time of ~20% (from 5 hours to 4 hours) to increase the daily array plate loading capacity.

Dispense, seal, shake, then centrifuge

For each stage of the Axiom™ Propel XPRES 384HT Workflow conducted at a Multidrop™ Combi Reagent Dispenser, the following steps are typically performed.

- Task name is provided in the table heading.
- Number of plates for the workflow is listed in the first row of the table.
- Each subsequent row in the table lists a step in the task/procedure, with specific details listed. The following image is a general example.

Multidrop™ Combi tasks		
		
①		Dispense • Method: 384-xyz-## • Dispense volume: ## µL
②		Seal • Settings: 150°C, 2.5 seconds
③		Shake • Settings: 1,100 rpm, 30 seconds
④		Centrifuge • Centrifuge at room temperature • Settings: 675 × *g*, 30 second

GeneTitan™ reagent tray barcodes

GeneTitan™ MC Instrument consumables and Applied Biosystems™ GeneChip™ Command Console™ (GCC) are required for the preparation of the Axiom™ 2.0 stain reagents. Each tray has a unique part number and barcode that offers traceability. These trays have the following labels and barcodes:

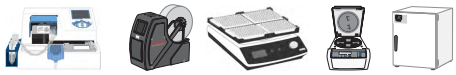
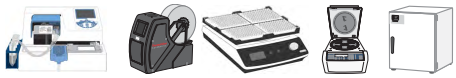
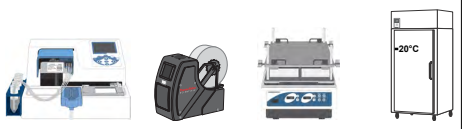
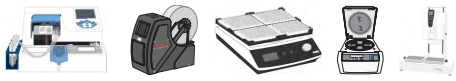
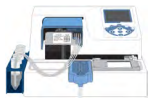
Figure 1 GeneTitan™ reagent tray barcodes and color-coded labels.

1. Stain 1 Tray—Part No. 501279
2. Stain 2 Tray—Part No. 501394
3. Ligation Tray—Part No. 501398
4. Stabilization Tray—Part No. 501396

The unique barcodes along with the GeneChip™ Command Console™ v3 or later software prevents users from making errors when placing the trays in the GeneTitan™ MC Instrument during array processing.

After the trays have been prepared, ensure that the trays are placed in the appropriate drawer location in the GeneTitan™ MC Instrument. Failure to place the proper tray in the correct location results in an error and the GeneTitan™ MC Instrument will not proceed with the processing of the trays. See “Proper tray alignment and placement” on page 150 for detailed instruction.

Axiom™ Propel XPRES 384HT Assay target preparation overview

Assay stage	Instruments required
Stage 1: Amplify the genomic DNA. • Three reagent additions with mixing—Denaturation Master Mix, Axiom™ Propel Neutral Solution, Amplification Master Mix. • 10-minute denature incubation at room temperature. • 4–24 hour amplification incubation at 37°C.	
Stage 2: Fragment the DNA. • Two reagent additions with mixing—Fragmentation Master Mix, Axiom™ Propel Frag Reaction Stop. • 30-minute fragmentation incubation at 37°C.	
Stage 3: Precipitate the DNA. • One reagent addition with mixing—384-Precipitation Master Mix. • 3–24 hour precipitation at –20°C.	
Stage 4: Centrifuge and dry DNA pellets. Purify amplified DNA into dried pellets.	
Stage 5: Resuspend the pelleted DNA and prepare for hybridization. • One reagent addition with mixing—Hybridization Cocktail. • 15 minute shaking to resuspend the DNA pellets. • Transfer from four 96-deepwell plates to one 384-format PCR plate.	
Stage 5A: In-process QC. • Three reagent dispenses—Dilution QC Plates, OD QC Plates, Gel QC Plates. • Transfer hybridization-ready target from 384 PCR plate to QC plates.	
Stage 6: Denature the target and transfer to hybridization tray. • Denature target in thermal cycler. • Transfer from 384-PCR plate to hybridization tray. • Off-line incubation of the array plate/hybridization tray stack at 48°C for 23.5–24 hours. .	
Stage 7: Preparing ligation, stain, stabilization reagent trays, and scan tray for the GeneTitan™ MC Instrument. • Five reagent dispenses—Ligation Master Mix, Stain 1 Master Mix, Stain 2 Master Mix, Stabilization Master Mix, Axiom™ Hold Buffer.	

Overview of the Axiom™ Propel XPRES 384HT Workflow

Genomic DNA preparation	
Chapter 2, Genomic DNA preparation.	
▼	
Target preparation	
Stage 1: Amplify the genomic DNA.	4–24 hour of Amplification Plate at 37°C. Optional stopping point. The post-amplification can be stored at –20°C for up to 1 week.
▼	
Stage 2: Fragment the DNA.	
▼	
Stage 3: Precipitate the DNA.	3–24 hour precipitation at –20°C.
▼	
Stage 4: Centrifuge and dry DNA pellets.	Optional stopping point. The pellets can be stored at –20°C for 1 day.
▼	
Stage 5: Resuspend the pelleted DNA and prepare for hybridization.	Optional stopping point. The hybridization-ready target can be stored at –20°C for up to 2 weeks.
▼	
Stage 6: Denature the target and transfer to hybridization tray.	23.5 to 24-hour array hybridization in the offline hybridization oven at 48°C.
▼	
Stage 7: Preparing ligation, stain, stabilization reagent trays, and scan tray for the GeneTitan™ MC Instrument.	
▼	
Array processing	
Chapter 5, Process array plates with the GeneTitan™ Multi-Channel (MC) Instrument. *Array processing is completed with the GeneTitan™ MC Instrument and GeneChip™ Command Console™ software v6.1 or later.*	Fluidics: ~4 hours Scan: ~4.75 hours

2 Genomic DNA preparation

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The general requirements for genomic DNA (gDNA) sources and extraction methods are described in this chapter. The success of this assay requires uniform amplification of the genome starting with relatively intact gDNA. To achieve uniform amplification, the gDNA must be of high quality, and must be free of contaminants that can affect the enzymatic reactions to be performed.

Sources of genomic DNA

The following sources of gDNA have been successfully tested in the laboratories at Thermo Fisher Scientific for DNA that meets the requirements for the Axiom™ Propel XPRES 384HT Assay.

Source	Sample type
Human	• Blood • Saliva • Cell line
Animal[1]	• Blood • Semen • Nasal swabs • Hair bulbs • Ear punch tissue
Plant	• Seeds • Leaves

[1] Success with sample types other than human depend on quality (degree of degradation, level of purity, and so on) and quantity of gDNA extracted.

Note: DNA derived from formalin-fixed paraffin-embedded (FFPE) blocks must not be used with this assay.

General requirements

- Starting DNA must be double-stranded for accurate concentration determination.
- DNA must be of high purity. DNA must be free of DNA polymerase inhibitors. Examples of inhibitors include high concentrations of heme (from blood) and high concentrations of chelating agents (that is, EDTA). The gDNA extraction/ purification method must create DNA that is salt-free because high concentrations of particular salts can also inhibit enzyme reactions. DNA purity indicated by OD_{260}/OD_{280} and OD_{260}/OD_{230} ratios. The OD_{260}/OD_{280} ratio should be between 1.8 and 2.0 and the OD_{260}/OD_{230} ratio should be greater than 1.5. We recommend that DNA samples that do not meet these criteria be cleaned up as described in "Clean up genomic DNA" on page 20.
- DNA must not be degraded. The average size of gDNA can be evaluated on a 1% agarose gel using an appropriate size standard control. Approximately 90% of the DNA must be greater than 10 Kb in size. Control DNA can be run on the same gel for comparison.

Note: DNA size integrity is important for successful assay performance. It is strongly advised to assess gDNA by gel electrophoresis as described in this chapter. This is of particular importance for DNA extracted from saliva and buccal cells, sample types prone to DNA degradation.

Special requirements

Preamplification area

Precautions are required when manipulating genomic DNA to avoid contamination with foreign DNA amplified in other reactions and procedures. It is recommended that genomic DNA manipulations are performed in a dedicated preamplification room or area separate from the main laboratory.

This preamplification area requires a dedicated set of pipettes and plasticware. If no dedicated area is available, use of a dedicated bench or a dedicated biosafety hood and dedicated pipettes is suggested. If no dedicated bench or biosafety hood is available, a set of dedicated pipettes is recommended.

Ideally, this preamplification area would be separate from the amplification staging area, however, these areas may be combined due to space and equipment limitations.

Evaluate the quality of genomic DNA with 1% agarose E-Gel™

We recommend this quality control step to evaluate the quality of the gDNA before starting the assay.

Equipment and reagents required

Unless otherwise indicated, all materials are available through **thermofisher.com**.

Item	Source
Invitrogen™ Mother E-Base™ Device	EBM03
Invitrogen™ Daughter E-Base™ Device (optional for running multiple gels in parallel)	EBD03
Invitrogen™ E-Gel™ *48* Agarose Gels, 1%	G800801
Invitrogen™ *Redi*Load™ Loading Buffer	750026
Invitrogen™ E-Gel™ 96 High Range DNA Marker	12352019

Guidelines for preparing the gDNA Sample Plate for gel analysis

The following guidelines are recommended when preparing the gDNA Sample Plate for gel analysis.

- Load a DNA mass of 10 ng to 20 ng per well (recommended). If lower amounts are loaded, omission of the loading dye is recommended to improve visualization. Loading ≥25-ng gDNA per well can improve the image.
- Add 3 µL of 0.1X of *Redi*Load™ Loading Buffer (*Redi*Load™ Loading Buffer dye diluted 10-fold with nuclease-free water) dye to each sample.
- Bring each sample to a total volume of 20 µL using nuclease-free water. For example, if the volume of genomic DNA is 5 µL, add 3 µL of *Redi*Load™ Loading Buffer, then bring to 20 µL total by adding 12 µL of water.
- Seal, vortex, and centrifuge briefly.

Run a 48-lane 1% agarose E-Gel™

1. Power on the E-Base™ device (red light).
2. Push **Power/Prg** to ensure that the gel base is in **EG** mode, not EP mode.
3. Insert the E-Gel™ *48* Agarose Gels, 1% into the slot.
4. Remove 2 combs.
5. Load 20 µL of gDNA samples onto the E-Gel™ *48* Agarose Gels, 1%.
6. If needed, load 15 µL of diluted E-Gel™ 96 High Range DNA Marker (1:3 dilution or ~0.34X from stock) into all marker wells.
7. Fill all empty wells with water.

8. Adjust the run time to ~27 minutes.

9. Push **Power/Prg** again.
   It changes from red to green.

When run time is reached (the ladder band reaches the end of the lane), the system automatically shuts off. The gel is ready for imaging.

E-Gel™ results

The following figure shows gel images of intact gDNA (that is appropriate for use in the Axiom™ Propel XPRES 384HT Assay) and degraded gDNA samples. For gDNA that is degraded perform a test experiment to investigate the performance of the samples in the Axiom™ Propel XPRES 384HT Assay before starting any large-scale genotyping projects.

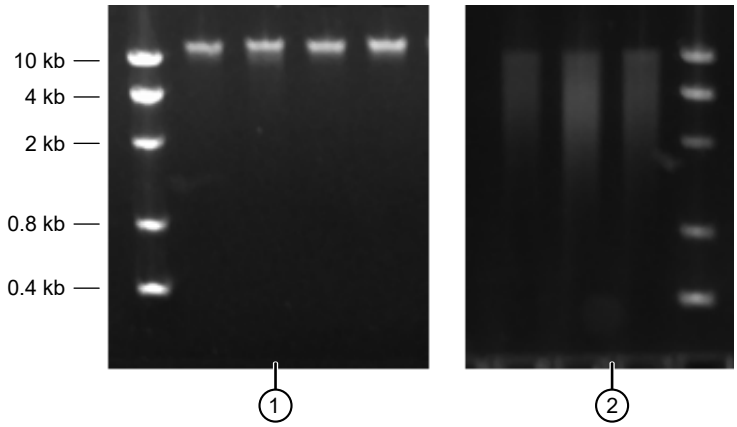


Figure 2 Gel images with intact gDNA and degraded gDNA.

① Intact samples ② Degraded samples

Genomic DNA extraction/purification methods

Genomic DNA extraction and purification methods that meet the general requirements that are outlined are expected to yield successful results. Methods that include boiling or strong denaturants are not acceptable because the DNA would be made single-stranded and can no longer be accurately quantified using a PicoGreen™-based assay.

Clean up genomic DNA

If a gDNA preparation is suspected to contain inhibitors, the following cleanup procedure can be used.

1. Add 0.5 volumes of 7.5 M NH_4OAc, 2.5 volumes of absolute ethanol (stored at –20°C), to gDNA.
2. Vortex, then incubate at –20°C for 1 hour.
3. Centrifuge at 12,000 × *g* in a microcentrifuge at room temperature for 20 minutes.
4. Remove supernatant, then wash pellet with 80% ethanol.
5. Centrifuge at 12,000 × *g* at room temperature for 5 minutes.
6. Remove the 80% ethanol, then repeat the 80% ethanol wash 1 more time.
7. Resuspend the pellet in Low EDTA TE Buffer (10-mM Tris-HCl pH 8.0, 0.1 mM EDTA).

Genomic DNA preparation

This step must be done before proceeding with the DNA amplification stages for Axiom™ Propel XPRES 384HT Workflow target preparation.

The genomic DNA (gDNA) you process using the Axiom™ Propel XPRES 384HT Assay must meet the general requirements that are listed earlier in this chapter. The amount of gDNA depends on which Axiom™ array is used in the downstream protocol. “Genomic DNA input requirements” on page 20 details the sample input requirements for Axiom™ Propel XPRES 384HT Workflow.

Genomic DNA input requirements

Sample type	Volume per well	Input mass per well	gDNA Concentration
Human	10 µL	100 ng	10 ng/µL
Diploid plants and animals	10 µL	150 ng	15 ng/µL
Polyploid plants and animals	10 µL	200 ng	20 ng/µL

Time required

Allow 30–60 minutes for reagents to thaw and 30 minutes for setup.

Equipment, consumables, and reagents required

Unless otherwise indicated, all materials are available through **thermofisher.com**.

Equipment and consumables required

Quantity	Item
As required	Adhesive seals for plates
1 each	Pipettes: single channel P10 or P20 Optional: multichannel P10 or P20
As required	Pipette tips
1	ABgene™ 96-well 2.2 mL Polypropylene Deepwell Storage Plate (AB0932). Referred to as the “96-deepwell plate” in this document.
1	Plate centrifuge
1	Plate spectrophotometer (required only if no OD measurements available for samples)
1	Vortexer

Reagents

Unless otherwise indicated, all materials are available through **thermofisher.com**. "MLS" indicates that the material is available from **fisherscientific.com** or another major laboratory supplier.

Reagent	Source
Axiom™ Genomic DNA Standard (Ref 103), –20°C (use as a positive control if genotyping human samples).	951957
Thermo Scientific™ Reduced EDTA TE Buffer (10 mM Tris-HCl pH 8.0, 0.1 mM EDTA).	Fisher Scientific™, AAJ75793AE
Positive control gDNA (if genotyping nonhuman samples).	
Ultra-pure water, from a purification system or equivalent.	**MLS**

Thaw samples and control

Thaw the following components to room temperature.

- gDNA samples
- gDNA positive control sample. For human studies, use Genomic DNA Standard (Ref 103).

To thaw, either:

- Place items on the bench top for 60 minutes.
- Thaw in a water bath.
  - Fill a small plastic dish with ultra-pure water. Do not overfill to prevent the level of the water overflowing when the sample tubes or plates are placed in the bath.
  - Thaw the sealed gDNA Sample Plate and reference sample for 30 minutes.
  - Wipe off the gDNA Sample Plate after removing from the water bath, and before removing the lid. Wiping off the gDNA Sample Plate minimizes the chances that the water enters the wells, then causes contamination or reaction failure.

Quantify and dilute test sample gDNA

1. Gently vortex (50% maximum), then centrifuge the gDNA and control DNA.
2. *Recommendation*: Quantify each sample (for example, using the Quant-iT™ PicoGreen™ dsDNA Assay Kit).
3. Using reduced EDTA TE buffer, dilute each sample to a concentration of:
   - 10 ng/µL for human DNA samples
   - 15 ng/µL for diploid plant and animal DNA samples
   - 20 ng/µL for polyploid plant and animal DNA samples
4. Seal, vortex, then centrifuge.

Note: Do *not* dilute the Genomic DNA Standard (Ref 103) control.

Aliquot the diluted samples and the control

Aliquot the diluted samples and control gDNA to the 96-deepwell plate as follows:

1. Aliquot 10 µL of each diluted gDNA sample to the appropriate well.
   This amount is the equivalent of 100 ng to 200 ng of gDNA, as required by the sample type.
2. Positive control: 10 µL of control gDNA. For genotyping arrays, we recommend including at least one positive gDNA control on each plate. For human array designs, the Genomic DNA Standard (Ref 103) control (Cat. No. 951957) *must* be used.
3. Seal, then centrifuge.

Freeze or proceed

Do one of the following:

- Store the gDNA Sample Plate at –20°C.
- Proceed to DNA amplification for Axiom™ Propel XPRES 384HT Workflow target preparation (see Chapter 4, “Target preparation with Multidrop™ Combi Reagent Dispensers for two 384HT array format plates”).

Note: If proceeding immediately to DNA amplification, you can leave the gDNA Sample Plate at room temperature.

GeneTitan™ Array Plate Registration file

Each array plate has a barcode for tracking and each row and column number identifies an individual array. The GeneTitan™ Array Plate Registration file is where you enter the sample information for each individual array of the array plate to be run. It is important to create and upload a GeneTitan™ Array Plate Registration file *before* loading the array plate and hybridization tray onto the GeneTitan™ Multi-Channel (MC) Instrument. We recommend that you create, but not upload, this file while you prepare your plate of genomic DNA. When samples are ready for hybridization, scan the array plate barcode and upload the file to Applied Biosystems™ GeneChip™ Command Console™ (GCC).

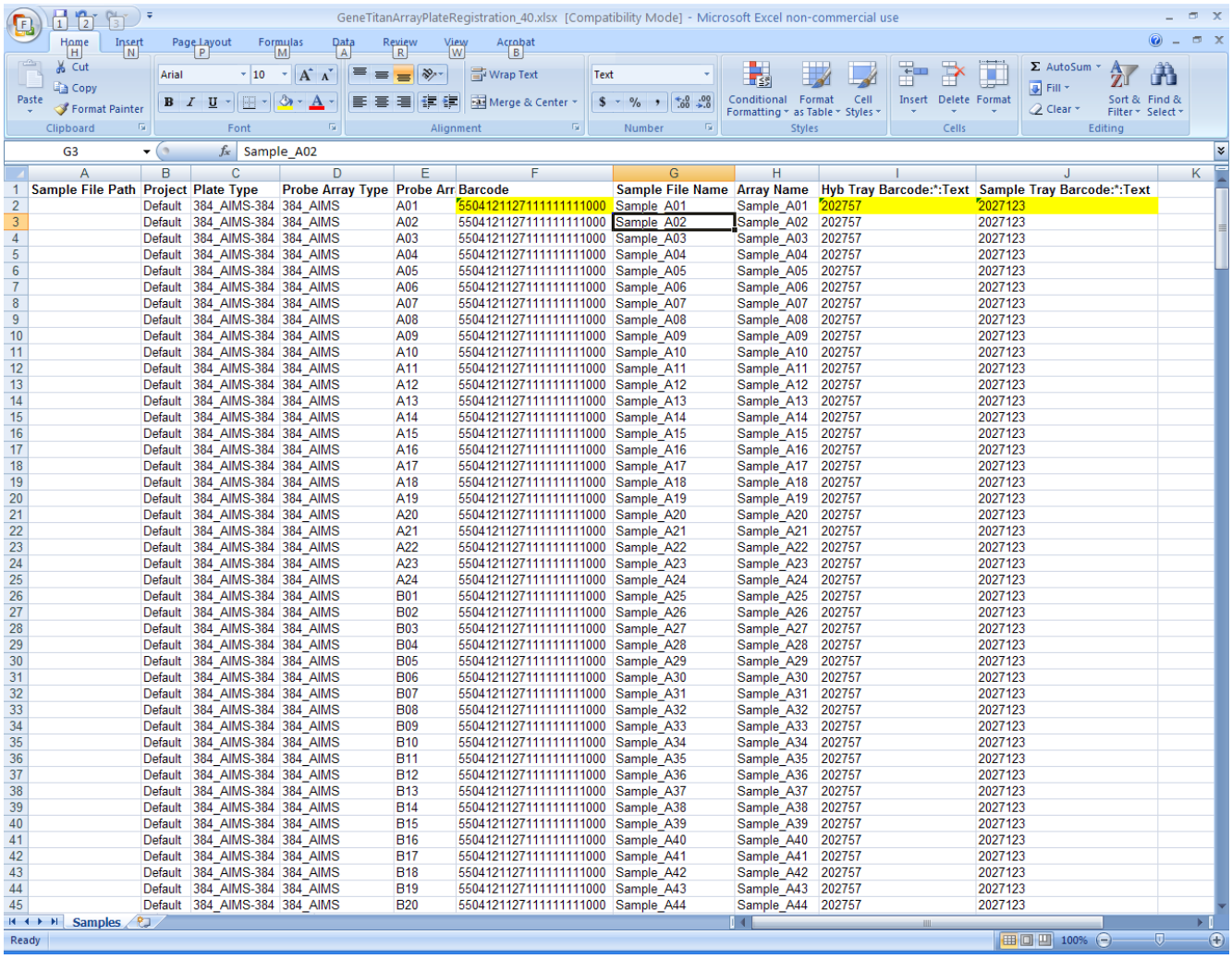
Create and save a GeneTitan™ Array Plate Registration file

This procedure creates and saves a GeneTitan™ Array Plate Registration file but does not upload the file to GeneChip™ Command Console™. The array plate and hybridization tray are scanned, and the GeneTitan™ Array Plate Registration file is uploaded when you are ready to load the plate and samples onto the GeneTitan™ MC Instrument for processing.

1. From the **Launcher** window, open **GCC Portal** ▸ **Samples** ▸ **GeneTitan™ Array Plate Registration**.
2. In the **GeneTitan Array Plate Registration** window, click to select a registration file template to use.
3. Select the **GeneTitan™ Array Plate Type** from the dropdown list.
4. Select the project for the sample files.
5. Click **Download**.
6. In the **Samples** tab of the **GeneTitan™ Array Plate Registration** window, enter a unique name for each sample and any additional information.

   For more information on the **GeneTitan™ Array Plate Registration** file, see *GeneChip™ Command Console™ User Guide*.
7. Save the file. Do not upload the file at this point.

Details for the array plate and hybridization tray scanning steps, and the GeneTitan™ Array Plate Registration file uploading steps are in Chapter 5, “Process array plates with the GeneTitan™ Multi-Channel (MC) Instrument”.



	A	B	C	D	E	F	G	H	I	J	K
1	Sample File Path	Project	Plate Type	Probe Array Type	Probe Arr	Barcode	Sample File Name	Array Name	Hyb Tray Barcode:*:Text	Sample Tray Barcode:*:Text	
2		Default	384_AIMS-384	384_AIMS	A01	550412112711111111111000	Sample_A01	Sample_A01	202757	2027123	
3		Default	384_AIMS-384	384_AIMS	A02	550412112711111111111000	Sample_A02	Sample_A02	202757	2027123	
4		Default	384_AIMS-384	384_AIMS	A03	550412112711111111111000	Sample_A03	Sample_A03	202757	2027123	
5		Default	384_AIMS-384	384_AIMS	A04	550412112711111111111000	Sample_A04	Sample_A04	202757	2027123	
6		Default	384_AIMS-384	384_AIMS	A05	550412112711111111111000	Sample_A05	Sample_A05	202757	2027123	
7		Default	384_AIMS-384	384_AIMS	A06	550412112711111111111000	Sample_A06	Sample_A06	202757	2027123	
8		Default	384_AIMS-384	384_AIMS	A07	550412112711111111111000	Sample_A07	Sample_A07	202757	2027123	
9		Default	384_AIMS-384	384_AIMS	A08	550412112711111111111000	Sample_A08	Sample_A08	202757	2027123	
10		Default	384_AIMS-384	384_AIMS	A09	550412112711111111111000	Sample_A09	Sample_A09	202757	2027123	
11		Default	384_AIMS-384	384_AIMS	A10	550412112711111111111000	Sample_A10	Sample_A10	202757	2027123	
12		Default	384_AIMS-384	384_AIMS	A11	550412112711111111111000	Sample_A11	Sample_A11	202757	2027123	
13		Default	384_AIMS-384	384_AIMS	A12	550412112711111111111000	Sample_A12	Sample_A12	202757	2027123	
14		Default	384_AIMS-384	384_AIMS	A13	550412112711111111111000	Sample_A13	Sample_A13	202757	2027123	
15		Default	384_AIMS-384	384_AIMS	A14	550412112711111111111000	Sample_A14	Sample_A14	202757	2027123	
16		Default	384_AIMS-384	384_AIMS	A15	550412112711111111111000	Sample_A15	Sample_A15	202757	2027123	
17		Default	384_AIMS-384	384_AIMS	A16	550412112711111111111000	Sample_A16	Sample_A16	202757	2027123	
18		Default	384_AIMS-384	384_AIMS	A17	550412112711111111111000	Sample_A17	Sample_A17	202757	2027123	
19		Default	384_AIMS-384	384_AIMS	A18	550412112711111111111000	Sample_A18	Sample_A18	202757	2027123	
20		Default	384_AIMS-384	384_AIMS	A19	550412112711111111111000	Sample_A19	Sample_A19	202757	2027123	
21		Default	384_AIMS-384	384_AIMS	A20	550412112711111111111000	Sample_A20	Sample_A20	202757	2027123	
22		Default	384_AIMS-384	384_AIMS	A21	550412112711111111111000	Sample_A21	Sample_A21	202757	2027123	
23		Default	384_AIMS-384	384_AIMS	A22	550412112711111111111000	Sample_A22	Sample_A22	202757	2027123	
24		Default	384_AIMS-384	384_AIMS	A23	550412112711111111111000	Sample_A23	Sample_A23	202757	2027123	
25		Default	384_AIMS-384	384_AIMS	A24	550412112711111111111000	Sample_A24	Sample_A24	202757	2027123	
26		Default	384_AIMS-384	384_AIMS	B01	550412112711111111111000	Sample_A25	Sample_A25	202757	2027123	
27		Default	384_AIMS-384	384_AIMS	B02	550412112711111111111000	Sample_A26	Sample_A26	202757	2027123	
28		Default	384_AIMS-384	384_AIMS	B03	550412112711111111111000	Sample_A27	Sample_A27	202757	2027123	
29		Default	384_AIMS-384	384_AIMS	B04	550412112711111111111000	Sample_A28	Sample_A28	202757	2027123	
30		Default	384_AIMS-384	384_AIMS	B05	550412112711111111111000	Sample_A29	Sample_A29	202757	2027123	
31		Default	384_AIMS-384	384_AIMS	B06	550412112711111111111000	Sample_A30	Sample_A30	202757	2027123	
32		Default	384_AIMS-384	384_AIMS	B07	550412112711111111111000	Sample_A31	Sample_A31	202757	2027123	
33		Default	384_AIMS-384	384_AIMS	B08	550412112711111111111000	Sample_A32	Sample_A32	202757	2027123	
34		Default	384_AIMS-384	384_AIMS	B09	550412112711111111111000	Sample_A33	Sample_A33	202757	2027123	
35		Default	384_AIMS-384	384_AIMS	B10	550412112711111111111000	Sample_A34	Sample_A34	202757	2027123	
36		Default	384_AIMS-384	384_AIMS	B11	550412112711111111111000	Sample_A35	Sample_A35	202757	2027123	
37		Default	384_AIMS-384	384_AIMS	B12	550412112711111111111000	Sample_A36	Sample_A36	202757	2027123	
38		Default	384_AIMS-384	384_AIMS	B13	550412112711111111111000	Sample_A37	Sample_A37	202757	2027123	
39		Default	384_AIMS-384	384_AIMS	B14	550412112711111111111000	Sample_A38	Sample_A38	202757	2027123	
40		Default	384_AIMS-384	384_AIMS	B15	550412112711111111111000	Sample_A39	Sample_A39	202757	2027123	
41		Default	384_AIMS-384	384_AIMS	B16	550412112711111111111000	Sample_A40	Sample_A40	202757	2027123	
42		Default	384_AIMS-384	384_AIMS	B17	550412112711111111111000	Sample_A41	Sample_A41	202757	2027123	
43		Default	384_AIMS-384	384_AIMS	B18	550412112711111111111000	Sample_A42	Sample_A42	202757	2027123	
44		Default	384_AIMS-384	384_AIMS	B19	550412112711111111111000	Sample_A43	Sample_A43	202757	2027123	
45		Default	384_AIMS-384	384_AIMS	B20	550412112711111111111000	Sample_A44	Sample_A44	202757	2027123	

Figure 3 Example of a GeneTitan™ Array Plate Registration file for Axiom™ 384HT Array Plate.

3 Set up for the Axiom™ Propel XPRES 384HT Workflow

- Required materials 26
- Guidelines for handling plates and reagents 44
- Best practices for Axiom™ Propel XPRES 384HT Workflow 45
- Multidrop™ Combi Reagent Dispenser operations 49
- Start up the Multidrop™ Combi 50
- Perform gravimetric checks 51
- Prime the cassette 54
- Flush the Multidrop™ Combi Reagent Dispenser between batches of plates 55
- Shut down the Multidrop™ Combi Reagent Dispenser 56

This chapter contains information describing the procedures, equipment, and materials required for running the Axiom™ Propel XPRES 384HT Workflow. To ensure operator safety and assay performance, operators must be familiar with this content before starting target preparation. (See Chapter 4, “Target preparation with Multidrop™ Combi Reagent Dispensers for two 384HT array format plates”.) Additional information for the Multidrop™ Combi Reagent Dispenser is in the *Multidrop™ Combi User Manual*. See “Documentation and support” on page 225.

Required materials

Equipment and materials required

The following table lists the equipment, labware, and consumables required for the assay. Exact quantities of equipment, labware, and consumables that are required is dependent on the number of plates that are processed.

Item	Used in stage
Instruments	
Multidrop™ Combi Reagent Dispenser	1, 2, 3, 5, 5A, 7
ALPS™ 3000 Automated Microplate Heat Sealer	1, 2, 3, 5
Compact Digital Microplate Shaker	1, 2, 5
One of the following ovens: • Thermo Scientific™ Heratherm™ Advanced Protocol Microbiological Incubator, capacity 66 L • BINDER™ ED 56 Drying and Heating Chamber • BINDER™ BD 56 Standard-Incubator with natural convection Oven requirements: • Must maintain a constant temperature of 37°C for at least 24 hours with a temperature accuracy of ±1°C (amplification, fragmentation, pellet drying). • Must maintain a constant temperature of 48°C for at least 24 hours with a temperature accuracy of ±1°C (off-line hybridization).	1, 2, 4, 6
Sorvall™ Legend™ XTR Centrifuge	1, 2, 4, 5, 5A, 6
VWR Signature™ High-Speed Microplate Shaker	3
96 Channel Pipetting Head (5–125 µL) with the VIAFLO 96 or VIAFLO™ 384 Base Unit and 2 single-position plate holders.	5
384 Channel Pipetting Head (2–50 µL) with the VIAFLO™ 384 Base Unit and 2 single-position plate holders.	5A, 6
Thermal cycler, ProFlex™ 2 x 384-well PCR System	6
Vortex mixer	1, 2, 3, 5, 5A, 6, 7
Mini centrifuge	2, 7
Freezer, –20°C	3
Electronic pipettor for serological pipettes	1, 2, 3, 5, 7
P1000 pipette	1, 2, 3, 5, 7
Fume hood	3, 4, 5, 5A, 6

(continued)

Item	Used in stage
Balance with: • readability of 0.01g or finer. • a maximum capacity of at least 300g.	Gravimetric checks
minION™2 Ionizing Air Blower	1, 2, 3, 5, 5A, 7
GeneTitan™ ZeroStat AntiStatic Gun	7
Digital timer	1, 2
Labware and consumables	
Multidrop™ Combi SMART 2 Standard tube dispensing cassette[1]	1, 2, 3, 5, 5A, 7
Multidrop™ Combi SMART 2 Small tube plastic tip dispensing cassette[1]	1, 2
384-Well PCR Plate Collar	5, 5A, 6
ABgene™ 96-well 2.2 mL Polypropylene Deepwell Storage Plate (Referred to as "96-deepwell plate" in this document.)	1[2]
PCR Plate, 384-well, raised chimney	5
ABgene™ 384-Well 250 µL Polypropylene Storage Plate	5A
Greiner Bio-One™ UV-Star™ 384-Well Microplate Flat Bottom	5A
Plate Alignment Tool	6
GeneTitan™ consumables for 384 format	7
50-mL conical tube	Multidrop™ Combi setup procedure, and for master mix preparation
250-mL conical bottle (Nunc™ 250 mL Wide Mouth Conical Centrifuge Tube)	Master mix preparation
500-mL bottle (Nalgene™ Wide-Mouth HDPE Economy Bottles with Closure), or equivalent glass or polypropylene bottle, minimum 500 mL capacity (for Precipitation Master Mix dispensing).	3
Matrix™ Reagent Reservoir, 25 mL	Multidrop™ Combi cleaning procedure
Serological pipettes (5, 10, 25 and 50 mL)	1, 2, 3, 5, 7
P1000 pipette tips	1, 2, 3, 5, 7
Integra VIAFLO XYZ GripTips™, 125 µL, 5 XYZ Racks of 384 Tips, Sterile, Filter	5, 5A, 6
Easy Peel Seal (for ALPS™ 3000 Automated Microplate Heat Sealer)	1, 2, 3, 5

(continued)

Item	Used in stage
MicroAmp™ Clear Adhesive Film	5, 5A
Laboratory tissues	as needed

[1] Each reagent must use a dedicated cassette.
[2] The same 96-deepwell plate is used through Stage 5.

Labware and consumable ordering information

Unless otherwise indicated, all materials are available through **thermofisher.com**. "MLS" indicates that the material is available from **fisherscientific.com** or another major laboratory supplier.

Table 1 Labware and consumable images and ordering information.

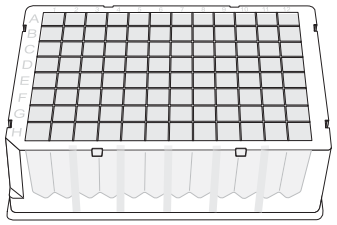
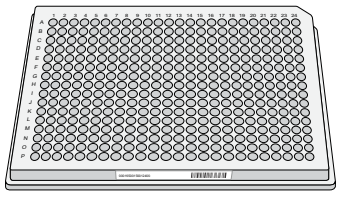
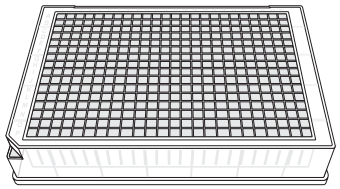
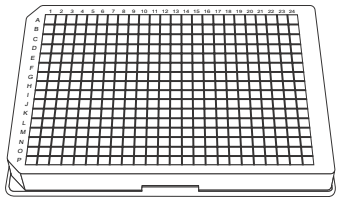
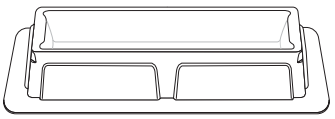
Item	Source	Image
ABgene™ 96-well 2.2 mL Polypropylene Deepwell Storage Plate, square wells, V-bottom **Note:** This plate is referred to as "96-deepwell plate" throughout this document.	Fisher Scientific™, AB0932	
PCR Plate, 384-well, raised chimney	Fisher Scientific™, AB0937	
ABgene™ 384-Well 250 µL Polypropylene Storage Plate	Fisher Scientific™, AB1178	
Greiner Bio-One™ UV-Star™ 384-Well Microplate Flat Bottom	Axiom™ 384HT Consumables Kit for QC, 902289	
Matrix™ Reagent Reservoir, 25 mL	Fisher Scientific™, 809311	

Table 1 Labware and consumable images and ordering information. *(continued)*

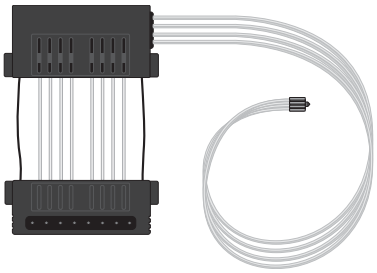
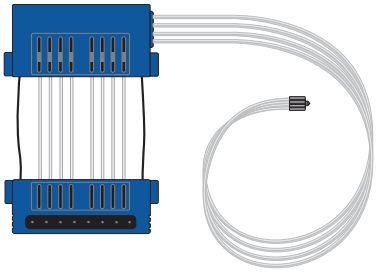
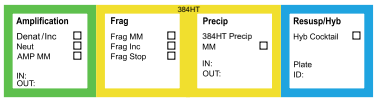
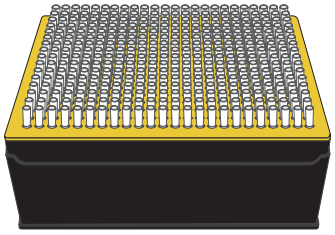
Item	Source	Image
SMART 2 Standard tube dispensing cassette	N15137, single cassette N15138, 5-pack	
SMART 2 Small tube plastic tip dispensing cassette	N15133, single cassette N15134, 5-pack	
Pipette tips, 1,000 µL	**MLS**	
Serological pipettes, following sizes: • 5 mL • 10 mL • 25 mL • 50 mL	**MLS**	
Electronic pipettor (for serological pipettes)	**MLS**	
Axiom™ Propel 384HT Tracker Label	952388	
XYZ GripTips™, 125 µL, 5 XYZ Racks of 384 Tips, Sterile, Filter	INTEGRA Biosciences, 6465	
50-mL centrifuge tubes	**MLS**	

Table 1 Labware and consumable images and ordering information. *(continued)*

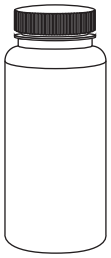
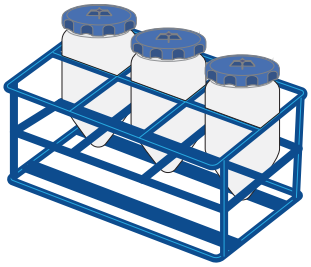
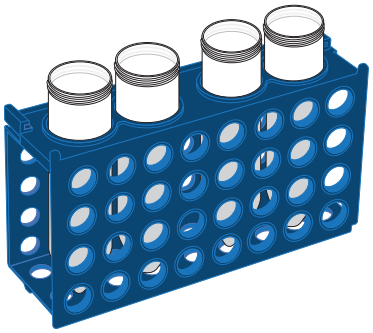
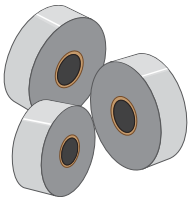
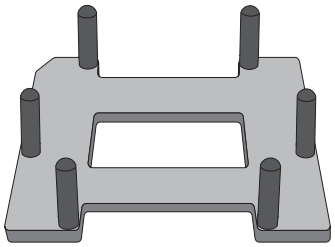
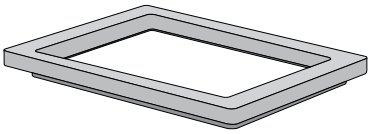
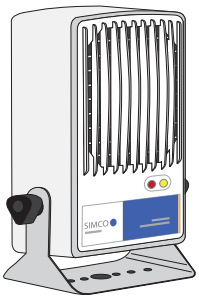
Item	Source	Image
Nunc™ 250 mL Wide Mouth Conical Centrifuge Tube	376814	
Nalgene™ Wide-Mouth HDPE Economy Bottles with Closure	332189-0016	
Nunc™ Conical Tube Rack	374179	
Fisherbrand™ 4-Way Tube Rack	Fisher Scientific™, 03-448-12	
BTL Safety Carrier, black **Note:** This carrier is recommended as the secondary liquid waste container for the Multidrop™ Combi.	Fisher Scientific™, 50-109-4650	

Table 1 Labware and consumable images and ordering information. *(continued)*

Item	Source	Image
Easy Peel Seal **Note:** The Easy Peel Seal is the sealing material used in the ALPS™ 3000 Automated Microplate Heat Sealer.	AB-3739	
MicroAmp™ Clear Adhesive Film	4306311	
Plate Alignment Tool	13-0401	
384-Well PCR Plate Collar	952400	
minION™2 Ionizing Air Blower (or equivalent)	Simco-Ion™ Technology, 4011425, or **MLS**	
GeneTitan™ ZeroStat AntiStatic Gun and Ion-Indicator Cap **Note:** The GeneTitan™ ZeroStat AntiStatic Gun can be used as an alternative if the minION™2 Ionizing Air Blower is not available.	74-0014	

Axiom™ Propel XPRES Reagent Kit components

IMPORTANT! The Applied Biosystems™ Axiom™ Propel XPRES Reagent Kit, 2x384HT is for single use only. Large fill reagent kits are configured to include priming volumes for the Multidrop™ Combi cassettes and is incorporated into the master mix formulations. Discard all excess reagents after use.

Each Axiom™ Propel XPRES Reagent Kit, 2x384HT, Cat. No. 952352, is sufficient for 2 Axiom™ Array Plates (384HT array format).

Table 2 Components of the Axiom™ Propel XPRES Reagent Kit, 2x384HT.

Component	Part No.[1]	Storage
Axiom™ Propel XPRES Reagent Kit Module 1 for 384HT only	**952351**	–25°C to –15°C
Axiom™ Propel 10X Denat Solution	952176	
Axiom™ Propel Neutral Solution	952173	
Axiom™ Propel Water	952177	
Axiom™ Propel Amp Solution	952174	
Axiom™ XPRES Amp Enzyme	952349	
Axiom™ Propel Reagent Kit Module 2-1 for 96F or 384HT—Box 1 of 2	**952263**	–25°C to –15°C
Axiom™ Propel Frag Enzyme	952181	
Axiom™ Propel 10X Frag Buffer	952179	
Axiom™ Propel Precip Solution 2	952178	
Axiom™ Propel Hyb Buffer	952182	
Axiom™ Propel Hyb Solution 1	952183	
Axiom™ Propel Reagent Kit Module 2-2 for 96F or 384HT—Box 2 of 2	**952265**	2°C to 8°C
Axiom™ Propel Frag Diluent	952184	
Axiom™ Propel Frag Reaction Stop	952190	
Axiom™ Propel Precip Solution 1	952203	
Axiom™ Propel Resuspension Buffer	952206	
Axiom™ Propel Hyb Solution 2	951979	
Module 3	—	Room temperature
Axiom™ Wash Buffer A	901446	
Axiom™ Wash Buffer B	901447	
Axiom™ Water	901578	

Table 2 Components of the Axiom Propel XPRES Reagent Kit, 2x384HT. *(continued)*

Component	Part No.[1]	Storage
Axiom™ Propel Fast Wash Reagent Kit Module 4-1 for 96F or 384HT—Box 1 of 2	**952369**	–25°C to –15°C
Axiom™ Propel Ligation Buffer	952208	
Axiom™ Fast Ligation Enzyme	952367	
Axiom™ Propel Ligation Solution 1	952212	
Axiom™ Propel Probe Mix 1	952213	
Axiom™ Propel Stain Buffer	952214	
Axiom™ Propel Stabilize Solution	952215	
Axiom™ Propel Reagent Kit Module 4-2 for 96F or 384HT—Box 2 of 2	**952268**	2°C to 8°C
Axiom™ Propel Wash A	952218	
Axiom™ Propel Probe Mix 2	952217	
Axiom™ Propel Ligation Solution 2	952216	
Axiom™ Propel Stain 1-A	952219	
Axiom™ Propel Stain 2-A	952231	
Axiom™ Propel Stabilize Diluent	952248	
Axiom™ Water	952177	
Axiom™ Propel Hold Buffer	952254	
Axiom™ Propel Stain 1-B	952258	
Axiom™ Propel Stain 2-B	952260	

[1] Component Part Numbers are for identification puproses only. Kit components are not available for purchase separately.

Master mix preparation for large-fill reagent kits

IMPORTANT! The Applied Biosystems™ Axiom™ Propel XPRES Reagent Kit, 2x384HT is for single use only. This large fill reagent kit is configured to include priming volumes for the Multidrop™ Combi cassettes and is incorporated into the master mix formulations. Discard all excess reagents after use.

Additional reagents and materials required

Unless otherwise indicated, all materials are available through **thermofisher.com**. "MLS" indicates that the material is available from **fisherscientific.com** or another major laboratory supplier.

Reagent	Source	Where used
2-Propanol, anhydrous, 99.5% (Isopropanol)	Sigma-Aldrich™, 278475	Stage 3: Precipitate the DNA
E-Gel™ *48* Agarose Gels, 4%	G800804	Stage 5A: In-process QC
TrackIt™ Cyan/Orange Loading Buffer	10482028	
25 bp DNA Ladder	931343	
UltraPure™ DNase/RNase-Free Distilled Water	10977023	
Reagent Alcohol, Certified, 70% (v/v) (Ethanol solution 70%, reagent grade)	Fisher Scientific™, LC222102	Multidrop™ Combi cassette cleaning

GeneTitan™ 384HT bulk consumables

GeneTitan™ 384HT consumables are available in bulk quantities sufficient to process 10 Axiom™ 384HT array format plates. These trays are required for processing Axiom™ 384HT array format plates on the GeneTitan™ MC Instrument.

See Appendix A, “Recommended techniques for GeneTitan™ MC Instrument operation” for information on aligning and loading trays onto the GeneTitan™ MC Instrument.

IMPORTANT! All covers must have barcodes. Discard any cover without a barcode.

Table 3 GeneTitan™ 384HT bulk consumable kits available.

Contents	Quantity	Source
Axiom™ GeneTitan™ Scan Trays and Covers, Bulk, 384HT	10	952381
Axiom™ GeneTitan™ Hybridization Trays, Bulk, 384HT	10	952382
Axiom™ GeneTitan™ Covers for Stain Trays, Bulk, 384HT	50	952383
Axiom™ GeneTitan™ Barcoded Stain Trays, Bulk, 384HT	50	952387

Table 4 GeneTitan™ 384HT consumables identification.

Contents[1]	Part number
384-Layout GeneTitan™ Stain Tray (Stain 1)	501279
384-Layout Axiom™ Stain 2 Tray	501394
384-Layout Axiom™ Stabilization Tray	501396
384-Layout Axiom™ Ligation Tray	501398
384-Layout GeneTitan™ Hybridization Tray	501278
384-Layout GeneTitan™ Scan Tray	501280
384-Layout GeneTitan™ Scan and Stain Tray Cover	501315

[1] See Table 5 for detailed descriptions of each component.

Table 5 GeneTitan™ tray consumables.

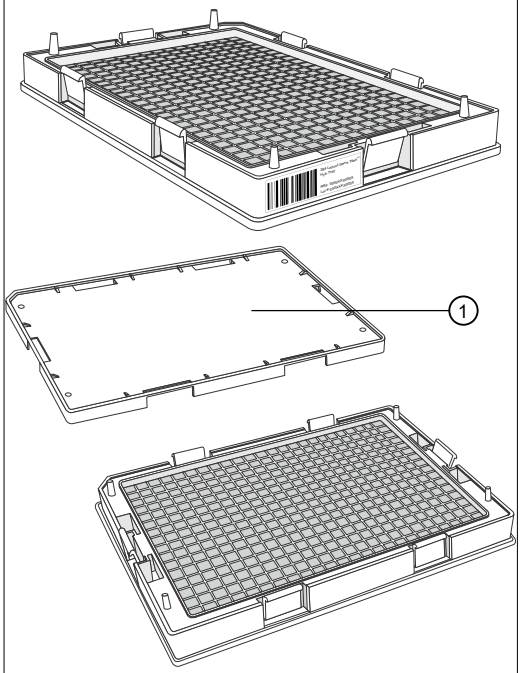
Item	Part No.	Image	Details
384-Layout GeneTitan™ Hybridization Tray	501278	 ① Hybridization tray cover to be discarded.	The 384-Layout GeneTitan™ Hybridization Trays are packaged in white pouches with the label "384 Layout GeneTitan™ Hyb Tray" ref# 501278 (pouch)/902278 (box) The hybridization trays are packaged with a protective cover that should be discarded before use. 384 hybridization tray cover, Part No. 203006

Table 5 GeneTitan tray consumables. *(continued)*

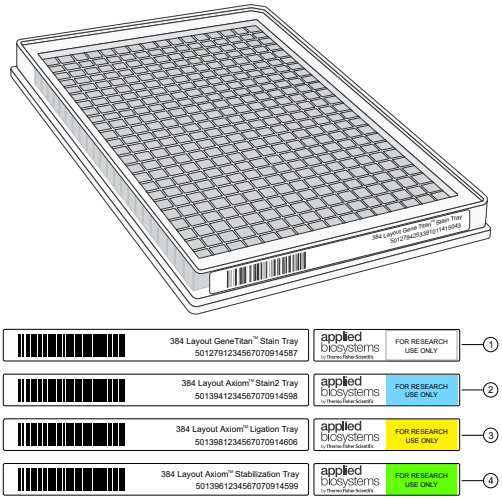
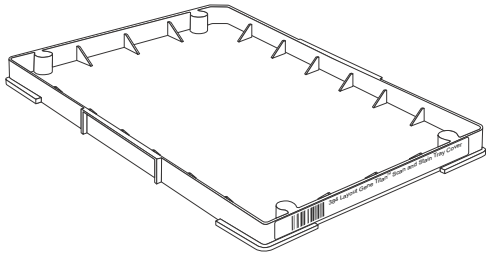
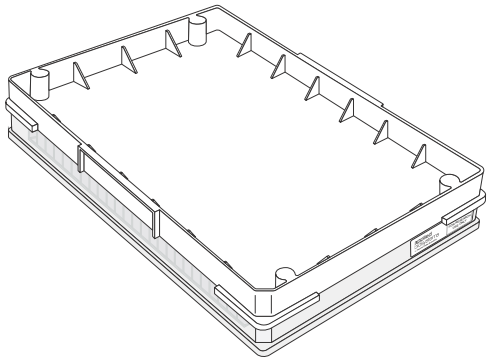
Item	Part No.	Image	Details
384 layout stain trays[1]	501279 - Stain 1 501394 - Stain 2 501398 - Ligation 501396 - Stabilization	 ① Stain 1 tray ② Stain 2 tray ③ Ligation tray ④ Stabilization tray	The stain trays are packaged in zip-top bags to keep them free of dust. Each stain tray is uniquely barcoded. **IMPORTANT!** Each stain tray is labeled with a name and an individual barcode. Ensure that you always use the appropriate tray with the correct reagent. Failure to do so can result in the wrong stain in the wrong location on the GeneTitan™ MC Instrument and assay failure. When transferring the trays to the GeneTitan™ MC Instrument, ensure that the trays are placed in the proper location in the drawer. Failure to do so results in an error and the GeneTitan™ MC Instrument will not proceed with processing trays.
384-Layout GeneTitan™ Scan and Stain Tray Cover	501315		The 384-Layout GeneTitan™ Scan and Stain Tray Covers are provided to prevent evaporation of the GeneTitan™ reagents in stain trays and the array holding buffer in the scan tray. The GeneTitan™ scan and stain tray covers are barcoded.
Stain tray cover, shown on top of the stain tray	Cover 501315		The stain trays must be placed in the GeneTitan™ MC Instrument with the stain tray cover.

Table 5 GeneTitan tray consumables. *(continued)*

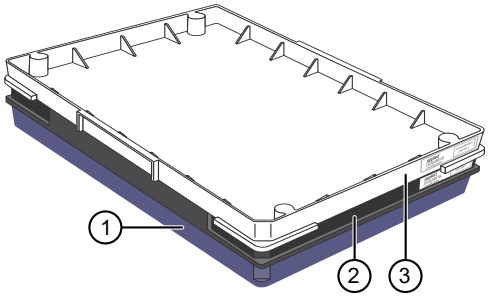
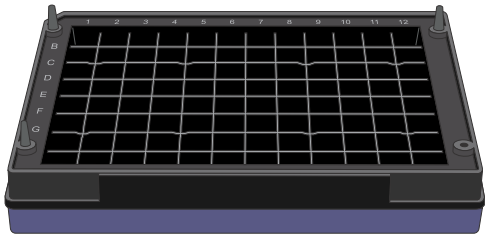
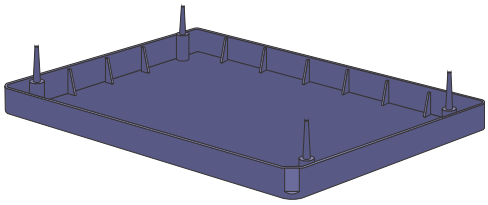
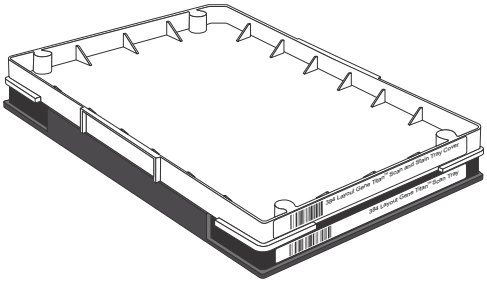
Item	Part No.	Image	Details
GeneTitan™ scan tray[1]		 ① Scan tray protective base ② 384-Layout GeneTitan™ Scan Tray ③ Barcoded scan tray cover	The Axiom™ scan tray package includes the following: • The GeneTitan™ scan tray includes a scan tray cover. The tray cover should be used to cover the scan tray before placing the tray in the GeneTitan™ MC Instrument. • The scan tray must be protected at all times from damage or exposure to dust. The scan tray must be in the blue scan tray protective base at all times except when loaded into the GeneTitan™ MC Instrument. • The blue scan tray protective base in the package is used to protect the bottom of the scan tray glass from damage. Remove the protective base from the scan tray before loading the scan tray with the scan tray cover in the GeneTitan™ MC Instrument.
GeneTitan™ scan tray on blue scan tray protective base			This combination of the GeneTitan™ scan tray on the protective blue scan tray protective base is to be used during the Multidrop™ dispensing step.
Blue scan tray protective base	202096		The blue scan tray protective base in the package is used to protect the bottom of the scan tray glass from damage. The blue scan tray protective base is distinct from the blue array plate protective base and must not be used with the array plate. Remove the protective base from the scan tray before loading in the GeneTitan™ MC Instrument

Table 5 GeneTitan tray consumables. *(continued)*

Item	Part No.	Image	Details
GeneTitan™ scan tray with cover	Scan tray 501280 Cover 501315		The GeneTitan™ scan tray must be loaded with the scan tray cover into the GeneTitan™ MC Instrument. **Do not** load the scan tray with the protective base.

[1] After aliquoting the appropriate solution to each tray type, the tray should be loaded into the GeneTitan™ MC Instrument with the barcode facing away from the operator. That is, the barcode should be on the back side.

Other equipment, consumables, and reagents required

Microplate dispenser

Unless otherwise indicated, all materials are available through **thermofisher.com**. "MLS" indicates that the material is available from **fisherscientific.com** or another major laboratory supplier.

Item	Source
(Recommended) Multidrop™ Combi Reagent Dispenser with SMART 2 option	5840320
Multidrop™ Combi Reagent Dispenser	5840300

Plate centrifuge

The plate centrifuges listed are recommended for the Axiom™ Propel XPRES 384HT Workflow. (See Table 6.) When centrifuging and drying pellets, the centrifuge must be able to centrifuge plates at:

- Rcf: 3,200 × *g* with an appropriate rotor-bucket combination
- Temperature: 4°C

Relative centrifugal force (rcf) is calculated using the following formula:

$rcf = (1.118 \times 10^{-5}) R S^2$

Where R is the radius of the rotor in centimeters, and S is the speed of the centrifuge in revolutions per minute.

In addition, the bottom of the rotor buckets must be soft rubber to ensure that the 96-deepwell plates do not crack. Do not use buckets where the plates sit directly on a metal or hard plastic bottom.

Table 6 Plate centrifuge recommendations for the Axiom™ Propel XPRES 384HT Workflow.

Item	Source
Sorvall™ Legend™ XT/XF Centrifuge Series[1]	
Sorvall™ Legend™ XTR Centrifuge (refrigerated), with:	75004521 (120 V, 60 Hz) 75004523 (230 V, 50–60 Hz, USA and Canada) 75004520 (230 V, 50 Hz)
• TX-1000 Swinging Bucket Rotor Body • Adapter for TX-1000 Swinging Bucket Rotor • Buckets for TX-1000 Rotor	• 75003017 • 75007303 (pack of 4) • 75003001 (set of 4)
Sorvall™ Legend™ XFR Centrifuge (refrigerated), with:	75004539 (120 V, 60 Hz) 75004541 (230 V, USA and Canada) 75004538 (230 V, 50–60 Hz)
• TX-1000 Swinging Bucket Rotor Body • Adapter for TX-1000 Swinging Bucket Rotor • Buckets for TX-1000 Rotor	• 75003017 • 75007303 (pack of 4) • 75003001 (set of 4)
Other	
Sorvall™ X4R Pro-MD Centrifuge, with:	75009520 (220 V-240 V 50 Hz/230 V, 60 Hz) 75009521 (120 V, 50–60 Hz) 75009620 (220 V, 60 Hz)
• TX-1000 Swinging Bucket Rotor Body • Adapter for TX-1000 Swinging Bucket Rotor • Buckets for TX-1000 Rotor	• 75003017 • 75007303 (pack of 4) • 75003001 (set of 4)
Eppendorf™ Centrifuge 5810 R, with:	Fisher Scientific™, 022625551 (230 V, 50–60 Hz) Fisher Scientific™, 022625501 (120 V, 50–60 Hz, 15 A) Fisher Scientific™, 022625101 (120 V, 50–60 Hz, 20 A)
• Rotor A-4-81, with 4 MTP/Flex buckets	• Fisher Scientific™, 022638807 (rotor)

[1] XFR = floor model and XTR = bench model

Plate sealer

The following plate sealer meets the requirements for the Axiom™ Propel XPRES 384HT Workflow.

Item	Source
Thermo Scientific™ ALPS™ 3000 Automated Microplate Heat Sealer[1]	AB3000

[1] The Easy Peel Seal is the sealing material used in the ALPS™ 3000 Automated Microplate Heat Sealer.

Oven requirements

We recommend using either the Thermo Scientific™ Heratherm™ Advanced Protocol Microbiological Incubator, BINDER™ ED 56 Drying and Heating Chamber, or the BINDER™ BD 56 Standard-Incubator with natural convection that are listed in the following table. If another oven is used, it must meet the following requirements.

- Be able to maintain a constant temperature of 37°C for at least 24 hours, and have a temperature accuracy of ±1°C, and
- Be able to maintain a constant temperature of 48°C for at least 24 hours, and have a temperature accuracy of ±1°C.

Item	Source
Thermo Scientific™ Heratherm™ Advanced Protocol Microbiological Incubator, capacity 66 L • 120V, 60 Hz • 230V, 50/60 Hz	 • 51028066 • 51028133
BINDER™ ED 56 Drying and Heating Chamber • ED056UL-120V Voltage: 120 V 1~60 Hz • ED056-230V Voltage: 230 V 1~50/60 Hz	 • BINDER™, 9010-0334 • BINDER™, 9010-0333
BINDER™ BD 56 Standard-Incubator with natural convection • BD056UL-120V Voltage: 120 V 1~60 Hz • BD056-230V Voltage: 230 V 1~50/60 Hz	 • BINDER™, 9010-0324 • BINDER™, 9010-0323

Shakers

The following shakers are required for use in the Axiom™ Propel XPRES 384HT Workflow.

Unless otherwise indicated, all materials are available through **thermofisher.com**. "MLS" indicates that the material is available from **fisherscientific.com** or another major laboratory supplier.

IMPORTANT! Both types of shakers (Thermo Scientific™ Digital Microplate Shaker and VWR Signature™ High-Speed Microplate Shaker) are required and are not interchangeable. Use only the shaker specified in the instructions for the assay stage.

Item	Source
Either: • Thermo Scientific™ Compact Digital Microplate Shaker • Thermo Scientific™ Digital Microplate Shaker	 • 88880023 or 88880024 • 88882005 or 88882006
VWR Signature™ High-Speed Microplate Shaker	VWR, 10027-220

Vortex mixer

A vortex mixer is required for use in the Axiom™ Propel XPRES 384HT Workflow.

Unless otherwise indicated, all materials are available through **thermofisher.com**. "MLS" indicates that the material is available from **fisherscientific.com** or another major laboratory supplier.

Item	Source
Vortex mixer	**MLS**

Mini centrifuge

A mini centrifuge is required for use in the Axiom™ Propel XPRES 384HT Workflow.

Unless otherwise indicated, all materials are available through **thermofisher.com**. "MLS" indicates that the material is available from **fisherscientific.com** or another major laboratory supplier.

Item	Source
Mini centrifuge	**MLS**

Liquid handler

Two liquid handler configurations are recommended to perform the 384-well PCR plate merge (Stage 5), the in-process QC step (Stage 5A), and the transfer to hybridization tray step (Stage 6).

Table 7 Recommended liquid handler setup.

Item	Source
VIAFLO™ 384 Base Unit, with:	INTEGRA Biosciences, 6031
☐ 384 Channel Pipetting Head (2 µL to 50 µL)	INTEGRA Biosciences, 6136
☐ Spring Loaded Plate Holder A with slide function—384 well plate offset	INTEGRA Biosciences, 6215
☐ Spring Loaded Plate Holder B with slide function—384 well plate offset	INTEGRA Biosciences, 6220
☐ Installation and Training VIAFLO 384 (required)	INTEGRA Biosciences, 999111
VIAFLO™ 96 Base Unit or VIAFLO™ 384 Base Unit, with:	INTEGRA Biosciences, 6001 or 6031
☐ 96 Channel Pipetting Head (5 µL to 125 µL)	INTEGRA Biosciences, 6102
☐ Spring Loaded Plate Holder A with slide function—384 well plate offset	INTEGRA Biosciences, 6215
☐ Spring Loaded Plate Holder B with slide function—384 well plate offset	INTEGRA Biosciences, 6220
☐ Installation and Training VIAFLO 96 (required)	INTEGRA Biosciences, 999110

The minimum requirement for the Axiom™ Propel XPRES 384HT Workflow is a VIAFLO™ 384 Base Unit and both the 96 Channel Pipetting Head (5–125 µL) and 384 Channel Pipetting Head (2–50 µL). This option requires changing to the appropriate pipetting head that is used for the selected VIAFLO™ method.

For instructions on how to change the pipetting head on the VIAFLO™ Instrument:

- See the VIAFLO 96/384 Operating Instructions (Pub. No. 125950) posted on the INTEGRA Biosciences website.
- Contact INTEGRA Biosciences support.

Table 8 Minimum liquid handler requirement.

Item	Source
VIAFLO™ 384 Base Unit, with:	INTEGRA Biosciences, 6031
☐ 384 Channel Pipetting Head (2 µL to 50 µL)	INTEGRA Biosciences, 6136
☐ 96 Channel Pipetting Head (5 µL to 125 µL)	INTEGRA Biosciences, 6102
☐ Spring Loaded Plate Holder A with slide function—384 well plate offset	INTEGRA Biosciences, 6215
☐ Spring Loaded Plate Holder B with slide function—384 well plate offset	INTEGRA Biosciences, 6220
☐ Installation and Training VIAFLO 384 (required)	INTEGRA Biosciences, 999111

Thermal cycler recommendation and protocol

Verified thermal cyclers	Source
Applied Biosystems™ ProFlex™ 2 x 384-well PCR System[1]	4484077

[1] The ramp rate on the ProFlex™ 2 x 384-well PCR System can be programmed to 3.0C/sec (maximum).

IMPORTANT! Always use the heated lid option when programming a protocol. See the appropriate thermal cycler user guide for programming information.

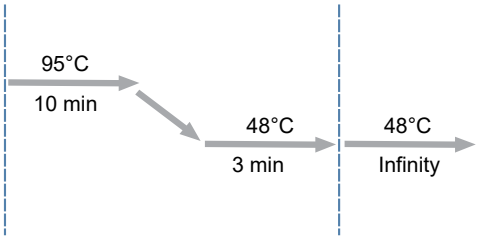


Figure 4 Axiom Denature thermal cycler protocol (Stage 6).

WARNING! Evaporation during denaturation can negatively affect assay performance. Use the recommended thermal cycler consumables and sealing film to eliminate condensation and evaporation.

Spectrophotometer

Specifications: Must be able to read DNA samples using UV/VIS absorbance setting at 260 nm, 280 nm and 320 nm wavelengths.

We recommend using the following spectrophotometer, or equivalent.

Item	Source
Multiskan™ Sky Microplate Spectrophotometer	51119600

Fume hood

Some procedures in the assay require the use of adequate local or general ventilation to keep airborne concentrations low. A fume hood is a way to achieve the desired concentration. Thus, a fume hood is strongly recommended for several steps of this assay.

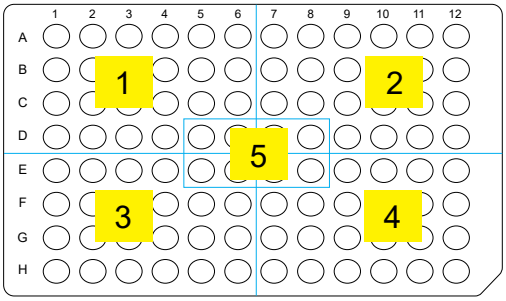
Guidelines for handling plates and reagents

Blot-dry

- Before sealing plates, check the top of the plate to ensure that there are no droplets. If droplets are present, blot-dry the top of the plate before sealing to ensure a tight seal.
  - To remove droplets before sealing, overlay a sheet of laboratory tissue across the top of the plate and gently pat down to dry. Discard the tissue after blotting.
  - Ensure that the top of the plate is dry and seal the plate as usual.

Vortex

- Plates: Vortex 1 second each corner, and 1 second in the center at the maximum setting.



- Reagent vials and bottles:
  - Reagent vials with a capacity of less than 2 mL, vortex the vials 3 times, 1 second each time at the maximum setting.
  - Reagent bottles with a capacity larger than 2 mL, vortex the bottle for ~20–30 seconds. If precipitates are seen on the inside the bottle, follow the instructions in the reagent handling section for the appropriate stage.

Centrifuge

When instructed to perform a brief centrifugation step of plates or reagent vials, follow these guidelines, unless otherwise instructed.

- Plates:
  - Centrifuge plates to 675 × *g* for 30 seconds at room temperature.
  - Do not centrifuge for more than 1 minute.
- Reagent vials:
  - Briefly centrifuge for 3 seconds on the microcentrifuge.

Best practices for Axiom™ Propel XPRES 384HT Workflow

Preamplification/amplification staging area

Precautions are required when manipulating genomic DNA to avoid contamination with foreign DNA amplified in other reactions and procedures. It is recommended that genomic DNA manipulations are performed in a dedicated preamplification room or in an area separate from the main laboratory.

This preamplification area must have a dedicated set of pipettes and plasticware. If no dedicated area is available, use of a dedicated bench or a dedicated biosafety hood and dedicated pipettes is suggested. If no dedicated bench or biosafety hood is available, a set of dedicated pipettes is recommended.

General guidelines

CAUTION! To obtain optimal performance, the use of recommended instruments and procedures described in this user guide is required. Using instruments other than those recommended, such as a different microplate sealer or high-speed shaker, can affect assay performance. Additional optimization and verification is required if non-recommended instruments are used.

- Read and follow all safety precautions that are described in Appendix G, “Safety”.
- Practice standard pre- and post-lab segregation.
- Follow manufacturer's recommendations on calibration and preventive maintenance schedules for all instruments used in the workflow.
- Use a temperature monitoring system to track the performance of incubators, freezers, and refrigerators.
- To prevent liquid from overflowing from the reagent bottles, always insert the pipette slowly into the bottle.
- Ensure that the plastic consumables used in the procedures are free of defects (such as cracks) before using them.
- Mitigate static electricity by using the de-ionizing device where specified in the user guide.
- Maintain the same order of sample plate processing throughout the target preparation stages for consistency.
- Maintain proper traceability of the source hybridization-ready targets when merging the four 96-deepwell plates into one 384-format PCR plate.
- Mark the plates to indicate that the reagent addition by the Multidrop™ Combi has occurred. A label, such as the one shown in the following image, is used to help tracking.

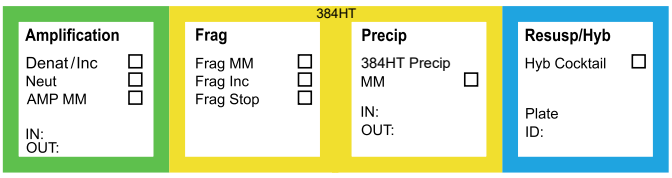


Figure 5 Axiom™ Propel 384HT Tracker Label, Cat. No. 952388.

- Maintain a clear work area and designate appropriate space to help identify the plates before and after reagent dispensing.
- It is strongly recommended to use the GeneTitan™ Barcoded Stain Trays on the GeneTitan™ MC Instrument. The unique barcodes on the stain trays prevent errors when placing the trays in the GeneTitan™ MC Instrument during array processing.

Master mix preparation guidelines

- Follow the specified order of reagent addition when preparing master mixes.
- Use the recommended transfer apparatus (such as serological pipette or pipette) listed in the master mix preparation sections.
- Always use pipettes that have been calibrated.
- If multiple transfers are required to pipet the reagent, we recommend dividing the transfers into equal or simple volumes for ease of operation.

  Example: To transfer 1.13 mL of stain 1A using a P1000 pipette, pipet 565 µL of stain 1A from the reagent tube 2 times.
- Use conical bottles as reagent reservoirs to minimize the dead volumes in the reagent reservoir, unless otherwise instructed.

Multidrop™ Combi use guidelines

- Use dedicated cassettes for each type of master mix/reagent to prevent cross-contamination. Trace amounts of chemicals can be absorbed by the tubing.
- Before priming the Multidrop™ cassettes with reagents, ensure that the startup and gravimetric check procedures have been performed.
- Follow the recommended cleaning procedure for cassette to prevent clogging and contamination.
- Handle the dispense tips (nozzles) of the Multidrop™ cassette with care to prevent damage of the tips. Damaged tips could cause liquid stream to split and cause inaccuracy of dispense.
- Place the cassette tubing bundle at the bottom of the reagent bottle to maximize reagent usage.
- Avoid having the cassette be in contact with dust or lint—excessive exposure can cause clogging of the tubing or tips. Cover the tips with the lid when storing.
- Avoid touching any part of cassette that comes in contact with the reagent.
- When cassette is not in used for more than 1 hour, put the cassette in the "Rest" position.
- The small tubing cassettes have a smaller internal diameter than the standard tubing cassettes. Always follow the recommended Multidrop™ reagent priming and shutdown procedures.
- Use a sturdy tube rack or secondary container to hold the reagent tube or bottle to prevent spillage.
- Use clean labware at the start of every run.
- Ensure that the sample plate is seated properly into the Multidrop™ stage before dispensing.
- Place a minION™2 Ionizing Air Blower at each dispensing station and power on before dispensing reagent into the sample plates.

- If droplets are seen on the top grid of the plate, carefully blot off the droplets by laying a piece of clean laboratory tissue over the top of the plate to absorb the droplets.
- Avoid large spillage or splashing of liquid which can get into the electronic components of the instrument and cause malfunction.

IMPORTANT!

- Static electricity can cause the liquid stream to stray and result in inaccurate delivery of the reagent into the samples. The use and proper placement of the ion blower is critical to the success of the assay.
- Place consumables (96-deepwell plates, QC plates, scan trays, and stain trays) to be deionized within a distance of 12" x 36" from the ion blower for at least 10 seconds before using.

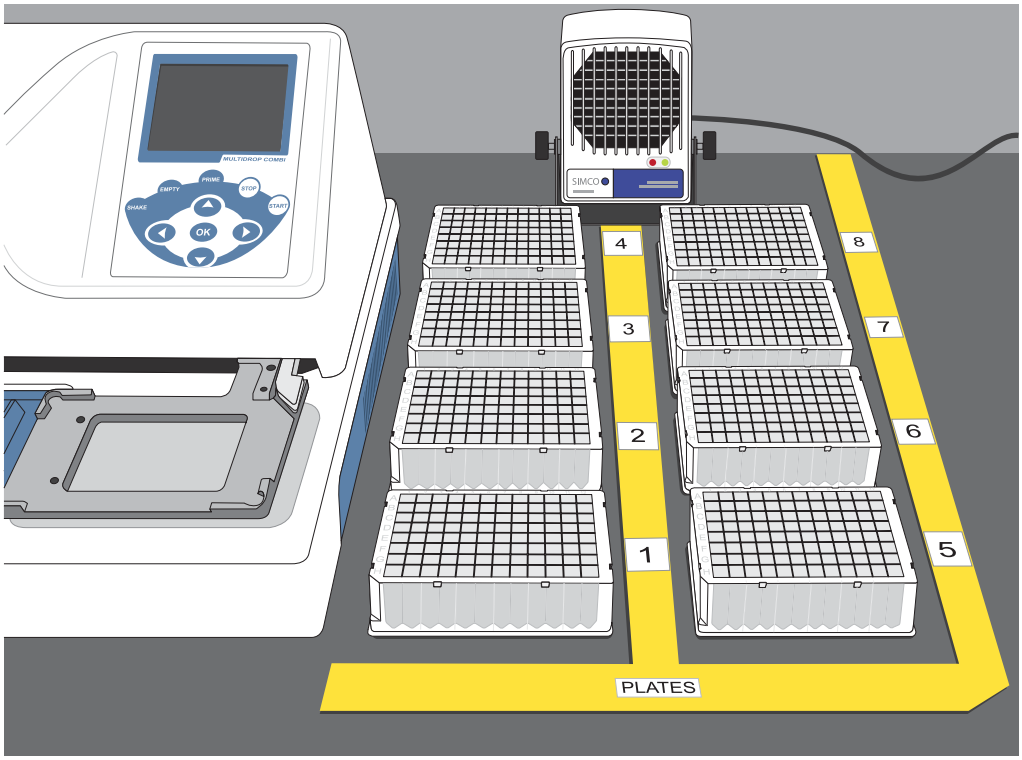


Figure 6 minION™2 Ionizing Air Blower placement.

Plate heat sealer guidelines

- Power on the plate heat sealer, then allow the sealer to reach set temperature before starting the target preparation workflow.
- When the sealer is first powered on for the day, run a few sealing operations to ensure seal quality and proper seal alignment to the plate.
- After the sealing operation, check the seal to ensure all the grids of the wells have visible imprints on the sealing material. If misalignment is observed, follow the guidelines on correcting alignment of the seal to the plate and adjusting seal quality. (See Appendix E, “Troubleshooting”.)
- If voids (incomplete melting between the seal material and the plate) are observed, remove the seal, blot dry any visible droplets on the grids, then reseal the plate.
- Follow recommendation for cleaning and preventive maintenance that is outlined in the instrument user guide.

Shaker guidelines

- Ensure that the plates are seated properly into the shaker before running the shaker.
- Always balance the shaker when mixing.
- Ensure that you use the correct type of shaker that is specified in the protocol. Shaker types and parameters are not interchangeable.

Centrifuge guidelines

- Always balance the rotor when performing the centrifugation step.
- Ensure that the centrifuge temperature is set to the correct temperature before the start of a procedure.
- Ensure that there is sufficient time for the centrifuge to cool down to 4°C before centrifuging the Precipitation Plates.
- The bottom of the rotor buckets must be soft rubber to ensure that the 96-deepwell plates do not crack. Do not centrifuge plates in metal or hard plastic buckets.

Incubator guidelines

- Put the plates on the racks of the incubator and do not stack plates directly on top of each other.
- Do not place the plates directly on the bottom surface of the incubator.
- After the batch of 8 plates are placed into oven for fragmentation, do not disturb the oven by opening and closing the door during the 30-minute incubation period.

Multidrop™ Combi Reagent Dispenser operations

Multidrop™ Combi protocol names and parameters

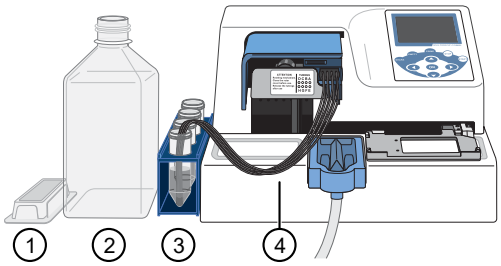
Table 9 Protocol names and parameters for the Axiom™ Propel XPRES 384HT Workflow.

Protocol name	Cassette	Plate type	Set point	Speed	Plate name
384-Den-10	Small tubing	96 DW (44mm)	10 µL	High	96-deepwell plate
384-Neu-65	Standard tubing	96 DW (44mm)	65 µL	Medium	96-deepwell plate
384-Amp-115	Standard tubing	96 DW (44mm)	115 µL	Medium	96-deepwell plate
384-Frag-30	Small tubing	96 DW (44mm)	30 µL	Medium	96-deepwell plate
384-Stop-10	Small tubing	96 DW (44mm)	10 µL	High	96-deepwell plate
384-Pre-440	Standard tubing	96 DW (44mm)	440 µL	Medium	96-deepwell plate
384-ResHyb-50	Standard tubing	96 DW (44mm)	50 µL	Medium	96-deepwell plate
384-Scan-170	Standard tubing	384_Scan_Tray	170 µL	Medium	GeneTitan™ 384-scan tray
384-Stain-40	Standard tubing	384_Stain_Tray	40 µL	High	GeneTitan™ 384-stain tray
384-QC-Dil-95	Standard tubing	384 DW (22mm)	95 µL	Medium	ABgene™ 384-Well 250 µL Polypropylene Storage Plate
384-QC-OD-25	Standard tubing	384 standard (15mm)	25 µL	Medium	Greiner Bio-One™ UV-Star™ 384-Well Microplate Flat Bottom
384-QC-Gel-60	Standard tubing	384 DW (22mm)	60 µL	Medium	ABgene™ 384-Well 250 µL Polypropylene Storage Plate

Materials required

- 50-mL conical tubes, labeled "Water 1", "Water 2", "Air", and "70% ethanol"
- Tube rack for the four 50-mL conical tubes
- 25-mL reservoir
- 70% ethanol
- DI water
- Large clean DI water bottle for cleaning the exterior of the tubing. The size of bottle must be sufficient to submerge the tubing bundle and >80% of the length of the input tubing. The 1 L Axiom™ Water container is sufficient.

IMPORTANT! Only use DI water one time to clean the exterior of *one* cassette. Discard the rinse water after use. Use fresh DI water for each individual cassette. Rinse the bottle with copious amount of DI water then fill with DI water for the cleaning step.



① 25-mL reservoir
② D. I. water bottle
③ Tube rack with four 50-mL prepared conical tubes
④ Multidrop™ Combi tubing bundle submerged at the bottom of the conical tube.

Start up the Multidrop™ Combi

The following procedure is required when the Multidrop™ cassette is used for the first time that day.

1. Install the Multidrop™ cassette and power on the Multidrop™ Combi instrument.
2. Place the tubing bundle into the "Water 1" conical tube, then press the **PRIME** button for ~5 seconds (10 mL for the standard tubing cassette and 3 mL for the small tubing cassette).
3. Observe the fluid dispense to ensure that the liquid is being dispensed through all the nozzles and that there are no air bubbles in the tubing.

Perform gravimetric checks

Gravimetric checks on the Multidrop™ Combi Reagent Dispenser must be conducted daily before running the target preparation stages. Perform the check on all the dedicated reagent cassettes used for that day.

Perform gravimetric checks after the Multidrop™ Combi is primed with water.

1. On the Multidrop™ Combi, select the protocol to be used. (See “Multidrop™ Combi protocol names and parameters” on page 49.)
2. Obtain an empty plate that is specified in the protocol.
3. Place the empty plate on a scale, then tare the scale.
4. Load the tared plate onto the Multidrop™ plate carrier, then run the protocol.
5. Observe the fluid dispense to ensure that the liquid is being dispensed through all the nozzles.
6. After the dispensing is complete, remove the plate then weigh the plate containing water.
   The weight must be within the specified range that is listed. (See “Range guidelines for gravimetric tests” on page 52.)
7. Repeat step 2—step 6 to obtain 2 more readings from the same protocol.
8. Do one of the following:
   - If the weights are within the specified range, proceed to priming the cassette with the appropriate Axiom™ reagent or reagent master mix.
   - If the weights are outside of the specified range, adjust the set using a 5-µL increment for standard tubing cassettes or 0.5-µL increment for small tubing cassettes, then repeat the gravimetric check. See the *Thermo Scientific™ Multidrop™ Combi User Manual* for instructions on applying the set point. (See “Related documentation” on page 225.)

Range guidelines for gravimetric tests

Table 10 Water weight checks for "Stage 1: Amplify the genomic DNA" (weight per plate, g).

	Protocol name		
	384-Den-10	**384-Neu-65**	**384-Amp-115**
Plate type	96-deepwell plate	96-deepwell plate	96-deepwell plate
Liquid to use	DI water	DI water	DI water
Volume per well	**10 µL**	**65 µL**	**115 µL**
Lower limit	0.91	5.93	10.5
Target, g	0.96	6.24	11.0
Upper limit	1.01	6.55	11.6

Table 11 Water weight checks for "Stage 2: Fragment the DNA", "Stage 3: Precipitate the DNA", and "Stage 5: Resuspend the pelleted DNA and prepare for hybridization" (weight per plate, g).

	Protocol name			
	384-Frag-30	**384-Stop-10**	**384-Pre-440**	**384-ResHyb-50**
Plate type	96-deepwell plate	96-deepwell plate	96-deepwell plate	96-deepwell plate
Liquid to use	DI water	DI water	DI water	DI water
Volume per well	**30 µL**	**10 µL**	**440 µL**	**50 µL**
Lower limit	2.85	1.00	38.4	4.60
Target, g	3.00	1.06	42.2	4.90
Upper limit	3.14	1.11	46.0	5.20

Table 12 Water weight checks for "Stage 5A: In-process QC" (weight per plate, g).

	Protocol name		
	384-QC-Dil-95	**384-QC-OD-25**	**384-QC-Gel-60**
Plate type	ABgene™ 384-Well 250 µL Polypropylene Storage Plate	Greiner Bio-One™ UV-Star™ 384-Well Microplate Flat Bottom	ABgene™ 384-Well 250 µL Polypropylene Storage Plate
Liquid to use	DI water	DI water	DI water
Volume per well	**95 µL**	**25 µL**	**60 µL**
Lower limit	34.66	8.64	21.89
Target, g	36.48	9.60	23.04
Upper limit	38.30	10.56	24.19

Table 13 Water weight checks for "Stage 7: Preparing ligation, stain, stabilization reagent trays, and scan tray for the GeneTitan™ MC Instrument" (weight per plate, g).

	Protocol name	
	384-Scan-170	**384-Stain-40**
Plate type	GeneTitan™ 384-scan tray	GeneTitan™ 384-stain tray
Liquid to use	DI water	DI water
Volume per well	**170 µL**	**40 µL**
Lower limit	15.5	13.82
Target, g	16.3	15.36
Upper limit	18.0	17.20

Prime the cassette

For each stage conducted at a Multidrop™ Combi Reagent Dispenser, the following steps are performed.

- Reagents and reagent master mixes are prepared before these steps.
- Before priming a reagent on the Multidrop™ Combi, always clean the cassette tubing with DI water and purge with air.

1. Lift the tubing bundle from the liquid surface, then press the **PRIME** button for ~5 seconds until DI water is purged from the tubing.

2. Prime the cassettes.
   a. Place the standard tubing bundle into the bottom of the reagent reservoir, then press the **PRIME** button until the reagent is seen flowing through all the tubing and liquid is dispensed out of all the tips. Continue holding the **PRIME** button for another 3–5 seconds to allow ~6–10 mL of reagent purging through the standard tubing cassette.

   Note: Do not exceed 6 seconds of priming after the reagent starts to come out from the nozzles. Excessive priming can cause loss of reagent and lead to insufficient reagent volume for the target preparation stages.

   b. Place the small tubing bundle into the bottom of the reagent reservoir, then press the **PRIME** button until the reagent is seen flowing through all the tubing and liquid is dispensed out of all the tips. Continue holding the **PRIME** button for another 5–8 seconds to allow ~3–5 mL of reagent purging through the small tubing cassette.
      - Immediately before dispensing the reagent into the first sample plate of a batch (that is, Plate 1 or Plate 5), perform another 1–2 seconds of priming, then begin the reagent dispensing.

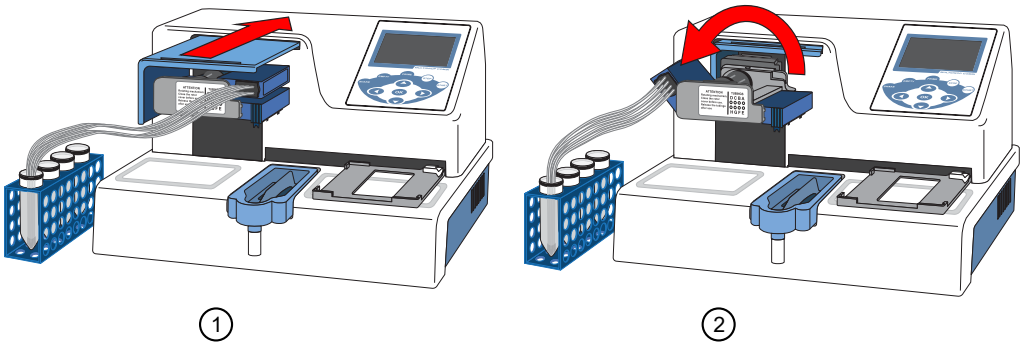
   Note: Do not exceed 10 seconds of priming after the reagent starts to come out from the nozzles. Excessive priming can cause loss of reagent and lead to insufficient reagent volume for the target preparation stages.

Flush the Multidrop™ Combi Reagent Dispenser between batches of plates

The following procedure is required to flush reagents from the Multidrop™ Combi cassette between processing plate batches.

1. Lift the tubing bundle from the liquid surface, then press the **PRIME** button for ~5 seconds or until the reagent is purged out.
2. Move the tubing bundle to the "Water 1" conical tube, then press the **PRIME** button for ~5 seconds (~10 mL) for a standard tubing cassette, and ~10 seconds (~6 mL) for a small tubing cassette.
3. Lift the tubing bundle from the liquid surface then press **PRIME** for ~5 seconds or until the DI water is purged out.
4. Move the tubing bundle to the "Water 2" conical tube, then press the **PRIME** button for ~5 seconds (~10 mL) for a standard tubing cassette, and ~10 seconds (~6 mL) for a small tubing cassette.
5. Lift the tubing bundle from the liquid surface, then press the **PRIME** button for ~5 seconds or until the DI water is purged out. Move the tubing bundle into the "Air" conical tube.
6. For a small tubing cassette, press the **EMPTY** button to raise the cassette. Immerse the tips into a 25-mL reservoir filled with DI water, then gently move the reservoir up and down a few times to rinse out any residual reagent around the outside of the tips.

   This action prevents salt build-up around the tips.
7. Place the cassette in the "Rest" position.



① Push the cover back into the instrument.
② Move the pipetting head up and to the left into the rest position.

Shut down the Multidrop™ Combi Reagent Dispenser

After the last batch of plates has completed the Axiom™ Propel XPRES 384HT Workflow, the Multidrop™ Combi must be shut down.

1. Prime the reagent out by pressing the **PRIME** button for ~5 seconds while lifting tubing bundle from solution.
2. Rinse the external tubing by submerging the tubing bundle into a large bottle that is filled with DI water, then dunk up and down 10 times. Let the tubing bundle hang inside the bottle above the liquid surface, then proceed to the next step.

   IMPORTANT! Dedicate a large bottle for each reagent cassette to prevent cross-contamination.

3. Backflush the cassette.
   a. Press the **EMPTY** button to raise the cassette.
   b. Fill a 25-mL reservoir with DI water.
   c. Submerge all tips into the filled 25-mL reservoir, then press the **EMPTY** button for ~5 seconds.
   d. Remove the reservoir from the tips. Then press the **EMPTY** button for ~5 seconds until all the water is out of the tubing.
   e. Remove the tubing bundle, then shake off excess water.
   f. For small tubing cassette, discard the DI water from the 25-mL reservoir, replenish with fresh DI water, then repeat substep 3c—substep 3e.
4. Place the tubing bundle into the “Water 1” conical tube, then press the **PRIME** button for ~10 seconds (~20 mL) for a standard tubing cassette and ~20 seconds (~12 mL) for a small tubing cassette.
5. Lift the tubing bundle from the liquid surface then press the **PRIME** button for ~5 seconds until the DI water is purged out.
6. Place the tubing bundle into the “Water 2” conical tube, then press the **PRIME** button for ~10 seconds (~20 mL) for a standard tubing cassette and ~20 seconds (~12 mL) for a small tubing cassette.
7. Lift the tubing bundle from the liquid surface, then press the **PRIME** button for ~5 seconds until the DI water is purged out.
8. Move the tubing bundle to the “70% EtOH” conical tube, then press the **PRIME** button for ~5 seconds (10 mL for a standard tubing cassette and 3 mL for a small tubing cassette).
9. Lift the tubing bundle from the liquid surface then, press the **PRIME** button for ~5 seconds until the ethanol is purged out.
10. Remove the cassette, place the lid to cover the tips (nozzles), then store it.

11. Turn the power off.

12. Discard all remaining reagents, water, ethanol, and labware used for the target preparation and cleaning procedures.

13. Rinse the DI water bottle with copious amounts of DI water.

4 Target preparation with Multidrop™ Combi Reagent Dispensers for two 384HT array format plates

- Stage 1: Amplify the genomic DNA 59
- Stage 2: Fragment the DNA 67
- Stage 3: Precipitate the DNA 73
- Stage 4: Centrifuge and dry DNA pellets 77
- Stage 5: Resuspend the pelleted DNA and prepare for hybridization 79
- Stage 5A: In-process QC 96
- Stage 6: Denature the target and transfer to hybridization tray 105
- Stage 7: Preparing ligation, stain, stabilization reagent trays, and scan tray for the GeneTitan™ MC Instrument 112

The instructions that are provided in this chapter are for processing two 384HT array format plates or less. When working with fewer than 2 plates, stop the dispensing after the sample containing plates are processed.

Ensure that the centrifuge and shaker are balanced.

IMPORTANT! Four 96-well gDNA Sample Plates must be prepared per 384HT array plate before starting DNA amplification. See Chapter 2, “Genomic DNA preparation”.

Stage 1: Amplify the genomic DNA

Equipment and labware required

Quantity	Item
Instruments	
3	Multidrop™ Combi Reagent Dispenser
2	Multidrop™ Combi SMART 2 Standard tube dispensing cassette[1]
1	Multidrop™ Combi SMART 2 Small tube plastic tip dispensing cassette[1]
1	ALPS™ 3000 Automated Microplate Heat Sealer
1	Compact Digital Microplate Shaker
1	Sorvall™ Legend™ XT Centrifuge (Set to room temperature.)
1	Heratherm™ Advanced Protocol Microbiological Incubator, capacity 66 L, BINDER™ ED 56, or BINDER™ BD 56 (set to 37°C) • Must maintain a constant temperature of 37°C for at least 24 hours with a temperature accuracy of ±1°C.
1	Vortex mixer
1	Electronic pipettor for serological pipettes
1	P1000 pipette
2	Digital timer
Labware and consumables	
As required	Easy Peel Seal (for ALPS™ 3000 Automated Microplate Heat Sealer)
1	50-mL conical tube
1	250-mL conical bottle (Nunc™ 250 mL Wide Mouth Conical Centrifuge Tube)
As required	P1000 pipette tips
As required	Serological pipettes (25 mL and 50 mL)

[1] Each reagent must use a dedicated cassette.

Input samples

Number of plates	Plate name	Content
8 (maximum)	gDNA Sample Plate	Genomic DNA samples prepared in Chapter 2, "Genomic DNA preparation" (in a 96-deepwell plate).

Note: The same eight 96-deepwell plates are processed in the workflow until the "Option 1: Perform QC after merging (recommended)" step in Stage 5.

Reagent handling

Fast Module 1, 2x384 (Part No. 952351) reagents are required from the Axiom™ Propel XPRES Reagent Kit, 2x384HT.

Thaw and prepare reagents according to the following table.

Table 14 Reagent handling for Stage 1 reagents.

Module	Reagent	Component Part No.	Treatment
Axiom™ Propel XPRES Reagent Kit Module 1 for 384HT only, –20°C	Axiom™ Propel 10X Denat Solution	952176	• Thaw at room temperature. This can be done as early as one day before use. • Vortex before master mix preparation.
	Axiom™ Propel Neutral Solution	952173	• Thaw at room temperature. This can be done as early as one day before use. • Vortex before master mix preparation.
	Axiom™ Propel Amp Solution	952174	• Thaw at room temperature. This can be done as early as one day before use.[1] • Vortex before master mix preparation.
	Axiom™ Propel Water	952177	• Thaw at room temperature. This can be done as early as one day before use.
	Axiom™ XPRES Amp Enzyme	952349	• Keep at –20°C until ready to use. • Flick 5 times before master mix preparation.

[1] Note: The thawed Axiom™ Propel Amp Solution solution may have white precipitates on the bottom of the bottle. Ensure that the precipitates dissolve into the solution after vortexing.

Prepare Denaturation Master Mix

Ensure that all components are treated according to the reagent handling table before preparing the master mix. (See Table 14.)

1. In a 50-mL conical tube, add reagents in the order shown.

Table 15 Denaturation Master Mix for 2x384HT array format plates.

Order of addition	Reagent	Quantity	Transfer method
1	Axiom™ Propel Water	25.0 mL	25-mL serological pipette
2	Axiom™ Propel 10X Denat Solution	2.8 mL	P1000
	Total volume	**27.8 mL**	

2. Ensure that the cap of the conical tube is closed tightly, then vortex the Denaturation Master Mix for 5 seconds. Leave at room temperature until use.

Note: The Denaturation Master Mix can be stored at room temperature and used within 12 hours of its preparation.

Prepare the Axiom™ Propel Neutral Solution

Ensure that the Axiom™ Propel Neutral Solution is vortexed before use.

1. Use the Axiom™ Propel Neutral Solution bottle for Multidrop™ dispensing.

Table 16 Axiom™ Propel Neutral Solution for 2x384HT array format plates.

Reagent	Quantity	Transfer method
Axiom™ Propel Neutral Solution	Entire bottle	Use straight from bottle[1]

[1] The Axiom™ Propel Neutral Solution reagent bottle can be used for Multidrop™ Combi dispensing.

2. Leave at room temperature. Vortex before use.

Prepare the Amplification Master Mix

Ensure that all components are treated according to the reagent handling table before preparing the master mix. (See Table 14.)

1. In a 250-mL conical bottle, add reagents in the order shown.

Table 17 Amplification Master Mix for 2x384HT array format plates.

Order of addition	Reagent	Quantity	Transfer method
1	Axiom™ Propel Amp Solution	114.0 mL	50-mL serological pipette
2	Axiom™ XPRES Amp Enzyme	2.53 mL	P1000 pipette
	Total volume	**116.53 mL**	

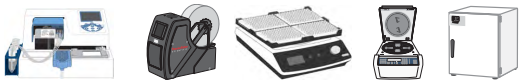
Note: The thawed Axiom™ Propel Amp Solution solution may have white precipitate on the bottom of the bottle. Ensure that the precipitates dissolve into the solution after vortexing.

Note: The Axiom™ XPRES Amp Enzyme is a viscous solution. Pipet the solution slowly when aspirating from the reagent bottle to avoid bubble formation.

2. Ensure that the cap of the conical bottle is closed tightly, then invert the Amplification Master Mix at least 15 times. Leave at room temperature.

Note: The Amplification Master Mix can be stored at room temperature and used within 2 hours of its preparation.

Stage 1 summary

Stage activities	Instruments required
• Three reagent additions with mixing—Denaturation Master Mix, Axiom™ Propel Neutral Solution, Amplification Master Mix • 10-minute denature incubation at room temperature • 4–24 hour amplification incubation at 37°C	

Perform the pre-run checklist

Before starting the workflow, the following tasks must be completed.

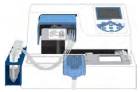
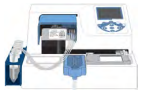
1. Multidrop™ Combi Reagent Dispenser:
   a. Ensure that the Multidrop™ Combi startup procedure has been performed. (See "Start up the Multidrop™ Combi" on page 50.)
   b. Perform gravimetric checks before running the workflow. (See "Perform gravimetric checks" on page 51.)
   c. Prime the Multidrop™ cassette with reagent. (See "Prime the cassette" on page 54.)
   d. Move the ion blowers to each dispensing station, then power them on. (Figure 6 on page 47.)
2. ALPS™ 3000 Automated Microplate Heat Sealer:
   a. Power on the ALPS™ 3000 Automated Microplate Heat Sealer, then allow it to reach 150°C.
   b. If the sealer is being used at the start of the day for this stage, do the following:
      - Allow 15 minutes for the sealer to reach 150°C.
      - Perform the sealing operation 2 to 3 times, then check to ensure that the seal quality and seal alignment are correct before starting the workflow.
3. If the gDNA Sample Plates are frozen, thaw them on the benchtop at room temperature, then centrifuge at 675 × *g* for 30 seconds to get all the droplets down.

IMPORTANT! The gDNA samples must be brought to room temperature before proceeding with denaturation.

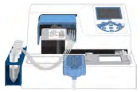
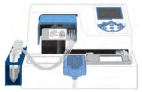
Stage 1: Amplify the genomic DNA

To process 8 sample plates, complete 2 cycles of 4 plates per cycle.

Denaturation Master Mix

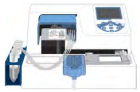
Plates 1 2 3 4	Plates 5 6 7 8
①  Dispense Denaturation Master Mix • Method: 384-Den-10 • Dispense volume: 10 µL • Prime for 1–2 seconds before dispensing.	⑥  Dispense Denaturation Master Mix • Method: 384-Den-10 • Dispense volume: 10 µL • Prime for 1–2 seconds before dispensing.
②  Seal • Settings: 150°C, 2.5 seconds	⑦  Seal • Settings: 150°C, 2.5 seconds
③  Shake • Settings: 1,100 rpm, 30 seconds	⑧  Shake • Settings: 1,100 rpm, 30 seconds
④  Time • Start 10-minute timer	⑨  Time • Start 10-minute timer
⑤  Centrifuge • Centrifuge at room temperature • Settings: 675 × *g*, 30 seconds • Leave plates at room temperature after centrifugation until the timer alarms. • Continue to Step 6.	⑩  Centrifuge • Centrifuge at room temperature • Settings: 675 × *g*, 30 seconds • Leave plates at room temperature after centrifugation until the timer alarms.
When the 10-minute timer alarms, proceed *immediately* to the Axiom™ Propel Neutral Solution step.	• When the 10-minute timer alarms, proceed *immediately* to the "Axiom™ Propel Neutral Solution" step. • Perform the appropriate Multidrop™ procedure: Flush between batches, on page 55; Shutdown, on page 56.[1]

[1] **IMPORTANT!** Follow the special instructions for small tubing cassettes, required for this step.

Axiom™ Propel Neutral Solution	
Plates 1 2 3 4	Plates 5 6 7 8
①  Dispense Axiom™ Propel Neutral Solution • Method: 384-Neu-65 • Dispense volume: 65 µL	⑤  Dispense Axiom™ Propel Neutral Solution • Method: 384-Neu-65 • Dispense volume: 65 µL
②  Seal • Settings: 150°C, 2.5 seconds	⑥  Seal • Settings: 150°C, 2.5 seconds
③  Shake • Settings: 1,100 rpm, 30 seconds	⑦  Shake • Settings: 1,100 rpm, 30 seconds
④  Centrifuge • Centrifuge at room temperature • Settings: 675 × *g*, 30 seconds • Continue to the Amplification Master Mix step	⑧  Centrifuge • Centrifuge at room temperature • Settings: 675 × *g*, 30 seconds
	• Continue to the Amplification Master Mix step. • Perform the appropriate Multidrop™ procedure: – Flush between batches. See page 55. – Shutdown. See page 56.

Amplification Master Mix

Plates 1 2 3 4

① 
Dispense Amplification Master Mix
- Method: 384-Amp-115
- Dispense volume: 115 µL

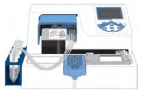
② 
Seal
- Settings: 150°C, 2.5 seconds

③ 
Shake
- Settings: 1,100 rpm, 30 seconds

④ 
Centrifuge
- Centrifuge at room temperature
- Settings: 675 × *g*, 30 seconds

⑤ 
Incubate
- Temperature: 37°C
- Time: 4–24 hours
- Volume = 200 µL/well

Plates 5 6 7 8

⑥ 
Dispense Amplification Master Mix
- Method: 384-Amp-115
- Dispense volume: 115 µL

⑦ 
Seal
- Settings: 150°C, 2.5 seconds

⑧ 
Shake
- Settings: 1,100 rpm, 30 seconds

⑨ 
Centrifuge
- Centrifuge at room temperature
- Settings: 675 × *g*, 30 seconds

⑩ 
Incubate
- Temperature: 37°C
- Time: 4–24 hours
- Volume = 200 µL/well

1 2 3 4 5 6 7 8

⑪
- Proceed directly to "Stage 2: Fragment the DNA" on page 67, or
- Store the Amplification Plates at –20°C. The frozen post-amplification plates can be stored at –20°C for up to 1 week.
- Perform the appropriate Multidrop™ procedure:
  - Flush between batches. See page 55.
  - Shutdown. See page 56.

Stage 2: Fragment the DNA

Equipment and labware required

Quantity	Item
Instruments	
2	Multidrop™ Combi Reagent Dispenser
2	Multidrop™ Combi SMART 2 Small tube plastic tip dispensing cassette[1]
1	ALPS™ 3000 Automated Microplate Heat Sealer
1	Compact Digital Microplate Shaker
1	Sorvall™ Legend™ XTR Centrifuge (set to room temperature)
1	Heratherm™ Advanced Protocol Microbiological Incubator, capacity 66 L, BINDER™ ED 56, or BINDER™ BD 56 (set to 37°C) • Must maintain a constant temperature of 37°C for at least 24 hours with a temperature accuracy of ±1°C.
1	Vortex mixer
1	Mini centrifuge
1	Electronic pipettor for serological pipettes
1	P1000 pipette
2	Digital timer
Labware and consumables	
As required	Easy Peel Seal (for ALPS™ 3000 Automated Microplate Heat Sealer)
As required	P1000 pipette tips
As required	Serological pipettes (10 mL and 50 mL)
1	50-mL conical tube

[1] Each reagent must use a dedicated cassette.

Input samples

Number of plates	Plate name	Content
8 (maximum)	Amplification Plate	Amplified gDNA samples prepared in “Stage 1: Amplify the genomic DNA” (in a 96-deepwell plate).

Reagent and plate handling

Module 2-1, 4x96F and Module 2-2, 4x96F reagents are required from the Axiom™ Propel XPRES Reagent Kit, 2x384HT.

- Axiom™ Propel Reagent Kit Module 2-1 for 96F or 384HT, Part No. 952263
- Axiom™ Propel Reagent Kit Module 2-2 for 96F or 384HT, Part No. 952265

Thaw and prepare reagents according to the following table.

Table 18 Reagent and plate handling for Stage 2 and Stage 3.

Module	Reagent	Component Part No.	Treatment
Axiom™ Propel Reagent Kit Module 2-1 for 96F or 384HT, –20°C	Axiom™ Propel 10X Frag Buffer	952179	• Thaw and equilibrate to room temperature on the day of use, or • Thaw at 4°C the day before then equilibrate to room temperature on the day of use. • Vortex before master mix preparation.
	Axiom™ Propel Frag Enzyme	952181	• Keep at –20°C until ready to use. • Flick 5 times, then perform brief centrifuge before master mix preparation.
	Axiom™ Propel Precip Solution 2	952178	• Thaw and equilibrate to room temperature on the day of use, or • Thaw at 4°C the day before then equilibrate to room temperature on the day of use. • Vortex before master mix preparation.
Axiom™ Propel Reagent Kit Module 2-2 for 96F or 384HT, 4°C	Axiom™ Propel Frag Diluent	952184	• Remove from 4°C and equilibrate to room temperature on day of use. • Vortex before master mix preparation.
	Axiom™ Propel Frag Reaction Stop	952190	• Remove from 4°C and equilibrate to room temperature on day of use. • Vortex before use.
	Axiom™ Propel Precip Solution 1	952203	• Remove from 4°C and equilibrate to room temperature on day of use. • Vortex before master mix preparation.
Amplification Plates			
• If proceeding directly from the 4—24-hour amplification, continue with the fragmentation step that is described in this chapter. • If the Amplification Plates have been frozen, place the frozen plates in an incubator at 37°C for 2 hours, then continue to the fragmentation step that is described in this chapter. **Note:** Place plates on the rack in a single layer—do not stack plates directly on top of one another.			

Prepare the Fragmentation Master Mix

Ensure that all components are treated according to the reagent handling table before preparing the master mix. (See Table 18.)

1. In a 50-mL conical tube, add reagents in the order shown.

Table 19 Fragmentation Master Mix for 2x384HT array format plates.

Order of addition	Reagent	Quantity	Transfer method
1	Axiom™ Propel 10X Frag Buffer	36.0 mL	50-mL serological pipette
2	Axiom™ Propel Frag Diluent	8.1 mL	10-mL serological pipette
3	Axiom™ Propel Frag Enzyme	0.79 mL	P1000 pipette
	Total volume	**44.89 mL**	

2. Ensure that the cap of the conical tube is closed tightly, then invert the Fragmentation Master Mix at least 15 times to mix. Leave at room temperature until use.

Note: Use the Fragmentation Master Mix within 1 hour of its preparation.

Prepare the Axiom™ Propel Frag Reaction Stop

Ensure that the Axiom™ Propel Frag Reaction Stop is vortexed before use.

1. Use the Axiom™ Propel Frag Reaction Stop bottle for Multidrop™ dispensing.

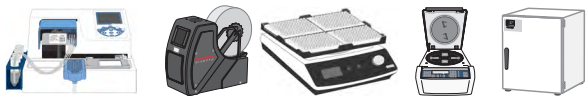
Table 20 Axiom™ Propel Frag Reaction Stop for 2x384HT array format plates.

Reagent	Quantity	Transfer method
Axiom™ Propel Frag Reaction Stop	Entire bottle	Use straight from bottle[1]

[1] The Axiom™ Propel Frag Reaction Stop reagent bottle can be used for Multidrop™ Combi dispensing.

2. Leave at room temperature. Vortex before use.

Stage 2 summary

Stage activities	Instruments required
• Two reagent additions with mixing—Fragmentation Master Mix, Axiom™ Propel Frag Reaction Stop • 30-minute fragmentation incubation at 37°C	

Perform the pre-run checklist

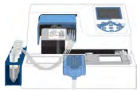
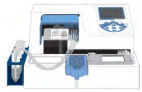
Before starting the workflow, the following tasks must be completed.

1. Multidrop™ Combi Reagent Dispenser:
   a. Ensure that the Multidrop™ Combi startup procedure has been performed. (See "Start up the Multidrop™ Combi" on page 50.)
   b. Perform gravimetric checks before running the workflow. (See "Perform gravimetric checks" on page 51.)
   c. Prime the Multidrop™ cassette with reagent. (See "Prime the cassette" on page 54.)
   d. Move the ion blowers to each dispensing station, then power them on. (Figure 6 on page 47.)
2. ALPS™ 3000 Automated Microplate Heat Sealer:
   a. Power on the ALPS™ 3000 Automated Microplate Heat Sealer, then allow it to reach 150°C.
   b. If the sealer is being used at the start of the day for this stage, do the following:
      - Allow 15 minutes for the sealer to reach 150°C.
      - Perform the sealing operation 2 to 3 times, then check to confirm that the seal quality and seal alignment are correct before starting the workflow.

Stage 2: Fragment the DNA

To process 8 sample plates, complete 2 cycles of 4 plates per cycle.

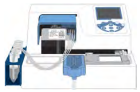
Fragmentation Master Mix

Plates 1 2 3 4	Plates 5 6 7 8
①  Dispense Fragmentation Master Mix • Method: 384-Frag-30 • Dispense volume: 30 µL • Prime for 1–2 seconds before dispensing.	⑧  Dispense Fragmentation Master Mix • Method: 384-Frag-30 • Dispense volume: 30 µL • Prime for 1–2 seconds before dispensing.
②  Seal • Settings: 150°C, 2.5 seconds	⑨  Seal • Settings: 150°C, 2.5 seconds
③  Shake • Settings: 1,100 rpm, 30 seconds	⑩  Shake • Settings: 1,100 rpm, 30 seconds
④  Time • Start 30 minute timer	⑪  Time • Start 30 minute timer
⑤  Centrifuge • Centrifuge at room temperature • Settings: 675 × *g*, 30 seconds	⑫  Centrifuge • Centrifuge at room temperature • Settings: 675 × *g*, 30 seconds
⑥  Incubate • Temperature: 37°C • Time: 30 minutes (Time begins at completion of shaking step.)	⑬  Incubate • Temperature: 37°C • Time: 30 minutes (Time begins at completion of shaking step.)
⑦ When the 30-minute timer alarms, proceed *immediately* to "Axiom™ Propel Frag Reaction Stop" step.	⑭ • When the 30-minute timer alarms, proceed *immediately* to "Axiom™ Propel Frag Reaction Stop" step. • Perform the appropriate Multidrop™ procedure: Flush between batches, on page 55; Shutdown, on page 56.[1]

[1] **IMPORTANT!** Follow the special instructions for small tubing cassettes, required for this step.

Axiom™ Propel Frag Reaction Stop

Plates 1 2 3 4

① 

Dispense Axiom™ Propel Frag Reaction Stop
- Method: 384-Stop-10
- Dispense volume: 10 µL
- Prime for 1–2 seconds before dispensing.

② 

Seal
- Settings: 150°C, 2.5 seconds

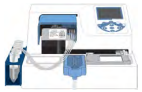
③ 

Shake
- Settings: 1,100 rpm, 30 seconds

④ 

Centrifuge
- Centrifuge at room temperature
- Settings: 675 × *g*, 30 seconds

Plates 5 6 7 8

⑤ 

Dispense Axiom™ Propel Frag Reaction Stop
- Method: 384-Stop-10
- Dispense volume: 10 µL
- Prime for 1–2 seconds before dispensing.

⑥ 

Seal
- Settings: 150°C, 2.5 seconds

⑦ 

Shake
- Settings: 1,100 rpm, 30 seconds

⑧ 

Centrifuge
- Centrifuge at room temperature
- Settings: 675 × *g*, 30 seconds

1 2 3 4 5 6 7 8

⑨

- Volume = 240 µL
- Continue to “Stage 3: Precipitate the DNA” on page 73.
- Perform the appropriate Multidrop™ procedure:
  - Flush between batches. See page 55.
  - Shutdown. See page 56.

IMPORTANT! Follow the special instructions for small tubing cassettes, required for this step.

Stage 3: Precipitate the DNA

Equipment and labware required

Quantity	Item
Instruments	
1	Multidrop™ Combi Reagent Dispenser
1	Multidrop™ Combi SMART 2 Standard tube dispensing cassette[1]
1	ALPS™ 3000 Automated Microplate Heat Sealer
1	VWR Signature™ High-Speed Microplate Shaker
1	Vortex mixer
1	Freezer, –20°C
1	P1000 pipette
1	Electronic pipettor for serological pipettes
1	Fume hood
Labware and consumables	
As required	Serological pipettes (50 mL)
As required	Easy Peel Seal (for ALPS™ 3000 Automated Microplate Heat Sealer)
1	500-mL bottle

[1] Each reagent must use a dedicated cassette.

Input samples

Number of plates	Plate name	Content
8 (maximum)	Fragmentation Plate	Fragmented DNA samples prepared in "Stage 2: Fragment the DNA" on page 67 (in a 96-deepwell plate).

Prepare 384-Precipitation Master Mix

Ensure that all components are treated according to the reagent handling table before preparing the master mix. (See Table 18.)

CAUTION! It is recommended that the steps in this stage be performed under a fume hood.

1. Use a 500-mL bottle for Multidrop™ dispensing.

Table 21 384-Precipitation Master Mix for 2x384HT array format plates.

Order of addition	Reagent	Quantity	Transfer method
1	Axiom™ Propel Precip Solution 1	Entire bottle	Use straight from the bottle
2	Axiom™ Propel Precip Solution 2	Entire tube. Transfer contents into Axiom™ Propel Precip Solution 1 bottle, then vortex for 5 seconds.	P1000 pipette
3	Mixture of Axiom™ Propel Precip Solution 1 and Axiom™ Propel Precip Solution 2	Transfer 110 mL of the mixture into a 500-mL bottle.	50-mL serological
4	Isopropanol	330 mL	50-mL serological
	Total volume	**440 mL**	

2. Ensure that the cap of the 500-mL bottle is closed tightly, then invert the 384-Precipitation Master Mix at least 15 times to mix. Leave at room temperature until use.

Note: The 384-Precipitation Master Mix can be stored at room temperature and used within 2 hours of its preparation.

Stage 3 summary

Stage activities	Instruments required
• One reagent addition with mixing—384-Precipitation Master Mix. • 3–24 hour precipitation incubation at –20°C.	

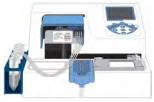
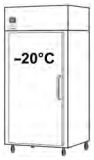
Perform the pre-run checklist

Before starting the workflow, the following tasks must be completed.

1. Multidrop™ Combi Reagent Dispenser:
   a. Ensure that the Multidrop™ Combi startup procedure has been performed. (See "Start up the Multidrop™ Combi" on page 50.)
   b. Perform gravimetric checks before running the workflow. (See "Perform gravimetric checks" on page 51.)
   c. Prime the Multidrop™ cassette with reagent. (See "Prime the cassette" on page 54.)
   d. Move the ion blowers to each dispensing station, then power them on. (Figure 6 on page 47.)
2. ALPS™ 3000 Automated Microplate Heat Sealer:
   a. Power on the ALPS™ 3000 Automated Microplate Heat Sealer then allow it to reach 150°C.
   b. If the sealer is being used at the start of the day for this stage, do the following:
      • Allow 15 minutes for the sealer to reach 150°C.
      • Perform the sealing operation 2 to 3 times, then check to confirm that the seal quality and seal alignment are correct before starting the workflow.

Stage 3: Precipitate the DNA

To process 8 sample plates, complete 1 cycle of 8 plates.

Precipitation		
	Plates 1 2 3 4 5 6 7 8	
①		Dispense 384-Precipitation Master Mix • Method: 384-Pre-440 • Dispense volume: 440 µL
③		Seal • Settings: 150°C, 2.5 seconds
④		Shake • Settings: 2,450 rpm, 60 seconds **Note:** The 60-second shaking time is different than the shaking times in other stages. **IMPORTANT!** Do not centrifuge the plates after shaking. Continue directly to freezing the plates at –20°C.
⑤		Freeze • –20°C, 3–24 hours • Volume = 680 µL/well • Perform the appropriate Multidrop™ procedure: – Flush between batches. See page 55. – Shutdown. See page 56.

Stage 4: Centrifuge and dry DNA pellets

Equipment required

Quantity	Item
2	Sorvall™ Legend™ XTR Centrifuge (set at 4°C)
1	Heratherm™ Advanced Protocol Microbiological Incubator, capacity 66 L, BINDER™ ED 56, or BINDER™ BD 56 (set to 37°C) • Must maintain a constant temperature of 37°C for at least 24 hours with a temperature accuracy of ±1°C.
1	Fume hood

Input samples

Number of plates	Plate name	Content
8 (maximum)	Precipitation Plate	Samples, post precipitation incubation, prepared in “Stage 3: Precipitate the DNA” (in a 96-deepwell plate).

Stage 4 summary

Stage activities	Instruments required
• Purify amplified DNA into dried pellets.	

Perform the pre-run checklist

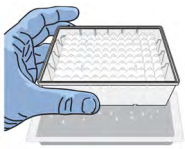
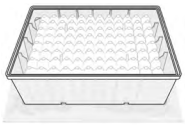
Before starting the workflow, the following task must be completed.

Pre-chill the centrifuge at 4°C.

Stage 4: Centrifuge and dry pellets

To process 8 sample plates, complete 1 cycle of 8 plates. Decant under a ventilated fume hood.

CAUTION! During this stage, handle the plate gently to avoid disturbing the pellets. Do not bump, tap, or bang the plate against another object.

Centrifugation		
	Plates: 1 2 3 4 5 6 7 8	
①		Centrifuge at 4°C. • 3,200 × *g* for 40 minutes.
②		Gently pour off solution over a waste container and allow the liquid to drain.
③		Keep inverted for 5 minutes over laboratory tissues. Transfer the plate to a new pile of laboratory tissues twice during the 5-minute period.
④		• Dry in 37°C oven for 20 minutes. • Place the plates in the oven with wells facing up.
⑤		• Do one of the following: – Proceed directly to "Stage 5: Resuspend the pelleted DNA and prepare for hybridization" on page 79, even if some droplets of liquid remain. Leave the sample plates at room temperature. It is helpful to start preparing reagents for stage 5 during centrifuging and drying pellets. – Store the plates for resuspension later in the same day. Tightly seal the plates. – If resuspension is carried out in 4 hours, keep the plates at room temperature. – If resuspension is carried out in more than 4 hours, store the plates in a refrigerator (2–8°C). The refrigerated pellets must be equilibrated to room temperature for at least 30 minutes before proceeding to Stage 5. • To process the plates for resuspension on another day, tightly seal the plates and store at –20°C. – The frozen pellets must be equilibrated to room temperature for at least 90 minutes before proceeding to Stage 5.

Stage 5: Resuspend the pelleted DNA and prepare for hybridization

Equipment and labware required

Table 22 Equipment and labware required for Stage 5.

Quantity	Item
Instruments	
1	Multidrop™ Combi Reagent Dispenser
1	Multidrop™ Combi SMART 2 Standard tube dispensing cassette[1]
1	ALPS™ 3000 Automated Microplate Heat Sealer
1	Compact Digital Microplate Shaker
1	Sorvall™ Legend™ XTR Centrifuge (Set to room temperature)
1	Vortex mixer
1	Electronic pipettor for serological pipettes
1	P1000 pipette
1	Fume hood
Labware and consumables	
As required	Easy Peel Seal (This is used for the ALPS™ 3000 Automated Microplate Heat Sealer.)
1	250-mL conical tube (Nunc™ 250 mL Wide Mouth Conical Centrifuge Tube)
As required	Serological pipettes (5 mL, 25 mL, and 50 mL)
As required	P1000 pipette tips

[1] Each reagent must use a dedicated cassette.

Table 23 Equipment and labware required for the Integra transfer.

Quantity	Item
Instruments	
1	VIAFLO™ 96 Base Unit or VIAFLO™ 384 Base Unit with 2 single-position plate holders and a 96 Channel Pipetting Head (5–125 µL) installed
Labware and consumables	
768 tips	Integra VIAFLO XYZ GripTips™, 125 µL
2	PCR Plate, 384-well, raised chimney

Table 23 Equipment and labware required for the Integra transfer. *(continued)*

Quantity	Item
1	384-well PCR plate collar
As required	MicroAmp™ Clear Adhesive Film

Input samples

Number of plates	Plate name	Content
8 (maximum)	Pelleted Precipitation Plate	Dried DNA pellets prepared in "Stage 4: Centrifuge and dry DNA pellets" on page 77 (in a 96-deepwell plate).

Reagent handling

Module 2-1 and Module 2-2 reagents are required from the Axiom™ Propel XPRES Reagent Kit, 2x384HT.

- Axiom™ Propel Reagent Kit Module 2-1 for 96F or 384HT, Part No. 952263
- Axiom™ Propel Reagent Kit Module 2-2 for 96F or 384HT, Part No. 952265

Thaw and prepare reagents according to the following table.

Table 24 Reagent handling for Stage 5 reagents.

Module	Reagent	Component Part No.	Treatment
Axiom™ Propel Reagent Kit Module 2-1 for 96F or 384HT –20°C	Axiom™ Propel Hyb Buffer	952182	• Thaw and equilibrate to room temperature on the day of use, or • Thaw at 4°C the day before then equilibrate to room temperature on the day of use. • Vortex before master mix preparation.
	Axiom™ Propel Hyb Solution 1	952183	• Thaw and equilibrate to room temperature on the day of use, or • Thaw at 4°C the day before then equilibrate to room temperature on the day of use. • Vortex before master mix preparation.

Table 24 Reagent handling for Stage 5 reagents. *(continued)*

Module	Reagent	Component Part No.	Treatment
Axiom™ Propel Reagent Kit Module 2-2 for 96F or 384HT 4°C	Axiom™ Propel Hyb Solution 2	951979	• Remove from 4°C, then equilibrate to room temperature on day of use. • Vortex before master mix preparation.
	Axiom™ Propel Resuspension Buffer	952206	• Remove from 4°C and equilibrate to room temperature on day of use. • Vortex before master mix preparation.

Guidelines for pellet preparation

The equilibration of the plate of pelleted DNA, Axiom™ Propel Resuspension Buffer, and Axiom™ Propel Hyb Buffer to room temperature (18-25°C) is very critical for the success of the assay. If any of these are cooler than room temperature, pellets may not resuspend completely. This can result in compromised assay performance. Follow the guidelines below on how to work with plates with fresh, cold, or frozen pellets.

- **Fresh pellets:** A plate with fresh pellets can be kept at room temperature if proceeding with Stage 5 within 4 hours.
- **Cold pellets:** A plate with fresh pellets that are not processed within 4 hours can be transferred to a refrigerator (2-8°C) if processed during the same day. However, it is critical to equilibrate the plate to room temperature for at least 30 minutes before proceeding with Stage 5.
- **Frozen pellets:** A plate with frozen pellets must be pre-equilibrated at room temperature for at least 1.5 hours before proceeding with Stage 5.

Prepare Hybridization Cocktail

Ensure that all of the components are treated according to the reagent handling table before preparing the master mix. (See Table 24.)

CAUTION! It is recommended that the remainder of the steps in this stage be performed under a fume hood.

IMPORTANT! The plate of pelleted DNA and resuspension reagents must be at room temperature before proceeding with this step.

1. In a 250-mL conical bottle, add reagents in the order shown.

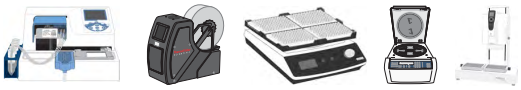
Table 25 Hybridization Cocktail for 2x384HT array format plates.

Order of addition	Reagent	Quantity	Transfer method
1	Axiom™ Propel Resuspension Buffer	18.3 mL	25-mL serological pipette
2	Axiom™ Propel Hyb Buffer	37 mL	50-mL serological pipette
3	Axiom™ Propel Hyb Solution 1	0.265 mL	P1000 pipette
4	Axiom™ Propel Hyb Solution 2	4.7 mL (3.0 mL + 1.7 mL)	5-mL serological pipette
	Total volume	**60.3 mL**	

2. Ensure that the cap of the conical bottle is closed tightly, then vortex the Hybridization Cocktail for 5 seconds. Leave at room temperature until use.

Note: The Hybridization Cocktail can be stored at room temperature and used within 2 hours of its preparation.

Stage 5 summary

Stage activities	Instruments required
• One reagent addition with mixing—Hybridization Cocktail • 15 minute shaking to resuspend the DNA pellets • Transfer from four 96-deepwell plates to one 384-format PCR plate	

Perform the pre-run checklist

Before starting the workflow, the following tasks must be completed.

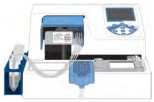
1. Multidrop™ Combi Reagent Dispenser:
   a. Ensure that the Multidrop™ Combi startup procedure has been performed. (See "Start up the Multidrop™ Combi" on page 50.)
   b. Perform gravimetric checks before running the workflow. (See "Perform gravimetric checks" on page 51.)
   c. Prime the Multidrop™ cassette with reagent. (See "Prime the cassette" on page 54.)
   d. Move the ion blowers to each dispensing station, then power them on. (Figure 6 on page 47.)
2. ALPS™ 3000 Automated Microplate Heat Sealer:
   a. Power on the ALPS™ 3000 Automated Microplate Heat Sealer then allow it to reach 150°C.
   b. If the sealer is being used at the start of the day for this stage, do the following:
      • Allow 15 minutes for the sealer to reach 150°C.
      • Perform the sealing operation 2 to 3 times, then check to ensure that the seal quality and seal alignment are correct before starting the workflow.
3. Ensure that the VIAFLO™ 96 Base Unit or the VIAFLO™ 384 Base Unit is clean, then power on the unit.
4. If the pellets are refrigerated or frozen, equilibrate them to room temperature before the resuspension step. (See "Guidelines for pellet preparation" on page 82.)

Note: The equilibration of pelleted DNA and Hybridization Cocktail to room temperature (18°C to 25°C) is critical for the resuspension process. When either is cooler than room temperature, pellets may not resuspend completely, which can result in lower performance.

Stage 5: Resuspend the pelleted DNA and prepare for hybridization

To process 8 sample plates, complete 1 cycle of 8 plates.

CAUTION! It is recommended that the steps in this stage be performed under a fume hood.

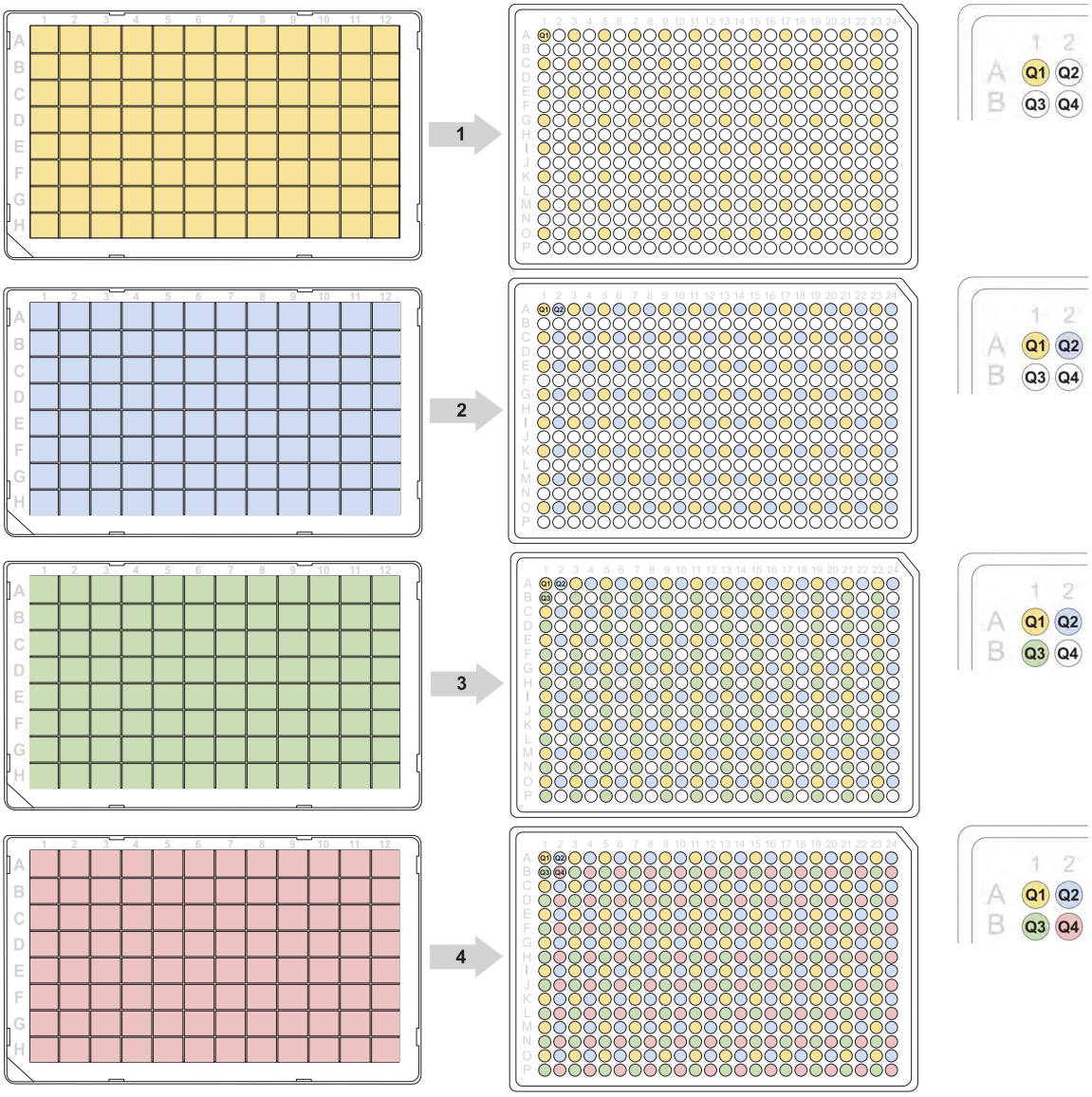
Hybridization Cocktail		
	Plates 1 2 3 4 5 6 7 8	
①		Dispense Hybridization Cocktail • Method: 384-ResHyb-50 • Dispense volume: 50 µL
②		Seal • Settings: 150°C, 2.5 seconds
③		Shake • Settings: 1,100 rpm, 15 minutes
④		Centrifuge • Centrifuge at room temperature • Settings: 675 × *g*, 30 seconds
⑤		• Continue to “Merge: Transfer the hybridization-ready target from four 96-deepwell plates into one 384-well PCR plate”. • Perform the appropriate Multidrop™ procedure: – Flush between batches. See page 55. – Shutdown. See page 56.

Concepts of plate format transfer

IMPORTANT!

- It is essential to understand the concepts required to successfully transfer the hybridization-ready target from four 96-deepwell plates into one 384-format PCR plate. Review this section before starting the procedure.
- Maintain proper traceability of source hybridization-ready targets when merging the four 96-deepwell plates into one 384-format PCR plate.

Quadrants of the 384-format PCR plate



The hybridization-ready target in the four 96-deepwell plates is represented by the colors yellow, blue, green, and pink (plates 1–4).

The entire contents of each 96-format plate is transferred into one "quadrant" of the 384-well PCR plate, one plate at a time. After the transfer from the final 96-format plate (represented in pink), each quadrant in the 384-well PCR plate contains the target from each of the four 96-deepwell plates (yellow, blue, green, and pink).

VIAFLO™ Instrument configuration

The Axiom™ Propel XPRES 384HT Workflow uses 2 models of the VIAFLO™ liquid handler: the VIAFLO™ 96 Base Unit and the VIAFLO™ 384 Base Unit. Both models require 2 single-position plate holders and the appropriate pipetting head (5–125 µL 96 Channel Pipetting Head or 2–50 µL 384 Channel Pipetting Head), respectively.

The minimum requirement for the Axiom™ Propel XPRES 384HT Workflow is a VIAFLO™ 384 Base Unit and both the 96 Channel Pipetting Head (5–125 µL) and 384 Channel Pipetting Head (2–50 µL). This option requires changing to the appropriate pipetting head that is used for the selected VIAFLO™ method.

For instructions on how to change the pipetting head on the VIAFLO™ Instrument:

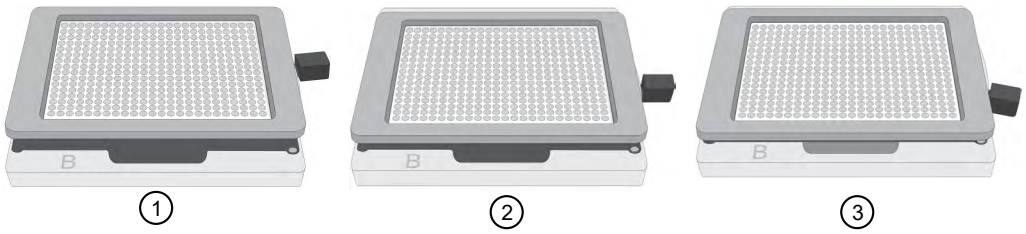
- See the VIAFLO 96/384 Operating Instructions (Pub. No. 125950) posted on the INTEGRA Biosciences website.
- Contact INTEGRA Biosciences support.

Figure 7 VIAFLO™ 96 and VIAFLO™ 384 configuration.

1. Position A
2. Position B
3. Stage orientation sliders

Stage orientation

To accomplish the transfer of content from one 96-well plate into the specified quadrant, the stage is shifted in one of 3 different orientations to move the plate and transfer the target. Both the A and B positions include the stage sliding lever.



① Up
② Center
③ Down

Merge: Transfer the hybridization-ready target from four 96-deepwell plates into one 384-well PCR plate

Two VIAFLO method options exist for merging the four 96-deepwell plates containing the hybridization-ready target into one 384-well PCR plate.

- Option 1: Perform QC after merging (recommended).

  The **96 TO 384_48µL** VIAFLO method is used when in-process quality controls are performed. The 2 recommended in-process quality controls are:

  – QC quantification: an OD quantification of each sample to check DNA yield.

  – Fragmentation QC: a gel to view the fragmentation of the DNA.

  This method transfers 48 µL/well of hybridization-ready target from the four 96-deepwell plates to one 384-well PCR plate.

- Option 2: Performing only hybridization after merging (No QC).

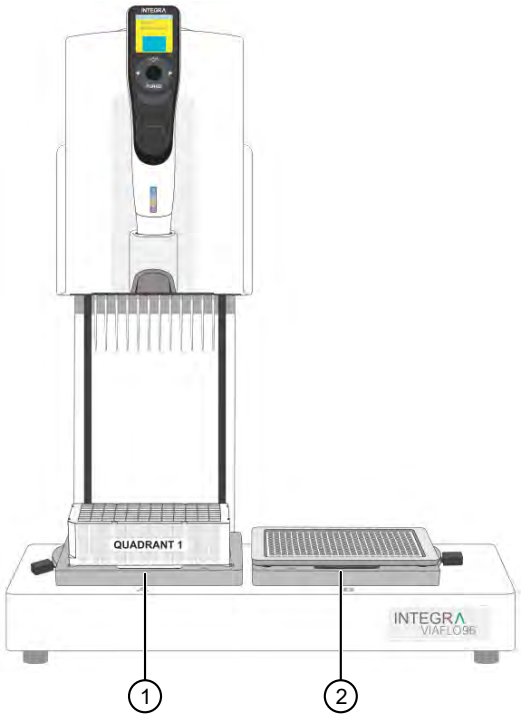
  The **96 TO 384_43µL** VIAFLO method is used when proceeding directly to Stage 6 with no in-process quality controls performed. This method transfers 43 µL/well of the hybridization-ready target from the four 96-deepwell plates to one 384-well PCR plate. A lower transfer volume is used to prevent the samples from making contact with the adhesive seal during target denaturation step due to volume expansion.

Option 1: Perform QC after merging (recommended)

The *96 TO 384_48µL* VIAFLO method merges the four 96-deepwell plates by transferring 48 µL/well of the hybridization-ready target from each plate to one 384-well PCR plate.

1. Select **Custom Programs** ▸ **96 TO 384_48µL** from the VIAFLO main menu.
2. Load the tips onto the VIAFLO with a 96 Channel Pipetting Head.
3. Press **Run** to start the method.

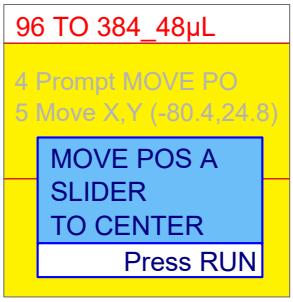
4. Follow the prompts and place the plates on the correct stage.

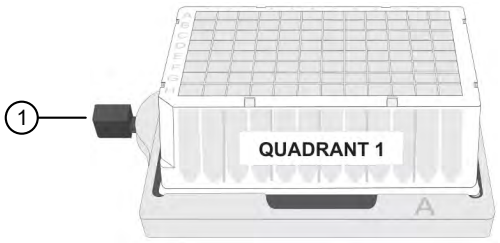


① Position A: Quadrant 1 96-deepwell plate
② Position B: 384-well PCR plate with plate collar

5. Slide the Position A slider to the center orientation, then press **RUN**.

   The Position A slider in the center orientation allows the tips to go to the middle of the 96-deepwell plate, then aspirates from the Quadrant 1 plate.





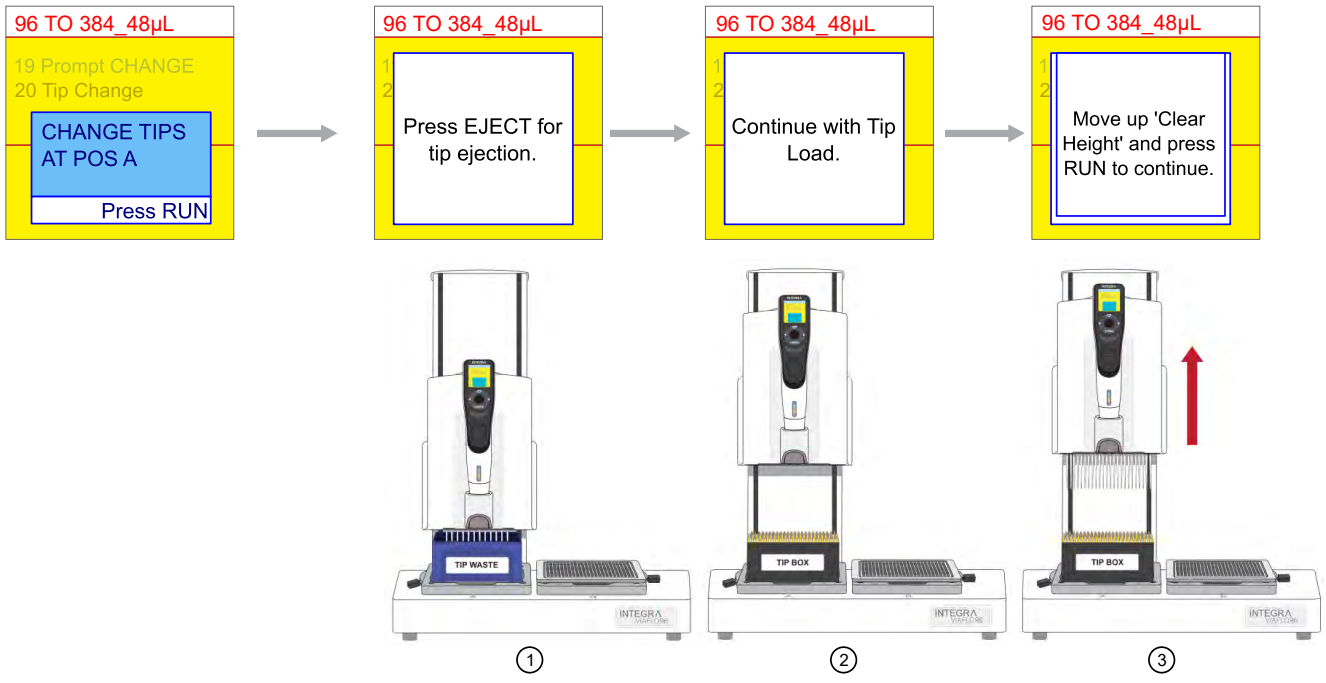
① Slider in the Center orientation.

6. Follow the prompt and move the Position B slider to the down orientation, then press **RUN**.

   The Position B slider in the down orientation moves the tips to dispense to quadrant 1 of the 384-well PCR plate.

① Slider in the Down orientation.

7. Follow the prompts to change tips at Position A.
   a. Remove the 96-deepwell plate in Position A, then replace with a tip waste container in Position A.
   b. Press **EJECT**.
   c. Remove the tip waste container, then replace with a box of pipette tips.
   d. Load the tips onto the VIAFLO with 96 Channel Pipetting Head.
   e. Move the pipette head up to a safe distance above the tip box, then press **RUN**.

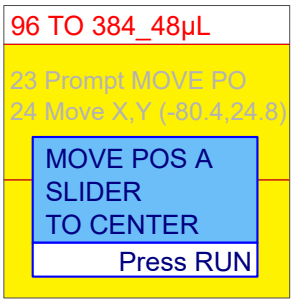


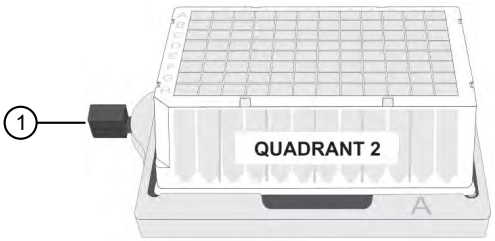
① Swap out the Quadrant 1 plate with a tip waste container.
② Swap out the tip waste container with a tip box.
③ Move the pipette head to a safe position above the tip box.

8. Remove the tip box, place the Quadrant 2 plate in Position A, then press **RUN**.

9. Slide the Position A slider to the center orientation, then press **RUN**.

   The Position A slider in the center orientation allows the tips to go to the middle of the 96-deepwell plate, then aspirates from the Quadrant 2 plate.

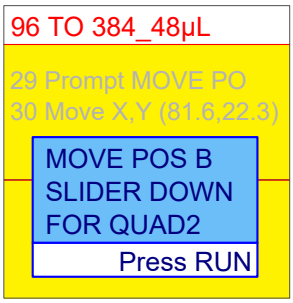




① Slider in the Center orientation.

10. Follow the prompt and move the Position B slider to the down orientation, then press **RUN**.

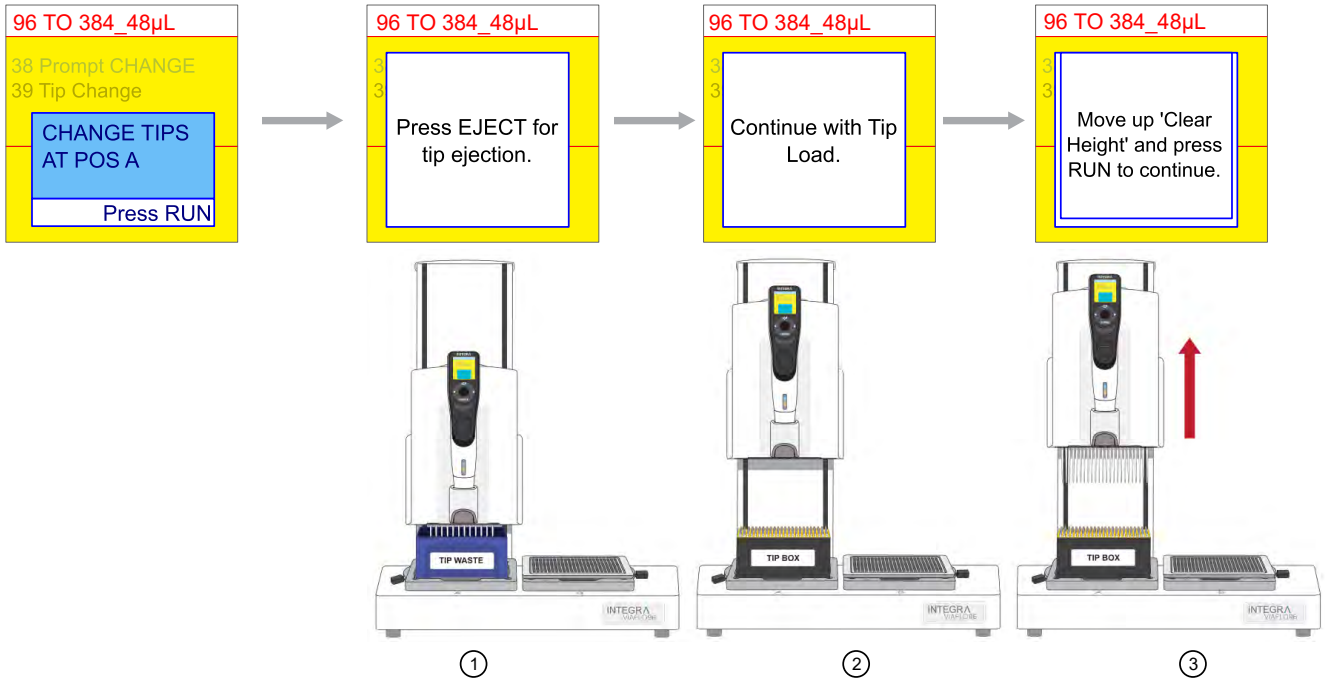
    The Position B slider in the down orientation moves the tips to dispense to quadrant 2 of the 384-well PCR plate.



① Slider in the Down orientation.

11. Follow the prompts to change tips at Position A.
    a. Remove the 96-deepwell plate in Position A, then replace with a tip waste container in Position A.
    b. Press **EJECT**.
    c. Remove the tip waste container, then replace with a box of pipette tips.
    d. Load the tips onto the VIAFLO with 96 Channel Pipetting Head.

e. Move the pipette head up to a safe distance above the tip box, then press **RUN**.

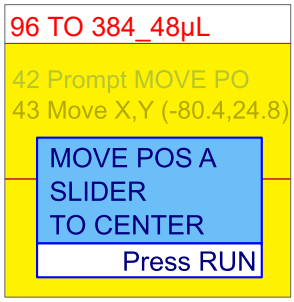


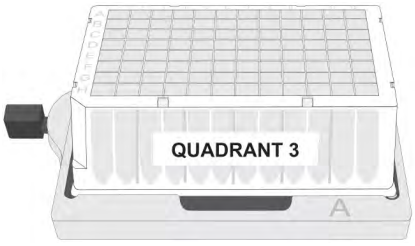
① Swap out the Quadrant 2 plate with a tip waste container.
② Swap out the tip waste container with a tip box.
③ Move the pipette head to a safe position above the tip box.

12. Remove the tip box, place the Quadrant 3 plate in Position A, then press **RUN**.

13. Slide the Position A slider to the center orientation, then press **RUN**.

    The Position A slider in the center orientation allows the tips to go to the middle of the 96-deepwell plate, then aspirates from the Quadrant 3 plate.

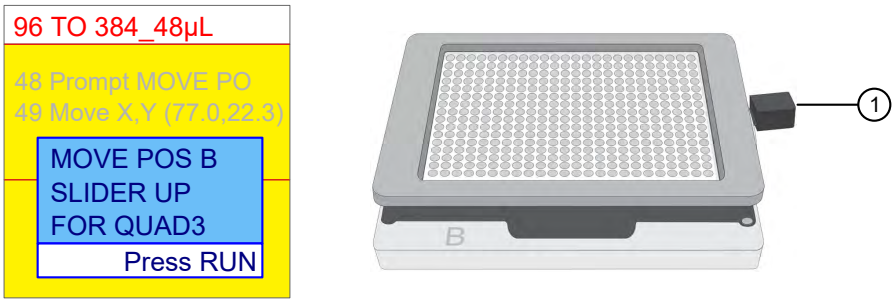




① Slider in the Center orientation.

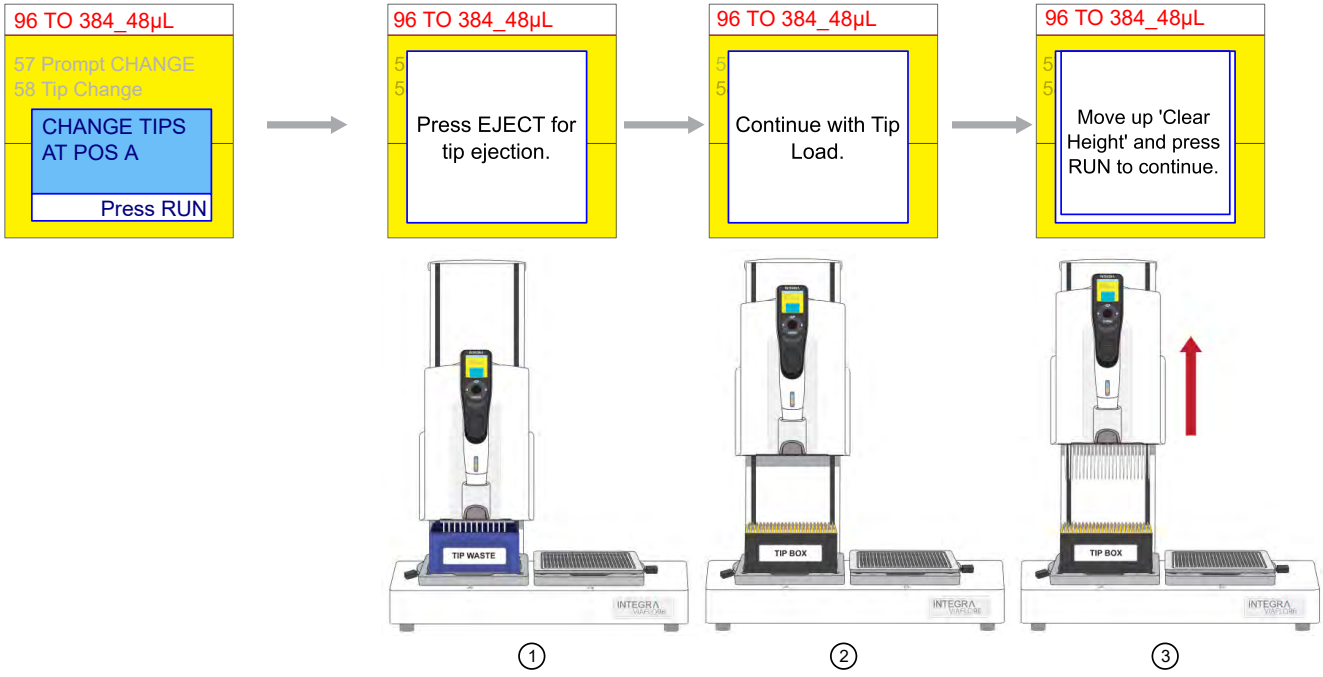
14. Follow the prompt and move the Position B slider to the up orientation, then press **RUN**.

    The Position B slider in the up orientation moves the tips to dispense to quadrant 3 of the 384-well PCR plate.



① Slider in the Up orientation.

15. Follow the prompts to change tips at Position A.
    a. Remove the 96-deepwell plate in Position A, then replace with a tip waste container in Position A.
    b. Press **EJECT**.
    c. Remove the tip waste container, then replace with a box of pipette tips.
    d. Load the tips onto the VIAFLO with 96 Channel Pipetting Head.
    e. Move the pipette head up to a safe distance above the tip box, then press **RUN**.

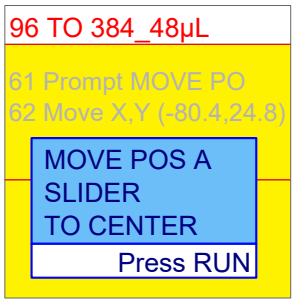


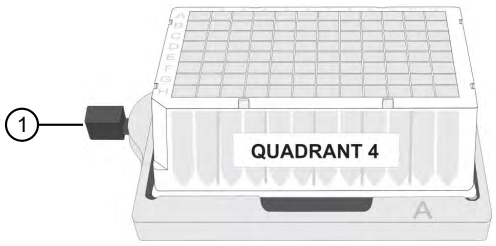
① Swap out the Quadrant 3 plate with a tip waste container.
② Swap out the tip waste container with a tip box.
③ Move the pipette head to a safe position above the tip box.

16. Remove the tip box, place the Quadrant 4 plate in Position A, then press **RUN**.

17. Slide the Position A slider to the center orientation, then press **RUN**.

    The Position A slider in the center orientation allows the tips to go to the middle of the 96-deepwell plate, then aspirates from the Quadrant 4 plate.

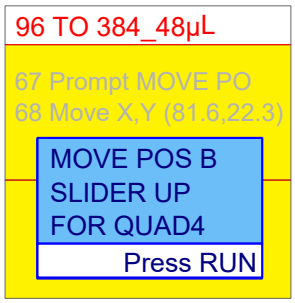




① Slider in the Center orientation.

18. Follow the prompt and move the Position B slider to the up orientation, then press **RUN**.

    The Position B slider in the up orientation moves the tips to dispense to quadrant 4 of the 384-well PCR plate.



① Slider in the Up orientation.

19. Remove the 384-well PCR plate from Position B, then seal the plate with an adhesive seal.

    Tip: After unpeeling a new adhesive seal, use the ion fan to reduce the amount of static on the sticky side of the seal. This minimizes the droplets jumping up onto the seal due to static.

20. Centrifuge the sealed 384-well PCR plate at 675 × *g* for 30 seconds at room temperature.

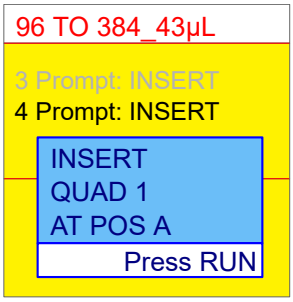
21. Discard the used tips into the waste container.

22. Proceed to “Stage 5A: In-process QC” on page 96 if running in-process QC.

Option 2: Perform only hybridization after merging (no QC)

The *96 TO 384_43µL* VIAFLO method merges the four 96-deepwell plates by transferring 43 µL/well of the hybridization-ready target from each plate to one 384-well PCR plate.

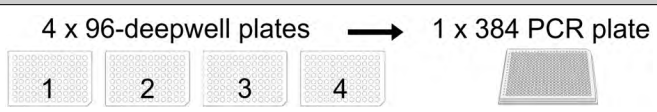
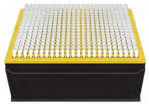
1. Select **Custom Programs ▸ 96 TO 384_43µL** from the VIAFLO main menu.



2. Continue by following the instructions for Option 1 procedure, starting at step 2.
   See “Option 1: Perform QC after merging (recommended)” on page 87.

Merge 96 TO 384 VIAFLO method workflow

To process 8 sample plates, complete 2 cycle of 4 plates.

Plate format transfer		
		
①		Integra VIAFLO with 96 head configured with 2 single stages.
②		Load tips onto the VIAFLO with 96 Channel Pipetting Head.
③		Transfer hybridization-ready target from four 96-deepwell plates to one 384-PCR plate. • Method: 96 TO 384_48µL or 96 TO 384_43µL • Transfer volume: 48 µL or 43 µL
④		Follow the method prompt instructions displayed on the VIAFLO with 96 head.
⑤		Seal the 384-well PCR plate with an adhesive seal. **Tip:** After unpeeling a new adhesive seal, use the ion fan to reduce the amount of static on the sticky side of the seal. This minimizes the droplets jumping up onto the seal due to static.
⑥		Centrifuge the 384-well PCR plate. • Setting: 675 x *g* for 30 seconds
⑦		Continue to “Stage 5A: In-process QC” on page 96 using the VIAFLO 384.

Stage 5A: In-process QC

Equipment and labware required

Quantity	Item
Instruments	
1	VIAFLO™ 384 Base Unit with 2 single-position plate holders and a 384 Channel Pipetting Head (2–50 µL) installed
1	Multidrop™ Combi Reagent Dispenser
2	Multidrop™ Combi SMART 2 Standard tube dispensing cassette[1] for: • Nuclease-free water • Gel diluent
1	Sorvall™ Legend™ XTR Centrifuge set to room temperature
1	Vortex mixer
1	minION™2 Ionizing Air Blower
1	Fume hood
Labware and consumables	
768 tips	Integra VIAFLO XYZ GripTips™, 125 µL
1	384-well PCR plate collar
4	ABgene™ 384-Well 250 µL Polypropylene Storage Plate
2	Greiner Bio-One™ UV-Star™ 384-Well Microplate Flat Bottom
As required	MicroAmp™ Clear Adhesive Film

[1] Each reagent must use a dedicated cassette.

Input samples

Number	Plate name	Content
2 (maximum)	Hyb-Ready Plate	Hybridization-ready target DNA prepared in “Stage 5: Resuspend the pelleted DNA and prepare for hybridization” on page 79 (in aPCR Plate, 384-well, raised chimney).

Reagents required

Reagent	Volume
UltraPure™ DNase/RNase-Free Distilled Water	500 mL
Gel diluent	Prepare as instructed. (See "Prepare the gel diluent" on page 178.)

Stage 5A summary

Stage activities	Instruments required
• Three reagent dispenses—Dilution QC Plates, OD QC Plates, Gel QC Plates • Transfer hybridization-ready target from 384 PCR plate to QC plates	

Perform the pre-run checklist

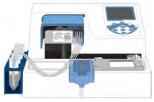
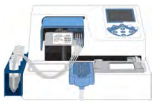
Before starting the workflow, the following tasks must be completed.

Multidrop™ Combi Reagent Dispenser:

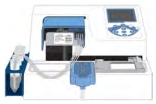
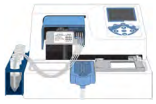
1. Ensure that the Multidrop™ Combi startup procedure has been performed. (See "Start up the Multidrop™ Combi" on page 50.)
2. Perform gravimetric checks before running the workflow. (See "Perform gravimetric checks" on page 51.)
3. Prime the Multidrop™ cassette with reagent. (See "Prime the cassette" on page 54.)
4. Move the ion blowers to each dispensing station, then power them on. (Figure 6 on page 47.)
5. Ensure that the VIAFLO™ 384 Base Unit with a 384 Channel Pipetting Head (2–50 µL) installed is clean and is powered on.

Stage 5A: In-process QC

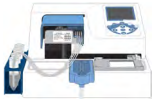
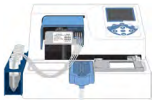
To process 2x384-format sample plates, complete 2 cycles of 1 plate per cycle.

Dilution QC Plates[1]		
	Plates 1 2	
1		Dispense UltraPure™ DNase/RNase-Free Distilled Water • Method: 384-QCDil-95 • Dispense volume: 95 µL
2		After dispensing, seal the plate. Tip: After unpeeling a new adhesive seal, use the ion fan to reduce the amount of static on the sticky side of the seal. This minimizes the droplets jumping up onto the seal due to static.
3		Repeat steps 1 and 2 for the second plate.
4		Centrifuge both plates • Setting: 675 × *g* for 30 seconds
5		Perform the appropriate Multidrop™ procedure: • Flush between batches. See page 55. • Shutdown. See page 56.

[1] The ABgene™ 384-Well 250 µL Polypropylene Storage Plate is used as the Dilution QC Plate.

OD QC Plates[1]		
		Plates 1 2
1		Dispense UltraPure™ DNase/RNase-Free Distilled Water • Method: 384-QC-OD-25 • Dispense volume: 25 µL
2		After dispensing, cover the plate with an unopened adhesive seal.
3		Repeat steps 1 and 2 for the second plate.
4		Continue with Integra VIAFLO 384
5		Perform the appropriate Multidrop™ procedure: • Flush between batches. See page 55. • Shutdown. See page 56.

[1] The Greiner Bio-One™ UV-Star™ 384-Well Microplate Flat Bottom is used as the OD QC Plate.

Gel QC Plates[1]		
	Plates 1 2	
1		Dispense gel diluent • 384-QCGel-60 • Dispense volume: 60 µL
2		After dispensing, cover the plate with an unopened adhesive seal.
3		Repeat steps 1 and 2 for the second plate.
4		Continue with Integra VIAFLO 384
5		Perform the appropriate Multidrop™ procedure: • Flush between batches. See page 55. • Shutdown. See page 56.

[1] The ABgene™ 384-Well 250 µL Polypropylene Storage Plate is used as the Gel QC Plate.

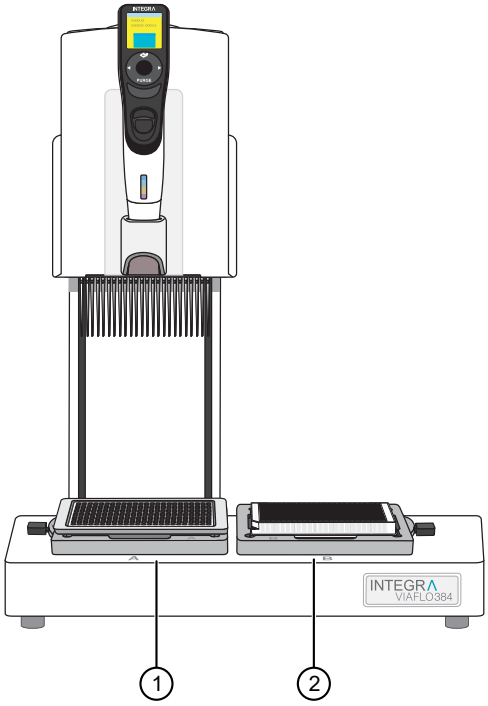
Transfer hybridization-ready target in 384 PCR plate to QC plates

Before running the *384PCR TO QC* method on the VIAFLO 384, the QC plates must be prepared using the Multidrop™ Combi described in stage 5A.

The *384PCR TO QC* method transfers 5 µL of sample from the Hyb-Ready Plate to the Dilution QC Plate, then 5 µL of diluted sample from the Dilution QC Plate to the OD QC Plate, and finally 2.5 µL of diluted sample from the Dilution QC Plate to the Gel QC Plate.

1. Load the tips onto the VIAFLO 384 with a 384 Channel Pipetting Head.
2. Select **Custom Programs ▸ 384PCR TO QC** from the VIAFLO main menu.
3. Ensure that both the Position A and Position B stage sliders are in the center orientation.
4. Press **RUN** to start the method.

5. Follow the prompts and place the plates on the correct stage.



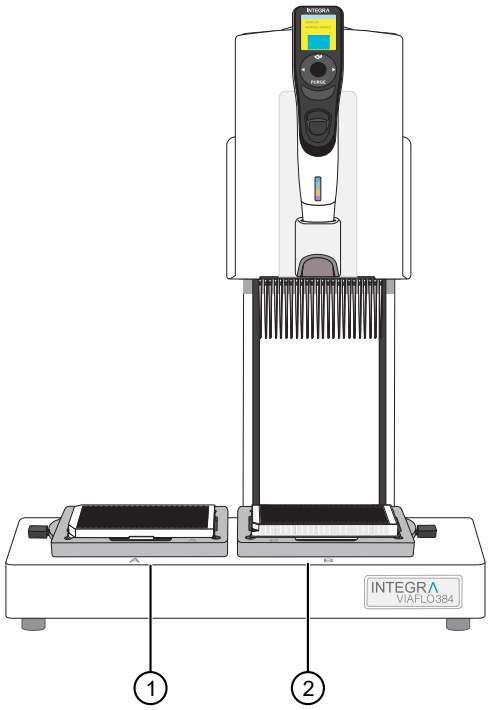
① Position A: Hyb-Ready Plate with plate collar
② Position B: Dilution QC Plate

The VIAFLO 384 aspirates 5 µL/well from Hyb-Ready Plate containing the hybridization-ready samples, then dispenses and mixes into the Dilution QC Plate.

6. When prompted, remove the Hyb-Ready Plate from Position A, then seal with an adhesive seal.

Tip: After unpeeling a new adhesive seal, use the ion fan to reduce the amount of static on the sticky side of the seal. This minimizes the droplets jumping up onto the seal due to static.

7. Place the OD QC Plate in Position A.



① Position A: OD QC Plate
② Position B: Dilution QC Plate

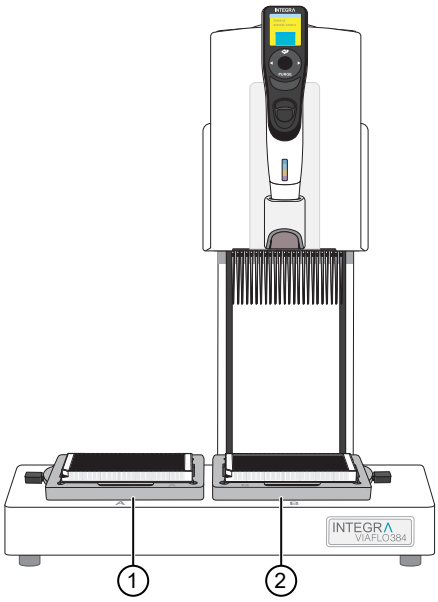
8. Press **RUN** to continue with the method.
   The VIAFLO 384 aspirates 5 µL/well from the Dilution QC Plate, then dispenses and mixes into the OD QC Plate.

9. When prompted, remove the OD QC Plate from Position A, then seal with an adhesive seal.

   Tip: After unpeeling a new adhesive seal, use the ion fan to reduce the amount of static on the sticky side of the seal. This minimizes the droplets jumping up onto the seal due to static.

   OD QC Plates are later vortexed, then centrifuged at 675 × *g* for 30 seconds for QC quantification.

10. Place the Gel QC Plate in Position A.



① Position A: Gel QC Plate
② Position B: Dilution QC Plate

11. Press **RUN** to continue with the method.
The VIAFLO 384 aspirates 2.5 µL/well from the Dilution QC Plate, then dispenses and mixes into the Gel QC Plate.

12. Remove the Gel QC Plate from Position A, then seal with an adhesive seal.

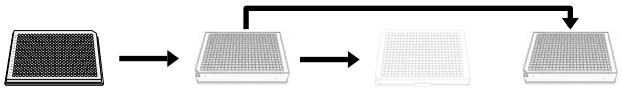
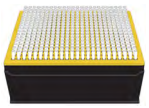
Tip: After unpeeling a new adhesive seal, use the ion fan to reduce the amount of static on the sticky side of the seal. This minimizes the droplets jumping up onto the seal due to static.

Gel QC Plates are later vortexed, then centrifuged at 675 × *g* for 30 seconds for Fragmentation QC.

13. Discard tips into a waste container.

384PCR TO QC VIAFLO method workflow

To process two 384-format sample plates, complete 2 cycles of 1 plate per cycle.

Make QC plates		
	 Hyb-Ready Plate → Dilution QC Plate → OD QC Plate → Gel QC Plate	
1		Integra VIAFLO 384 with a 384 Channel Pipetting Head configured with 2 single stages.
2		Load tips onto the VIAFLO 384.
3		Transfer hybridization-ready target from Hyb-Ready Plate to QC plates. • Method: 384PCR TO QC • Transfer volumes: – 5 µL from 384-Well PCR Plate to Dilution QC Plate – 5 µL from QC Dilution Plate to OD QC Plate – 2.5 µL from QC Dilution Plate to Gel QC Plate
4		Follow the method prompt instructions displayed on the VIAFLO 384.
5		After plate is removed from the VIAFLO 384: • **Hyb-Ready Plate**: Seal with an adhesive seal. • **Dilution QC Plate**: Seal with an adhesive seal. Store in 4°C. • **OD QC Plate**: Seal with an adhesive seal. This plate is later vortexed and centrifuged at 675 × *g* for 30 seconds for QC quantification on a plate reader. (See "Use a Multiskan™ Sky session" on page 189.) • **Gel QC Plate**: Seal with an adhesive seal. This plate is later vortexed and centrifuged at 675 × *g* for 30 seconds for Fragmentation QC. **Tip:** After unpeeling a new adhesive seal, use the ion fan to reduce the amount of static on the sticky side of the seal. This minimizes the droplets jumping up onto the seal due to static.

Stage 6: Denature the target and transfer to hybridization tray

Equipment and labware required

Quantity	Item
Instruments	
1	VIAFLO™ 384 Base Unit with 2 single-position plate holders and a 384 Channel Pipetting Head (2–50 µL) installed
1	Fume hood
1	Thermal cycler, Applied Biosystems™ ProFlex™ 2 x 384-well PCR System
1	Heratherm™ Advanced Protocol Microbiological Incubator, capacity 66 L, BINDER™ ED 56, or BINDER™ BD 56 (Set to 48°C.) • Must maintain a constant temperature of 48°C for at least 24 hours with a temperature accuracy of ±1°C.
1	Sorvall™ Legend™ XTR Centrifuge set to room temperature. (As needed.)
1	Vortex mixer (As needed.)
Labware and consumables	
1	384-Well PCR Plate Collar
1	Plate Alignment Tool
2	384-Layout GeneTitan™ Hybridization Tray
768 tips	Integra VIAFLO XYZ GripTips™, 125 µL

Input samples

Number	Plate name	Content
2	Hyb-Ready Plate	Hybridization-ready target DNA prepared in “Stage 5: Resuspend the pelleted DNA and prepare for hybridization” (PCR Plate, 384-well, raised chimney)

Stage 6 summary

Stage activities	Instruments required
• Denature target in thermal cycler • Transfer from 384-format PCR plate to hybridization tray • Off-line incubation of the array plate/hybridization tray stack at 48°C for 23.5–24 hours	

Perform the pre-run checklist

Before starting the workflow, the following tasks must be completed.

1. Ensure that the VIAFLO™ 384 Base Unit with a 384 Channel Pipetting Head (2–50 µL) installedis clean and is powered on.
2. Ensure that the oven is powered on and has warmed up to 48°C.
3. Ensure that the Proflex™ thermal cycler is powered on and loaded with the **Axiom Denature** thermal cycler protocol.

Warm the array plate to room temperature

The array plate must be at room temperature before setting up hybridization.

1. Remove the array plate packaging from the 4°C refrigerated storage.
2. Open the array plate box, then remove the pouch containing the array plate and protective base. Do not open the pouch.
3. Equilibrate the unopened pouch on the bench for at least 25 minutes.
4. During, or at the end of the array warm up time, open the pouch and scan the array plate barcode into the GeneTitan™ Array Plate Registration file.
   See Appendix D, “Register samples in GeneChip™ Command Console™”.

WARNING! Do not remove the array plate from the protective base or touch the surface of any of the arrays.

Prepare hybridization-ready samples stored at –20°C

1. Warm the Hyb-Ready Plate at room temperature for 5 minutes. It is not necessary to equilibrate the plate for longer than 5 minutes.
2. Ensure that the Hyb-Ready Plate is sealed well. If the plate is not sealed well:
   a. Centrifuge the plate, then carefully remove the old seal.
   b. If there is condensation on the top of the plate, blot dry gently with a laboratory tissue.

c. Tightly reseal the plate with a fresh seal.

d. Vortex the newly sealed plate, then proceed to the next step.

3. Centrifuge for 30 seconds, then proceed to Stage 6.

Stage 6: Denature the target and transfer to hybridization tray

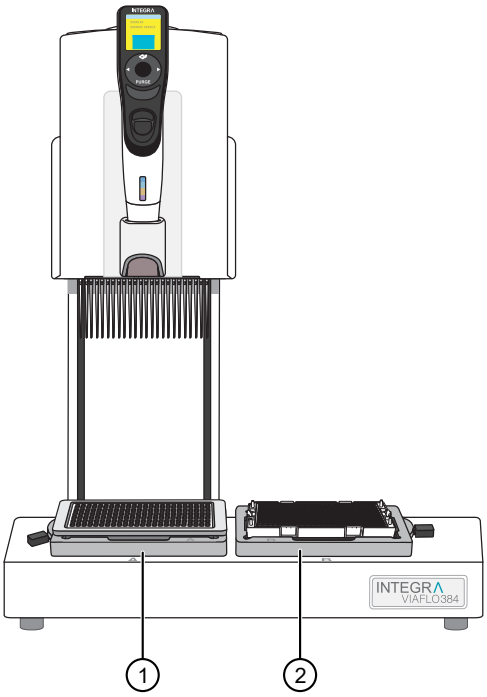
To process 2x384-format sample plates, complete 2 cycles of 1 plate per cycle.

Denature and transfer to hybridization tray		
	Plates 1 2	
①		Denature target • Protocol: **Axiom Denature**
②		Transfer from PCR plate to hybridization tray • Method: 384PCR TO HYB • Dispense volume: 37 µL
③		Incubate • Settings: 48°C, 23.5–24 hours

Transfer hybridization-ready target in Hyb-Ready Plate to hybridization tray

The *384PCR TO HYB* method transfers 37 µL of denatured hybridization-ready target from the 384-well PCR plate to the hybridization tray.

1. Load the tips onto the VIAFLO 384 with a 384 Channel Pipetting Head.
2. Select **Custom Programs ▸ 384PCR TO HYB** from the VIAFLO main menu.
3. Ensure that both the Position A and Position B stage sliders are in the center orientation.
4. Press **RUN** to start the method.
5. Follow the prompts and place the plates on the correct stage.



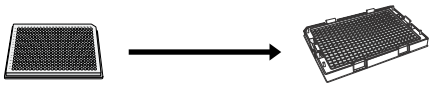
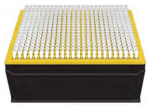
① Position A: Hyb-Ready Plate with plate collar
② Position B: 384-Layout GeneTitan™ Hybridization Tray

The VIAFLO 384 aspirates from the Hyb-Ready Plate containing the denatured hybridization-ready samples, then dispenses to the 384-Layout GeneTitan™ Hybridization Tray.

6. When complete, inspect the Hyb-Ready Plate to ensure that the hybridization-ready target was transferred to the hybridization tray.
7. After the method is complete:
   a. Continue to “Perform array plate clamping and off-line hybridization” on page 109.
   b. Discard tips into a waste container.

384PCR TO HYB VIAFLO method workflow

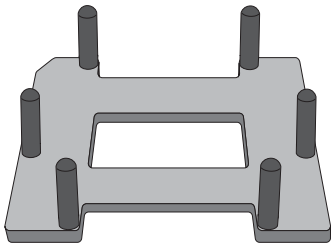
To process two 384-format sample plates, complete 2 cycles of 1 plate per cycle.

384-well PCR plate to 384-Layout GeneTitan™ Hybridization Tray		
	 384-well PCR plate 384 Layout Hybridization Tray	
1		Integra VIAFLO 384 with a 384 Channel Pipetting Head configured with 2 single stages.
2		Load tips onto the VIAFLO 384.
3		Transfer hybridization-ready target from 384-well PCR plate to hybridization tray. • Method: 384PCR TO HYB • Transfer volume: 37 µL
4		Follow the method prompt instructions displayed on the VIAFLO 384.
5		After transfer is complete, clamp the array plate to the hybridization tray using the Plate Alignment Tool.
6		Incubate the array plate/hybridization tray in a hybridization oven set at 48°C for 23.5–24 hours.

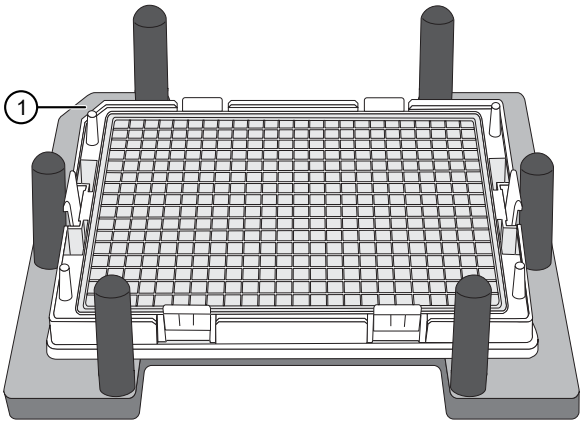
Perform array plate clamping and off-line hybridization

This procedure is performed for all plates in the workflow.

1. Place the Plate Alignment Tool flat on the table top.



2. Carefully place the hybridization tray containing the denatured hybridization target into the Plate Alignment Tool as shown in the following figure. Ensure that the hybridization tray is sitting inside the tool and that the notched corner of the hybridization tray is aligned with the notched corner of the Plate Alignment Tool.



(1) Notched corner of the Plate Alignment Tool and hybridization tray aligned.

3. Hold the array plate with the arrays facing down, then slowly align the array plate into the Plate Alignment Tool until the entire plate is positioned within the 6 alignment posts of the tool. Ensure that the notched corner of the arrays plate is aligned with the notched corner of the hybridization tray. (See Figure 8.)

IMPORTANT! Do not lower the array plate to the hybridization tray if the plate is not aligned within the alignment posts on the tool.

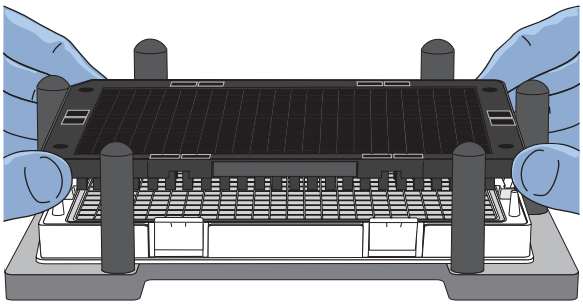
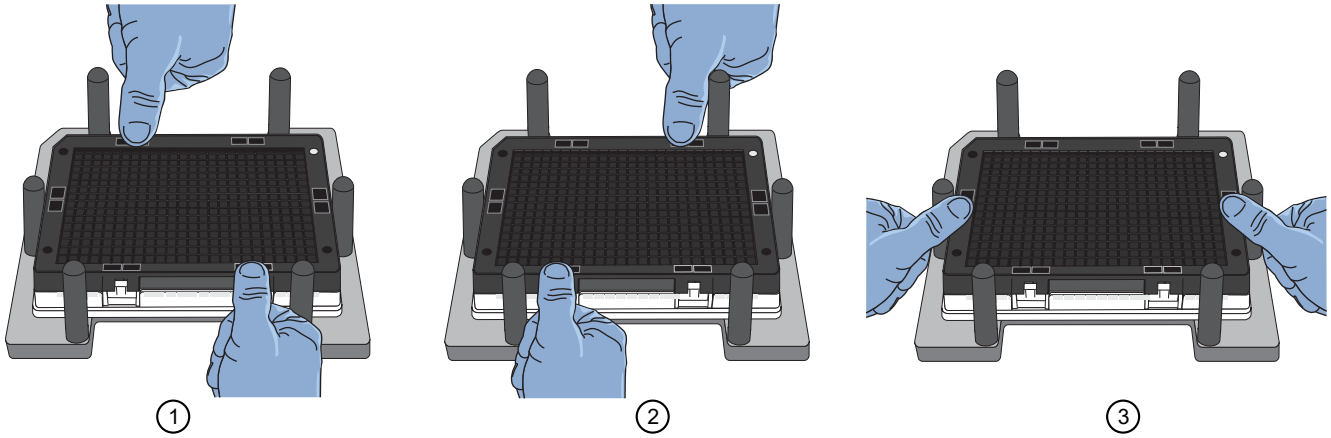


Figure 8 Place the array plate on the hybridization tray in the Plate Alignment Tool.

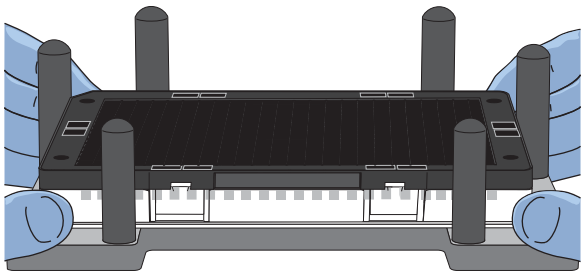
4. After the array plate is aligned inside the tool, slowly lower the plate towards the hybridization tray until a snapping sound is heard. The snapping sound indicates that the array plate and hybridization tray have been properly latched together.

IMPORTANT! Lowering the array plate to the hybridization tray as slowly as possible prevents air bubbles from generating inside the hybridization tray.

5. Position your hands as shown in the following figures, then lightly push the array plate downward to ensure that each latching mechanism is tightly closed.



6. Grip the array plate/hybridization tray stack from the hybridization tray at the bottom of the stack, then lift to remove the entire stack from the Plate Alignment Tool.



7. Inspect the stack to ensure that all clamps are fastened.

8. Make a note of the array plate barcode number for later use.

   The barcode number is used to create the GeneTitan™ Array Plate Registration file and for the GeneTitan™ instrument **Wash-Scan** setup.

   Tip: Scan the barcode directly into a blank registration file or into a text program, such as Microsoft™ Notepad, Word™, or Excel™ applications.

9. Place the array plate/hybridization tray stack onto the wire racks of the hybridization oven set at 48°C.

   Note: Do not stack the array plate/hybridization tray stacks on top of one another.

   Note: Do not place the array plate/hybridization tray stack on the bottom of the hybridization oven.

10. Incubate at 48°C for 23.5–24 hours. Set a timer or record the time.

Stage 7: Preparing ligation, stain, stabilization reagent trays, and scan tray for the GeneTitan™ MC Instrument

Equipment and labware required

Quantity	Item
Instruments	
5	Multidrop™ Combi Reagent Dispenser
5	Multidrop™ Combi SMART 2 Standard tube dispensing cassette[1]
1	Vortex mixer
1	Mini centrifuge
1	Electronic pipettor for serological pipettes
1	P1000 pipette
Labware and consumables	
4	250-mL conical bottle (Nunc™ 250 mL Wide Mouth Conical Centrifuge Tube)
As required	Serological pipette (5 mL, 10 mL, and 50 mL)
As required	P1000 pipette tips
4	384-Layout GeneTitan™ Stain Tray (Stain 1)
2	384-Layout Axiom™ Stain 2 Tray
2	384-Layout Axiom™ Stabilization Tray
2	384-Layout Axiom™ Ligation Tray
2	384-Layout GeneTitan™ Scan Tray
12	384-Layout GeneTitan™ Scan and Stain Tray Cover

[1] Each reagent must use a dedicated cassette.

Reagent handling

Module 4-1, 4x96F and Module 4-2, 4x96F reagents from the Axiom™ Propel XPRES Reagent Kit, 2x384HT:

- Axiom™ Propel Fast Wash Reagent Kit Module 4-1 for 96F or 384HT, Part No. 952369
- Axiom™ Propel Reagent Kit Module 4-2 for 96F or 384HT, Part No. 952268

Thaw and prepare reagents according to the following table.

Table 26 Reagent handling for Stage 7 reagents.

Module	Reagent	Component Part No.	Treatment
Axiom™ Propel Fast Wash Reagent Kit Module 4-1 for 96F or 384HT –20°C	Axiom™ Propel Ligation Buffer	952208	• Thaw and equilibrate to room temperature on the day of use, or • Thaw at 4°C the day before then equilibrate to room temperature on the day of use. • Vortex before master mix preparation. **Note:** White precipitate is sometimes observed when the Axiom™ Propel Ligation Buffer is thawed. The presence of some precipitate is acceptable and does not adversely impact assay performance. Use the following instructions to resuspend most precipitate before use. **1.** Vortex for 30 seconds. **2.** Examine the buffer for precipitate. **3.** If precipitate is still present, warm the bottle with your hands, then vortex again for 30 seconds.
	Axiom™ Fast Ligation Enzyme	952367	• Keep in –20°C until ready for use. • Flick 5 times before master mix preparation.
	Axiom™ Propel Ligation Solution 1	952212	• Thaw and equilibrate to room temperature on the day of use, or • Thaw at 4°C the day before then equilibrate to room temperature on the day of use. • Vortex before master mix preparation.
	Axiom™ Propel Probe Mix 1	952213	• Thaw and equilibrate to room temperature on the day of use, or • Thaw at 4°C the day before then equilibrate to room temperature on the day of use. • Vortex before master mix preparation.
	Axiom™ Propel Stain Buffer	952214	• Thaw and equilibrate to room temperature on the day of use, or • Thaw at 4°C the day before then equilibrate to room temperature on the day of use. • Vortex before master mix preparation.

Table 26 Reagent handling for Stage 7 reagents. *(continued)*

Module	Reagent	Component Part No.	Treatment
Axiom™ Propel Fast Wash Reagent Kit Module 4-1 for 96F or 384HT –20°C	Axiom™ Propel Stabilize Solution	952215	• Thaw and equilibrate to room temperature on the day of use, or • Thaw at 4°C the day before then equilibrate to room temperature on the day of use. • Vortex, then centrifuge briefly before master mix preparation.
Axiom™ Propel Reagent Kit Module 4-2 for 96F or 384HT 4°C	Axiom™ Propel Ligation Solution 2	952216	• Remove from 4°C and let equilibrate to room temperature on day of use. • Vortex before master mix preparation.
	Axiom™ Propel Probe Mix 2	952217	• Remove from 4°C and let equilibrate to room temperature, but *keep it stored away from light*. • Flick 5 times before master mix preparation.
	Axiom™ Propel Wash A	952218	• Remove from 4°C and equilibrate to room temperature. • Vortex before master mix preparation. If precipitate is present, vortex again to dissolve the precipitate.
	Axiom™ Propel Stain 1-A	952219	• Remove from 4°C and let equilibrate to room temperature on day of use. • Flick 5 times, then centrifuge briefly before master mix preparation.
	Axiom™ Propel Stain 1-B	952258	• Remove from 4°C and let equilibrate to room temperature on day of use. • Flick 5 times, then centrifuge briefly before master mix preparation.
	Axiom™ Propel Stain 2-A	952231	• Remove from 4°C and let equilibrate to room temperature on day of use. • Flick 5 times, then centrifuge briefly before master mix preparation.
	Axiom™ Propel Stain 2-B	952260	• Remove from 4°C and let equilibrate to room temperature on day of use. • Flick 5 times, then centrifuge briefly before master mix preparation.

Table 26 Reagent handling for Stage 7 reagents. *(continued)*

Module	Reagent	Component Part No.	Treatment
Axiom™ Propel Reagent Kit Module 4-2 for 96F or 384HT 4°C	Axiom™ Propel Stabilize Diluent	952248	• Remove from 4°C and let equilibrate to room temperature on day of use. • Vortex before master mix preparation.
	Axiom™ Water	952177	• Remove from 4°C and let equilibrate to room temperature on day of use.
	Axiom™ Propel Hold Buffer	952254	• Remove from 4°C and let equilibrate to room temperature on day of use. • Vortex before use.

Reagent preparation for Stage 7: Prepare GeneTitan™ reagents

You can add flexibility to the timing of array processing by making the following GeneTitan™ reagents up to 2 days (48 hours) in advance.

- Stain 1 and Stain 2 Master Mixes can be prepared up to 2 days in advance, wrapped in foil, then stored at 4°C.
- Stabilization Master Mix can be prepared up to 2 days in advance and stored at 4°C.
- Ligate Master Mix can be prepared with all the components listed up to 2 days in advance, except the Axiom™ Fast Ligation Enzyme. Wrap the master mix bottle in foil, then store at 4°C. Axiom™ Fast Ligation Enzyme must be kept at –20°C and added only when ready to be dispensed into the ligation tray and loaded into the GeneTitan™ MC Instrument.

IMPORTANT! Add the Axiom™ Fast Ligation Enzyme (#6) only if 2 GeneTitan™ MC Instruments instruments (GTMC) are available to process *and* stain trays for both array plates can be loaded into GTMC within 1 hour. If there is only 1 GeneTitan™ MC Instrument available to process (only 1 array plate can be processed at a time), see “Ligate Master Mix—advance preparation procedure” on page 116. Consult with your Field Application Scientist for further guidance.

The following sections provide detailed procedures for master mix preparation.

Prepare Ligate Master Mix

Ensure that all components are treated according to the reagent handling table before preparing the master mix. (See Table 26.)

1. In a 250-mL conical bottle, add reagents in the order shown.

Table 27 Ligate Master Mix for 2x384HT array format plates.

Order of addition	Reagent	Quantity	Transfer method
1	Axiom™ Propel Ligation Buffer	41.0 mL	50-mL serological pipette
2	Axiom™ Propel Ligation Solution 1	8.1 mL	10-mL serological pipette
3	Axiom™ Propel Probe Mix 1	6.5 mL	10-mL serological pipette
4	Axiom™ Propel Probe Mix 2	6.5 mL	10-mL serological pipette
5	Axiom™ Propel Ligation Solution 2	1.95 mL	P1000 pipette
6	Axiom™ Fast Ligation Enzyme[1]	2.45 mL	P1000 pipette
	Total volume	**66.5 mL**	

[1] See the IMPORTANT note in the section, "Reagent preparation for Stage 7: Prepare GeneTitan™ reagents" on page 115.

2. Ensure that the cap of the conical bottle is closed tightly, then invert the Ligate Master Mix at least 15 times to mix.

3. Protect the Ligate Master Mix from direct light by covering with aluminum foil.

4. Leave the master mix at room temperature and use within 1 hour of its preparation.

Ligate Master Mix—advance preparation procedure

To provide flexibility for the workflow and to accommodate the number of GeneTitan™ MC Instruments that are available to process array plates, the Ligation Master Mix can be made up to 2 days (48 hours) in advance, but without the Axiom™ Fast Ligation Enzyme.

Note: The Axiom™ Fast Ligation Enzyme must be added within 1 hour before dispensing into the ligation tray and then loading intoGeneTitan™ MC Instrument.

1. Add the reagents 1–5 from Table 27. *Do not* add the Axiom™ Fast Ligation Enzyme.
2. Invert the conical tube at least 15 times to mix.

   IMPORTANT! If there is only 1 GTMC available to process array plates (only 1 array plate can be processed at a time), divide the Ligation Master Mix into 2 aliquots in 50-mL conical tubes with 32 µL volume each, then label the tubes to indicate “No Enzyme”.
3. Protect the Ligate Master Mix from direct light by covering with aluminum foil, then store at 4°C.

   IMPORTANT! Label the 250-mL conical tube to indicate that the content contains "*No Enzyme*".
4. On the day of use, remove from the 4°C storage, then allow the Ligate Master Mix to equilibrate to room temperature.

   Note: If there is only 1 GTMC available to process array plates (only 1 array plate can be processed at a time), remove only 1 of the 50-mL conical tubes with 32 µL Ligation Master Mix.

   Equilibration can take up to 30 minutes.
5. Within 1 hour before dispensing to the ligation tray, add the Axiom™ Fast Ligation Enzyme to the conical tube.

   Note: If there is only 1 GTMC available to process array plates (only 1 array plate can be processed at a time), add 1.23 mL of Axiom™ Fast Ligation Enzyme to 1 of the 50-mL conical tubes with 32 µL Ligation Master Mix.
6. Invert the tube at least 15 times to mix.
7. Leave the master mix at room temperature and dispense into the ligation tray, then Load into the GTMC within 1 hour of preparation.

Prepare Stain 1 Master Mix

Ensure that all components are treated according to the reagent handling table before preparing the master mix. (See Table 26.)

Note: Two stain trays of Stain 1 Master Mix are required per 384 samples, therefore, a higher volume of master mix is required.

1. In a 250-mL conical bottle, add reagents in the order shown.

Table 28 Stain 1 Master Mix for 2x384HT array format plates.

Order of addition	Reagent	Quantity	Transfer method
1	Axiom™ Propel Wash A	108 mL	50-mL serological pipette
2	Axiom™ Propel Stain Buffer	2.3 mL	5-mL serological pipette
3	Axiom™ Propel Stain 1-A	1.13 mL	P1000 pipette
4	Axiom™ Propel Stain 1-B	1.13 mL	P1000 pipette
	Total volume	**112.56 mL**	

2. Ensure that the cap of the conical bottle is closed tightly, then invert the Stain 1 Master Mix at least 15 times to mix.

3. Protect the Stain 1 Master Mix from direct light by covering with aluminum foil.

4. Store the prepared master mix in one of the following ways:
   - Store at room temperature and use within 2 hours of its preparation.
   - **Optional:** To provide flexibility for the workflow, the master mix can be made up to 2 days in advance, then stored in the dark at 4°C. On the day of use, remove from the 4°C storage, then allow the master mix to equilibrate to room temperature. Equilibration can take up to 30 minutes.

Prepare Stain 2 Master Mix

Ensure that all components are treated according to the reagent handling table before preparing the master mix. (See Table 26.)

1. In a 250-mL conical bottle, add reagents in the order shown.

Table 29 Stain 2 Master Mix for 2x384HT array format plates.

Order of addition	Reagent	Quantity	Transfer method
1	Axiom™ Propel Wash A	63 mL	50-mL serological pipette
2	Axiom™ Propel Stain Buffer	1.31 mL	P1000 pipette
3	Axiom™ Propel Stain 2-A	0.66 mL	P1000 pipette
4	Axiom™ Propel Stain 2-B	0.66 mL	P1000 pipette
	Total volume	**65.63 mL**	

2. Ensure that the cap of the conical bottle is closed tightly, then invert the Stain 2 Master Mix at least 15 times to mix.
3. Protect the Stain 2 Master Mix from direct light by covering with aluminum foil.
4. Store the prepared master mix in one of the following ways:
   - Store at room temperature and use within 2 hours of its preparation.
   - **Optional:** To provide flexibility for the workflow, the master mix can be made up to 2 days in advance, then stored in the dark at 4°C. On the day of use, remove from the 4°C storage, then allow the master mix to equilibrate to room temperature. Equilibration can take up to 30 minutes.

Prepare Stabilization Master Mix

Ensure that all components are treated according to the reagent handling table before preparing the master mix. (See Table 26.)

1. In a 250-mL conical bottle, add reagents in the order shown.

Table 30 Stabilization Master Mix for 2x384HT array format plates.

Order of addition	Reagent	Quantity	Transfer method
1	Axiom™ Water	58 mL	50-mL serological pipette
2	Axiom™ Propel Stabilize Diluent	6.5 mL	10-mL serological pipette
3	Axiom™ Propel Stabilize Solution	0.815 mL	P1000 pipette
	Total volume	**65.3 mL**	

2. Ensure that the cap of the conical bottle is closed tightly, vortex for 5 seconds, then leave at room temperature until use.

3. Store the prepared master mix in one of the following ways:
   - Store at room temperature and use within 2 hours of its preparation.
   - **Optional:** To provide flexibility for the workflow, the master mix can be made up to 2 days in advance, then stored at 4°C. On the day of use, remove from the 4°C storage, then allow the master mix to equilibrate to room temperature. Equilibration can take up to 30 minutes.

Prepare the Axiom™ Propel Hold Buffer

Ensure that the Axiom™ Propel Hold Buffer is vortexed before use.

1. Use the Axiom™ Propel Hold Buffer bottle for Multidrop™ dispensing.

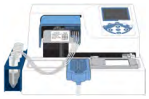
Table 31 Axiom™ Propel Hold Buffer for 2x384HT array format plates.

Reagent	Quantity	Transfer method
Axiom™ Propel Hold Buffer	Entire bottle	Use straight from bottle[1]

[1] The Axiom™ Propel Hold Buffer reagent bottle can be used for Multidrop™ Combi dispensing.

2. Leave at room temperature until use.

Stage 7 summary

Stage activities	Instruments required
Five reagent dispenses—Ligation Master Mix, Stain 1 Master Mix, Stain 2 Master Mix, Stabilization Master Mix, Axiom™ Hold Buffer	

Prepare the GeneTitan™ trays

Familiarity with handling GeneTitan™ is required. If needed, review the proper techniques provided in this document. See Appendix A, "Recommended techniques for GeneTitan™ MC Instrument operation".

1. Collect and label the stain trays as described in the following table.

Quantity per array plate	Tray type	Label[1]
1	Ligation Tray 	*Lig*
2	Stain 1 Tray 	*Stain 1* or *S1*
1	Stain 2 Tray 	*Stain 2* or *S2*
1	Stabilization Tray 	*Stbl*

[1] It is critical that you write on the proper location of the stain/reagent trays. See "Label a GeneTitan™ 384 Layout Reagent Tray" on page 156.

2. Collect enough covers for the trays (5 times the number of array plates).

3. Place the labeled stain trays and covers next to the Multidrop™ Combi. Ensure that each labeled stain tray is placed next to the correct Multidrop™ Combi dispensing the appropriate/corresponding reagent.

   For example, stain trays labeled "Stain 2" are placed next to the Multidrop™ Combi dispensing the Stain 2 Master Mix.

4. Deionize the stain trays by placing them directly in front of the ion blower and allowing the air to stream across them for at least 10 seconds.

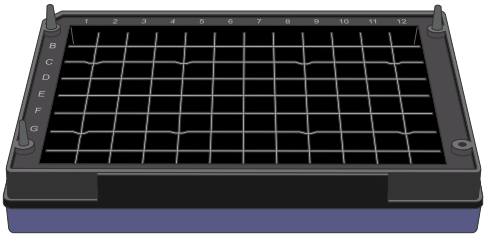
   Alternatively, trays can be deionized manually using the GeneTitan™ ZeroStat AntiStatic Gun. See "Manual deionization of GeneTitan™ trays" on page 159.

Perform the pre-run checklist

Before starting the workflow, the following tasks must be completed.

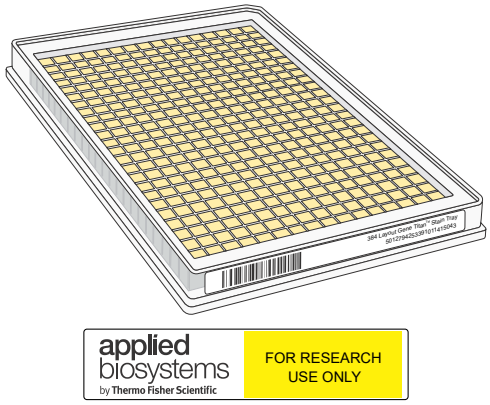
1. Ensure that the Multidrop™ Combi startup procedure has been performed. (See “Start up the Multidrop™ Combi” on page 50.)
2. Perform gravimetric checks before running the workflow. (See “Perform gravimetric checks” on page 51.)
3. Prime the Multidrop™ cassette with reagent. (See “Prime the cassette” on page 54.)
4. Ensure that each labeled stain tray is placed next to the correct Multidrop™ Combi dispensing the corresponding reagent.
5. Move the ion blowers to each dispensing station, then power them on. (Figure 6 on page 47.)

Stage 7: Prepare GeneTitan™ reagents for two 384HT array plates

Scan tray dispense		
Plates 1 2		
		
⚠ **CAUTION!** Do not remove the Scan Tray from the blue scan tray protective base until loading onto the GeneTitan™ MC Instrument.		
①		Dispense Axiom™ Hold Buffer • Method: 384-Scan-170 • Dispense volume: 170 µL
②	✓	Examine the Scan Tray. • If bubbles are present, puncture them with a pipette tip. • If droplets of liquid splashed onto the well dividers, use a laboratory tissue and gently dab the droplet on top of the tray divider to remove the droplet without disturbing the liquid inside the tray.
③	ⓘ	Place the Scan Tray cover onto the Scan Tray.
④	ⓘ	• Continue to "Stain tray dispense—Ligation Master Mix" • Perform the appropriate Multidrop™ procedure: – Flush between batches. See page 55. – Shutdown. See page 56.

Stain tray dispense—Ligation Master Mix

Plates 1 2



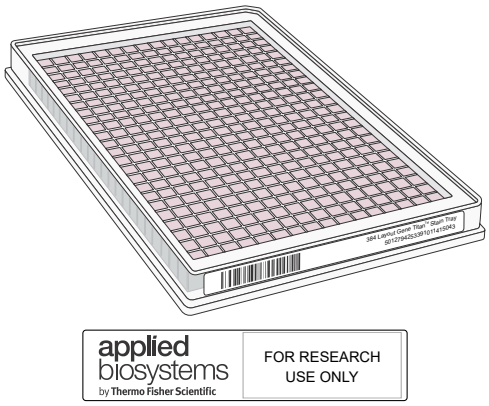
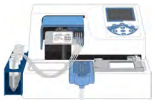
- Place the ion blowers at the dispensing station. If ion blowers are not available, the GeneTitan™ ZeroStat AntiStatic Gun can be used to individually deionize the stain trays. (See “Deionization of GeneTitan™ stain trays” on page 158.)

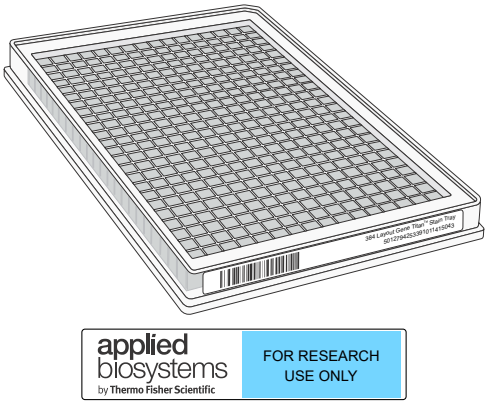
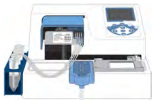
IMPORTANT! Deionze the trays with ion blowers or GeneTitan™ ZeroStat AntiStatic Gun before dispensing the reagents.

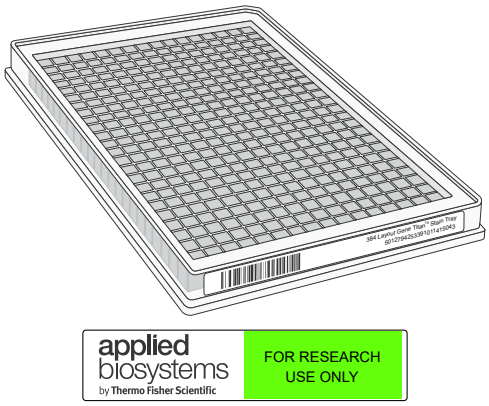
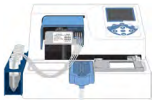
IMPORTANT! Ensure that the stain trays labeled “Lig” are used for the Multidrop™ Combi dispensing the Ligation Master Mix.

IMPORTANT! If the Ligation Master Mix was made a day in advance, ensure that the Axiom™ Fast Ligation Enzyme is added to the mixture before dispensing.

①		Dispense Ligation Master Mix • Method: 384-Stain-40 • Dispense volume: 40 µL
②	✓	Examine the Ligation Tray. • If bubbles are present, puncture them with a pipette tip. • If droplets of liquid splashed onto the well dividers, use a laboratory tissue and gently dab the droplet on top of the tray divider to remove the droplet without disturbing the liquid inside the tray.
③	ⓘ	• Place a stain tray cover on top of each Ligation stain tray. Continue to "Stain tray dispense—Stain 1 Master Mix" • Protect the trays from light if not immediately loading onto the GeneTitan™ MC Instrument. • Perform the appropriate Multidrop™ procedure: – Flush between batches. See page 55. – Shutdown. See page 56.

Stain tray dispense—Stain 1 Master Mix		
Plates 1 2 3 4		
		
IMPORTANT! Deionze the trays with ion blowers or GeneTitan™ ZeroStat AntiStatic Gun before dispensing the reagents. **IMPORTANT!** Ensure that the stain trays labeled “Stain 1” or “S1” are used for the Multidrop™ Combi dispensing the Stain 1 Master Mix.		
①		Dispense Stain 1 Master Mix • Method: 384-Stain-40 • Dispense volume: 40 µL
②	✓	Examine the Stain 1 Tray. • If bubbles are present, puncture them with a pipette tip. • If droplets of liquid splashed onto the well dividers, use a laboratory tissue and gently dab the droplet on top of the tray divider to remove the droplet without disturbing the liquid inside the tray.
③	ⓘ	• Place a stain tray cover on top of each Stain 1 stain tray. Continue to "Stain tray dispense—Stain 2 Master Mix" • Protect the trays from light if not immediately loading onto the GeneTitan™ MC Instrument. • Perform the appropriate Multidrop™ procedure: – Flush between batches. See page 55. – Shutdown. See page 56.

Stain tray dispense—Stain 2 Master Mix		
Plates 1 2		
		
IMPORTANT! Deionze the trays with ion blowers or GeneTitan™ ZeroStat AntiStatic Gun before dispensing the reagents. **IMPORTANT!** Ensure that the stain trays labeled “Stain 2” or “S2” are used for the Multidrop™ Combi dispensing the Stain 2 Master Mix.		
①		Dispense Stain 2 Master Mix • Method: 384-Stain-40 • Dispense volume: 40 µL
②	✓	Examine the Stain 2 Tray. • If bubbles are present, puncture them with a pipette tip. • If droplets of liquid splashed onto the well dividers, use a laboratory tissue and gently dab the droplet on top of the tray divider to remove the droplet without disturbing the liquid inside the tray.
③	ⓘ	• Place a stain tray cover on top of each Stain 2 stain tray. Continue to "Stain tray dispense—Stabilization Master Mix" • Protect the trays from light if not immediately loading onto the GeneTitan™ MC Instrument. • Perform the appropriate Multidrop™ procedure: – Flush between batches. See page 55. – Shutdown. See page 56.

Stain tray dispense—Stabilization Master Mix		
Plates 1 2		
		
IMPORTANT! Deionze the trays with ion blowers or GeneTitan™ ZeroStat AntiStatic Gun before dispensing the reagents. **IMPORTANT!** Ensure that the stain trays labeled “Stbl” are used for the Multidrop™ Combi dispensing the Stabilization Master Mix.		
①		Dispense Stabilization Master Mix • Method: 384-Stain-40 • Dispense volume: 40 µL
②	✓	Examine the Stabilization Tray. • If bubbles are present, puncture them with a pipette tip. • If droplets of liquid splashed onto the well dividers, use a laboratory tissue and gently dab the droplet on top of the tray divider to remove the droplet without disturbing the liquid inside the tray.
③	ⓘ	• Place a stain tray cover on top of each Stabilization stain tray. • Perform the appropriate Multidrop™ procedure: – Flush between batches. See page 55. – Shutdown. See page 56.

5 Process array plates with the GeneTitan™ Multi-Channel (MC) Instrument

- Create and upload a GeneTitan™ Array Plate Registration file 129
- Run Wash-Scan 130
- Continue the scan workflow 138
- Shut down the GeneTitan™ MC Instrument 139

IMPORTANT! For optimal GeneTitan™ MC Instrument performance, ensure that the maximum relative humidity is 80% for temperatures up to 75.2°F (24°C), with a minimum humidity of 30 ±7% relative humidity. Operating outside the working environment specifications leads to higher static levels, and results in the evaporation of reagents from stain trays.

IMPORTANT! Review Appendix A, "Recommended techniques for GeneTitan™ MC Instrument operation" for details on array processing setup options and consumable handling.

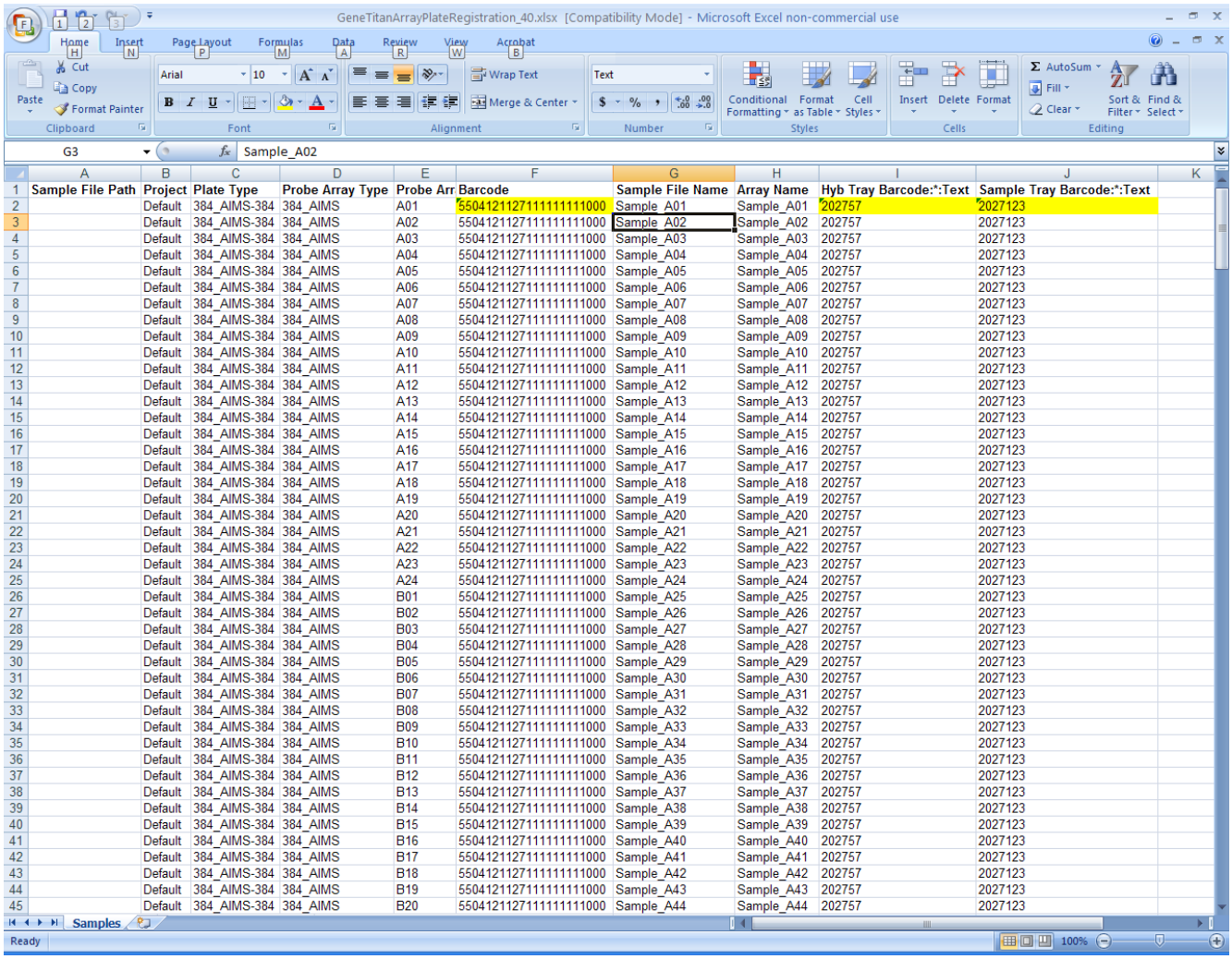
Create and upload a GeneTitan™ Array Plate Registration file

A batch registration file must be created and uploaded with GeneChip™ Command Console™ (GCC) software. This file contains information critical for data file generation during scanning, and tracking the experimental results for each sample loaded onto an array plate. This file can be created at any time before loading the array plate and hybridization tray onto the GeneTitan™ MC Instrument.

Note: When creating the GeneTitan™ Array Plate Registration file, you can scan the barcode of the hybridization tray to implement sample traceability. If you do not upload the sample file names before scanning the array plate barcode, the software assigns names to the samples.

The array plate barcode is scanned when you are ready to load the array plate and samples onto the GeneTitan™ MC Instrument for processing.

1. If you have already created a batch registration file but have not yet uploaded the file to GCC, open the file, then go to step 6.
2. From the **Launcher** window, open **GCC Portal** ▸ **Samples** ▸ **GeneTitan™ Array Plate Registration**.
3. In the GeneTitan™ Array Plate Registration window, select the **GeneTitan Array Plate Type** to be processed from the dropdown list.
4. Click **Download**.
5. In the **Samples** tab of the GeneTitan™ Array Plate Registration file, enter a unique name for each sample (Sample File Name) and any additional information. Additional information on the GeneTitan™ Array Plate Registration file is in the *GeneChip™ Command Console™ User Guide* (Pub. No. 702569)
6. Scan the array plate barcode into the yellow **Barcode** field, column **F**. See Figure 9.
7. Scan the barcode of the hybridization tray if the array plate registration file template includes a column for the hybridization tray barcode.
8. Save the file.
   By default, the file is saved in the **Applied Biosystems Download** folder.
9. Return to the GCC Portal GeneTitan™ Array Plate Registration page.
   a. Click **Browse**, navigate to the array plate registration file, then click **Open**.
   b. Under **Step 3**, click **Upload**, wait for the information to load, then click **Save** found at the bottom of the next window that is displayed.



GeneTitanArrayPlateRegistration_40.xlsx [Compatibility Mode] - Microsoft Excel non-commercial use

G3 | Sample_A02

Sample File Path	Project	Plate Type	Probe Array Type	Probe Arr	Barcode	Sample File Name	Array Name	Hyb Tray Barcode:*:Text	Sample Tray Barcode:*:Text
	Default	384_AIMS-384	384_AIMS	A01	55041211271111111111000	Sample_A01	Sample_A01	202757	2027123
	Default	384_AIMS-384	384_AIMS	A02	55041211271111111111000	Sample_A02	Sample_A02	202757	2027123
	Default	384_AIMS-384	384_AIMS	A03	55041211271111111111000	Sample_A03	Sample_A03	202757	2027123
	Default	384_AIMS-384	384_AIMS	A04	55041211271111111111000	Sample_A04	Sample_A04	202757	2027123
	Default	384_AIMS-384	384_AIMS	A05	55041211271111111111000	Sample_A05	Sample_A05	202757	2027123
	Default	384_AIMS-384	384_AIMS	A06	55041211271111111111000	Sample_A06	Sample_A06	202757	2027123
	Default	384_AIMS-384	384_AIMS	A07	55041211271111111111000	Sample_A07	Sample_A07	202757	2027123
	Default	384_AIMS-384	384_AIMS	A08	55041211271111111111000	Sample_A08	Sample_A08	202757	2027123
	Default	384_AIMS-384	384_AIMS	A09	55041211271111111111000	Sample_A09	Sample_A09	202757	2027123
	Default	384_AIMS-384	384_AIMS	A10	55041211271111111111000	Sample_A10	Sample_A10	202757	2027123
	Default	384_AIMS-384	384_AIMS	A11	55041211271111111111000	Sample_A11	Sample_A11	202757	2027123
	Default	384_AIMS-384	384_AIMS	A12	55041211271111111111000	Sample_A12	Sample_A12	202757	2027123
	Default	384_AIMS-384	384_AIMS	A13	55041211271111111111000	Sample_A13	Sample_A13	202757	2027123
	Default	384_AIMS-384	384_AIMS	A14	55041211271111111111000	Sample_A14	Sample_A14	202757	2027123
	Default	384_AIMS-384	384_AIMS	A15	55041211271111111111000	Sample_A15	Sample_A15	202757	2027123
	Default	384_AIMS-384	384_AIMS	A16	55041211271111111111000	Sample_A16	Sample_A16	202757	2027123
	Default	384_AIMS-384	384_AIMS	A17	55041211271111111111000	Sample_A17	Sample_A17	202757	2027123
	Default	384_AIMS-384	384_AIMS	A18	55041211271111111111000	Sample_A18	Sample_A18	202757	2027123
	Default	384_AIMS-384	384_AIMS	A19	55041211271111111111000	Sample_A19	Sample_A19	202757	2027123
	Default	384_AIMS-384	384_AIMS	A20	55041211271111111111000	Sample_A20	Sample_A20	202757	2027123
	Default	384_AIMS-384	384_AIMS	A21	55041211271111111111000	Sample_A21	Sample_A21	202757	2027123
	Default	384_AIMS-384	384_AIMS	A22	55041211271111111111000	Sample_A22	Sample_A22	202757	2027123
	Default	384_AIMS-384	384_AIMS	A23	55041211271111111111000	Sample_A23	Sample_A23	202757	2027123
	Default	384_AIMS-384	384_AIMS	A24	55041211271111111111000	Sample_A24	Sample_A24	202757	2027123
	Default	384_AIMS-384	384_AIMS	B01	55041211271111111111000	Sample_A25	Sample_A25	202757	2027123
	Default	384_AIMS-384	384_AIMS	B02	55041211271111111111000	Sample_A26	Sample_A26	202757	2027123
	Default	384_AIMS-384	384_AIMS	B03	55041211271111111111000	Sample_A27	Sample_A27	202757	2027123
	Default	384_AIMS-384	384_AIMS	B04	55041211271111111111000	Sample_A28	Sample_A28	202757	2027123
	Default	384_AIMS-384	384_AIMS	B05	55041211271111111111000	Sample_A29	Sample_A29	202757	2027123
	Default	384_AIMS-384	384_AIMS	B06	55041211271111111111000	Sample_A30	Sample_A30	202757	2027123
	Default	384_AIMS-384	384_AIMS	B07	55041211271111111111000	Sample_A31	Sample_A31	202757	2027123
	Default	384_AIMS-384	384_AIMS	B08	55041211271111111111000	Sample_A32	Sample_A32	202757	2027123
	Default	384_AIMS-384	384_AIMS	B09	55041211271111111111000	Sample_A33	Sample_A33	202757	2027123
	Default	384_AIMS-384	384_AIMS	B10	55041211271111111111000	Sample_A34	Sample_A34	202757	2027123
	Default	384_AIMS-384	384_AIMS	B11	55041211271111111111000	Sample_A35	Sample_A35	202757	2027123
	Default	384_AIMS-384	384_AIMS	B12	55041211271111111111000	Sample_A36	Sample_A36	202757	2027123
	Default	384_AIMS-384	384_AIMS	B13	55041211271111111111000	Sample_A37	Sample_A37	202757	2027123
	Default	384_AIMS-384	384_AIMS	B14	55041211271111111111000	Sample_A38	Sample_A38	202757	2027123
	Default	384_AIMS-384	384_AIMS	B15	55041211271111111111000	Sample_A39	Sample_A39	202757	2027123
	Default	384_AIMS-384	384_AIMS	B16	55041211271111111111000	Sample_A40	Sample_A40	202757	2027123
	Default	384_AIMS-384	384_AIMS	B17	55041211271111111111000	Sample_A41	Sample_A41	202757	2027123
	Default	384_AIMS-384	384_AIMS	B18	55041211271111111111000	Sample_A42	Sample_A42	202757	2027123
	Default	384_AIMS-384	384_AIMS	B19	55041211271111111111000	Sample_A43	Sample_A43	202757	2027123
	Default	384_AIMS-384	384_AIMS	B20	55041211271111111111000	Sample_A44	Sample_A44	202757	2027123

Samples | Ready | 100%

Figure 9 Example of a GeneTitan™ Array Plate Registration file.

Run Wash-Scan

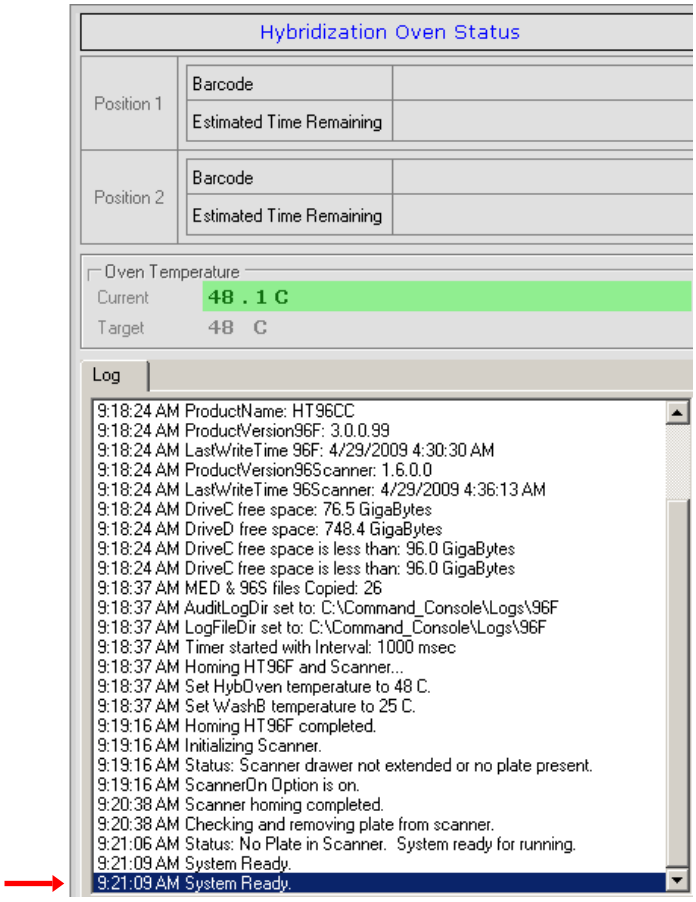
The following reagents from the Axiom™ Propel XPRES Reagent Kit, 2x384HT are required for this procedure.

- Axiom™ Wash Buffer A, Part No. 901446
- Axiom™ Wash Buffer B, Part No. 901447
- Axiom™ Water, Part No. 901578

1. Verify that the correct hybridization time for the “Array Plate/Hyb Tray Stack” has been achieved. Keep the “Array Plate/Hyb Tray Stack” in the off-line oven until you are ready to place it into the GeneTitan™ MC Instrument.

2. From the **GCC Launcher** and select **GCC GeneTitan Control**.

   The "System Ready" is indicated at the bottom of the status pane.



3. When the system has initialized and “System Ready” status appears, click to select the **System Setup** tab.

4. Select **Wash-Scan** from the **Setup Option** dropdown list.

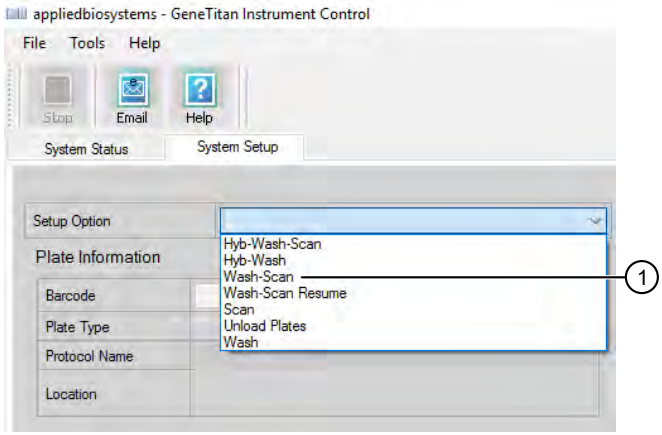


Figure 10 Wash–Scan setup option.

5. Enter the barcode of the array plate to be processed.

   During this step, the array plate must remain in the hybridization oven. The barcode can be retrieved from the registration file that was created before the start of the off-line hybridization.

6. Select a protocol from the **Protocol Name** dropdown list, then click **Next**.

   IMPORTANT! For executing the accelerated wash protocol, ensure that the selected **Protocol Name** has the format: "XXXXXX_fastwash.protocol", where "XXXXXX" is the array part number. Failure to select the correct protocol can result in compromised assay performance, and a delay with the timing of the array processing workflow.

7. Refill the GeneTitan™ bottles with the following reagents.

   Note: Invert the Axiom™ Wash Buffer A and Axiom™ Wash Buffer B bottles 2–3 times to mix before filling the GeneTitan™ bottles.

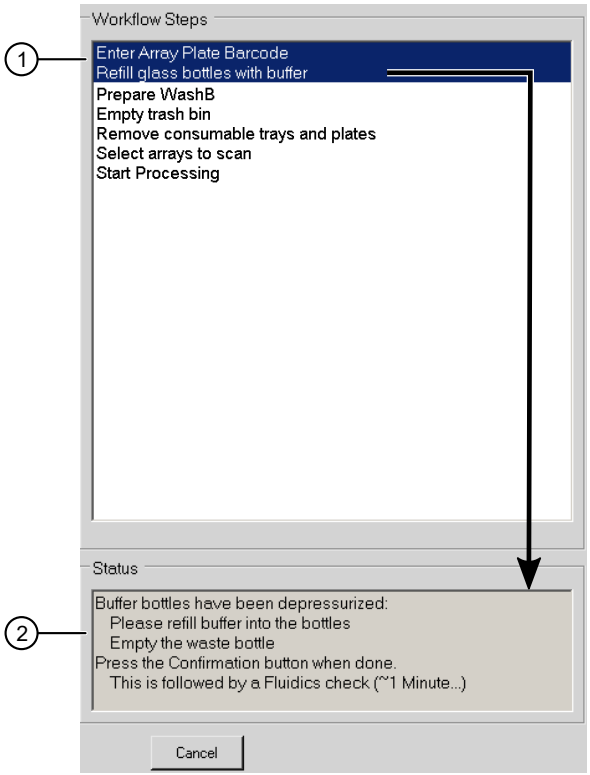
   - Wash A: fill with Axiom™ Wash Buffer A—keep at 2 L full.
   - Wash B: fill with Axiom™ Wash Buffer B—use all 600 mL of Axiom™ Wash Buffer B.
   - Rinse: fill with Axiom™ Water—keep at 1 L full.

   IMPORTANT! Always ensure that the GeneTitan™ bottles labeled Wash A, Wash B, and Rinse are above the 50% mark when setting up the system to process an array plate.

   We strongly recommend refilling these bottles every time you are prompted to do so.
   If the volume in any of the GeneTitan™ bottles become too low during a run, an error message is displayed. However, if you wait to fill the bottle at the time of the error message, the instrument cannot complete the step that was in progress.

8. Empty the GeneTitan™ bottle labeled Waste.

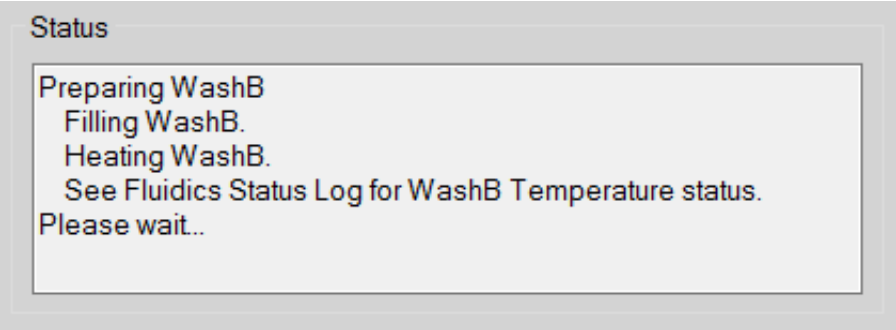
9. Press the blue confirmation button on GeneTitan™ MC Instrument to continue.
   A fluidics check is run (~1 minute).



① **Workflow Step**
② Specific instructions for the current workflow step appear in the **Status** pane.

10. Prepare the Axiom™ Wash Buffer B for the **Wash-Scan** workflow.

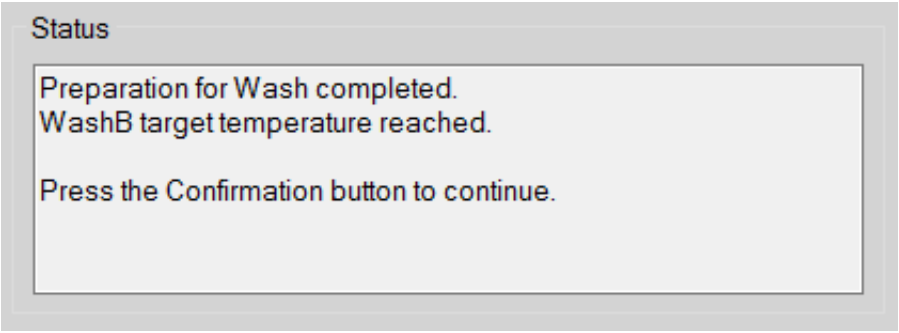
    Warming up the Axiom™ Wash Buffer B takes 25–30 minutes. Therefore, this step must be completed at least 30 minutes before to the completion of the 24-hour hybridization time.



Note: During this waiting time, we recommend that you prepare the reagents for "Stage 7: Preparing ligation, stain, stabilization reagent trays, and scan tray for the GeneTitan™ MC Instrument".

WARNING! *Do not* load reagents onto the GeneTitan™ MC Instrument until you are prompted by GeneTitan™ Instrument Control Software.

11. After the temperature has been reached, you are prompted to press the Confirmation button on GeneTitan™ MC Instrument to continue (blinking light blue button found at the top center of the GeneTitan™ Instrument).



12. Empty the trash bin.
    a. Open the trash bin and empty.
    b. If already empty, the trash bin remains locked and the **Status** pane reads "Trash bin is empty".
    c. Press the blue confirmation button to continue.
13. Remove consumable trays and plates.
    a. Remove used trays and plates when drawers open.
    b. If there are no consumables to remove, the **Status** window reads "Drawers are empty".
    c. Press the blue confirmation button on the GeneTitan™ Instrument to continue.

14. Load consumable trays and plates. Follow the prompts in the **Status** window to load the reagent trays (scan, stains, ligation, and stabilization).
    a. When drawer 2 opens:
        - Left side: Scan tray with cover. Remove the blue scan tray protective base from the scan tray immediately before loading. Do not load the blue scan tray protective base.
        - When complete, press the blue confirmation button on the GeneTitan™ Instrument to continue.

IMPORTANT! Before installing the consumables into the instrument, ensure that the fingers are retracted. Do not lay the consumables on top of the drawer fingers.

You must place the trays into the instrument drawers when a drawer is fully extended by the instrument. The fingers are retracted when the drawer is open, and are extended when the drawer is closed in order to restrain the consumable. When the drawer is open and the fingers are not retracted, the instrument is not functioning correctly. Notify a Field Service Engineer if the fingers do not retract automatically.

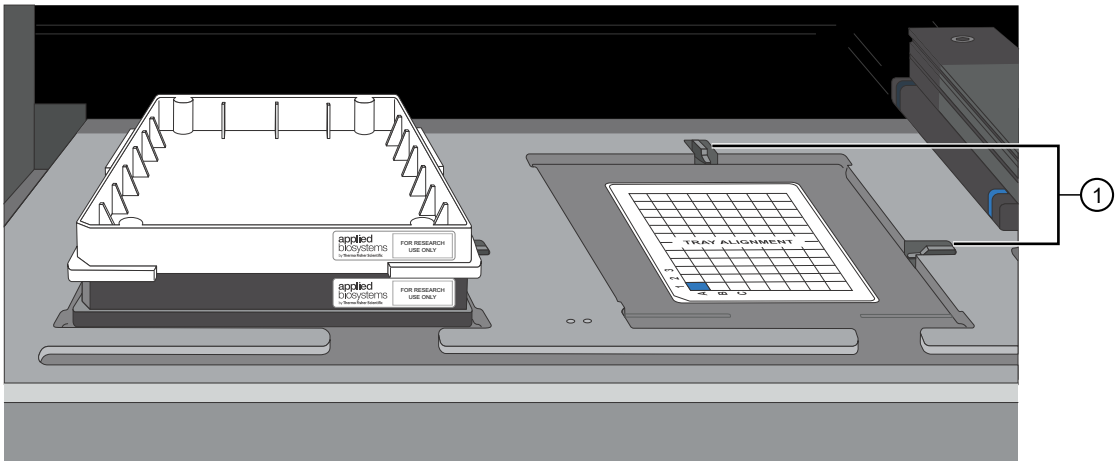


Figure 11 Drawer 2, left side: Scan tray with cover.

(1) Locations of drawer tabs, or "fingers".

b. When drawer 3 opens:

- Left side: Stain 1 tray with cover.
- Right side: Ligation tray with cover.
- Press the blue confirmation button on the GeneTitan™ Instrument to continue.

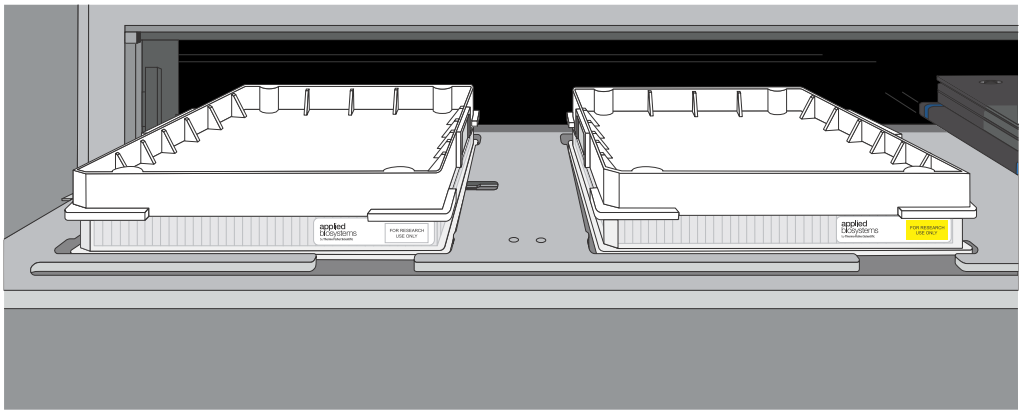


Figure 12 Drawer 3, left side: Stain 1 tray with cover. Drawer 3, right side: Ligation tray with cover.

c. When drawer 4 opens:

- Left side: Stain 2 tray with cover.
- Right side: Stabilization tray with cover.
- Press the blue confirmation button on the GeneTitan™ Instrument to continue.

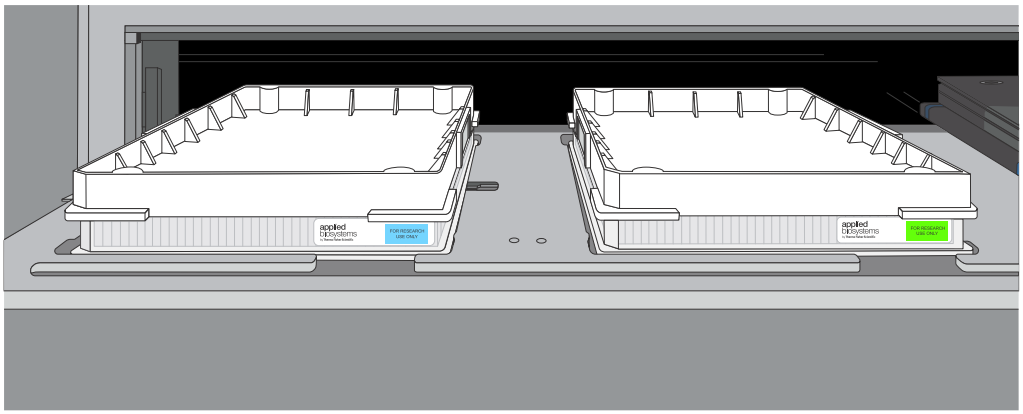


Figure 13 Drawer 4, left side: Stain 2 tray with cover. Drawer 4 right side: Stabilization tray with cover.

d. When drawer 5 opens:
   - Left side: Stain 1 tray with cover.
   - Press the blue confirmation button on the GeneTitan™ Instrument to continue.

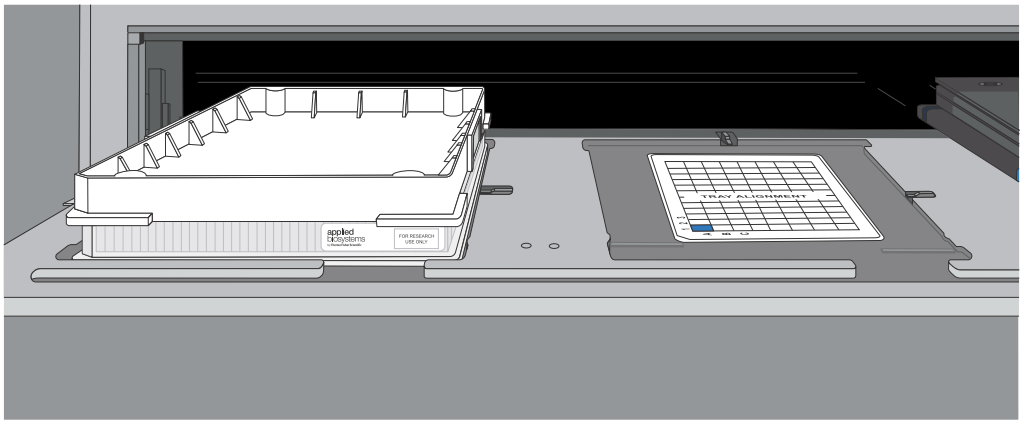


Figure 14 Drawer 5, left side: Stain 1 tray with cover.

e. The blue base and array plate/hybridization tray stack are loaded in drawer 6.

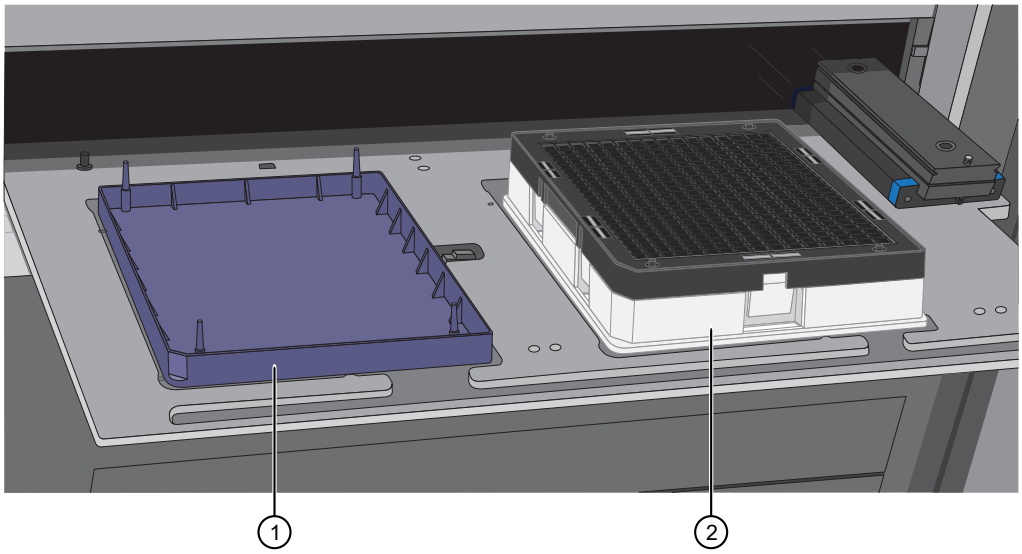


Figure 15 Blue protective base and the array plate/hybridization tray stack properly loaded into drawer 6.

(1) Blue protective base
(2) Array plate/hybridization tray stack

IMPORTANT! A blue protective base must be present on the left side of drawer 6 at all times when an array plate is being processed in the fluidics.

15. Select the arrays that you want to scan.

16. Start processing.

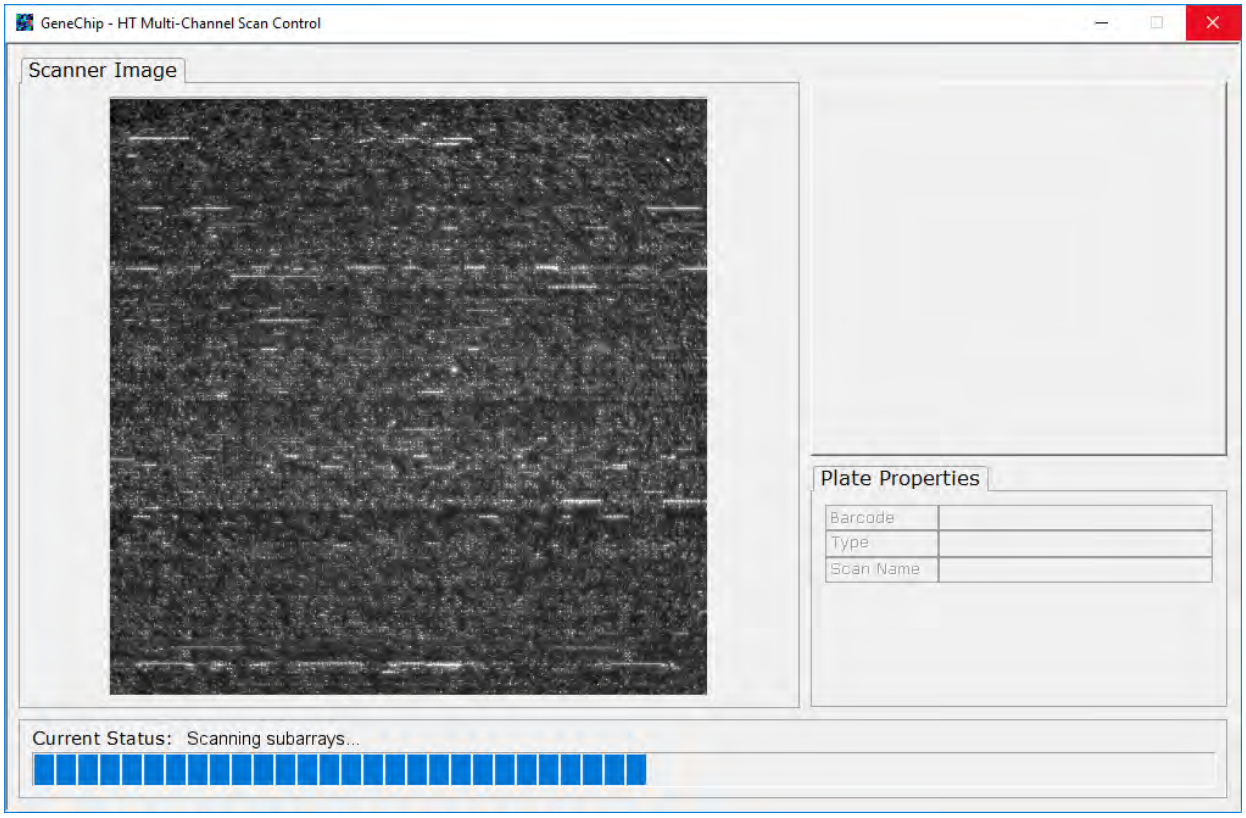
Continue the scan workflow

After a plate has completed the fluidics stage of the workflow, the GeneTitan™ Instrument moves the plate to the imaging device.

When the scanning process starts, a Scan Control window displaying the scan image appears. This window must remain open while the array plate is being scanned.

CAUTION! The **Scan Control** window must remain open while the array plate is being scanned. If the window is closed, the scanning process halts. If needed, this window can be minimized without interference to the imaging.

Do not manually, or through the GCC transfer utility, move data that are associated with the plate that is being processed/scanned. Transferring data dramatically slows scanning and can cause the computer to freeze.



Queue another plate for Wash-Scan

This technique can be used in high throughput workflows to maximize the capacity of the GeneTitan™ MC Instrument.

- Load the second plate to be washed and scanned immediately after the first plate has finished washing (~4 hours).
- Load the third and subsequent plates for **Wash-Scan** at a ~4.75-hour interval, or after the plate being processed in the scanner has completed.

Shut down the GeneTitan™ MC Instrument

This procedure assumes that all the array plates that are loaded onto the GeneTitan™ MC Instrument have been processed.

1. From the **System Setup** window, open the **Setup Options** dropdown list, then select **Unload Plates**.
2. Unload all the consumables as prompted.
3. Power off the GeneTitan™ MC Instrument by opening **Tools** ▸ **Shutdown**.
4. Exit the GCC software if it does not close automatically.

Note: If the instrument is processing an array plate, the software does not allow you to shut down the system.

6 High throughput with the Axiom™ Propel XPRES 384HT Workflow

- Overview 140
- Axiom™ Propel XPRES 384HT Workflow schedule flexibility 140
- Considerations for customization of the workflows 141
- 48-hour turnaround time schedule with workflow A 143
- 32 plates per week (~640,000 samples yearly) using workflow D 145

Overview

The Axiom™ Propel XPRES 384HT Workflow is uniquely designed to accommodate the growing demand for processing a higher number of Axiom™ 384HT array plates. The Axiom™ Propel XPRES Reagent Kit features the new Axiom™ XPRES Amp Enzyme and Axiom™ Fast Wash reagents reducing the total time required to obtain genotypes.

Axiom™ Propel XPRES Reagent Kit enables the end user to execute various workflows depending on their needs for sample-to-data turnaround time or yearly sample throughput. With this flexibility you can generate genotypes in 48 hours or process up to one million samples yearly. The workflows features the Thermo Scientific™ Multidrop™ Combi Reagent Dispenser and the Applied Biosystems™ GeneTitan™ MC Instrument, along with other common laboratory equipment, to achieve the tasks required for multi-array plate processing.

This chapter provides example equipment and labor schedules for 2 different production levels of Axiom™ 384HT array format plates.

IMPORTANT! The successful execution of this workflow requires experienced operators and careful timing.

The requirements and schedules presented in this chapter are intended as general guidelines for developing an individualized plan to fit your needs. Contact your Thermo Fisher Scientific FAS representative for help with planning the scale-up of the Axiom™ Propel XPRES 384HT Workflow.

Axiom™ Propel XPRES 384HT Workflow schedule flexibility

The Axiom™ Propel XPRES 384HT Workflow provides ranges of incubation times for the amplification and precipitation stages allowing customizable workflow schedules.

- Amplification incubation time with the Axiom™ XPRES Amp Enzyme: 4–24 hours.
- Precipitation incubation time: 3–24 hours.

Workflow type	Amplification incubation time	Precipitation incubation time	Hybridization-ready target completed	When array results are available
A	4 hours	3 hours	Day 1[1]	Beginning of day 3 (after 48 hours)
B	4 hours	Overnight to 24 hours	Day 2	Beginning of day 4
C	24 hours	3 hours	Day 2	Beginning of day 4
D[2]	24 hours	Overnight to 24 hours	Day 3	Beginning of day 5

[1] Facilitates a 48-hour turnaround time. Requires 2 shifts.
[2] Workflow D performs well for high throughput environments.

Considerations for customization of the workflows

GeneTitan™ MC Instrument throughput

The wash-stain, and scan steps of the Axiom™ Propel XPRES 384HT Workflow are completed with the GeneTitan™ Multi-Channel (MC) Instrument, with the following assumptions:

- Wash-stain time = 4 hours per plate
- Scan time = 4.75 hours per plate

Number of GeneTitan™ MC Instruments	Number of shifts[1]	Plates/day	Plates/week[2]	Samples/year
2	1	4	16	~320,000
	2	8	32	~640,000

[1] Nine hours per shift.
[2] Five working days per week.

Target preparation with the Axiom™ Propel XPRES 384HT Workflow

Use the following processing times for each of the Axiom™ Propel XPRES 384HT Workflow steps to estimate the number of instruments and man hours required to achieve a given throughput.

Table 32 Task duration for target preparation of 2x384HT array plates.

Task	Allotted time[1]	Unattended time
Amplification	1 hour	4–24 hours
Fragmentation and precipitation	1 hour, 15 minutes	3–24 hours
Centrifugation and drying	1 hour, 30 minutes	1 hour
Resuspension of hybridization-ready targets	45 minutes	N/A

Table 32 Task duration for target preparation of 2x384HT array plates. *(continued)*

Task	Allotted time[1]	Unattended time
Merge	30 minutes	N/A
In-process quality control	30 minutes	N/A
Hybridization transfer and hybridization	30 minutes	23.5–24 hours
GTMC preparation	1 hour	N/A
Washing on GTMC	N/A	4 hours
Scanning on GTMC	N/A	4 hours, 45 minutes

[1] The allotted time is the the amount of time provided to complete each task in the assay workflow schedule. However, the actual time needed to run each task is less than the allotted time.

Practices to promote effiency

When used, the following practices increase the efficiency of the workflow when running in an ultrahigh throughput environment.

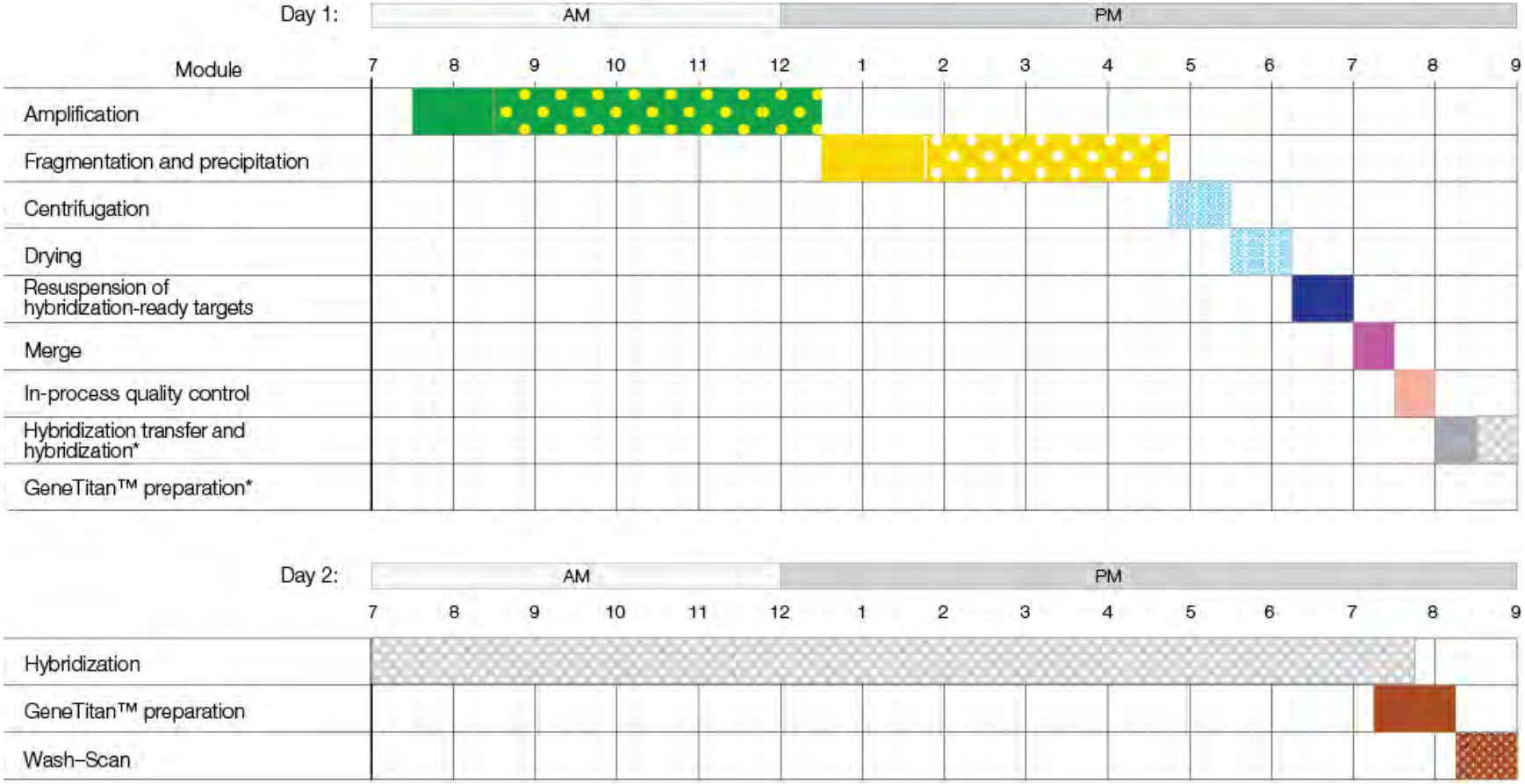
- Keep all required instruments in close proximity to their use in the laboratory, and position the instruments to enable a smooth workflow.
- Thaw the necessary reagents the day before use. See the appropriate stage "Reagent handling" table in the target preparation chapter. See Chapter 4, “Target preparation with Multidrop™ Combi Reagent Dispensers for two 384HT array format plates”.
- Prepare the master mixes ahead of time. For example, advanced preparation of some GeneTitan™ reagents reduces the morning start-up time for “Stage 7: Preparing ligation, stain, stabilization reagent trays, and scan tray for the GeneTitan™ MC Instrument” on page 112.
- Utilize the Wash B warm-up time (25–30 minutes) on the GeneTitan™ MC Instrument to perform tasks such as setting up the Multidrop™ Combi instruments to dispense the stain trays and scan trays.
- Maximize the GeneTitan™ MC Instrument throughput by loading subsequent plates as soon as the instrument is available. For further information, review the following procedure. (See “Queue a second plate for scanning” on page 165.)

48-hour turnaround time schedule with workflow A

- Requires 2 shifts of operation.
- Two plates of 384 hybridization-ready targets generated in a ~14-hour day.
- Using 2 GeneTitan™ MC Instruments, it is possible to hybridize 2 array plates on the same day to generate cel files at the beginning of day 3.
- Amplification incubation time = 4 hours
- Precipitation incubation time = 3 hours

Note: The following schedule is intended as a general guideline. Contact your Thermo Fisher Scientific FAS representative for help with planning an individualized scale-up of the Axiom™ Propel XPRES 384HT Workflow.

48-hour turnaround time schedule, workflow A



Patterned blocks are tasks that do not require a FTE to be present the entire time.

Note: CEL files are available at ~5:00 a.m. on day 3.

32 plates per week (~640,000 samples yearly) using workflow D

Workflow D with 2 shifts and 2 GeneTitan™ MC Instruments.

- Amplification incubation: overnight (23—24 hours)
- Precipitation incubation: overnight (23—24 hours)
- 2 shifts (total = 18 hours)
- 7 FTEs
- 2 GeneTitan™ MC Instruments

Note: The following schedules are intended as a general guideline. Contact your Thermo Fisher Scientific FAS representative for help with planning an individualized scale-up of the Axiom™ Propel XPRES 384HT Workflow.

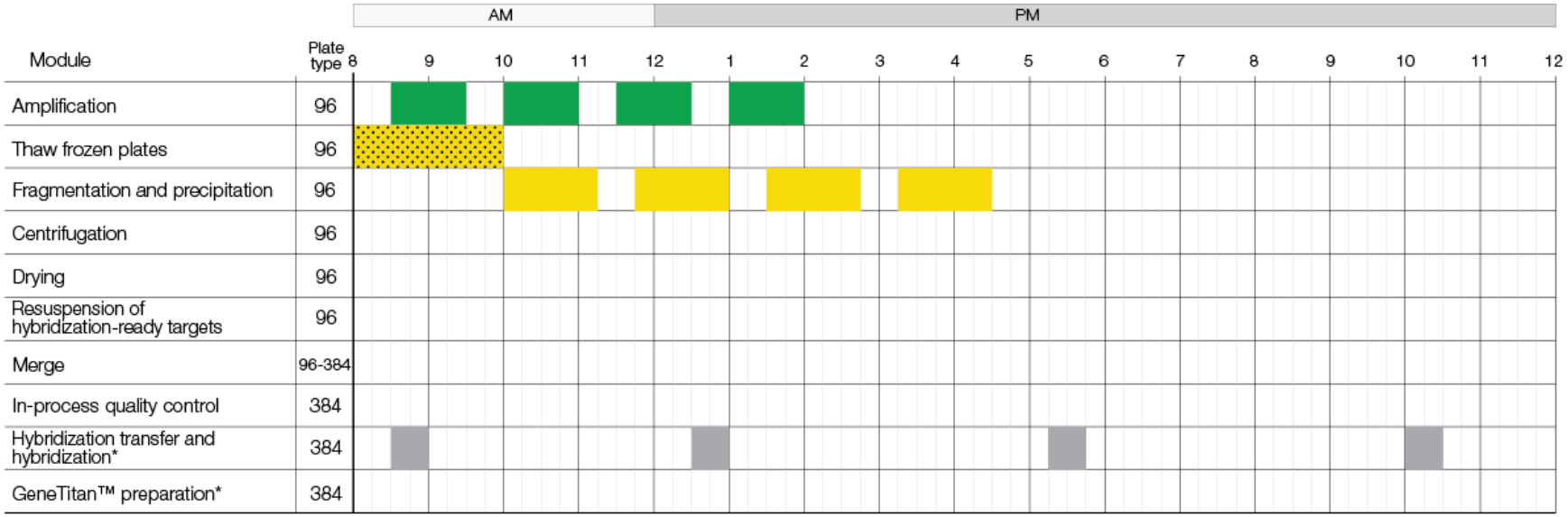
Instruments required

Instruments	32 plates/week, 2 shifts
GeneTitan™ MC Instrument	2
Multidrop™ Combi Reagent Dispenser	12
ALPS™ 3000 Automated Microplate Heat Sealer	3
Thermo Scientific™ Compact Digital Microplate Shaker	4
VWR Signature™ High-Speed Microplate Shaker	1
Sorvall™ Legend™ XTR Centrifuge, room temperature	3
Sorvall™ Legend™ XTR Centrifuge, refrigerated	2
Ovens (Heratherm™ and/or BINDER™ ovens)	5
Thermal cycler, ProFlex™ 2 x 384-well PCR System	1
INTEGRA Biosciences VIAFLO with 96-format dispense head	1
INTEGRA Biosciences VIAFLO with 384-format dispense head	1

Monday activities, 32 plates per week, workflow D

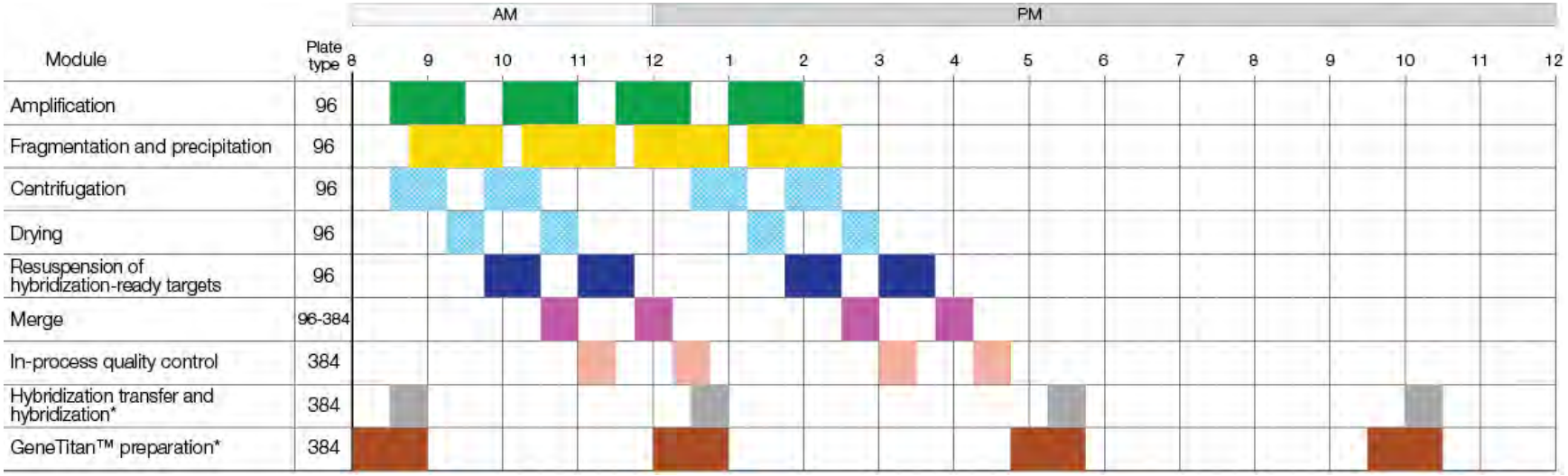
Note:

- Samples for fragmentation are from frozen Amplification Plates made the previous week. The frozen plates require 2-hour thawing time.
- Hybridization-ready targets are made the previous week.



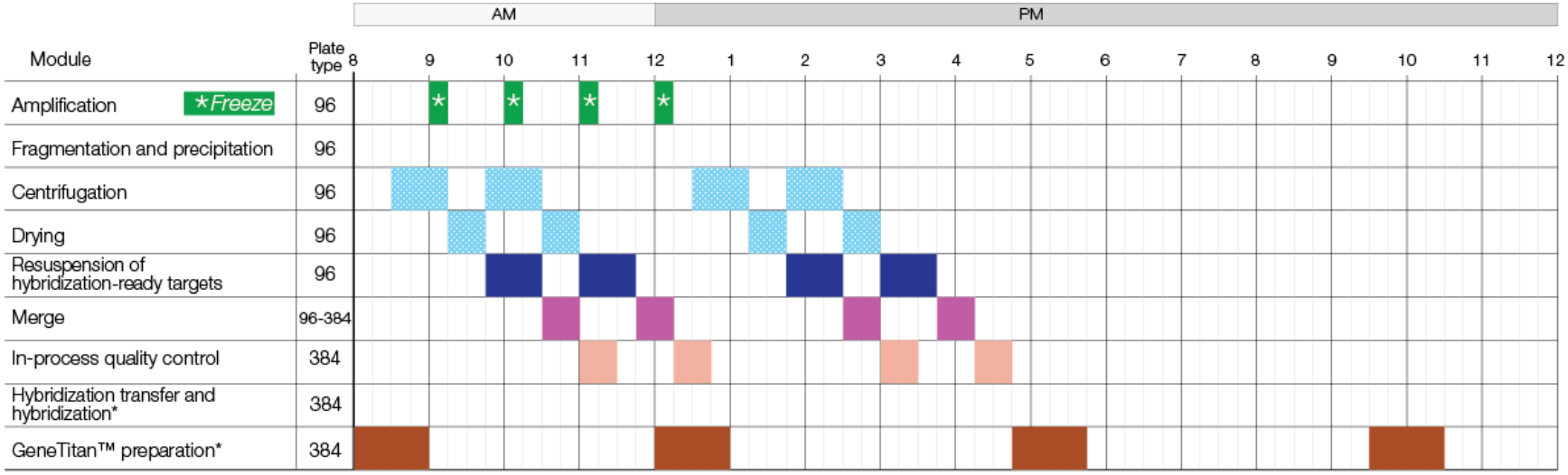
- 96: One batch of eight 96-deepwell plates to process.
- 96-384: One batch of eight 96-deepwell merge into two 384-format plates to process.
- 384: Two 384-format plates to process.
- Patterned blocks are tasks that do not require a FTE.
- (*): Two plates with 2 GeneTitan™ MC Instruments.

Tuesday—Thursday activities, 32 plates per week, workflow D



- 96: One batch of eight 96-deepwell plates to process.
- 96-384: One batch of eight 96-deepwell merge into two 384-format plates to process.
- 384: Two 384-format plates to process.
- Patterned blocks are tasks that do not require a FTE to be present the entire time.
- (*): Two plates with 2 GeneTitan™ MC Instruments.

Friday activities, 32 plates per week, workflow D



- 96: One batch of eight 96-deepwell plates to process.
- 96-384: One batch of eight 96-deepwell merge into two 384-format plates to process.
- 384: Two 384-format plates to process.
- Patterned blocks are tasks that do not require a FTE to be present the entire time.
- (*): Two plates with 2 GeneTitan™ MC Instruments.

A

Recommended techniques for GeneTitan™ MC Instrument operation

- Array plate packaging 150
- Proper tray alignment and placement 150
- Stain trays and covers 154
- Label GeneTitan™ hybridization and reagent trays 155
- Guidelines for aliquoting reagents to GeneTitan™ trays 157
- Deionization of GeneTitan™ stain trays 158
- Best practice guidelines for GeneTitan™ reagent bottles 161
- Setup options for array plate processing 161
- Load an array plate and hybridization tray into the GeneTitan™ MC Instrument (for Hyb-Wash-Scan or Hyb-Wash) 166
- Load a second array plate and hybridization tray onto the GeneTitan™ MC Instrument 171
- When to abort a process 173
- Email notifications from the GeneTitan™ MC Instrument 175
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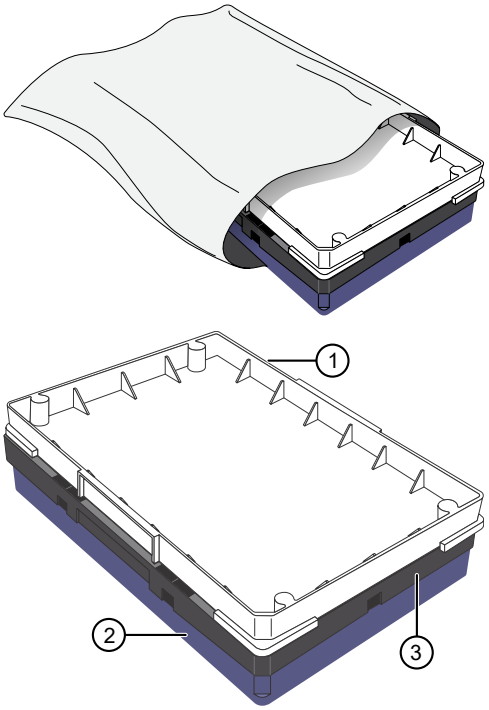
This chapter describes the recommended techniques and procedures to follow when using the GeneTitan™ MC Instrument for the fluidics processing and array scanning steps of the Axiom™ Propel XPRES 384HT Workflow. Being familiar with these techniques helps to ensure the success of the assay. Detailed safety information and instruction on using the GeneTitan™ MC Instrument is in the *GeneTitan™ Multi-Channel Instrument User Guide* and the *GeneChip™ Command Console™ User Guide*.

Array plate packaging

Item	Part No.	Image	Details
Axiom™ 384HT Array Plate or custom Axiom™ 384HT myDesign™ Array Plate	All array plates have the Part No. 202091 etched on the plastic	 ① Shipping cover (to be discarded) ② Array plate protective base ③ Array plate	The array plate package includes the following: • White plastic cover: The function of the white plastic cover for the array plate is to protect the array plate during transport. You can discard this after removing the array plate. • Array plate: The array plate must be protected at all times from damage or exposure to dust. The array plate must be in the blue array plate protective base at all times. • Protective base: The blue array plate protective base in the package must be used to protect the array plate from damage. • Desiccant pack: The desiccant pack can be discarded after the array plate is removed from the pouch.

Proper tray alignment and placement

Proper alignment and loading of plates, covers, and trays is critical when using the GeneTitan™ MC Instrument. Each plate, cover, and tray has one notched corner. The notched corner of plates, trays, covers, and bases must be in vertical alignment with each other and placed in position A1 per the Tray Alignment guide inside each GeneTitan™ MC Instrument drawer.

CAUTION! Be careful not to damage the consumables or bend the blue scan tray protective base cover posts or scan tray posts.

Note: Mark the notched corner of each plate, cover, and tray with permanent marker to help ensure proper alignment when loading onto the GeneTitan™ MC Instrument.

IMPORTANT! The drawer bed is *not* notched, so it is mechanically possible to load the tray in the wrong orientation. Be careful to avoid this mistake.

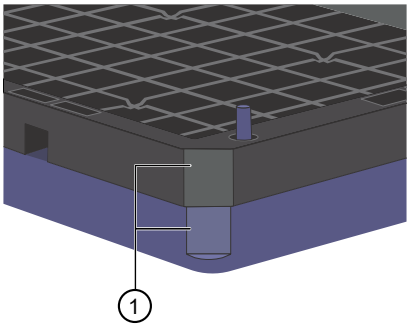


Figure 16 Notched corners aligned.

(1) The notched corner of array plate that is aligned with the notched corner of blue protective base.

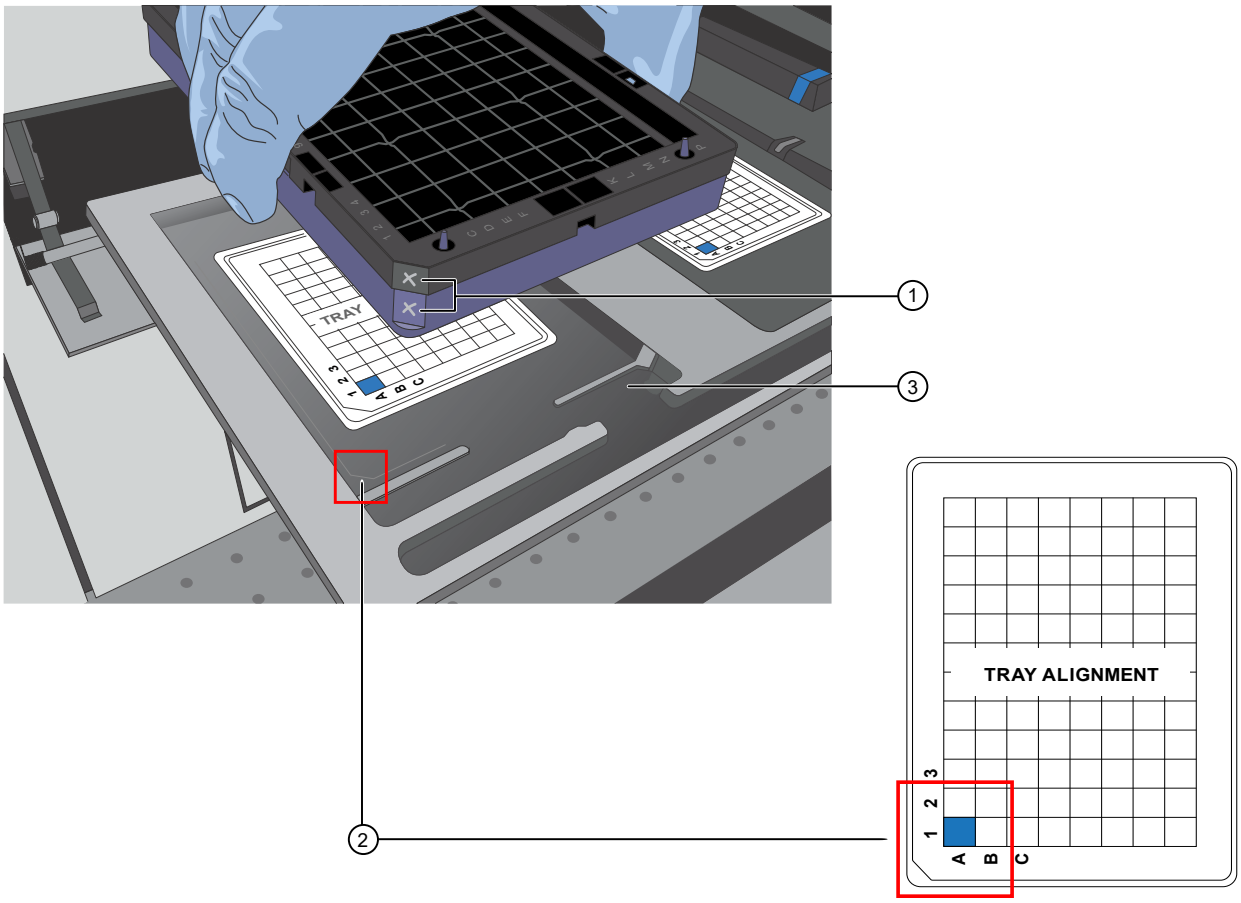


Figure 17 Notched corners marked and aligned with tray alignment guide.

(1) The notched corners of array plate and base that are marked with permanent marker.
(2) The notched corner of all plates, bases, and covers and must be seated in the front left corner of the drawer, as indicated in the Tray Alignment guide.
(3) Plates and trays must be seated in this groove.

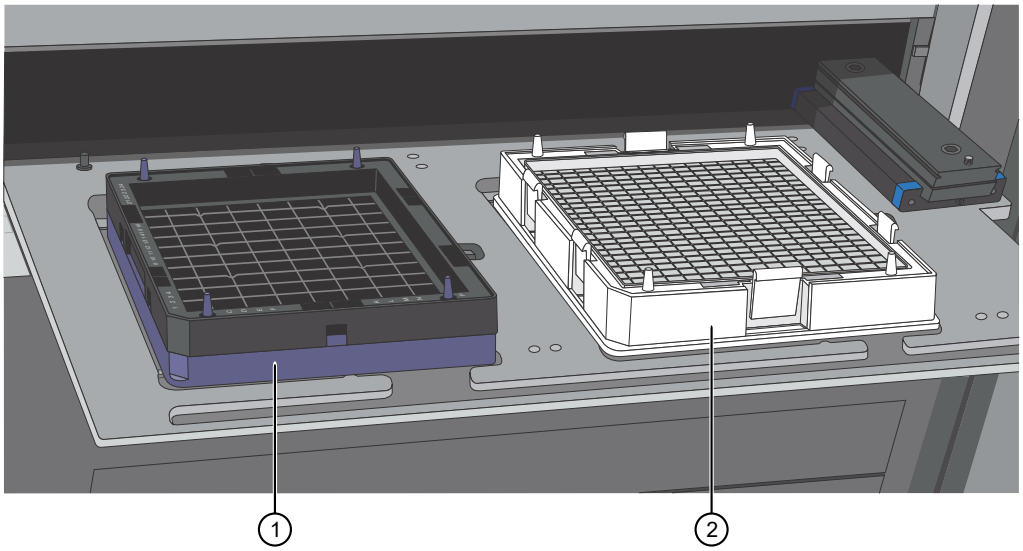


Figure 18 Array plate with blue protective base and the hybridization tray aligned properly loaded into drawer 6.

① Array plate on blue protective base
② Hybridization tray

Proper orientation of consumables

It is important that consumables be oriented properly when loaded into/onto the GeneTitan™ MC Instrument. The barcodes face into the instrument.

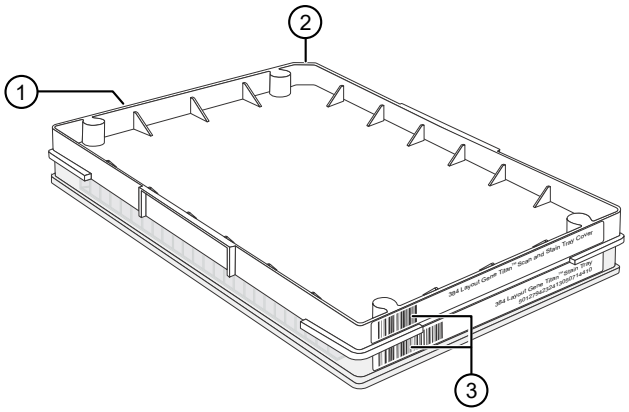


Figure 19 Example shows consumables that must be rotated and loaded on the drawer so that the barcodes face into the instrument.

① Front of instrument (facing you).
② Notched corners. The notched corners face out and left.
③ Barcodes. The barcodes face to the rear of the instrument where scanning by the internal barcode reader takes place.

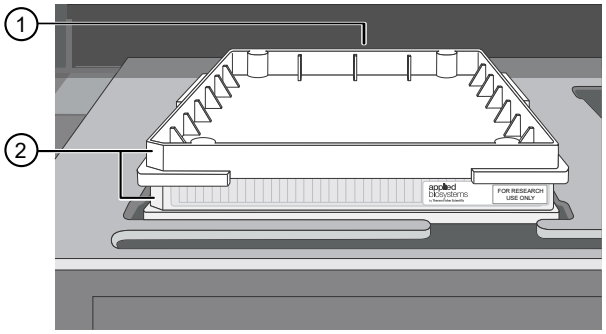


Figure 20 Example of properly loaded GeneTitan™ tray consumables.

A GeneTitan™ stain tray and the stain tray cover are shown in this example.

(1) Barcodes face the rear of the instrument.
(2) Notches face out and left. "For Research Use Only" faces out.

Drawer tabs in the GeneTitan™ MC Instrument

The GeneTitan™ MC Instrument drawers have tabs, or fingers, that restrain the consumable. The fingers retract when the drawer is open and extend when the drawer closes. When you load the plates or trays, ensure that the fingers are retracted and place trays onto the instrument drawers only after the drawer is fully extended. Ensure that the tray is not resting on the fingers. Notify your field service engineer if the fingers do not retract automatically.

IMPORTANT! Do not place the consumables on top of the drawer fingers—this position prevents the instrument from functioning correctly.

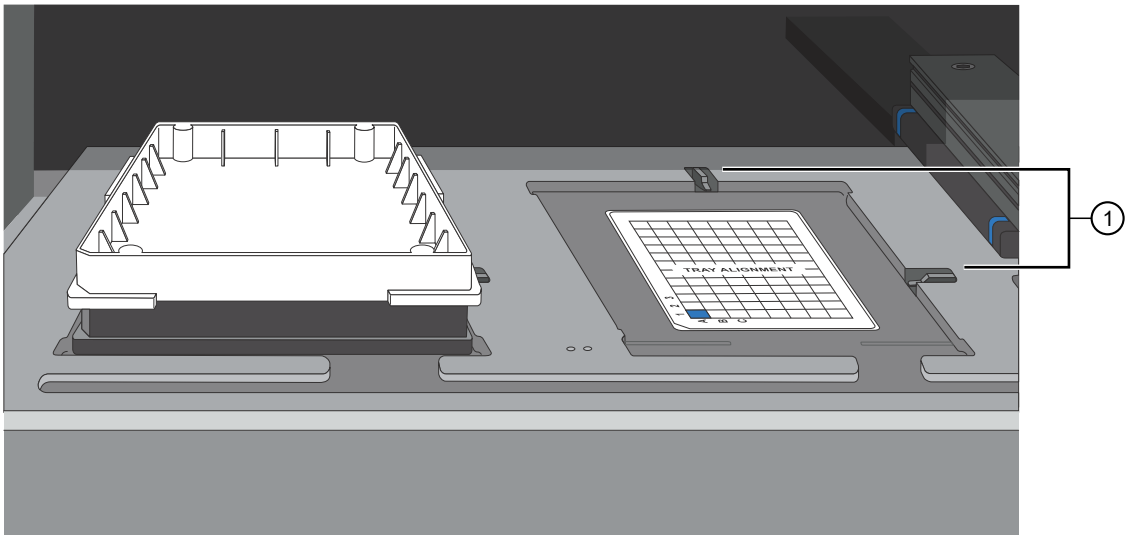


Figure 21 Location of drawer tabs, or fingers.

(1) Drawer tabs, or fingers, in the GeneTitan™ MC Instrument.

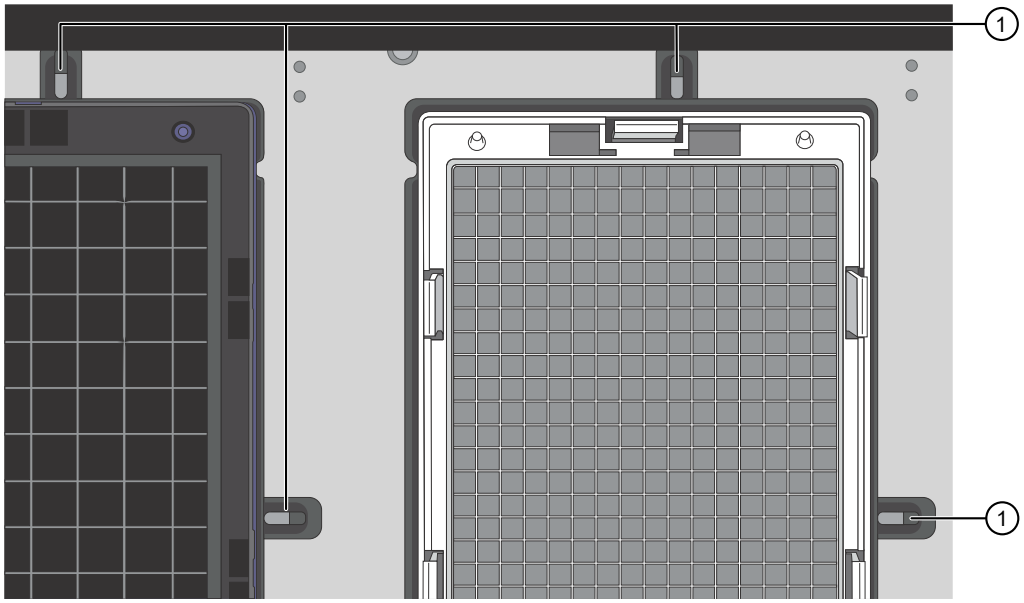


Figure 22 Place trays on the drawer when the tabs, or fingers, are retracted.

① Fingers retracted

Stain trays and covers

IMPORTANT! Always place the flat side of the cover against the stain tray.

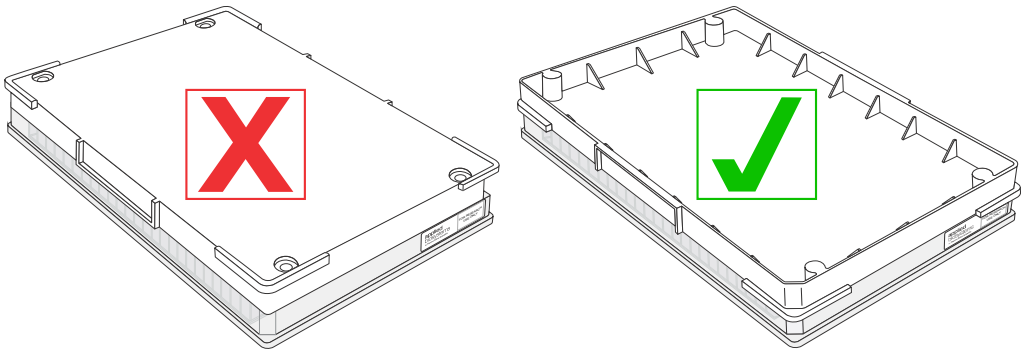


Figure 23 Placement of covers on trays.

Label GeneTitan™ hybridization and reagent trays

When preparing the hybridization and reagent trays to be loaded onto the GeneTitan™ MC Instrument, it is helpful to mark each tray in a way that identifies its contents.

IMPORTANT! It is critical that you write only on the proper locations of the proper sides of hybridization and stain trays. **Do not** write in any other location, because writing can interfere with sensors inside the GeneTitan™ MC Instrument and result in experiment failure. To ensure proper placement of lids onto stain trays, and trays onto the GeneTitan™ MC Instrument, you can also mark the notched corner of the trays and lids.

Label a GeneTitan™ 384 Layout Hybridization Tray

Label a GeneTitan™ 384 Layout Hybridization Tray on the front part of the short side of the tray, next to the notch at the left, as shown in the following image. The proper section for labeling is nearest to the notched corner, corresponding to the A1 through F1 wells.

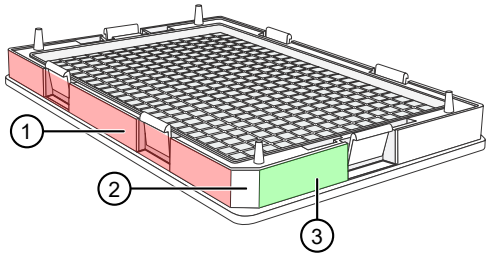


Figure 24 Correct area to label a GeneTitan™ 384 Layout Hybridization Tray.

1. Do not label hybridization tray on the long side.
2. Notched corner of the hybridization tray faces the front.
3. Label the hybridization tray here.

CAUTION! Writing on the wrong side of the hybridization tray can interfere with the operation of the sensors in the GeneTitan™ MC Instrument.

Label a GeneTitan™ 384 Layout Reagent Tray

You can label a GeneTitan™ 384 Layout Reagent Tray on the left side of the front of the tray as shown in the following image. The correct side is nearest to the notched corner, corresponding to the A1 through F1 wells.

IMPORTANT! This procedure is for noncolor-coded reagent trays. It is not necessary to label color-coded reagent trays.

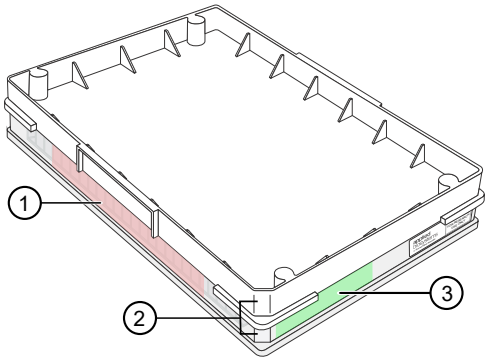
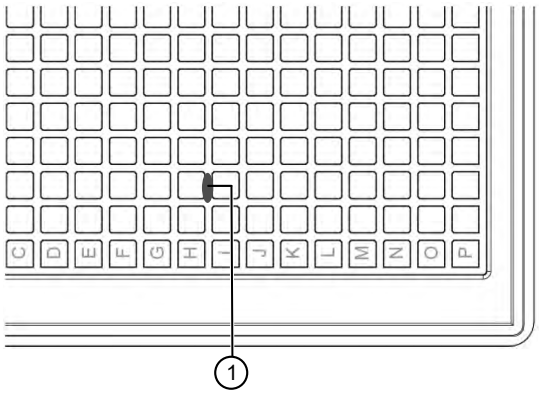


Figure 25 Correct area to label a GeneTitan™ 384 Layout Reagent Tray.

1. Do not label the reagent tray on the long side.
2. Notched corners of the reagent tray and cover must align and face the front.
3. Label the reagent tray here.

Guidelines for aliquoting reagents to GeneTitan™ trays

IMPORTANT! To prevent liquid droplets falling on the stain tray dividers due to electrostatic charges, deionize the stain trays before dispensing the GeneTitan™ reagents using the Multidrop™ Combi Reagent Dispenser.



① Example of a droplet of liquid that has splashed onto the well divider of a stain tray during reagent aliquoting. Ensure that no droplets of liquid are on top of the wells dividers. Blot with a laboratory tissue to remove.

- If the trays are not being used immediately, protect them from light by covering with foil or placing in a cabinet.

Deionization of GeneTitan™ stain trays

When dispensing the GeneTitan™ reagents using the Multidrop™ Combi Reagent Dispenser, we recommend using the minION™2 Ionizing Air Blower at the dispense station. The GeneTitan™ ZeroStat AntiStatic Gun (Cat. No. 74-0014) can also be used to manually deionize GeneTitan™ stain trays and scan trays before dispensing reagents with the Multidrop™ Combi.

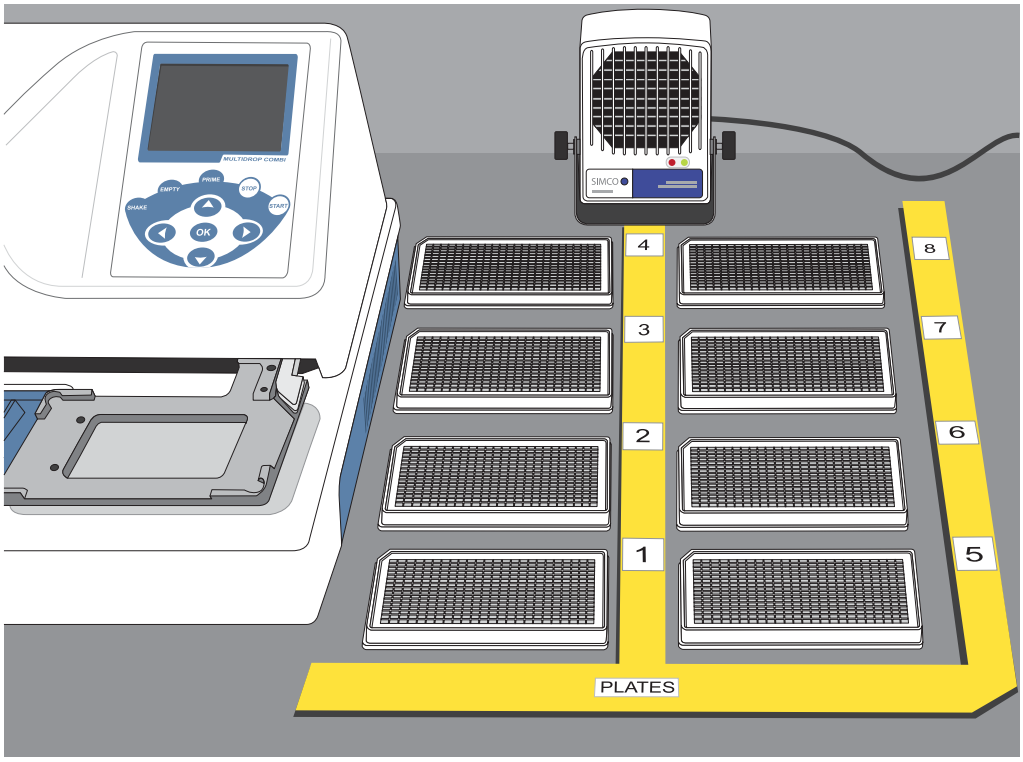


Figure 26 Placement and positioning of stain trays for deionization with the minION™2 Ionizing Air Blower.

Manual deionization of GeneTitan™ trays

Deionize the inner surface of each tray with the wells that hold reagents.

CAUTION! Do not deionize the hybridization tray.

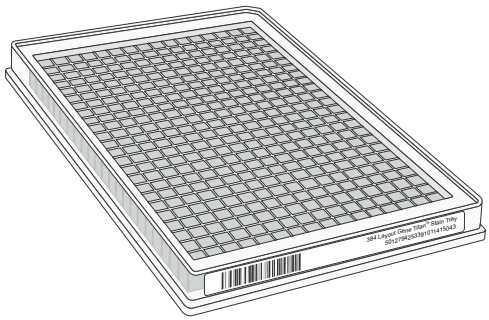


Figure 27 Deionize the 384-Layout GeneTitan™ Stain Tray.

Deionize GeneTitan™ trays

WARNING! The deionization steps damage the arrays on the plate. Before using the antistatic gun, ensure that the array plates remain in their protective pouch and placed away from the deionization area.

Place the scan tray and hybridization tray away from the area where you are performing deionization.

During this procedure, treat the plate as if it were divided into 6 sections. See Figure 28 for details.

1. Place a laboratory tissue on the benchtop.
2. Place the stain tray on a table top. Use the antistatic gun, squeeze, then release the trigger slowly 3 times over the center of each section, squeezing for approximately 2 seconds, then releasing for approximately 2 seconds.

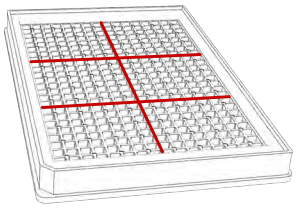


Figure 28 Deionization zones for 384-Layout GeneTitan™ Stain Tray.

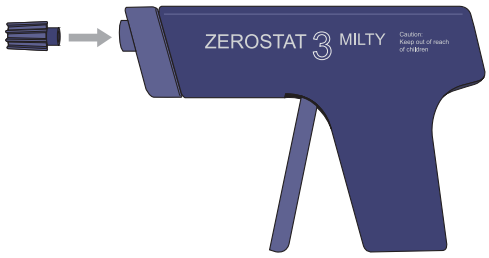
Ensure that a stream of ionized particles settles on all wells of the stain tray to dissipate the static electricity.

Ion-indicator cap

The GeneTitan™ ZeroStat AntiStatic Gun includes an ion-indicator cap. The cap is a device that is used to test the release of ions when the antistatic gun is in use.

Test the antistatic gun for ion release

1. Insert the ion-indicator cap into the nose of the GeneTitan™ ZeroStat AntiStatic Gun.



IMPORTANT! Do not leave the ion-indicator cap on the antistatic gun when deionizing trays.

2. Slowly squeeze the release trigger, then observe the discharge through the viewing window on the ion-indicator cap.

   A visible light is observed in the viewing window on the cap when charged ions are discharged.

3. If you cannot see a light through the viewing window, replace the antistatic gun as it is unusable.

   Each GeneTitan™ ZeroStat AntiStatic Gun produces approximately 50,000 trigger operations, which are sufficient for approximately 200-250 runs on the GeneTitan™ MC Instrument.

4. If you can see a light through the viewing window, then have determined that the gun is functional, remove the cap from the gun before deionizing a tray.

Best practice guidelines for GeneTitan™ reagent bottles

Follow these best practices for re-filling GeneTitan™ reagent bottles if processing 2 or more array plates on the same day.

1. When prompted to refill or replenish the glass bottles, follow these guidelines:
   - Axiom™ Wash Buffer A: Refill the GeneTitan™ Wash A bottle to a minimum of 2 L up to approximately 2/3rds of the glass bottle volume.
   - Axiom™ Wash Buffer B: Refill the GeneTitan™ Wash B bottle up to the 1-L mark.
   - Axiom™ Water: Refill the GeneTitan™ Rinse bottle up to the 1-L mark.
2. Do not overfill the bottles.

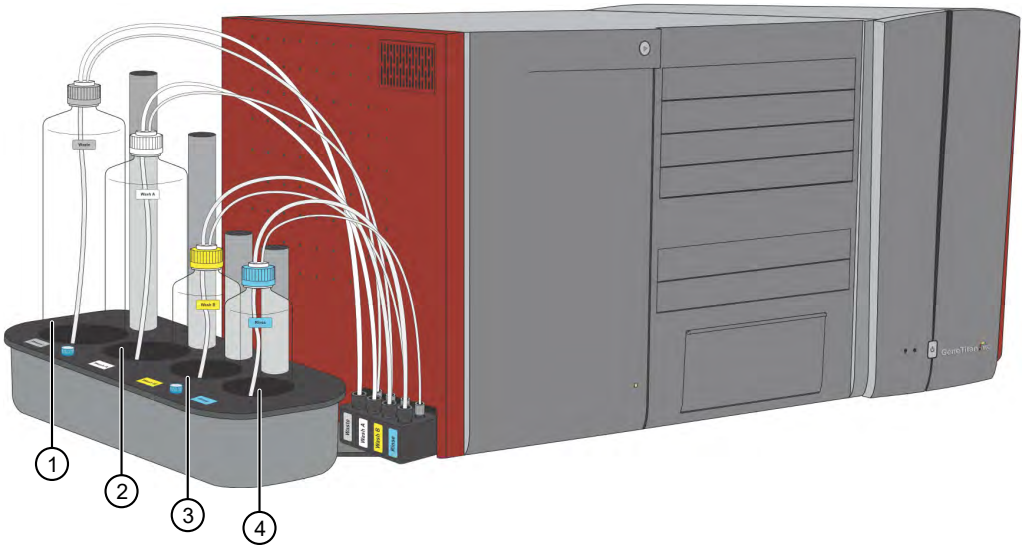


Figure 29 GeneTitan™ Multi-Channel (MC) Instrument reagent bottles.

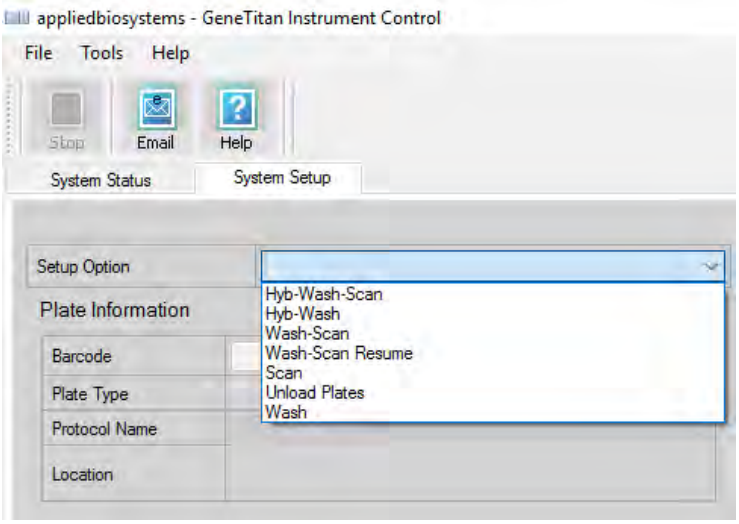
① Waste bottle
② Wash A bottle
③ Wash B bottle
④ Rinse bottle

Setup options for array plate processing

There are 3 steps performed by the GeneTitan™ MC Instrument for array plate processing:

- Hybridization
- Wash and Stain
- Imaging (Scan)

The GeneChip™ Command Console™ software provides options to perform all these steps, or only some of the steps. This section describes the **System Setup** options.



Hyb-Wash-Scan

The **Hyb-Wash-Scan** setup option enables you to hybridize, wash-ligate-stain-stabilize, and scan an array plate on the GeneTitan™ MC Instrument.

- **Hyb**: The array plate is moved to the hybridization oven inside the instrument. Each denatured sample in the hybridization tray is hybridized to an array on the array plate.
  - Time that is required for 384 samples = 23.5 hours
- **Wash**: Samples on arrays are ligated, washed, stained, and stabilized.
  - Time that is required for 384 samples = ~4 hours
- **Scan**: The array plate is moved to the imaging device in the GeneTitan™ MC Instrument and each array is scanned.
  - Time that is required for 384 samples = ~4.75 hours

Hyb-Wash

When the **Hyb-Wash** setup option is selected, processing stops after the array has gone through fluidics processing. Use this option if an array plate cannot be scanned on the same GeneTitan™ MC Instrument as the one used for hybridization and fluidics processing.

1. If the array plate cannot be scanned immediately after the **Hyb-Wash** process is complete, store the array plate following these steps:
   a. Wrap the array plate (in the scan tray with blue protective base) in aluminum foil to protect from light. No lid is required.

      Do not tilt or invert the plate stack. If tilted or inverted, the Hold Buffer spills out of the tray. To prevent liquid spillage, keep the plate stack level when handling it. Do not touch the bottom optical surface of the scan tray.

   b. Store at 4°C.

   c. Scan the array plate within 1 week.

2. When ready to scan, prepare the array plate following these steps:
   a. Protect the plate from light.
   b. Bring the plate to room temperature for approximately 50 minutes.
   c. Remove the aluminum foil, then load the plate onto the GeneTitan™ MC Instrument.

Wash-Scan

Note: The **Wash-Scan** option is available in GCC version 6.1 or later.

Use the **Wash-Scan** option if:

- The array plate was hybridized in an oven separate from the GeneTitan™ MC Instrument.
- To bypass the hybridization step and perform only the wash/stain and scan steps.

Note: If the **Wash-Scan** option is selected, it usually takes 25–30 minutes to warm up the Wash B.

Queue another plate for Wash-Scan

This technique can be used in high throughput workflows to maximize the capacity of the GeneTitan™ MC Instrument.

- Load the second plate to be washed and scanned immediately after the first plate has finished washing (~4 hours).
- Load the third and subsequent plates for **Wash-Scan** at a ~4.75-hour interval, or after the plate being processed in the scanner has completed.

Wash

The **Wash** workflow enables you to bypass the scan step, performing only wash and stain.

Note: When the **Wash** option is selected, allow 25–30 minutes to warm up the Wash B.

IMPORTANT! After the **Wash** workflow is complete, scan the array plate as soon as possible. Array plate data can be affected when the plate is not scanned immediately after washing.

Wash-Scan Resume

Use the **Wash-Scan Resume** option if fluidics processing has been interrupted (for example, a power failure at your facility). This allows you to resume an interrupted workflow at any point in the **Wash** stage.

If a run is aborted during fluidics processing, the instrument places the aborted array plate into the scan tray. To restart this process, remove the array plate from the scan tray then place the array in its blue protective base.

The step at which the run was aborted is identified by:

- Viewing the **System Status** window if you are aborting the last plate through the fluidics system.
- Starting the **Resume** process.

Select **Wash/Scan Resume** from the **System Setup** tab, then follow the prompts to unload and reload all drawers.

The trays are loaded. It is up to you to determine whether to load fresh reagents or reuse the trays already in the GeneTitan™ Multi-Channel (MC) Instrument. Base your decision on the step where the problem occurred.

To help ensure that the samples are processed correctly, we recommend that you:

- Load new stain trays with fresh reagents.
- Load a new scan tray.

We do not recommend the use of trays without reagents or holding buffers for steps that have already been executed.

Resume step

For **Resume**, select the step at which to resume plate processing. You can select any step that has not yet been started.

For specific steps, you can enter a length in seconds (even if the step requires >1 hour to run, you must enter the length in seconds). You can set a step for less time than normal length of time, but not for longer than normal.

Scan

Use the **Scan** option for the following circumstances.

- To rescan an entire array plate or specific arrays on a plate that failed to scan for reasons such as bubbles or gridding failure.
- To scan a plate that has already been hybridized, stained and washed on a different GeneTitan™ MC Instrument.

Queue a second plate for scanning

Use the **Scan** option in the **System Setup** tab to start a second scan workflow. The software automatically moves the second plate into the scanner when the first plate has completed scanning.

GeneChip™ Command Console™ v4.3 or later is required for this procedure.

1. Start the first scan workflow in the GeneTitan™ MC Instrument. Wait until the first plate is loaded into the imaging device and scanning starts.

2. Go to the **System Setup** tab, then select **Scan** from the **Setup Option** dropdown list. The **Setup Option** dropdown list is active only after the first plate starts scanning.

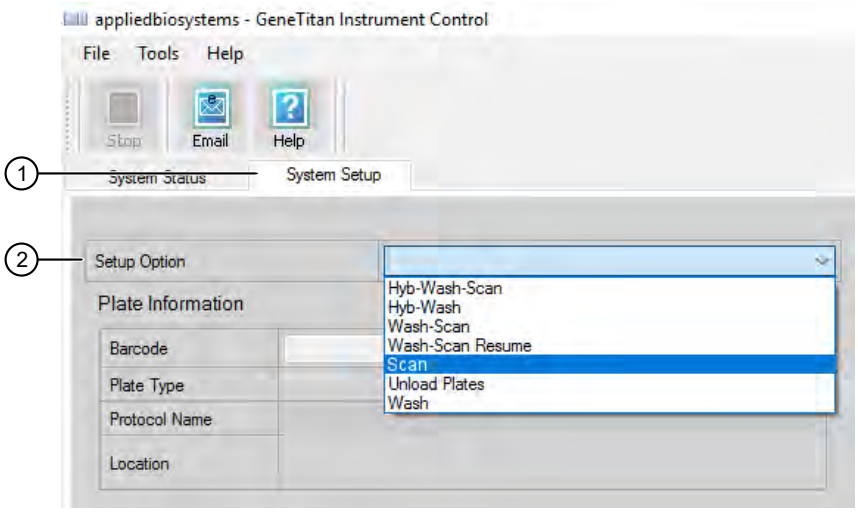


Figure 30 Scan setup option to process a second array plate.

① **System Setup** tab ② **Scan Option** dropdown list

3. Click **Next** in the lower left section of the window under the **Status** pane.

4. Scan or manually enter the array plate barcode, then click **Next**.

5. Follow the instructions in the **Status** pane and empty the trash bin if needed, then press the blue confirmation button on the instrument to continue.

6. Place the array plate on top of a scan tray in the correct orientation such that notched corner of the array plate and scan tray are aligned.

7. Load the array plate/scan tray combination in drawer 2 of the GeneTitan™ MC Instrument, on the left or right side, as instructed in the **Status** pane. Ensure that the array plate/scan tray combination is loaded in the correct orientation in the drawer. If needed, see Figure 16 for further information on the proper alignment and loading of plates, covers, and trays in the GeneTitan™ MC Instrument.

8. When ready, press the blue confirmation button on the instrument.

9. Select the arrays to scan in the **Array Selection** section in the upper right corner of the window, then click **Next**.

10. In the **Start Processing** confirmation message, click **OK** to continue.
    The second queued plate runs after the first scan finishes and the scanner is available.

Unload Plates

Use the **Unload Plates** option to unload plates and trays from the instrument when processing is complete or has been aborted.

Load an array plate and hybridization tray into the GeneTitan™ MC Instrument (for Hyb-Wash-Scan or Hyb-Wash)

1. When drawer 6 opens, load the array plate and hybridization tray in the following manner:
   a. Examine the wells of the hybridization tray for bubbles, then puncture any bubbles with a pipette tip.

   IMPORTANT! Removing bubbles at this step greatly reduces the chance of bubbles under the arrays when the hybridization tray and the array plate are clamped. Bubbles under an array can result in black spots on the array image.

   b. Load the uncovered hybridization tray on the right side of the drawer.

c. Remove the array plate and blue protective base from its package. For more information on the array packaging, see “Array plate packaging” on page 150.

To avoid dust or damage to the plate, leave the array plate packaged until ready to load onto the GeneTitan™ MC Instrument. The array plate must be loaded on its blue protective base. The white plastic shipping cover on top of the array plate *must not* be loaded in the GeneTitan™ MC Instrument.

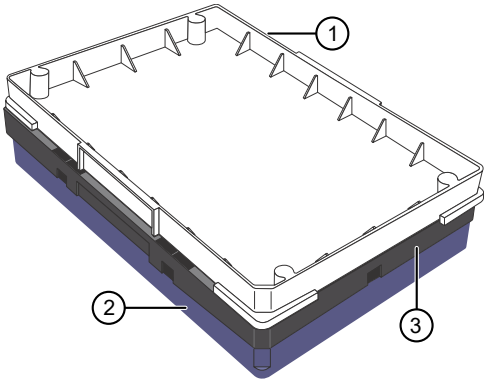


Figure 31 Array plate components, as shipped.

(1) White shipping cover to be discarded.
(2) Array plate blue protective base.
(3) Array plate.

d. Load the array plate with the blue protective base on the left side of the drawer.

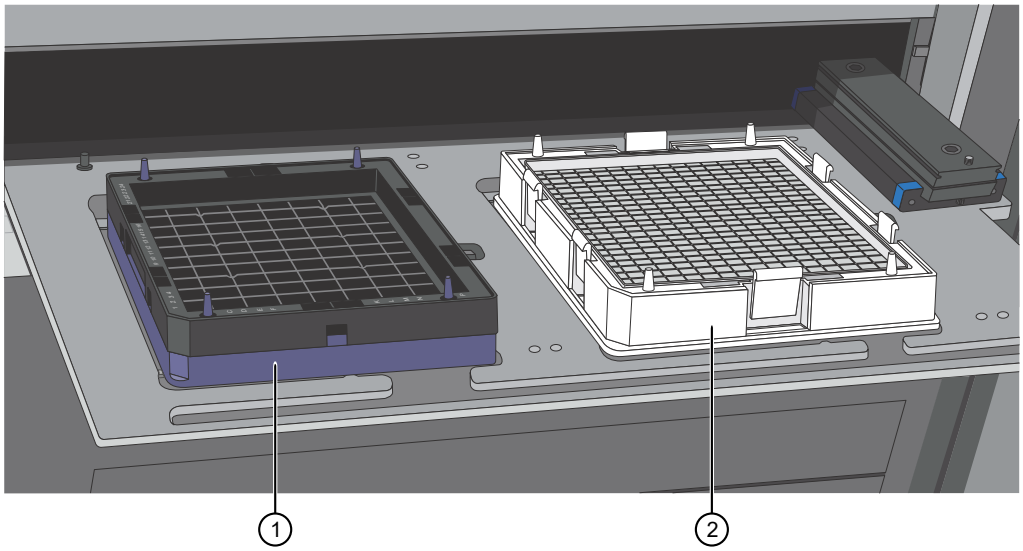


Figure 32 Array plate on blue protective base and the hybridization tray properly loaded into drawer 6.

① Array plate on blue protective base.
② Hybridization tray.

IMPORTANT! Do not install a three-plate stack of trays. Ensure that you have removed the white plastic shipping cover.

CAUTION! The notched corner of each plate, cover, and tray must be aligned. When loading onto the GeneTitan™ MC Instrument, the notched edge plates, covers, and trays must be aligned as indicated by the Tray Alignment guide in the drawer.

An error message is displayed if the instrument has difficulty reading the barcode on the plate. Plate barcodes must face the internal barcode reader in the back of the drawer. Improper tray positioning can crash the GeneTitan™ MC Instrument, resulting in substantial damage to the instrument, and loss of samples.

e. Press the blue confirmation button on the GeneTitan™ MC Instrument to continue.

Note: When an array plate is loaded on the left side of the drawer, the internal barcode reader reads the barcode of the array plate. The barcode is compared with the barcode and the plate type that is specified in the **Barcode** and **Plate Type** fields that were selected during the **Setup**. If the information is correct, the application allows you to proceed to the next step. If the instrument is unable to read the barcode, it pushes out the tray and prompts you to load the correct plate with the proper orientation into the instrument.

If an error occurs, check the loading of the array plate and click **OK** to retry. Alternatively, click **Skip** if the instrument continues to have problems after ensuring that the trays have been loaded in the proper orientation.

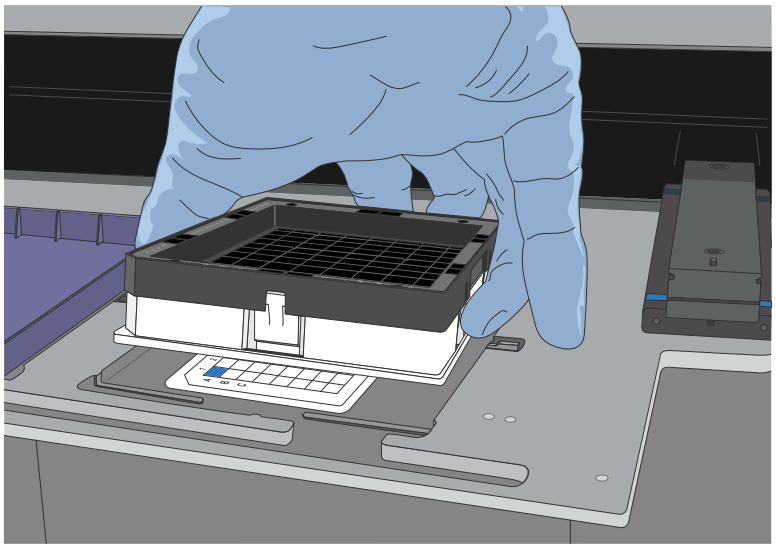
2. Select the arrays to scan. By default, all arrays are selected.

3. Click **Next**, then click **OK** in the **Start Processing** dialog box to start processing the samples.
   The GeneTitan™ MC Instrument places the array plate on top of the hybridization tray (now called the plate stack). The GCC software starts the process for placing the array plate onto the hybridization tray. A **Clamping in Progress** dialog appears.

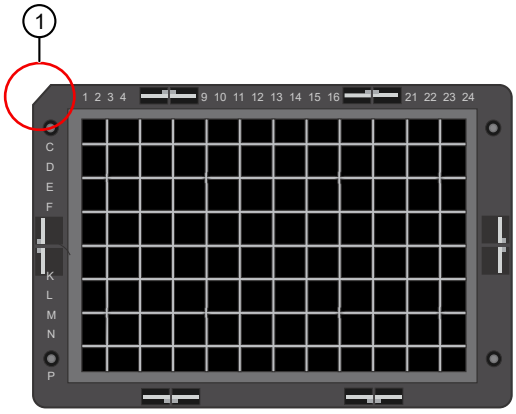
4. Press **OK**, then wait for the drawer to open completely before retrieving the array plate and hybridization tray combination for manual clamping and inspection.
   After clamping is complete in the instrument, drawer 6 opens and the **ensure Clamping** dialog appears. Do not click **OK** yet. The sandwich of the array plate and hybridization tray must be manually clamped and inspected before the array processing can start.

CAUTION! At this stage, the array plate does not latch securely to the hybridization tray. Always grip the plate stack from the hybridization tray, which is on the bottom of the plate stack. *Do not* grasp only the array plate to remove the plate stack from the drawer of the GeneTitan™ MC Instrument.

5. Complete the following steps to clamp the array plate manually to the hybridization tray.
   a. Grip the body of the hybridization tray by hand, then remove the plate stack from drawer 6 right location of the GeneTitan™ MC Instrument.

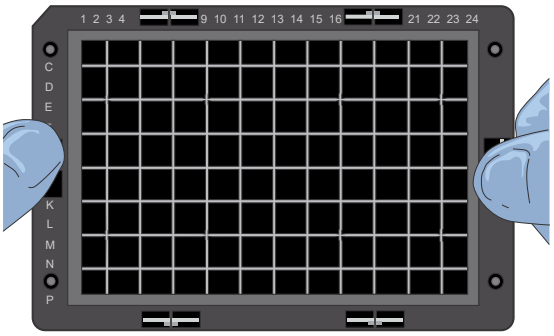


b. Place the plate stack on a flat surface of the table or the lab bench. Position the plate stack to match the orientation as shown in the figure.



1 Chamfer corner.

c. Position the left and right thumb fingers on the locations that are indicated in the picture. Press the array plate downward until the clicking sound is detected.



d. With the plate stack resting on a flat surface, rotate the stack 90° clockwise. Position the left and right thumbs on the locations that are indicated in the picture. Press the array plate downward until the clicking sound is detected.

6. Ensure the clamping of the plate stack to check that the array plate is securely fastened to the hybridization tray. Using your thumbs, press the array plate downward following the positions that are specified in Figure 33. *No clicking sound indicates proper clamping.*

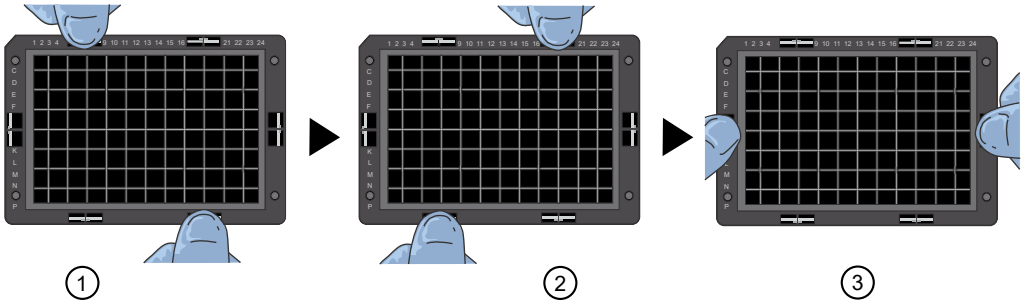


Figure 33 Clamping verification steps.

7. Inspect the array plate for bubbles.
   a. **Keeping the plate stack level**, inspect the bottom for bubbles under the arrays—*do not* tilt or invert the plates.
   b. If bubbles are present, gently tap the plate until the bubbles move out from under the arrays—*do not* unclamp the plate stack.
8. Return the plate stack to the drawer with the notched corner facing you, then press the blue confirmation button on the GeneTitan™ Instrument to proceed.
9. A message is displayed if plate orientation is not correct or if the hybridization tray barcode cannot be read. If this message appears, complete one or both of the following actions.
   - Check the loading of the array plate and click **OK**.
   - Click **Skip** if the instrument continues to have problems reading the barcode and after ensuring that the correct trays have been placed in the proper orientation.
10. Continue to “Load a second array plate and hybridization tray” on page 172.

Load a second array plate and hybridization tray onto the GeneTitan™ MC Instrument

When a second array plate and hybridization tray can be loaded

After processing starts on the first plate stack, you have a specific length of time during which you can load another array plate and hybridization tray. This length of time is displayed above the **Hybridization Oven Status** pane (Figure 34). You cannot load another hybridization tray before or after this time.

IMPORTANT! The next array plate and hybridization tray must be loaded during the time frame of that is displayed above the **Hybridization Oven Status** pane. You cannot load another hybridization tray before or after this time. You are required to wait until the current process is finished which results in disruption of the multi-plate plate workflow.

When the first plate is in the oven and the time spacing requirement is met, you can load another plate. This time spacing requirement is to ensure that the second plate does not have to wait for system resources in its workflow. The time spacing is approximately equal to the longer of the scan time of the first plate.

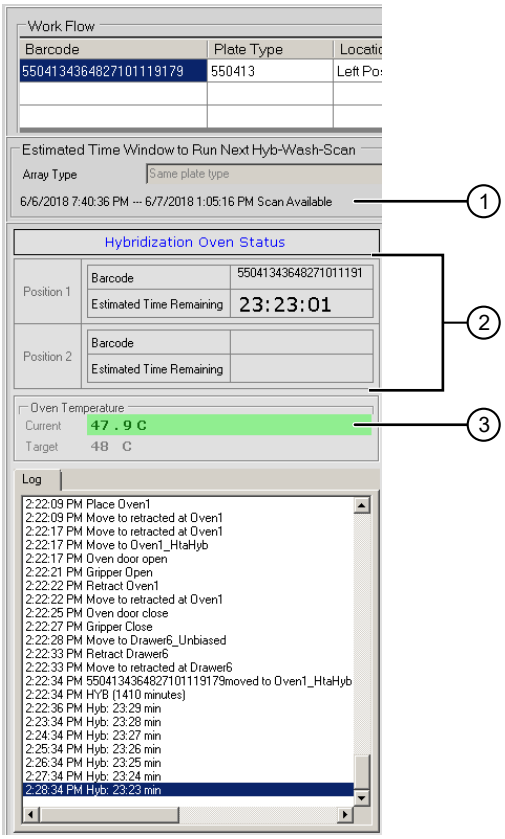


Figure 34 When to load a second array plate and hybridization tray based on oven status information.

① This pane displays the amount of time during which another array plate and hybridization tray can be loaded. Additional plates cannot be loaded before or after this time as the instrument is operating. In this figure, the system is currently available.

② Position of plate stack in the hybridization oven. Only 1 plate is being processed in this figure. As such, position 2 is blank.

Position 1—left side of the oven

Position 2—right side of the oven

③ Green indicates that the current oven temperature is in the target temperature range. Yellow indicates that oven temperature is outside of target temperature range.

Load a second array plate and hybridization tray

1. Select the **System Setup** tab.

2. Load an array plate and hybridization tray in the same manner as the previous plate and tray were loaded.

   a. Scan or manually enter the array plate barcode, then click **Next**.

b. Load the array plate with the blue protective base and the hybridization tray without the cover, then press the blue confirmation button.

c. Select the arrays to scan, then click **Next**.

d. Ensure that the plates are clamped securely when prompted, then press the blue confirmation button.

e. Click **OK** when prompted to resume plate processing.

3. Select the **System Status** tab to view the status of the array plates in the **Work Flow** pane.

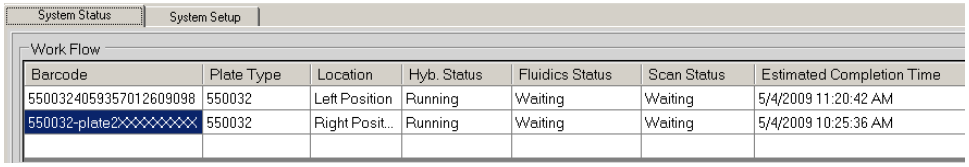


Figure 35 Example of the Work Flow pane when 2 plates are loaded and are in the hybridization oven.

When to abort a process

If needed, the processing of array plates can be aborted.

If a plate is in the fluidics station, the abort process can take up to 3 minutes. The status window displays “AbortRequested” and then changes to “Aborted”.

A clamped array plate/hybridization tray stack that is aborted while it is in the oven or in drawer 6 is moved to drawer 1.

To retrieve the array plate and related consumables after the instrument aborts a process, take the following actions as needed.

- Use the **Unload Plates** option.
- Start another run. That forces the system to unload the aborted plates.

An instrument-initiated abort can occur for the following reasons.

- The plates are improperly placed.
- The uninterruptible power supply (UPS) detects a long power interruption, draining the UPS to 75% power.
- The equipment malfunctions.

When the system aborts the processing, follow the instructions that are displayed in the user interface.

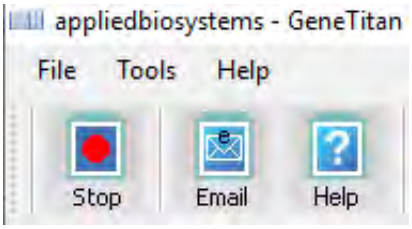
The operator can initiate an abort on 1 plate and the gripper can continue to process other plates in the instrument.

Abort a process

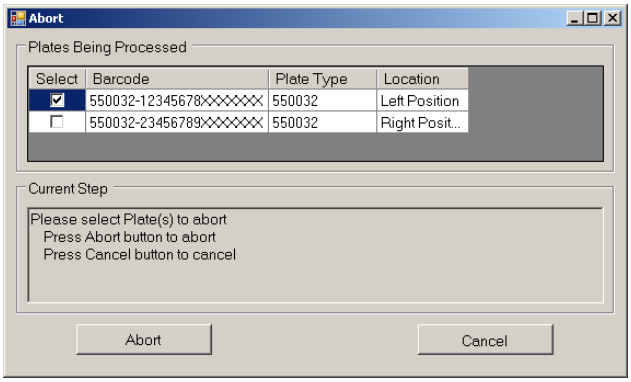
Use the following procedure to abort a process underway in the GeneTitan™ MC Instrument.

Note: If reagents are loading, do not use this method. Instead, click **Cancel** in the reagent load step.

1. Click **Stop** in the upper left corner of the **Instrument Control** window.



2. In the **Abort** dialog box, select the array plate to abort, then click **Abort**.



3. In the confirmation box, click **Yes**.

4. Wait until the status of the array plate in the **Work Flow** pane in the **Instrument Control** display changes from “AbortRequest” to “Aborted”.

Note: If reagents are loading, abort the plate by clicking **Cancel** in the reagent load step.

Note: If the gripper is required to complete the abort process, the plate remains in the “AbortRequest” state until the gripper becomes available.

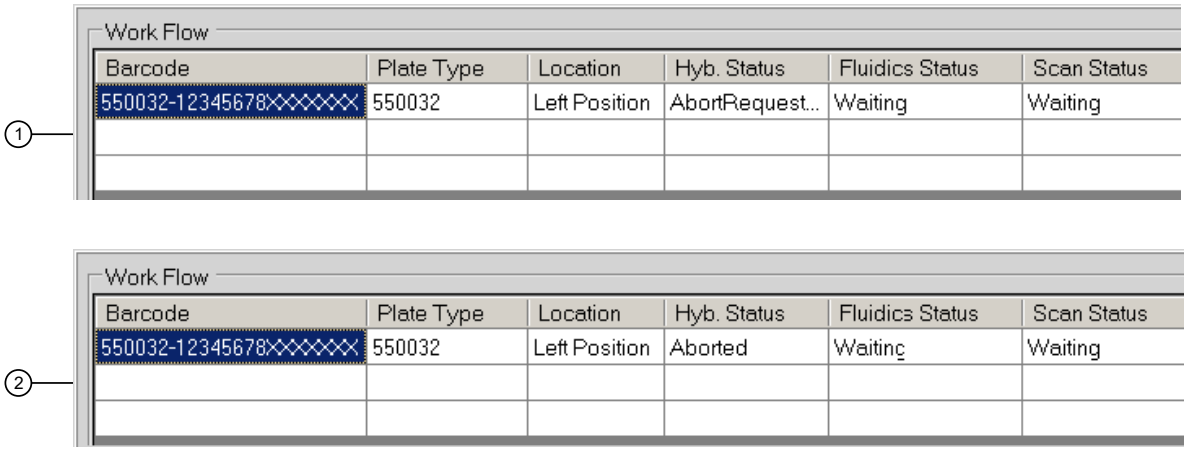


Figure 36 The Work Flow pane.

1. Shows that the abort has been requested.
2. Shows that the abort has been completed.

5. After the abort process is completed, do one of the following to retrieve the array plate and related consumables.
   - In the **Setup Option** list, select **Unload Plates**.
   - Start to load a new array plate.

Email notifications from the GeneTitan™ MC Instrument

You can configure the GeneChip™ Command Console™ software to send email notifications about the GeneTitan™ MC Instrument status. It is critical that you know when the instrument requires attention for sample handling or troubleshooting. Rapid notification can lessen the risk of sample loss.

The system can notify you when a process starts, completes, aborts, or encounters an error.

For instructions on setting up notifications, see the *GeneChip™ Command Console™ User Guide*.

GeneTitan™ MC Instrument lamp

The GeneTitan™ MC Instrument uses a xenon arc lamp system that is warranted to provide 500 hours of illumination for imaging the array at 2 wavelengths. The xenon lamp has a limited lifetime and must be replaced at regular intervals.

The GeneTitan™ Instrument Control software provides a timer that indicates the remaining useful life of the bulb and notifies you when it requires replacement. It is important to adhere to the warnings specified in the *GeneTitan™ Multi-Channel Instrument User Guide*.

See the user guide for the Lambda LS and Smart controller system. Never manually switch the lamp and the controller on or off. The GeneTitan™ Instrument Control software manages the lamp activity and switches the lamp on and off as required. It takes 10 minutes to warm up the lamp. In idle mode, the lamp remains on for 2 hours before it is automatically switched off if there are no more plates being transferred from the fluidics to the imaging station. This automatic switching is by design and is intended behavior. Do not try to save the lamp life by powering off the switch on the lamp.

Note: The power switch on the shutter box must always be ON. The OPEN/CLOSE switch on the shutter box must always be at the AUTO position.

B

Fragmentation quality control gel protocol

- Equipment required 177
- E-Gel™ and reagents required 177
- Consumables required 178
- Prepare the gel diluent 178
- Dilute the 25 bp DNA Ladder 178
- Run the fragmentation QC gel 179

Equipment required

"MLS" indicates that the material is available from **fisherscientific.com** or another major laboratory supplier.

Item	Source
Gel Imager	**MLS**
Pipette, multichannel or single-channel P20	**MLS**
Plate centrifuge	**MLS**
Vortexer	**MLS**

E-Gel™ and reagents required

Unless otherwise indicated, all materials are available through **thermofisher.com**.

Item	Source
Invitrogen™ Mother E-Base™ Device	EBM03
Invitrogen™ Daughter E-Base™ Device	EBD03
Invitrogen™ E-Gel™ *48* Agarose Gels, 4%	G800804
Applied Biosystems™ 25 bp DNA Ladder, or a similar product prepared as instructed by the manufacturer	931343

(continued)

Item	Source
Invitrogen™ TrackIt™ Cyan/Orange Loading Buffer	10482028
Invitrogen™ UltraPure™ DNase/RNase-Free Distilled Water	10977023

Consumables required

Unless otherwise indicated, all materials are available through **thermofisher.com**.

Item	Source
Adhesive film—use one of the following: • Applied Biosystems™ MicroAmp™ Clear Adhesive Film • Microseal™ 'B' PCR Plate Sealing Film	 • 4306311 • Bio-Rad™, MSB1001
Pipette tips	Same brand as pipettor

Prepare the gel diluent

A 100-fold dilution of the TrackIt™ Cyan/Orange Loading Buffer can be used in “Stage 5A: In-process QC” on page 96.

1. Add 500 µL of TrackIt™ Cyan/Orange Loading Buffer to 49.5-mL nuclease-free water. Total volume 50 mL.
2. Mix well.
3. Store at room temperature.

Dilute the 25 bp DNA Ladder

Applied Biosystems™ 25 bp DNA Ladder, Cat. No. 931343, is required for this procedure.

1. Add 25 µL of the 25 bp DNA Ladder to 125 µL of UltraPure™ DNase/RNase-Free Distilled Water.
2. Mix well.
3. Store at 4°C until use.

Run the fragmentation QC gel

Running one 48 lane E-Gel to sample a 384-well plate is recommended. A suggested sampling pattern is to load the gel with the following wells from the 384-well Gel QC Plate:

- Row A—Odd-numbered columns
- Row E—Even-numbered columns
- Row J—Odd-numbered columns
- Row N—Even-numbered columns

If processing multiple plates, sampling different wells from each plate can be helpful in monitoring assay and instrument performance.

1. Tightly seal the Gel QC Plate that is produced during "Stage 5A: In-process QC" on page 96.
2. Vortex the plate for 1 second each corner and 1 second in the center at the maximum setting, then centrifuge at 1,000 rpm for 30 seconds.
3. Power on the E-Base™ device.
4. Push the **Power/Prg** button on each to ensure the gel base is in **EG** mode, not EP mode.
5. Place the E-Gel™ *48* Agarose Gel onto the base unit.
6. Remove 2 combs from the gel.
7. Load 15 µL of samples from user-selected wells of the Gel QC Plate onto the gel.
8. Load 15 µL of 25 bp DNA ladder into the marker wells (M).
9. Load 15-µL nuclease-free water into any unused wells.

10. Run the gel for 19 minutes.

11. Capture a gel image.

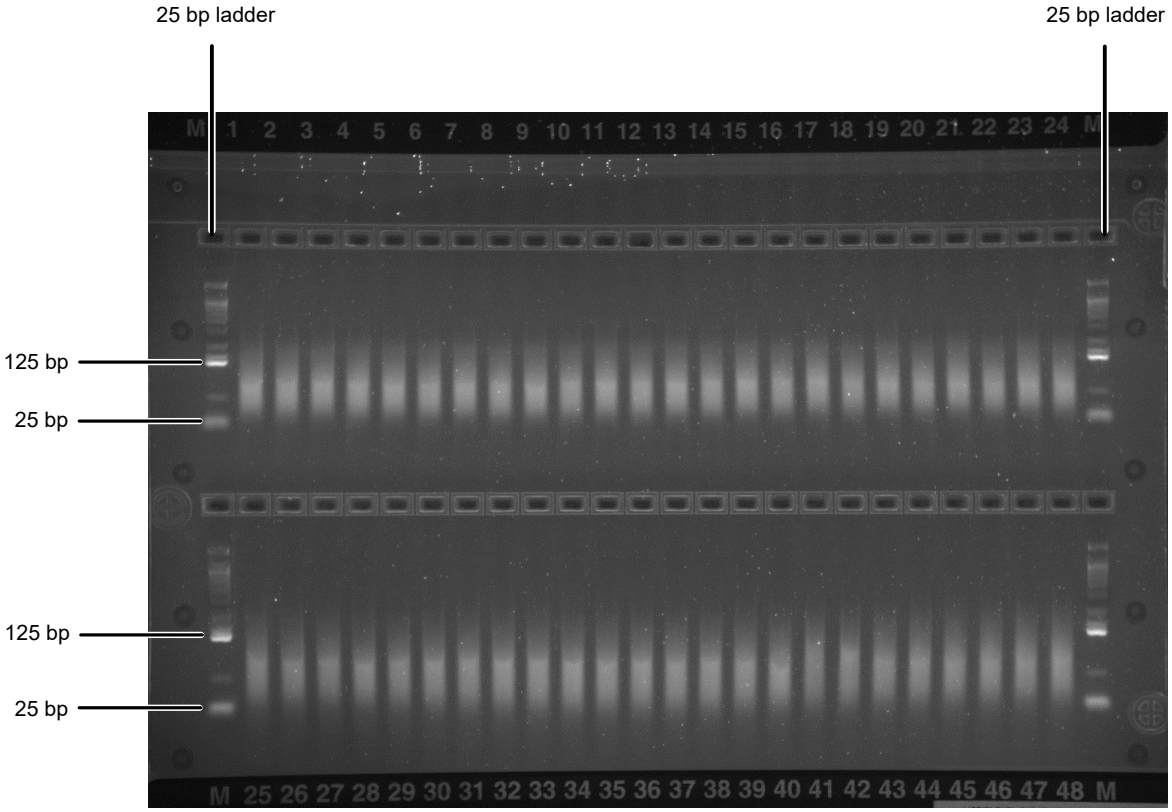


Figure 37 Fragments fall between 125 bp and 25 bp on a successful gel image.

C

Sample quantification after resuspension

- Equipment required 181
- Quantify the diluted samples 181
- Install Axiom™ OD methods on the Multiskan™ Sky Microplate Spectrophotometer 182
- Use a Multiskan™ Sky session 189
- OD yield evaluation guidelines 189
- Plate reader guidelines for sample quantification 190

Equipment required

Unless otherwise indicated, all materials are available through **thermofisher.com**.

Quantity	Item	Source
1	Multiskan™ Sky Microplate Spectrophotometer	51119600

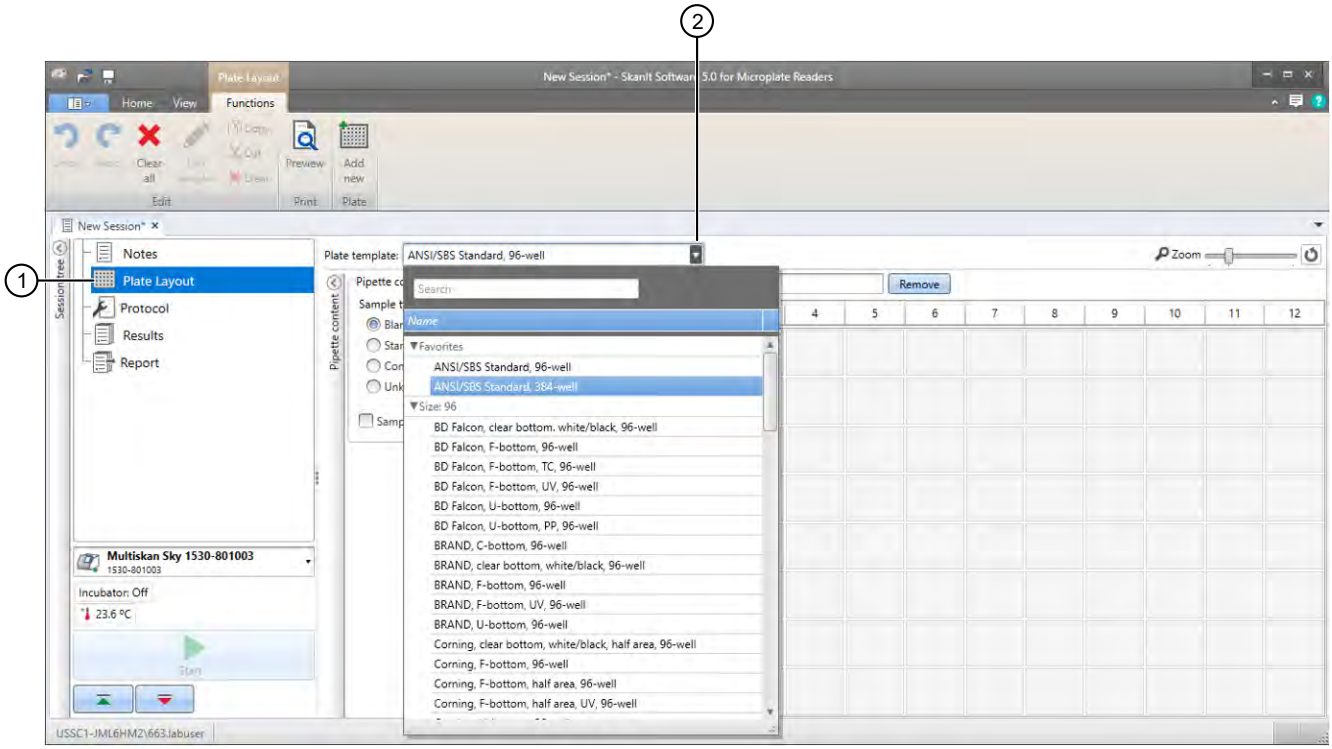
Quantify the diluted samples

During target preparation, 2 plates of diluted samples are prepared: 1 for OD quantification and 1 for a QC gel to check the fragmentation reaction.

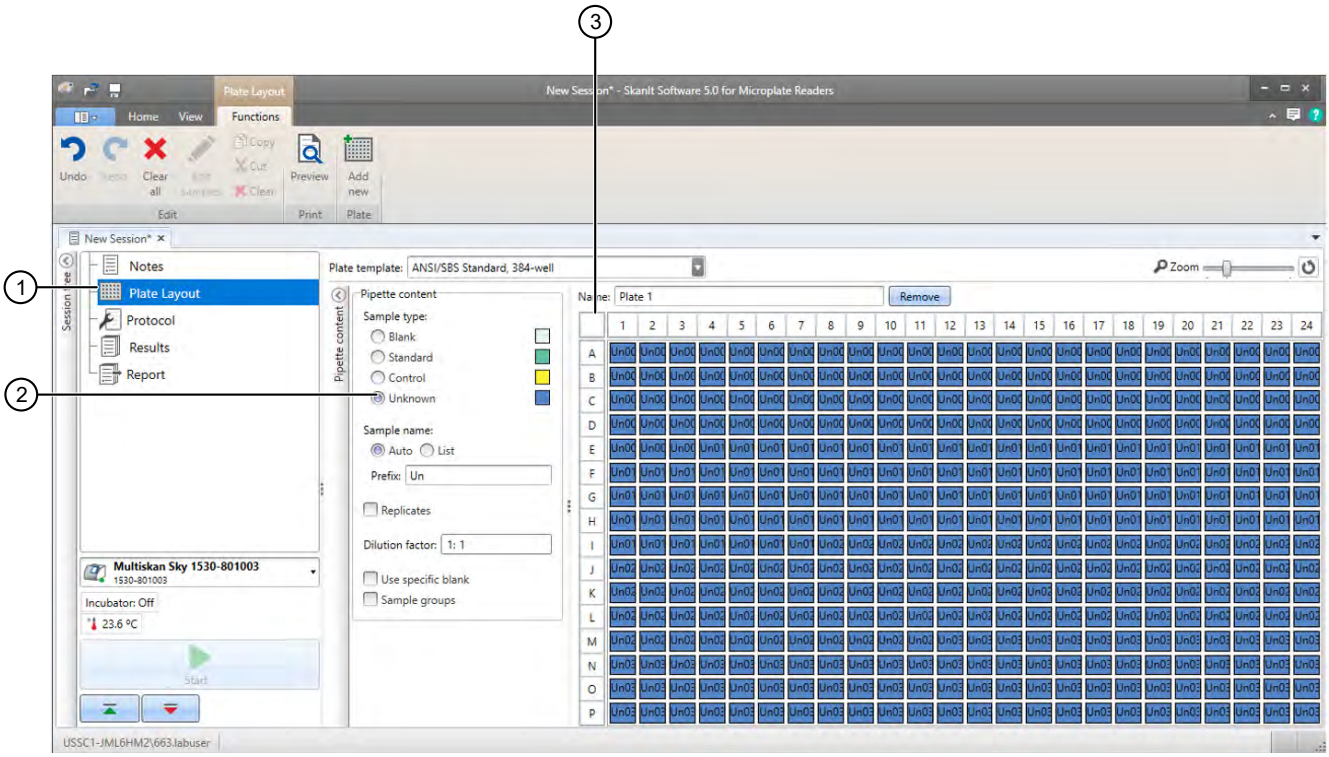
For OD quantification, readings must be taken at wavelengths of 260 nm, 280 nm, and 320 nm. See “Plate reader guidelines for sample quantification” on page 190.

Install Axiom™ OD methods on the Multiskan™ Sky Microplate Spectrophotometer

1. Launch the SkanIt™ software, then click **New session**.
2. In **New Session** pane, click **Plate Layout** then select **ANSI/SBS Standard, 384-well** from the **Plate template** dropdown list.



① Plate Layout.
② **Plate template** dropdown list box.



① **Plate Layout**.
② **Unknown**.
③ Select small square to define all wells as unknown.

3. In **New Session** window, click **Plate Layout**, then select **Unknown**.

4. Cick to select the small square above the **A** and to the left of the **1** to assign all the wells as "Unknown".

5. Click **Protocol** in the session tree pane on the left, then click **Absorbance** under the menu bar.

6. Assign 260 nm, 280 nm, and 320-nm wavelengths to be measured.

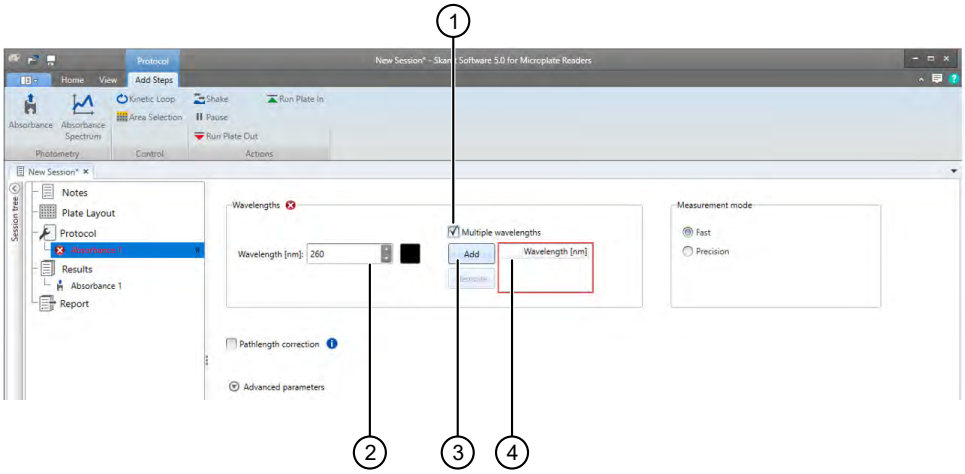


Figure 38 Assign wavelengths.

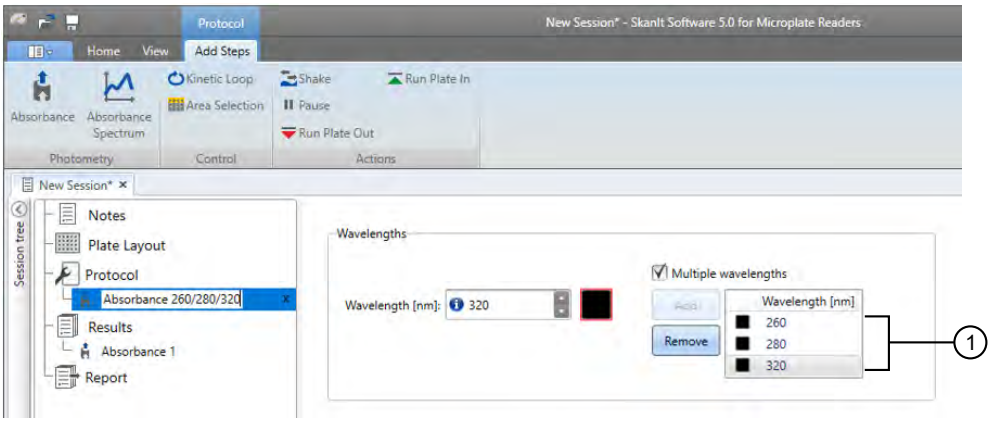
① **Multiple wavelengths** checkbox.
② **Wavelength [nm]** field.
③ **Add**.
④ **Wavelength [nm]** box.

a. Check the **Multiple wavelengths** checkbox.

b. Enter "260" in the **Wavelength [nm]** field, then click **Add**.

After clicking **Add**, 260 appears in the **Wavelengths [nm]** box in the middle of the screen.

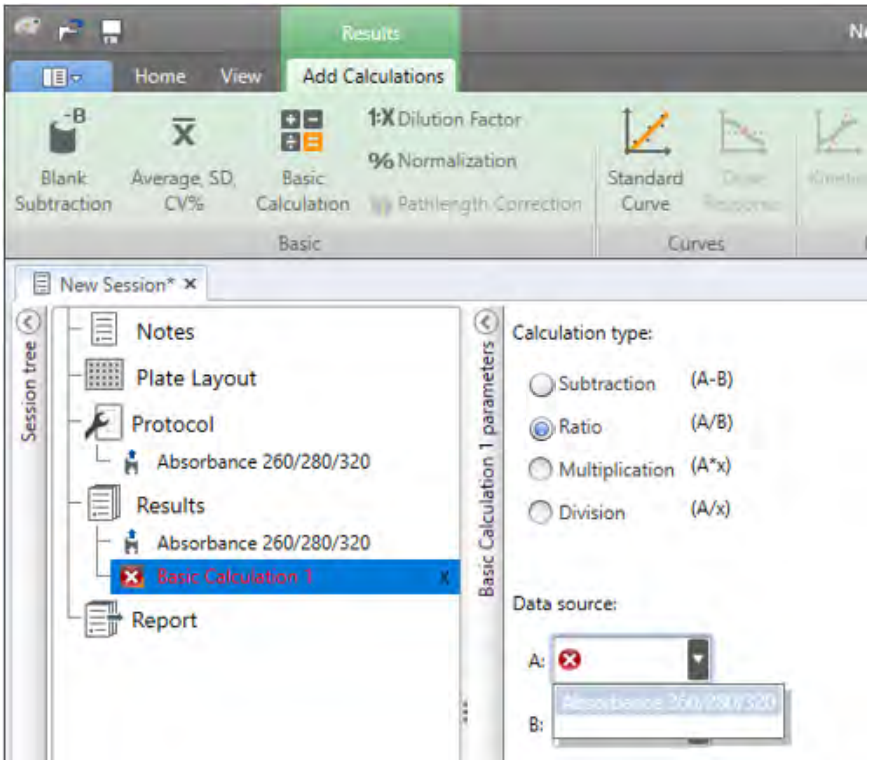
7. Repeat step 6 to add 280 nm and 320-nm wavelengths. When complete, 260, 280, and 320 appears in the **Wavelengths [nm]** box.



① Wavelengths added appear in **Wavelengths [nm]** box.

8. Click **Absorbance 1** in the session tree pane, then rename it "***Absorbance 260/280/320***".

9. Add a calculation to the **New Session**.

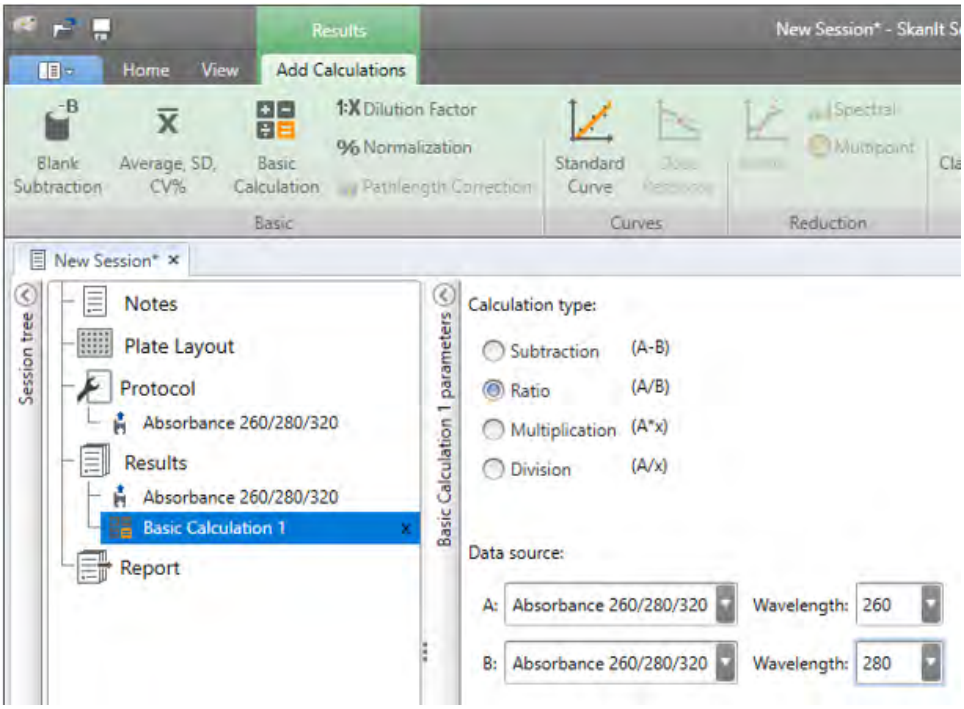


a. Under **Results** in the session tree pane, click **Absorance 260/280/320**.

b. Click **Basic Calculation** to calculate 260 nm/280 nm ratio for each sample, then select **Ratio (A/B)**.

c. Define the **Data source**:

- For **A**, select **Absorbance 260/280/320** and for **Wavelength** select **260**.
- For **B**, select **Absorbance 260/280/320** and for **Wavelength** select **280**.



10. Click **Basic Calculations 1** in the left pane, then rename it "***Ratio 260/280***".

11. Define the calculation for the DNA yields in each well.

    a. Click the **Custom Formula** button under the menu bar, then click the **Define Variables** button in the middle pane.

    b. In the **Define Variables** window, define a variable that is named "A260".

    1. In the **Variable Name** field, enter "***A260***".
    2. In the **Source Steps** dropdown list, select **Absorbance 260/280/320**.
    3. For **Wavelengths**, select **260**, then click **Add**.

    - After clicking **Add**, the new A260 variable and definition move to the right side of the **Define Variables** window.

c. Define a variable named "A320".

1. In the **Variable Name** field, enter "A320".
2. In the **Source Steps** dropdown list, select **Absorbance 260/280/320**.
3. For **Wavelengths**, select **320**, then click **Add**.
4. After clicking **Add**, the new A320 variable and definition move to the right side of the **Define Variables** window. Click **OK** to close the window and return to the **Custom Formula** screen.

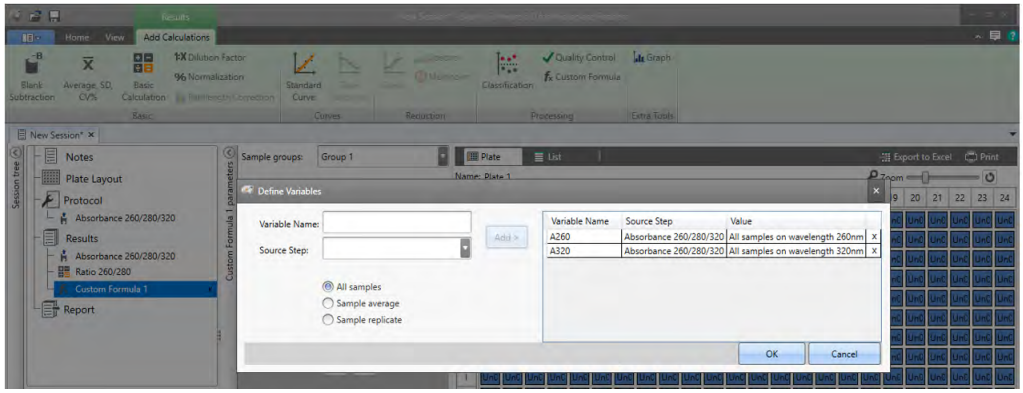
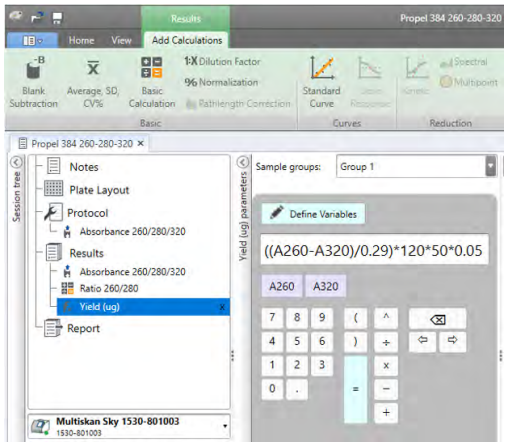


Figure 39 Define Variables window with 2 new variables added.

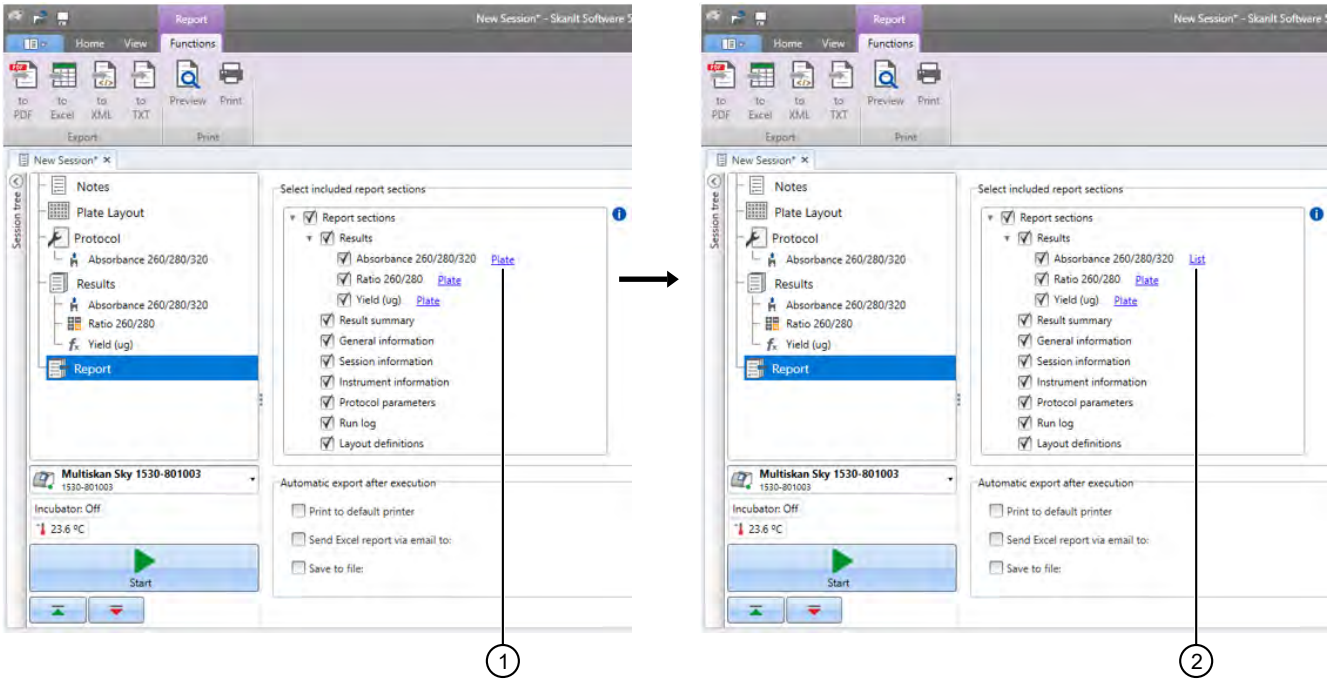
d. In the Custom Formula screen, enter the following DNA yield equation. Use the purple **A260** and **A320** buttons to enter them into the equation.

- *((A260-A320)/0.29)*120*50*0.05*

e. In the left pane, click **Custom Formula 1**, then rename it "***Yield (ug)***".



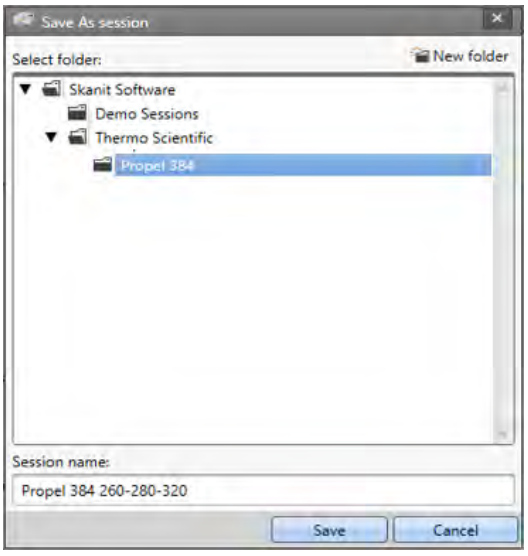
f. Click **Report** in the left pane. Results of the calculation are provided in either Plate or List format. Click **Plate** to change the results to **List** format.



① **Plate** format.
② **List** format.

12. Click the **Home** tab, then click **Save**.

13. In the **Save As session** window, select or create a folder to save it to, and then enter a **Session name**.



14. Click **Save**.
The session is available to be used to read the Axiom™ 384-well OD QC Plates.

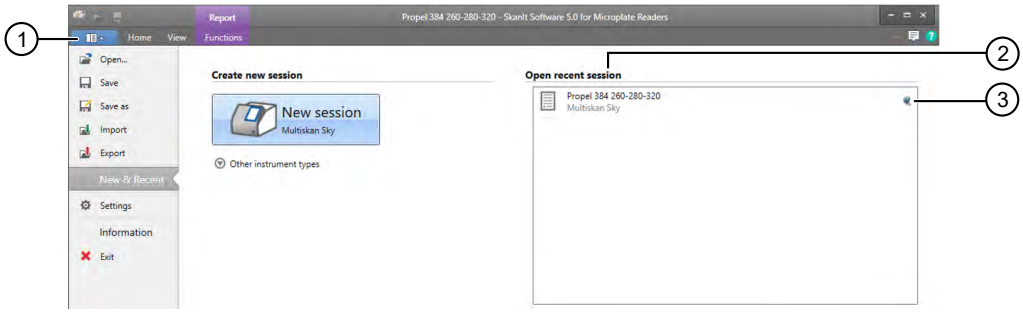
Use a Multiskan™ Sky session

See the *SkanIt™ Software for Microplate Readers User Manual*, Pub. No. N16243, for further details.

1. In the SkanIt™ software, click the file icon tab to the left of the **Home** tab in the upper left of the window.

2. Open a session using one of the following methods.

   - Click **Open** and navigate to the filepath of the session.
   - Click on the session name in the **Open recent session** section on the right side of the window. The session can be pinned to the Open recent session window by clicking on the pin icon on the right.



1. File icon tab
2. Saved session list
3. Pin

OD yield evaluation guidelines

The measurement of the yield of DNA after resuspension of the pellets is an important QC checkpoint in the Axiom™ Propel XPRES 384HT Workflow. If the median yield for the plate is <525 µg DNA per sample:

- Pause the protocol.
- Evaluate all steps that are performed to that point to determine the possible source of the low yields.

This DNA yield corresponds to an A_{260}-A_{320} value of approximately 0.53.

Plate reader guidelines for sample quantification

When performing sample quantification, the plate reader must be calibrated to ensure accurate readings.

The total yield in µg per well can be calculated as:

(A - C)*D*V*E/P

Where:

- A = the observed OD_{260}.
- C = the observed OD_{320} (an estimate of a blank reading).
- D = 120 (the net dilution factor when preparing the OD QC Plate).
- V = 50 (the volume of the sample in µL after the resuspension step).
- E = 0.05 (the extinction coefficient of duplex DNA at 260 nm).
- P = the optical path length for the plate type and plate reader used.

If your plate reader does not record the OD_{320}, the OD_{260} of a blank solution of water only must be used for the parameter "C".

The optical path length depends on the type of plate and can depend on the spectrophotometer used. Check the recommendations for the path length for your instrument and plate type or for recommendations on how to measure this quantity.

D

Register samples in GeneChip™ Command Console™

GeneTitan™ Array Plate Registration file

A GeneTitan™ Array Plate Registration file is a Microsoft™ Excel™ spreadsheet that includes information on the samples that you are processing on a single array plate. This information includes the array plate format, the array plate barcode, and the sample file names for tracking the samples that are loaded onto a particular array plate.

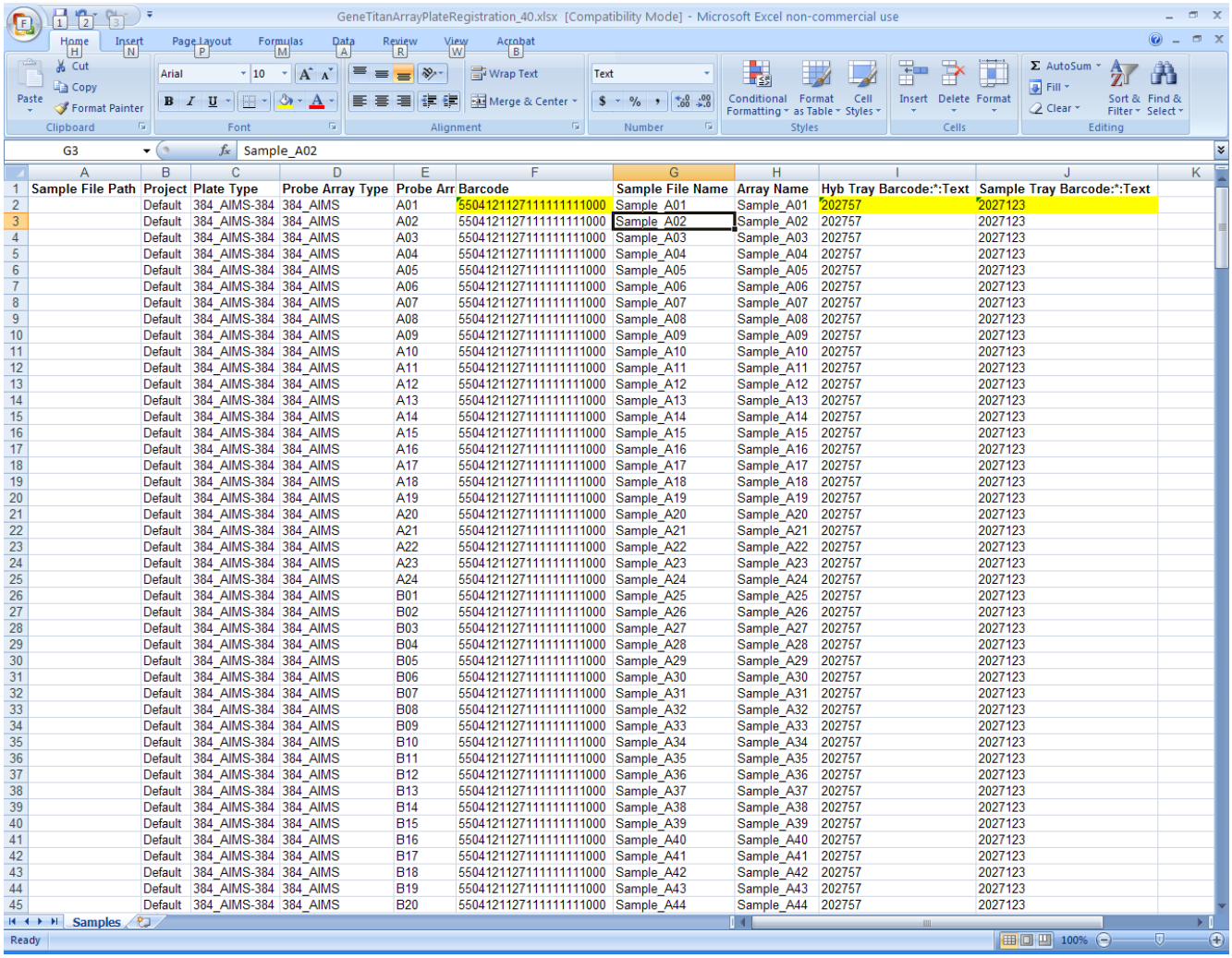
Note: The GeneTitan™ Array Plate Registration file uses the *.xls Microsoft™ Excel™ file extension. Do not use the *.xlsx file extension.

Create a GeneTitan™ Array Plate Registration file

1. In GCC Portal, click **Samples** ▸ **GeneTitan Array Plate Registration**.

2. Create a new template in GCC that includes fields required for sample traceability.
3. Select the array plate to be processed on the GeneTitan™ MC Instrument.
4. Select the newly created template that contains the fields that are required for traceability.
5. Select the **GeneTitan Array Plate Type** from the dropdown list.
6. Select the project where the sample registration data and all associated data files are saved.
7. Click **Download**.

8. Click the Microsoft™ Excel™ icon to open the spreadsheet.



	A	B	C	D	E	F	G	H	I	J	K
1	Sample File Path	Project	Plate Type	Probe Array Type	Probe Arr	Barcode	Sample File Name	Array Name	Hyb Tray Barcode:*:Text	Sample Tray Barcode:*:Text	
2		Default	384_AIMS-384	384_AIMS	A01	55041211271111111111000	Sample_A01	Sample_A01	202757	2027123	
3		Default	384_AIMS-384	384_AIMS	A02	55041211271111111111000	Sample_A02	Sample_A02	202757	2027123	
4		Default	384_AIMS-384	384_AIMS	A03	55041211271111111111000	Sample_A03	Sample_A03	202757	2027123	
5		Default	384_AIMS-384	384_AIMS	A04	55041211271111111111000	Sample_A04	Sample_A04	202757	2027123	
6		Default	384_AIMS-384	384_AIMS	A05	55041211271111111111000	Sample_A05	Sample_A05	202757	2027123	
7		Default	384_AIMS-384	384_AIMS	A06	55041211271111111111000	Sample_A06	Sample_A06	202757	2027123	
8		Default	384_AIMS-384	384_AIMS	A07	55041211271111111111000	Sample_A07	Sample_A07	202757	2027123	
9		Default	384_AIMS-384	384_AIMS	A08	55041211271111111111000	Sample_A08	Sample_A08	202757	2027123	
10		Default	384_AIMS-384	384_AIMS	A09	55041211271111111111000	Sample_A09	Sample_A09	202757	2027123	
11		Default	384_AIMS-384	384_AIMS	A10	55041211271111111111000	Sample_A10	Sample_A10	202757	2027123	
12		Default	384_AIMS-384	384_AIMS	A11	55041211271111111111000	Sample_A11	Sample_A11	202757	2027123	
13		Default	384_AIMS-384	384_AIMS	A12	55041211271111111111000	Sample_A12	Sample_A12	202757	2027123	
14		Default	384_AIMS-384	384_AIMS	A13	55041211271111111111000	Sample_A13	Sample_A13	202757	2027123	
15		Default	384_AIMS-384	384_AIMS	A14	55041211271111111111000	Sample_A14	Sample_A14	202757	2027123	
16		Default	384_AIMS-384	384_AIMS	A15	55041211271111111111000	Sample_A15	Sample_A15	202757	2027123	
17		Default	384_AIMS-384	384_AIMS	A16	55041211271111111111000	Sample_A16	Sample_A16	202757	2027123	
18		Default	384_AIMS-384	384_AIMS	A17	55041211271111111111000	Sample_A17	Sample_A17	202757	2027123	
19		Default	384_AIMS-384	384_AIMS	A18	55041211271111111111000	Sample_A18	Sample_A18	202757	2027123	
20		Default	384_AIMS-384	384_AIMS	A19	55041211271111111111000	Sample_A19	Sample_A19	202757	2027123	
21		Default	384_AIMS-384	384_AIMS	A20	55041211271111111111000	Sample_A20	Sample_A20	202757	2027123	
22		Default	384_AIMS-384	384_AIMS	A21	55041211271111111111000	Sample_A21	Sample_A21	202757	2027123	
23		Default	384_AIMS-384	384_AIMS	A22	55041211271111111111000	Sample_A22	Sample_A22	202757	2027123	
24		Default	384_AIMS-384	384_AIMS	A23	55041211271111111111000	Sample_A23	Sample_A23	202757	2027123	
25		Default	384_AIMS-384	384_AIMS	A24	55041211271111111111000	Sample_A24	Sample_A24	202757	2027123	
26		Default	384_AIMS-384	384_AIMS	B01	55041211271111111111000	Sample_A25	Sample_A25	202757	2027123	
27		Default	384_AIMS-384	384_AIMS	B02	55041211271111111111000	Sample_A26	Sample_A26	202757	2027123	
28		Default	384_AIMS-384	384_AIMS	B03	55041211271111111111000	Sample_A27	Sample_A27	202757	2027123	
29		Default	384_AIMS-384	384_AIMS	B04	55041211271111111111000	Sample_A28	Sample_A28	202757	2027123	
30		Default	384_AIMS-384	384_AIMS	B05	55041211271111111111000	Sample_A29	Sample_A29	202757	2027123	
31		Default	384_AIMS-384	384_AIMS	B06	55041211271111111111000	Sample_A30	Sample_A30	202757	2027123	
32		Default	384_AIMS-384	384_AIMS	B07	55041211271111111111000	Sample_A31	Sample_A31	202757	2027123	
33		Default	384_AIMS-384	384_AIMS	B08	55041211271111111111000	Sample_A32	Sample_A32	202757	2027123	
34		Default	384_AIMS-384	384_AIMS	B09	55041211271111111111000	Sample_A33	Sample_A33	202757	2027123	
35		Default	384_AIMS-384	384_AIMS	B10	55041211271111111111000	Sample_A34	Sample_A34	202757	2027123	
36		Default	384_AIMS-384	384_AIMS	B11	55041211271111111111000	Sample_A35	Sample_A35	202757	2027123	
37		Default	384_AIMS-384	384_AIMS	B12	55041211271111111111000	Sample_A36	Sample_A36	202757	2027123	
38		Default	384_AIMS-384	384_AIMS	B13	55041211271111111111000	Sample_A37	Sample_A37	202757	2027123	
39		Default	384_AIMS-384	384_AIMS	B14	55041211271111111111000	Sample_A38	Sample_A38	202757	2027123	
40		Default	384_AIMS-384	384_AIMS	B15	55041211271111111111000	Sample_A39	Sample_A39	202757	2027123	
41		Default	384_AIMS-384	384_AIMS	B16	55041211271111111111000	Sample_A40	Sample_A40	202757	2027123	
42		Default	384_AIMS-384	384_AIMS	B17	55041211271111111111000	Sample_A41	Sample_A41	202757	2027123	
43		Default	384_AIMS-384	384_AIMS	B18	55041211271111111111000	Sample_A42	Sample_A42	202757	2027123	
44		Default	384_AIMS-384	384_AIMS	B19	55041211271111111111000	Sample_A43	Sample_A43	202757	2027123	
45		Default	384_AIMS-384	384_AIMS	B20	55041211271111111111000	Sample_A44	Sample_A44	202757	2027123	

9. Enter a unique name for each sample in the **Sample File Name** column and any additional information, such as array plate barcode.

Note: The array plate's barcode can be scanned into the **Barcode** field. The barcode is stored in the sample file for each array.

---

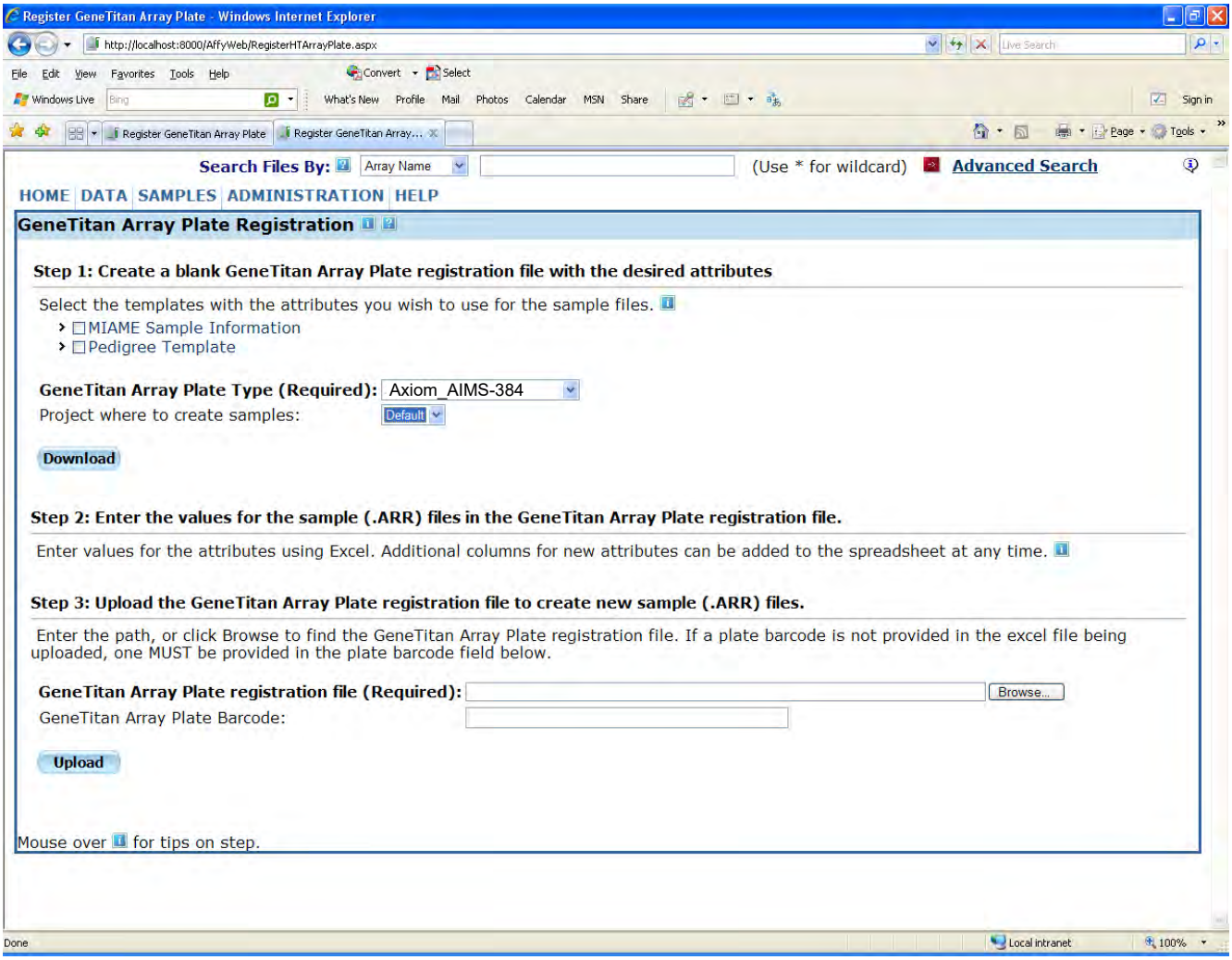
10. Complete one of the following:
    - If you are ready to load the array plate onto the GeneTitan™ MC Instrument, scan the array plate barcode into column F, then proceed to step 11.
    - If you are not ready to load the array plate onto the GeneTitan™ MC Instrument, proceed to step 11.

11. Follow these steps to save the file:
    a. Click **File** ▸ **Save As**.

    b. Enter a name for the array plate registration file.

c. Click **Save**.

12. Follow these steps when you are ready to load the array plate onto the GeneTitan™ MC Instrument.
    a. Click **Browse**, navigate to the GeneTitan™ Array Plate Registration file, then click **Open**.
    b. Scan the array plate barcode, if it has not already been scanned, and save the registration file.
    c. Click **Upload**, wait for the information to load, then click **Save** found at the bottom of the next window that is displayed.



E

Troubleshooting

- Multidrop™ Combi Reagent Dispenser 194
- Adjust Multidrop™ Combi dispense volume 195
- Thermo Scientific™ ALPS™ 3000 Automated Microplate Heat Sealer 196
- ALPS™ 3000 Automated Microplate Heat Sealer 197
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Multidrop™ Combi Reagent Dispenser

Observation	Possible cause	Recommended action
Gravimetric weight check out of range	• Cassette is not primed properly. • Wear of cassette tubing over time.	• Ensure that the cassette is primed properly before performing the gravimetric check. • Verify that each tip of the cassette is dispensing properly. • Adjust the Multidrop™ Combi dispense volume up or down, accordingly. See "Adjust Multidrop™ Combi dispense volume" on page 195.
	Wrong solution used for gravimetric checks.	Use the correct type of solution described in .
Dispensed liquid strays and does not go into the wells	Static electricity.	Ensure ion blowers are in place and powered on at the dispense station.
	Tips are clogged.	Perform the priming and backflush to remove the foreign materials inside the tips. ("Prime the cassette" on page 54. "Flush the Multidrop™ Combi Reagent Dispenser between batches of plates" on page 55.)
	Tips are damaged.	Replace the Multidrop™ Combi SMART 2 dispensing cassette.

Adjust Multidrop™ Combi dispense volume

1. Select the **Main** tab on the Multidrop™ Combi screen.
2. Select the protocol name then press **OK**.
3. Using the up/down arrows, select the dispense volume setting, then press **OK**.
4. Use the up/down arrows on the instrument to decrease or increase the dispense volume, then press **OK**.
   - For cassettes with standard tubing, the resolution of the volume adjustment is 5 µL.
   - For cassettes with small tubing, the resolution of the volume adjustment is 0.5 µL.
5. After adjusting the dispense volume on the Multidrop™ Combi, save the changes to the method.
   a. Press **OK** on the protocol name.
   b. Press the right arrow on the Multidrop to edit the protocol.
   c. Do not change the protocol name. Use the arrows on the Multidrop to select **Save** , then press **OK** to save.
   After pressing **OK**, the display goes back to the main screen.
6. Repeat the gravimetric testing to confirm that the dispense weight is within the specified limits. (See “Range guidelines for gravimetric tests” on page 52.)

Thermo Scientific™ ALPS™ 3000 Automated Microplate Heat Sealer

Observation	Possible cause	Recommended action
Seal is misaligned on the plate	The nut to position the roll of Easy Peel Seal is loose.	See "Align the ALPS™ 3000 seal roll" on page 197.
Bubble is seen on the film after sealing	Liquid spray is on the grids of the 96-deepwell plate, or the 96-deepwell plate is not flat.	Peel the seal off off of the plate, then inspect for liquid on the grid. If liquid is present, lay a clean laboratory tissue over the grid, then dab dry. Reseal the plate. If after several attempts bubbles persist, manually transfer the contents of the wells to a new 96-deepwell plate with a multichannel pipette, then continue the process. Use clean pipette tips for each transfer.
Seal is too light (light imprint or uneven imprint) or too heavy (seal material sticks to the webs of the plate during peel-off)	Temperature fluctuation of the heating element.	Increase or decrease the sealing temperature by 1°C, then check the qualitative seal strength. If proper sealing cannot be achieved within 145°C to 155°C for 2.5 seconds, contact customer support.

ALPS™ 3000 Automated Microplate Heat Sealer

Thermo Fisher Scientific recommends the ALPS™ 3000 Automated Microplate Heat Sealer for use with ultrahigh-throughput workflows, such as the Axiom™ Propel XPRES 384HT Workflow.

Align the ALPS™ 3000 seal roll

The roll of plate sealing material can drift out of alignment, resulting in a seal that doesn't cover the top or bottom of the plate.

A nut inside the spindle axle (2) holds the adjusting hub (1) in place, as shown in the left figure below. When the nut is loosened, the adjusting hub can be moved, as shown in the following figure on the right side.

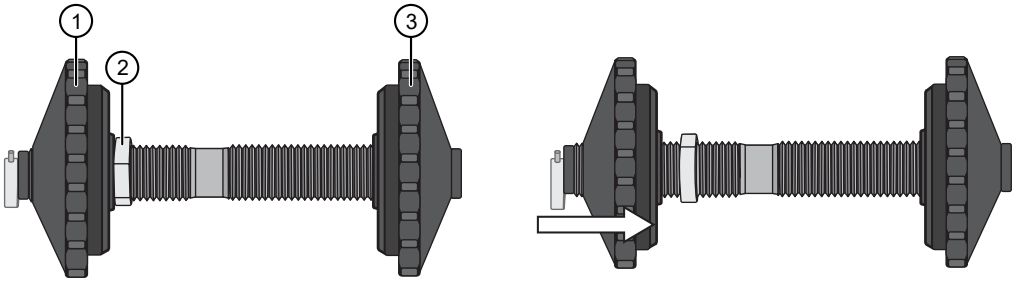


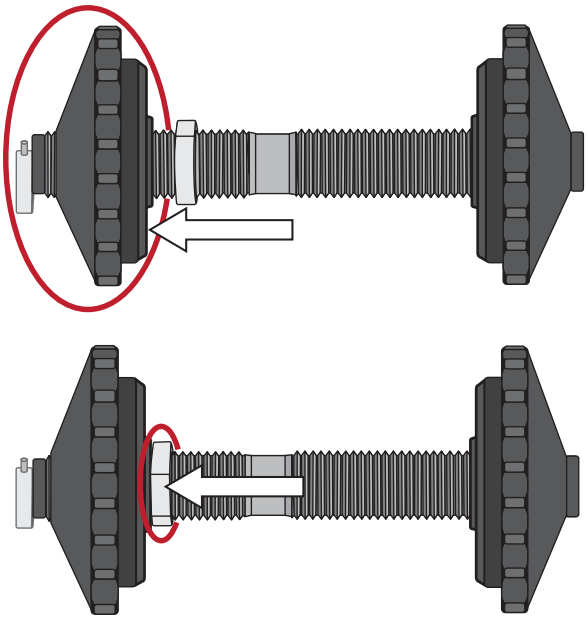
Figure 40 The spindle assembly, located in the back of the plate sealer.

1. Adjusting hub
2. Nut
3. Fixed hub

Align the seal roll—Seal does not cover bottom of the plate

If the seal does not cover the bottom of the plate (with the plate oriented so that A1 is at the back right corner), follow the steps below.

1. Remove the spindle from the back of the plate sealer.
2. Rotate the adjusting hub so that it moves to the left on the spindle axle. The nut might need to be loosened before the adjusting hub will rotate.
3. Using a wrench, lock the adjusting hub in place by tightening the nut against it.



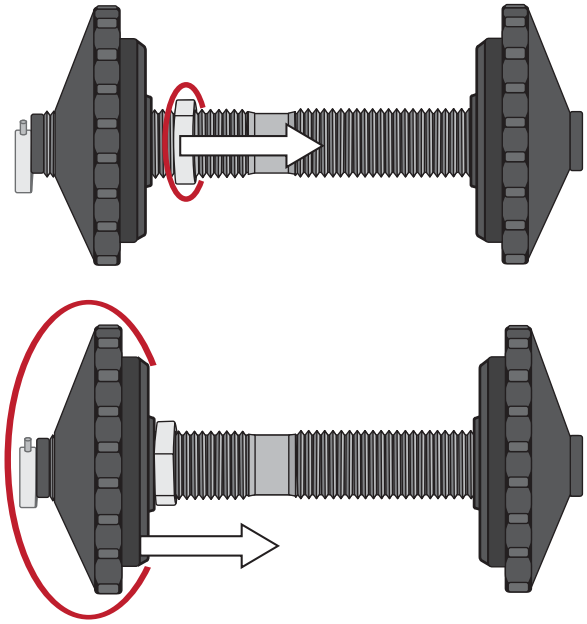
IMPORTANT! Move the adjusting hub in small increments. Test the seal after each adjustment.

Align the seal roll—Seal does not cover top of the plate

If the seal does not cover the top of the plate (with the plate oriented so that A1 is at the back right corner), follow the steps below.

1. Remove the spindle from the back of the plate sealer.
2. Rotate the adjusting hub so that it moves to the right on the spindle axle.
3. Move the locking nut to the right if necessary, as shown in left figure, before moving the adjusting hub.

4. Using a wrench, lock the adjusting hub in place by tightening the nut against it.



IMPORTANT! Move the adjusting hub in small increments. Test the seal after each adjustment.

Clean the ALPS™ 3000 Automated Microplate Heat Sealer

Periodically, clean the 2 linear slides and the 2 slide rests with 70% ethanol.

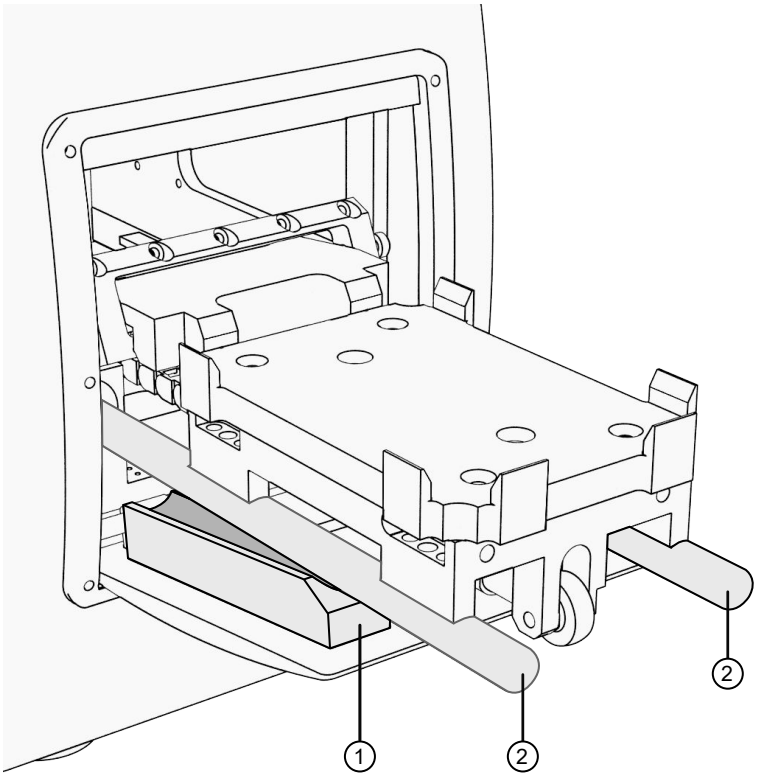


Figure 41 The ALPS™ 3000 slides and slide rests.

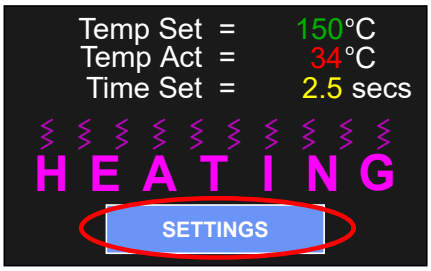
① Slide rest (left, right not shown)
② Linear slide (left and right)

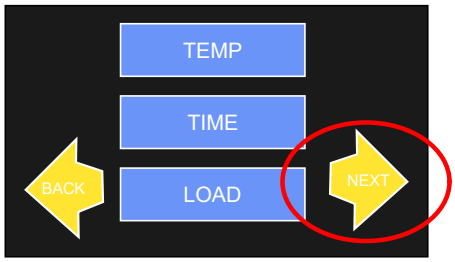
Park the ALPS™ 3000 Automated Microplate Heat Sealer

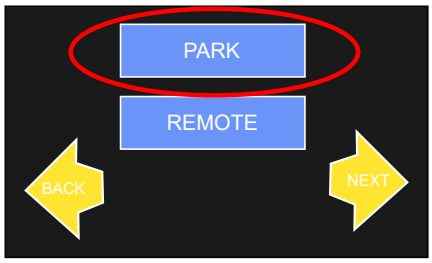
If the sealer will not be used for an extended period of time, put the sealer in the "Park" position to prevent the exposed parts from collecting dust and particles.

Before parking the instrument, ensure that there is not a plate in the plate shuttle.

1. Press **Settings** ▸ **Next** ▸ **Park**.







2. Remove the roll of sealing material, then press **NEXT**.

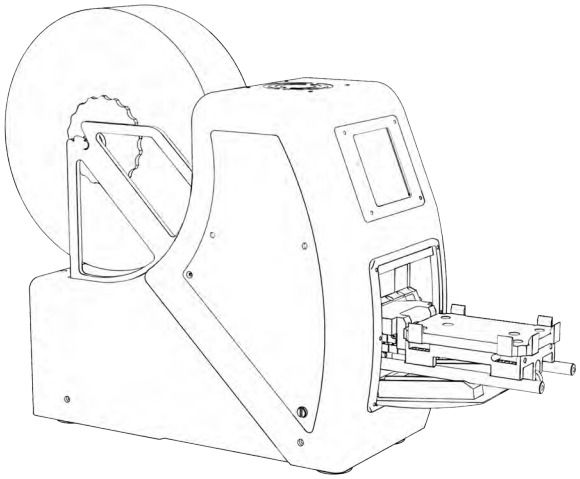
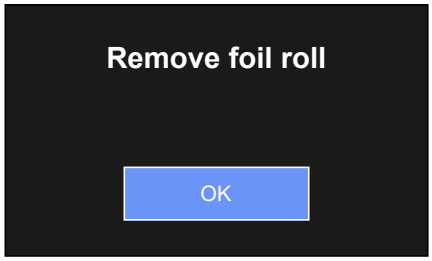


Figure 42 A roll of sealing material on the instrument.

3. After removing the roll, click **OK**.



After pressing **OK**, the plate seal nest retracts into the ALPS 3000, then the door closes.

GeneTitan™ MC Instrument support files for troubleshooting

Log files

The different GeneChip™ Command Console™ (GCC) components generate log files. The logs provide a record of the tasks performed by the different components, such as the migration tools and installer. These log files provide useful information for troubleshooting problems. Sometimes these files are required by your field application scientist (FAS), field service engineer (FSE), or the Thermo Fisher Scientific call center to help with troubleshooting.

GeneChip™ Command Console™ log files

The following files are generated by the GeneTitan™ MC Instrument. All the GCC log files are from the following path: `C:\Command_Console\Logs`.

Log file type	Description
`Systemlog.xml`	XML file with system information.
`DEC.log`	Text file with information on the use of the Data Exchange Console (DEC).
`DECError.log`	Text file with information on errors created while using DEC.

Other GeneChip™ Command Console™ files

The following GCC files and requests are sometimes used by FAS or FSE for troubleshooting.

- Library files (*.PARAMS, *.MASTER, *.WORKFLOW, *.SMD, *.MEDIA) in `C:\Command_Console\Library`, excluding the large analysis library files (CDF, PSI, GRC).
- Provide a list of all sub folders and their contents under the library files folder that is in `C:\Command_Console\Library`. Ensure that there are no duplicate library files, as these files can cause problems
- GCC system configuration file that is found at `C:\Command_Console\Configuration\Calvin.System.config`.
- Pending job order files that are in `C:\Command_Console\Jobs`
- Other GCC related information, such as
  - The number of files under `C:\Command_Console\Data`, including sub directory.
  - If the system is a networked system or a stand-alone system.
  - Other applications that are installed on the system, such as antivirus application, Microsoft™ Office™ , and Internet Explorer® versions.

GCC log files for GeneTitan™ MC Instrument systems

Log files for the GeneTitan™ MC Instrument control processes are placed in subdirectories of the `C:\Command_Console\Logs\` folder. Thermo Fisher Scientific sometimes requests the following files for troubleshooting.

GeneTitan™ MC Instrument fluidics

- `C:\Command_Console\Logs\96F\`
  - Subdirectories are named by date (for example, Log7-29-2016)

    Collect all dated directories and contents from the time the GeneTitan™ application was started, not just from the date of the event. Some logging goes into files from the date the application started so these files can be critical for troubleshooting.

    All the log directories from the date the run was started to the date of the event are essential.
- `C:\Command_Console\Logs\96F\FluidicErrorLog` - all files in this directory.

GeneTitan™ MC Instrument imaging device

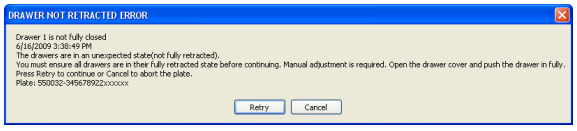
- `C:\Affymetrix\GeneChipHTScanControlMC\Log` - collect all dated directories and contents from the time the GeneTitan™ application was started.
- `C:\Affymetrix\GeneChipHTScanControlMC\RunLog` - collect all dated directories and contents from the time the GeneTitan™ application was started.

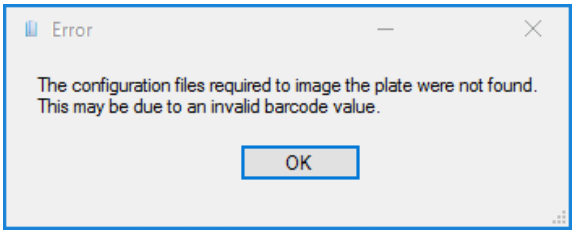
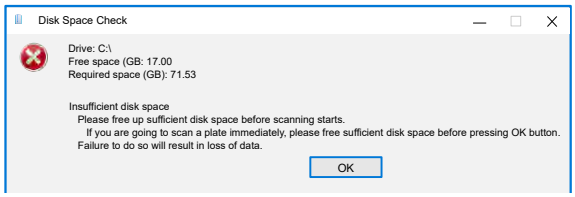
Troubleshooting the GeneTitan™ MC Instrument

This section provides instructions on how to identify and solve simple problems with the GeneTitan™ MC Instrument. If a problem or error occurs that is not listed in this chapter, contact Thermo Fisher Scientific Technical Support for help.

For software errors that do not involve hardware crashes, the most common solution is to close or exit the application, then restart it. If the same error occurs, close the application and power off the computer, then restart. If the error still occurs, power off the GeneTitan™ MC Instrument, then restart.

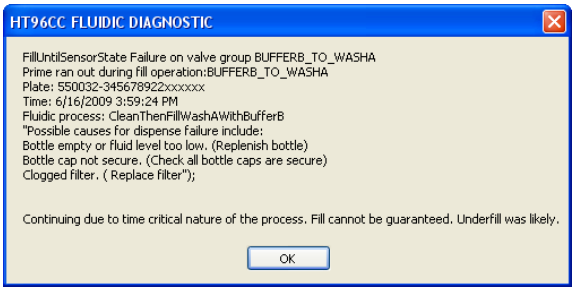
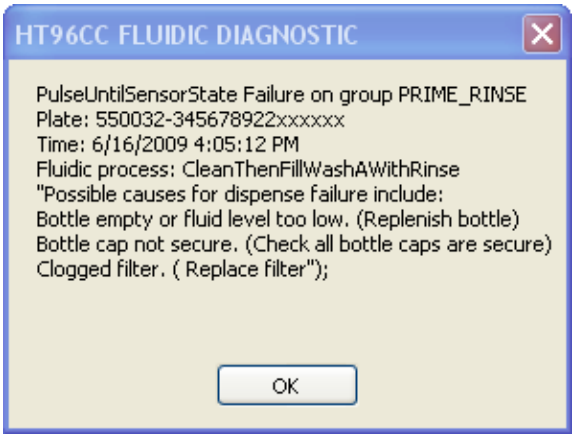
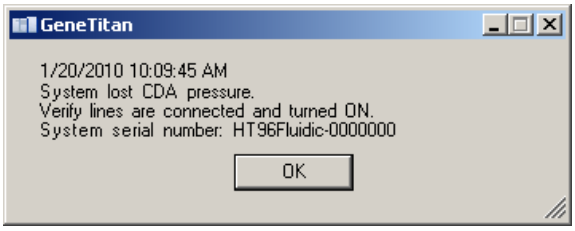
Observation	Possible cause	Recommended action
Plate trapped in the GeneTitan™ MC Instrument	• Plate (or plate with lid) not properly loaded in drawer. • Notched edge of lid and plate not aligned. • Gripper failed to retrieve plate. • System requires adjustment.	1. Restart the GeneTitan™ MC Instrument by unplugging and reconnecting power cord. 2. Run the **Unload Plates** setup option. 3. If the plate remains trapped in the instrument, call Thermo Fisher Scientific support.
Computer frozen	• Too many processes running. • Attempting to transfer data while an array plate is being scanned (imaged).	Restart the computer and unload all of the plates. • Plates in the hybridization station: finish hybridization off line. • Plate in the scanner: rescan using **Scan Only** function. • Plate in fluidics: use **Wash/Scan Resume** to resume the fluidics process. **IMPORTANT!** Do not manually, or through the GCC transfer utility, move any data associated with the current plate that is being processed/scanned.
Fluidics aborted	• System-initiated abort: power loss. • User-initiated abort: incorrect protocol selected.	Follow the recommendations and instructions under "Wash-Scan Resume" on page 163.

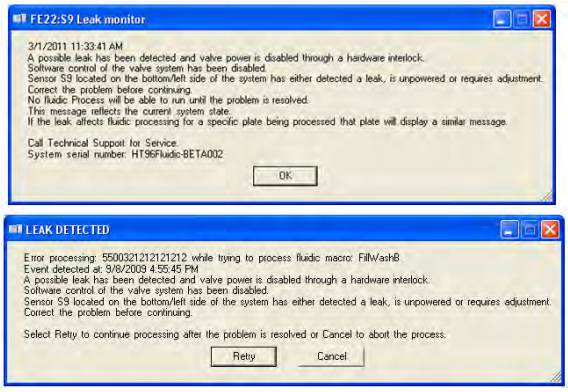
Observation	Possible cause	Recommended action
Homing recovery of gripped item 	Indicates that an item is in the gripper, and normal startup of the GeneTitan™ MC Instrument is not possible. The item must be removed from the instrument before you can start processing array plates.	Recommendation: click **Yes**. If you click **No**, nothing occurs. Homing will not complete and you will not be able to use the system. The item that is held by the gripper is moved to either: • Drawer 2—plates and trays • Trash Bin—covers The drawer names reflect the location (left or right) and the drawer number (1 through 6). Examples: Drawer2L_Hta_DOWN = Scan tray on left side of drawer 2 HtaHyb = Clamped hybridization tray and array plate Drawer(n)L/R_Hta_DOWN where n is the drawer number and L or R to indicate the left or right side. The _Hta_ (second term) indicates that the item held. An example is drawer1R_HtaHyb_DOWN indicating it is an array plate with a hybridization tray or Drawer2L_ScanHta_Pk_DOWN indicating it is an array plate with a scan tray
Drawer not retracted error 	The drawer that is listed in the message is not fully closed.	Manually push the drawer back into the instrument until it is fully closed. There are 2 stop positions with audible clicks. Push until you hear the second click and the drawer is fully seated.

Observation	Possible cause	Recommended action
Array registration error message 	The protocol file for the array plate barcode could not be found.	Check that the array plate barcode has been entered correctly. Ensure that the library files required for the type of array plate you are using have been installed, and are installed in the correct directory. Restart the GeneTitan™ Instrument control software after library files have been installed.
Insufficient disk space notice  This message appears when you first initialize the software and instrument, or when you select arrays for imaging.	There is not sufficient memory on the computer hard drive to save the data from an array plate.	Free up sufficient disk space before starting imaging with the GeneTitan™ MC Instrument.

GeneTitan™ MC Instrument fluidic diagnostic messages

Observation	Possible cause	Recommended action
Failed prime  If this message is displayed during a water wash step, array processing has been compromised. If this message is displayed during cleanup, array processing is okay, but cleanup will not be complete.	The fluid level is either too low or the bottle is empty.	Always ensure that the GeneTitan™ bottles containing Axiom™ Wash Buffer A and Axiom™ Water are above the 50% mark when setting up the system to process an array plate.
		We recommend that all 600 mL of the Axiom™ Wash Buffer B from the Axiom™ Propel XPRES Reagent Kit, 2x384HT be emptied into the GeneTitan™ Wash B bottle when setting up the system to process a plate. Using all 600 mL of the Axiom™ Wash Buffer B ensures that the bottle is filled to more than the requisite 35% of Wash B bottle volume.

Observation	Possible cause	Recommended action
Fluidics diagnostic dispense error  BUFFER*x* = Buffer bottle A, B, or Rinse. WASH*x* = Wash A or B reservoir in the fluidics station.  	Reagent bottle is empty or too low.	Replenish fluid level in the Rinse or Wash Bottle B to the 1-L mark. Do not overfill. **IMPORTANT!** Only replenish bottles when prompted by the UI. Replenishing during fluidic processing can cause system malfunction including overflowing inside the system and more problems. The only thing to do when a plate is running is to ensure that bottle caps are secure.
		Replenish fluid level in Wash Bottle A to 2 L.
	GeneTitan™ reagent bottle cap is loose.	Fasten the bottle cap.
	The GeneTitan™ reagent bottle filter is clogged.	Replace the filter. See “Bottle filter replacement” on page 212.
Loss in CDA pressure  	The instrument experienced a loss in Clean Dry Air (CDA) pressure.	Ensure that all lines are connected and turned on.
		Ensure that the facility CDA or the portable CDA compressor is in working condition. See the *GeneTitan™ Multi-Channel Instrument Site Preparation Guide* for the portable compressor model that has been verified with the GeneTitan™ MC Instrument.
		Contact your local field application specialist and notify the engineer about the error message.

Observation	Possible cause	Recommended action
Leak detected Leak checks are performed at application startup and any time a fluidic process, such as priming, filling, draining, is performed. Leak detection is a hard-wired sensor that shuts off fluid flow without software control. Leaks are normally confined to the drip pan found inside the system. 	• System malfunction. • The GCC application being manually closed using Windows™ Task Manager during a fill operation resulting in an application exit without stopping flow.	Contact Thermo Fisher Scientific support. The system cannot be used for fluidic processing until the problem is resolved.
Filter change required error message The software displays warning messages for the filter in each reagent bottle when it detects a problem or shows a trend of increased fill times during fluid fill operations. When an error is detected, a message box is displayed along with the information on the specific operation (dispense-related check or fill-related check).	One or more reagent bottle filters are clogged or worn out.	Change all 3 reagent bottle filters, even if only 1 is reported as problematic. See “Bottle filter replacement” on page 212.

GeneTitan™ Multi-Channel Instrument care

- Overview 210
- Maintenance 210
- Outer enclosure fan filters 211
- Bottle filter replacement 212
- Xenon lamp replacement in the GeneTitan™ MC Instrument 213

Overview

This chapter provides instructions on caring for and maintaining the instrument and on troubleshooting if problems arise.

- The GeneTitan™ Multi-Channel (MC) Instrument must be positioned on a sturdy level bench away from extremes in temperature and away from moving air.
- Always run a Shutdown protocol when the instrument is off or unused overnight or longer to prevent salt crystals from forming in the fluidics system.
- Always use deionized (DI) water to prevent contamination of the lines. Swap out old buffers with freshly prepared buffer at each system startup.

IMPORTANT! Before performing maintenance power off the instrument to avoid injury if an electrical malfunction occurs.

Maintenance

The GeneTitan™ family of instruments requires little in the way of customer maintenance. The instruments must be kept clean and free of dust. Dust buildup can degrade performance. Wipe the exterior surfaces clean using a mild dish detergent solution in water. Do not use ammonia-based cleaners or organic solvents such as alcohol or acetone to clean the system because they can damage the exterior surfaces.

The following tasks must be performed regularly to ensure that the imaging device remains in working order.

Monthly

Wipe down the outer surface of the imaging device with a dry cloth.

Every 6 months

1. Replace the cooling fan air filters at the rear of the instrument.
2. Replace the Micropore™ filters in the Wash A, Wash B, and Rinse bottles. If you run 4-8 plates/week, then replace the Micropore™ filters more frequently.

Outer enclosure fan filters

Cleaning schedule

The GeneTitan™ fan filter cartridge must be cleaned at least every 90 days of service. Note that in some service locations, the presence of excessive dust or particulate matter can require cleaning the cartridge more often than 90 days.

A plugged filter cartridge can cause excessive temperatures in the machine that can cause unwanted evaporation of GeneTitan™ reagents.

Figure 43 GeneTitan™ fan filter cartridge.

Clean the GeneTitan™ MC Instrument fan filter

Contact your field service engineer for GeneTitan™ fan filter ordering information when new filters are required.

Number of filters that are required per GeneTitan™ MC Instrument: 3

1. Slide the filter cartridge from the fan filter cartridge at the rear of the GeneTitan™ MC Instrument.
2. Submerse the filter in clean DI water. Rinse, then agitate gently to dislodge material.
3. Remove from water and dry with clean compressed air or towels.
4. When the filter cartridge is dry to the touch, reinstall the cartridge in the GeneTitan™ MC Instrument.

Bottle filter replacement

The bottles that are used in GeneTitan™ MC Instrument contain a filter to remove particulates that can exist in the buffers and DI water. The filters in the 3 GeneTitan™ fluidics bottles (Wash A, Wash B, and Rinse) must be replaced when the filters are clogged.

When the instrument detects an increase in the amount of time that is required to perform the fill operations, a **Filter Change Required** message window opens. The message window provides information on fluid dispense errors that were detected for any of the bottles during a dispense operation. All 3 filters must be changed when a warning is displayed for any of the 3 filters.

Note: The reagent bottles are depressurized when this warning message is displayed. It is safe to change the filters in all 3 fluidic bottles when this message is displayed.

After changing the filters in all 3 bottles using the procedure that is described in this section, press the **Yes** button to continue. If you select to ignore the error message, press the **No** button. This warning message is displayed each time GeneChip™ Command Console™ instrument control software is launched. You can also experience data quality problems when particulate matter is not trapped by the filters because they are clogged.

We recommend having 3 spare filters on hand in the event the filters must be replaced.

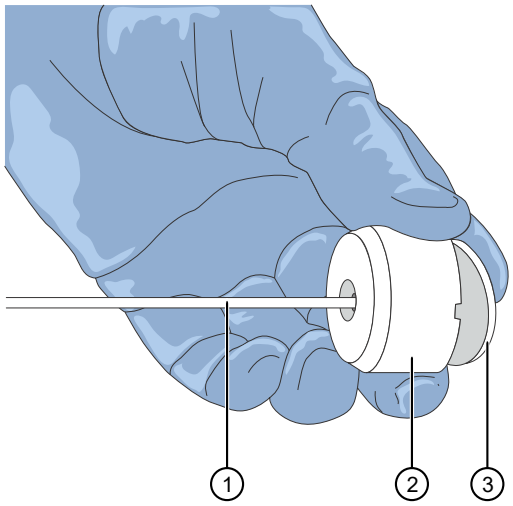


Figure 44 Components of the reagent buffer supply line and filter.

1. Buffer supply line
2. Filter holder
3. Filter

Remove and inspect the reagent bottle filters

1. Loosen, then remove the cap on the bottle.
2. Carefully remove the filter from the end of the filter body (see Figure 44).

3. Visually inspect the filter. If one of the filters appears to have a concentration of dirt or contaminate in it, discard it. Replace the filter in all 3 reagent bottles with a new one.
4. Replace the cap on the reagent bottle when finished.

Replace fluidics bottle filter

GeneTitan™ Fluidics Bottle Filter part details:

Thermo Fisher Scientific Cat. No. 01-0671

1. Loosen the reagent bottle cap, then remove the draw tube.
2. Carefully remove the filter from the end of the filter body.
3. Insert a new filter into the end of the filter holder.
4. Replace the cap on the reagent bottle, then tighten it.
5. Repeat these steps for each bottle.

IMPORTANT! Replace 1 filter at a time to ensure the correct connection of the buffer supply tube to its respective bottle. The color of the buffer supply tubing matches the bottle color code.

Xenon lamp replacement in the GeneTitan™ MC Instrument

This section applies to the GeneTitan™ MC Instrument.

After the normal life expectancy of the lamp has expired, the software application alerts you to the requirement to replace the lamp. The lamp replacement procedure is simple but good health and safety precautions must be followed.

CAUTION! Do not try to replace the lamp when a plate is being processed either in the fluidics or scanner system.

Lamp life/imaging device status notices

The **Imaging Device Status** pane displays lamp life and imaging device status notices for the GeneTitan™ MC Instrument.

In normal operation, the pane displays the hours of life that is left in the lamp.

Imaging Device Status	
Barcode	
Estimated Time Remaining	
Lamp Life Remaining	163 hours

A red or yellow notice is displayed when the lamp life is getting short.

Imaging Device Status	
Barcode	
Estimated Time Remaining	
Lamp Life Remaining	-1 hours -- Replace lamp as soon as possible

A red notice is also displayed when the imaging device is offline.

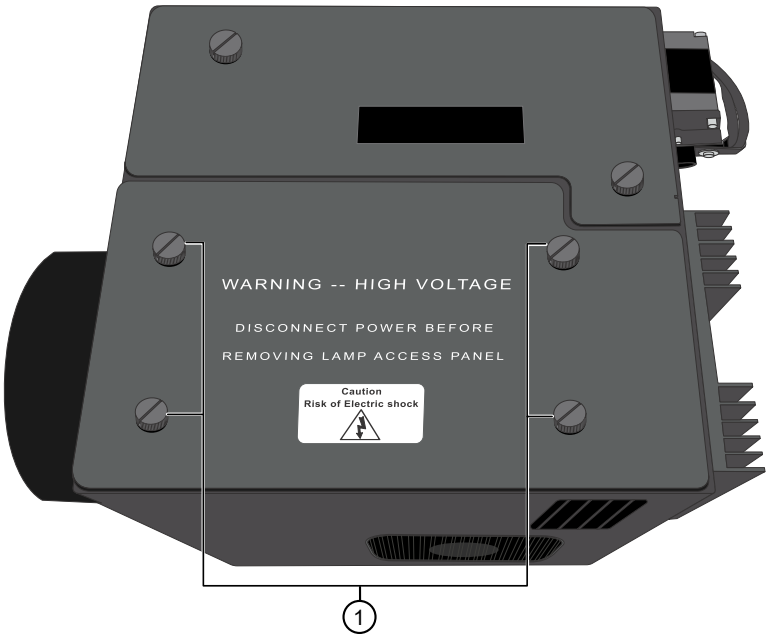
Imaging Device Status	
Barcode	
Estimated Time Remaining	
Scanner Status	Offline: scanning is not available.

Note: The 300-watt xenon lamp in the GeneTitan™ MC Instrument is warranted for 500 hours. The instructions to remove and replace the lamp are found in “Remove the xenon lamp” on page 215, and “Replace the xenon lamp” on page 216. After changing the lamp, you must manually reset the lamp life clock.

Remove the xenon lamp

WARNING! Power off the lamp using the switch in the rear of the unit and then disconnect the power cord. Allow to cool before attempting to replace the lamp.

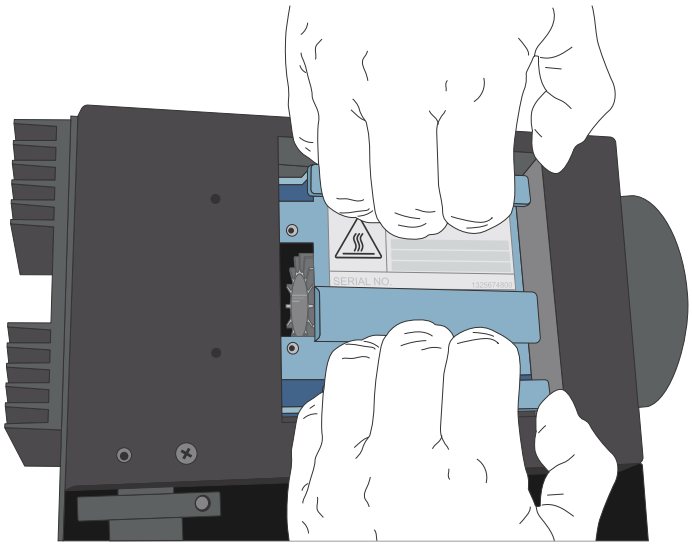
1. Unscrew the 4 retaining bolts with your fingers.



① Remove these 4 bolts.

2. Remove, then set aside the warning cover to reveal the xenon lamp that is contained inside.

3. Place a hand on each side of the blue plastic flange, then lift out the lamp in a vertical motion. Both hands must be used to remove the lamp. Apply equal pressure on each side of the lamp and gently lift.

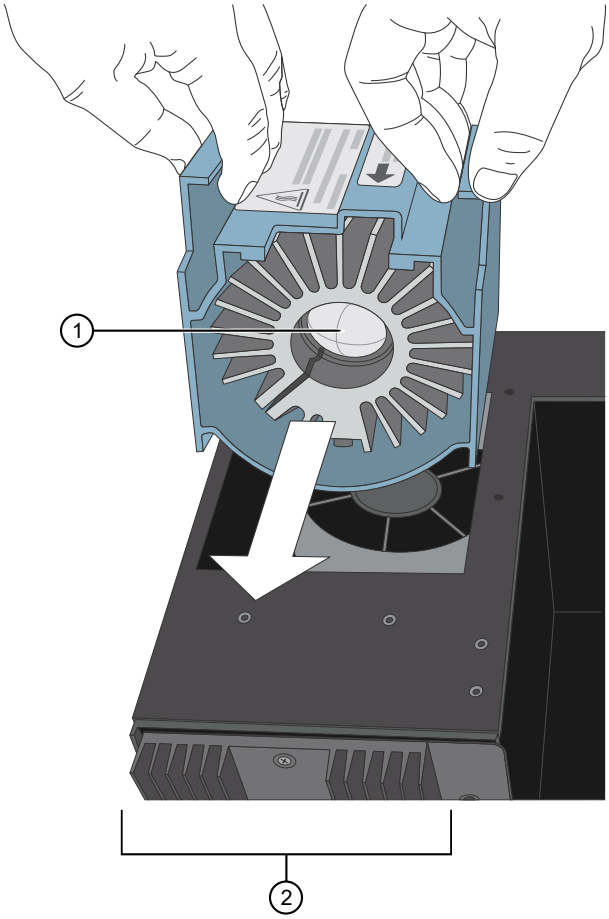


Replace the xenon lamp

A new Cermax™ Xenon Arc Lamp (Cat. No. 01-0740) is required for this procedure.

IMPORTANT! Ensure that you install the lamp in the correct orientation.

1. Hold the lamp by the blue plastic flanges. Ensure that the lamp bulb faces inward toward the rear heat sink on the unit, and then vertically lower the lamp to install.



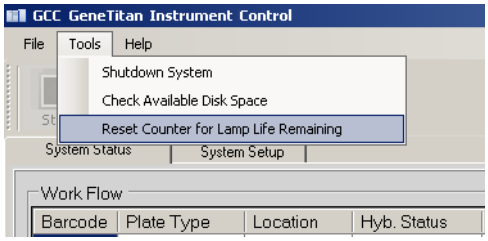
① Xenon bulb faces away from the fan and towards the heat sink.
② Heat sink on the Lambda LS unit.

2. Replace the warning cover, then hand tighten the bolts.

Reset the lamp life counter

Using the **GCC GeneTitan Instrument Control** module accessed from the **Launcher** window, you must alert the software that the lamp has been replaced so that the hours of the lamp counter are reset. This menu option is only available when the system is not processing any plates.

1. Select **Tools** ▸ **Reset Counter for Lamp Life Remaining**.



2. Click **Yes** in the message window to reset the counter.

Safety

- Symbols on this instrument 218
- Safety information for instruments not manufactured by Thermo Fisher Scientific 223
- Chemical safety 223
- Biological hazard safety 224

WARNING! GENERAL SAFETY. Using this product in a manner not specified in the user documentation may result in personal injury or damage to the instrument or device. Ensure that anyone using this product has received instructions in general safety practices for laboratories and the safety information provided in this document.

- Before using an instrument or device, read and understand the safety information provided in the user documentation provided by the manufacturer of the instrument or device.
- Before handling chemicals, read and understand all applicable Safety Data Sheets (SDSs) and use appropriate personal protective equipment (gloves, gowns, eye protection, and so on). To obtain SDSs, see the "Documentation and Support" section in this document.

Symbols on this instrument

Symbols may be found on the instrument to warn against potential hazards or convey important safety information. In this document, the hazard symbol is used along with one of the following user attention words.

- **CAUTION!**—Indicates a potentially hazardous situation that, if not avoided, may result in minor or moderate injury. It may also be used to alert against unsafe practices.
- **WARNING!**—Indicates a potentially hazardous situation that, if not avoided, could result in death or serious injury.
- **DANGER!**—Indicates an imminently hazardous situation that, if not avoided, will result in death or serious injury.

Standard safety symbols

Symbol and description
CAUTION! Risk of danger. Consult the manual for further safety information.
CAUTION! Risk of electrical shock.

(continued)

Symbol and description
CAUTION! Hot surface.
CAUTION! Potential biohazard.
CAUTION! Ultraviolet light.

Additional safety symbols

	Symbol and description
	CAUTION! Moving parts.
	CAUTION! Moving parts.
	CAUTION! Moving parts.
	CAUTION! Moving parts.
	CAUTION! Moving parts.
	CAUTION! Piercing hazard.
	CAUTION! Sharp edges.
	CAUTION! Potential slipping hazard.
	CAUTION! Potential overhead hazard.
	CAUTION!

Location of safety labels

Label and location

Figure 45 Detection cell cover (front view, door open)

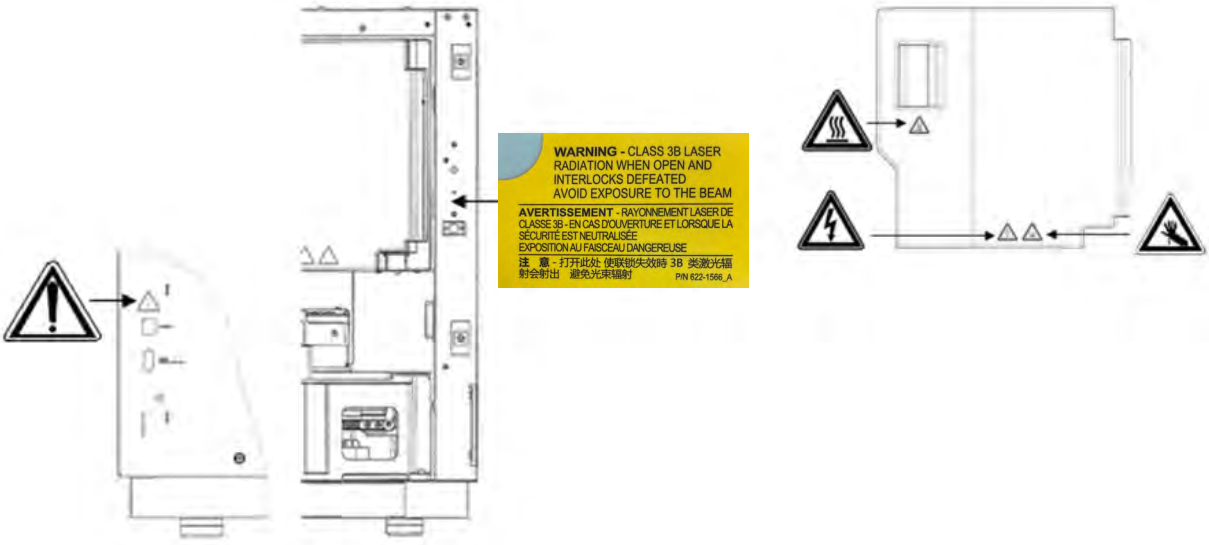



Figure 46 Rear panel (left) oven door (right)

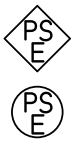
Control and connection symbols

Symbols and descriptions	
	On (Power)
	Off (Power)
	Earth (ground) terminal
	Protective conductor terminal (main ground)
	Direct current
	Alternating current
	Both direct and alternating current

Conformity symbols

Conformity mark	Description
	Indicates conformity with safety requirements for Canada and U.S.A.
	Indicates conformity with China RoHS requirements.
	Indicates conformity with European Union requirements.

(continued)

Conformity mark	Description
	Indicates conformity with Australian standards for electromagnetic compatibility.
	Indicates conformity with requirements for safety with Japanese requirements.
	Indicates conformity with the WEEE Directive 2012/19/EU. **CAUTION!** To minimize negative environmental impact from disposal of electronic waste, do not dispose of electronic waste in unsorted municipal waste. Follow local municipal waste ordinances for proper disposal provision and contact customer service for information about responsible disposal options.

Safety information for instruments not manufactured by Thermo Fisher Scientific

Some of the accessories provided as part of the instrument system are not designed or built by Thermo Fisher Scientific. Consult the manufacturer's documentation for the information needed for the safe use of these products.

Chemical safety

WARNING! GENERAL CHEMICAL HANDLING. To minimize hazards, ensure laboratory personnel read and practice the general safety guidelines for chemical usage, storage, and waste provided below. Consult the relevant SDS for specific precautions and instructions:

- Read and understand the Safety Data Sheets (SDSs) provided by the chemical manufacturer before you store, handle, or work with any chemicals or hazardous materials. To obtain SDSs, see the "Documentation and Support" section in this document.
- Minimize contact with chemicals. Wear appropriate personal protective equipment when handling chemicals (for example, safety glasses, gloves, or protective clothing).
- Minimize the inhalation of chemicals. Do not leave chemical containers open. Use only with sufficient ventilation (for example, fume hood).
- Check regularly for chemical leaks or spills. If a leak or spill occurs, follow the manufacturer cleanup procedures as recommended in the SDS.
- Handle chemical wastes in a fume hood.
- Ensure use of primary and secondary waste containers. (A primary waste container holds the immediate waste. A secondary container contains spills or leaks from the primary container. Both containers must be compatible with the waste material and meet federal, state, and local requirements for container storage.)
- After emptying a waste container, seal it with the cap provided.
- Characterize (by analysis if needed) the waste generated by the particular applications, reagents, and substrates used in your laboratory.
- Ensure that the waste is stored, transferred, transported, and disposed of according to all local, state/provincial, and/or national regulations.
- **IMPORTANT!** Radioactive or biohazardous materials may require special handling, and disposal limitations may apply.

WARNING! HAZARDOUS WASTE (from instruments). Waste produced by the instrument is potentially hazardous. Follow the guidelines noted in the preceding General Chemical Handling warning.

WARNING! 4L Reagent and Waste Bottle Safety. Four-liter reagent and waste bottles can crack and leak. Each 4-liter bottle should be secured in a low-density polyethylene safety container with the cover fastened and the handles locked in the upright position.

Biological hazard safety

WARNING! Potential Biohazard. Depending on the samples used on this instrument, the surface may be considered a biohazard. Use appropriate decontamination methods when working with biohazards.

WARNING! BIOHAZARD. Biological samples such as tissues, body fluids, infectious agents, and blood of humans and other animals have the potential to transmit infectious diseases. Conduct all work in properly equipped facilities with the appropriate safety equipment (for example, physical containment devices). Safety equipment can also include items for personal protection, such as gloves, coats, gowns, shoe covers, boots, respirators, face shields, safety glasses, or goggles. Individuals should be trained according to applicable regulatory and company/ institution requirements before working with potentially biohazardous materials. Follow all applicable local, state/provincial, and/or national regulations. The following references provide general guidelines when handling biological samples in laboratory environment.

- U.S. Department of Health and Human Services, *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, 5th Edition, HHS Publication No. (CDC) 21-1112, Revised December 2009; found at:
  https://www.cdc.gov/labs/pdf/CDC-BiosafetymicrobiologicalBiomedicalLaboratories-2009-P.pdf
- World Health Organization, *Laboratory Biosafety Manual*, 3rd Edition, WHO/CDS/CSR/LYO/2004.11; found at:
  www.who.int/csr/resources/publications/biosafety/Biosafety7.pdf

Documentation and support

Related documentation

Document	Publication number	Description
Axiom™ Propel XPRES 384HT Workflow Site Preparation Guide	MAN0018488	Provides guidance on reagents, instruments, and supplies required to run the Axiom™ Propel XPRES 384HT Workflow with the Thermo Scientific™ Multidrop™ Combi Reagent Dispenser.
Axiom™ 384HT and Mini 96 gDNA Sample Preparation Quick Reference	MAN0017719	An abbreviated reference on preparing the genomic DNA sample for 384HT and mini 96-format array plates.
GeneTitan™ MC Protocol for Axiom™ 384HT Array Plate Processing Quick Reference	MAN0017596	An abbreviated reference for processing Axiom™ 384HT Array Plates with the GeneTitan™ MC Instrument. This document is for experienced users.
Thermo Scientific™ *Multidrop™ Combi User Manual*	N05616	This document detailing the safety information, setup, use, maintenance, and troubleshooting for the Multidrop™ Combi Reagent Dispenser.
GeneTitan™ Multi-Channel Instrument User Guide	08-0308	The GeneTitan™ Multi-Channel (MC) Instrument automates array processing from target hybridization to data generation by combining a hybridization oven, fluidics processing, and state-of-the-art imaging device into a single benchtop instrument. This document detailing the use, care, and maintenance for the GeneTitan™ MC.
GeneTitan™ Multi-Channel Instrument Site Preparation Guide	08-0305	Provides guidance on creating and maintaining the proper environment required for the GeneTitan™ MC Instrument.
Thermo Scientific™ *ALPS™ 3000 Automated Laboratory Plate Sealer User Manual*	EXT0002597	Instructions on the setup and use of the ALPS™ 3000 Automated Microplate Heat Sealer.
Recommended Alternative Microarray Consumables Quick Reference	MAN0019853	A quick reference document identifying recommended alternative replacement consumables for use in microarray assays.

(continued)

Document	Publication number	Description
Software and analysis		
GeneChip™ Command Console™ User Guide	702569	This user guide provides instructions for using Applied Biosystems™ GeneChip™ Command Console™ software (GCC) used to control GeneChip™ instrument systems. GeneChip™ Command Console™ software provides an intuitive set of tools for instrument control and data management used in the processing of GeneChip™ arrays.
Axiom™ Analysis Suite User Guide	703307	Axiom™ Analysis Suite advances genotyping data analysis with a single-source software package to enable complete genotyping analysis of all Axiom™ arrays. This document provides instructions for using the software to automate the Best Practices Workflow to deliver accurate results in a single step for export in PLINK, VCF, or TXT formats.
Axiom™ Genotyping Solution Data Analysis User Guide	MAN0018363	This guide provides information and instructions for analyzing Axiom™ genotyping array data. It includes the use of Axiom™ Analysis Suite, Applied Biosystems™ Array Power Tools and SNPolisher R package to perform quality control analysis (QC) for samples and plates, SNP filtering before downstream analysis, and advanced genotyping methods.

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  - User guides, manuals, and protocols
  - Certificates of Analysis
  - Safety Data Sheets (SDSs; also known as MSDSs)

  Note: For SDSs for reagents and chemicals from other manufacturers, contact the manufacturer.

Limited product warranty

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