Cell Analysis

Advancements in multiplexed spatial phenotyping

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Abstract

Spatial omics is an expanding research area focused on integrating spatial knowledge of tissue with transcriptomics (RNA) and proteomics (protein). Understanding the complexity of the biological structure(s) is an important biological process in cancer immunotherapy research which requires accurate target classification. Translational profiling of 4+ targets on a single sample at one time can be difficult in many ways, because of the complexity involved with panel design, staining protocols, and data analysis. To design a reliable multi-target biomarker panel, the impact of protein abundance and localization, fluorophore compatibility, varying tissue types, and data characterization need to be considered, to name a few. To address the needs of high target multiplex-ability, labeling methods have advanced in cyclic detection enabling iterative labeling and detection using automated fluidics, however these approaches suffer from low simultaneous target throughput. Thermo Fisher Scientific can now provide a streamlined process for tissue labeling; a process that enables sufficient labeling with high plex panels completed within a couple of hours. We show successful detection of a wide range of various protein markers across numerous tissue organs used in various biological research applications with our uniform technique of labeling

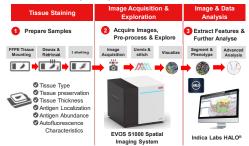
Introduction

Spatial Biology Provides Deep Insights

- Ability to analyze single cells in spatial context provides a variety of information Cellular identity in time and space
- · Cell types, cell states and cell functions
- Cell-cell interactions · Cellular neighborhoods
- Tissue microenvironments and architecture

Workflow

Immunology targets labeled with antibody-based detection, to characterize how cells interact across complex tissues imaged on a high-resolution spatial slide scanner



Cellular Phenotyping Characterization

Visualize and quantify cellular traits of innate and adaptive immune cell populations



Figure 1. Different types of FFPE Human Tonsil labeled using our IHC validated primary antibody conjugates. (A) Normal Human Tonsil consist of distinct tissue structure with a clear separation between the epithelial layer and B cells. (B) Human tonsil tumor exhibits a loss of structure organization and the and oparities and the centre of the centre o structure and a disarray of B cells.

Multiplex Labeling for Spatial Biology

Aluora Spatial Amplification Kits

Traditional signal amplification labeling technique for enhanced sensitivity. Combining the brightness of Invitrogen Aluora fluorescent dyes with polyHRP-mediated tyramide-like labeling permits high-fidelity multiplexing of a variety of validated antibody clones.

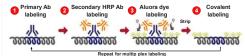


Figure 2. Multiplex Aluora spatial amlification labeling workflow uses horseradish peroxidase (HRP) to enzymatically deposit fluorophores on and surrounding protein epitope targeted primary antibody. Automated Stainer = ~12 hours: Manual staining = ~2-3 days

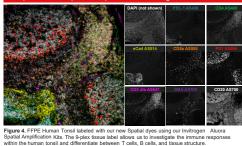
Spatial Biology Analyzing spatial relationships on tissue to investigate organization of biomolecules

communication, and how their function and behavior



Figure 3. FFPE Human Small Intestine labeled with our new Spatial dyes and Invitrogen Aluora Spatial Amplificat

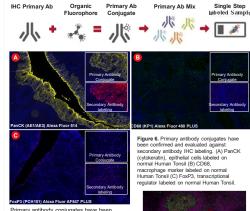
9-plex Aluora spatial amplification staining with UNMIXING





Antibodies and Fluorophores for Spatial Biology Primary Antibody Conjugate Mix

Convenient labeling method offered in a variety of fluorophores, including Alexa Fluor and Alexa Fluor PLUS dyes conjugated to highly validated IHC antibody clones for single-step labeling



including normal and certain cancerous

Primary antibody conjugates yield comparable results to those achieved with Aluora

Spatial Amplification labeling

Primary antibody conjugates have the same staining pattern with same positive and negative regions on designated tissue when compared to established IHC unconjugated antibody of the same clone

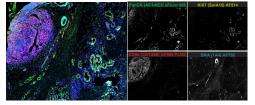


Figure 8. FFPE Human colon adenocarcinoma labeled using our Primary IHC validated antibody conjugates. The 5-plex (DAPI included) tissue label allows us to investigate the immune response within the human colon and differentiate between T cells, tissue structure, and proliferative cells.

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Primary antibody conjugates can be used in combination with signal amplification labeling methods

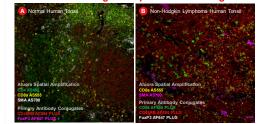


Figure 9. Both Aluora spatial amplification and primary antibody conjugates mix labeling methods were used to multiplex on a single tissue sample. The labeling process began with multiple rounds Alura dye labeling to achieve covalent fluorophore labeling on the sample, ensuring no crossreactivity with upcoming primary antibody labeling; subsequently, the primary antibody labeling mix was added in a single step. The figure above illustrates the contrasting cellular phenotypes between two tissue types (A) Normal Human Tonsil (B) Non-Hodgkin lymphoma Human Tonsil.

Consideration	Aluora Spatial Amplification	Primary Antibody Conjugate
1. Labeling time for 8-plex	12+ hours	Single step labeling ~1 hour
2. Antigen expression level	Low/Medium Abundance	Medium/High Abundance
3. Workflow Order	First	Last
4. Amplification over autofluorescence	e Yes	Wavelength dependent
5. Stripping Compatibility	No (Covalent)	Yes (Can be stripped)

Steps in determining fluorophore selection

1. Identify main target of interest 2. Determine antigen expression leve 3. Save brightest fluorophores for dimmest markers · Use on the most important target · Use on worst resolved targets · Low/unknown expression Poor access to antigen

4. Minimize spillover using kno expression patterns

Space out co-expressed marker · Mutually exclusive markers in adjacent channels

- 5. Plan for autofluorescence
- 6. Avoid using dim or low expressing targets in channels with wide spectrums The ability to resolve populations is a function of autofluorescence, background, and coexpressing markers

EVOS S1000 Spatial Imaging System

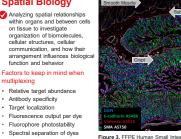
Designed to capture high-resolution images, utilizing advanced fluorescence microscopy technology for spectral imaging. Capturing images across a wide variety of channels enabling spectral unmixing for high multiplex sample analysis.

- Multiplex spectral fluorescence, transmitted brightfield.
- phase-contrast, and color brightfield Fast and Robust Imaging (<1 hour/cm²)
- 9-plex spectral unmixing (8 targets plus DAPI)
- Image up to 4 tissue slides in one sitting.
- · Robust and fast laser based autofocus
- Precise visualization of cellular structures
- · Highly intuitive graphical user interface Trademarks/licensing

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Kit reagents

Primary antibody conjugates have been validated across various tissue types.

human tissues: spleen, appendix, duodenum, tonsil, thymus, cerebellum, liver, colon etc.. Right Figure 7. Multiple primary antibody

conjugates can be combined in a single mixture for multiplexing on a single sample. Human Tonsil Multiplex staining with Primary antibody conjugates