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# Chromatography for Foods and Beverages

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## Chapter 2: Sugar Substitutes

### Introduction

A sugar substitute is a food additive that duplicates the sweetness of sugar in taste, but with fewer calories. Some sugar substitutes are natural while others are synthetic. Those that are not natural are, in general, called artificial sweeteners.

Six intensely sweet sugar substitutes are approved for use in the United States: acesulfame potassium, aspartame, neotame, saccharin, stevia, and sucralose.

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# Food Compendium: Analytical Technologies



## High-Performance Liquid Chromatography

Thermo Scientific™ Dionex™ UltiMate™ 3000 UHPLC+ systems offer excellent chromatographic performance, operational simplicity and unrivaled flexibility. Choose from a wide range of standard and unique specialty detectors to extend your laboratory's analytical capabilities.

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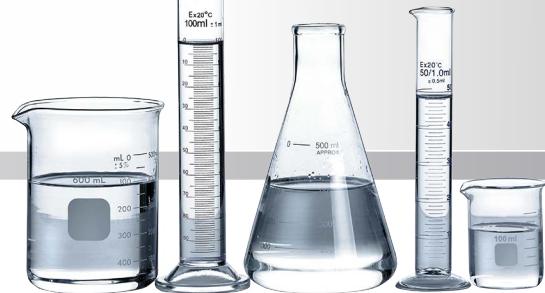
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## UltiMate 3000 UHPLC<sup>+</sup> Systems

### Best-in-class HPLC systems for all your chromatography needs

UltiMate 3000 UHPLC<sup>+</sup> Systems provide excellent chromatographic performance while maintaining easy, reliable operation. The basic and standard analytical systems offer ultra HPLC (UHPLC) compatibility across all modules, ensuring maximum performance for all users and all laboratories.

Covering flow rates from 20 nL/min to 10 mL/min with an industry-leading range of pumping, sampling, and detection modules, UltiMate 3000 UHPLC<sup>+</sup> Systems provide solutions from nano to semipreparative, from conventional LC to UHPLC.

### Superior chromatographic performance

- UHPLC design philosophy throughout nano, standard analytical, and rapid separation liquid chromatography (RSLC)
- 620 bar (9,000 psi) and 100 Hz data rate set a new benchmark for basic and standard analytical systems
- RSLC systems go up to 1000 bar and data rates up to 200 Hz
- ×2 Dual System for increased productivity solutions in routine analysis
- Fully UHPLC compatible advanced chromatographic techniques
- Thermo Scientific™ Dionex™ Viper™ and nanoViper™ fingertight fittings—the first truly universal, fingertight fitting system even at UHPLC pressures

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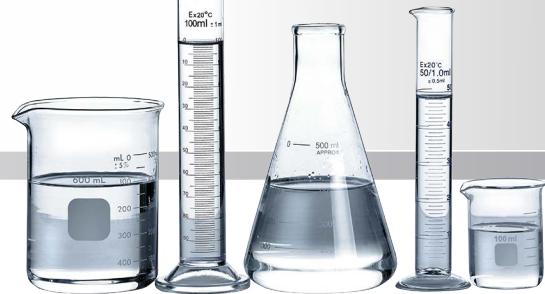
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## UltiMate 3000 UHPLC<sup>+</sup> Systems

We are uniquely focused on making UHPLC technology available to all users, all laboratories, and for all analytes.



### Rapid Separation LC Systems

The extended flowpressure footprint of the RSLC system provides the performance for ultrafast high-resolution and conventional LC applications.



### Standard LC Systems

Choose from a wide variety of standard LC systems for demanding LC applications at nano, capillary, micro, analytical, and semipreparative flow rates.



### RSLCnano Systems

The Rapid Separation nano LC System (RSLCnano) provides the power for high resolution and fast chromatography in nano, capillary, and micro LC.



### Basic LC Systems

UltiMate 3000 Basic LC Systems are UHPLC compatible and provide reliable, high performance solutions to fit your bench space and your budget.

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## UltiMate 3000 Variable Wavelength Detectors

The Thermo Scientific Dionex UltiMate 3000 VWD-3000 is a variable wavelength detector (VWD) series for industry leading UV-Vis detection. The forward optics design and wide range of available flow cells ensure optimal performance over a flow rate range of five orders of magnitude. Automated qualification, performance optimization, and instrument wellness monitoring deliver maximum uptime, simplify work-flow, and give you full confidence in your analytical results. The detector is available in a standard 100 Hz (VWD-3100) and a 200 Hz Rapid Separation version (VWD-3400RS) for the most challenging UHPLC applications.

### High-Performance UV-Vis Detection

- The VWD-3400RS variant provides data collection rates of up to 200 Hz for optimal support of today's and tomorrow's UHPLC separations
- The VWD-3100 standard detector operates at up to 100 Hz data rate for optimum support of 62 MPa (9000 psi) UltiMate 3000 Standard systems
- Superior detection of trace analytes with low noise (< -2.0 µAU) and drift (< 100 µAU/h)
- The detector's large linearity range of up to 2.5 AU is ideal for applications with widely varying analyte concentrations
- Up to four absorption channels (VWD-3400RS) and spectral scans support effective method development
- Active temperature control of optics and electronics for data acquisition independent of ambient conditions

## Standard HPLC Detectors

- Front panel access for quick and easy lamps and flow cells changes
- Automated qualification monitoring for full regulatory compliance
- Large front panel display for monitoring the detector status even from a distance
- Maximize uptime using predictive performance—based on monitoring the life cycle of detector lamps
- The detector can be upgraded with the Thermo Scientific Dionex pH/Conductivity Monitor (PCM-3000) for accurate and precise pH- and conductivity monitoring
- Unique 45 nL ultra-low dispersion UV monitor for dispersion-free UV detection in LC/MS



UltiMate 3000 VWD-3400 Variable Wavelength Detector.

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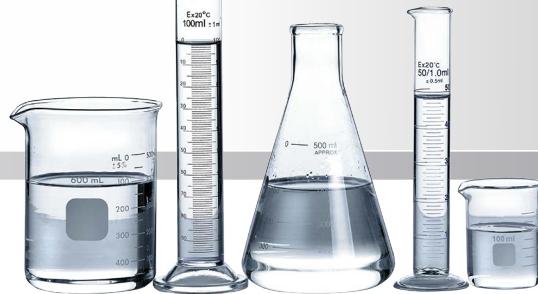
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## UltiMate 3000 Diode Array and Multiple-Wavelength Detectors

The Thermo Scientific Dionex UltiMate DAD 3000 detector is a high-resolution, 1024-element diode array detector (DAD) available in Rapid Separation (200 Hz) and Standard (100 Hz) versions. It operates with the Thermo Scientific™ Dionex™ Chromeleon™ Chromatography Data System (CDS) software to provide a variety of spectra views, including 3-D plotting and automated chromatogram handling. The high resolution and low-noise performance of the DAD-3000 family makes it ideal for the most sensitive and accurate library searches and peak purity analyses.

The detector is also available as a multiple wavelength detector (MWD) in Standard (100 Hz) and Rapid Separation (200 Hz) versions.

- Data collection at up to 200 Hz using a maximum of eight single-wavelength data channels and one 3-D field (3-D only with DAD-3000 (RS)) for best support of ultrafast separations
- Standard versions operate at up to 100 Hz data collection rate for optimum support of 62 MPa (9000 psi) UltiMate 3000 Standard systems
- Accurate compound confirmation with a 1024-element, high resolution photodiode array
- Flexibility in both UV and Vis applications with 190–800 nm wavelength range
- Low-noise over the full spectral range using deuterium and tungsten lamps
- Fast and accurate wavelength verification using a built-in holmium oxide filter

## Standard HPLC Detectors

- The detector can be upgraded with the UltiMate PCM 3000 for accurate monitoring pH gradients
- Excellent reliability and reproducibility with low baseline drift (typically < 500 µAU/h)
- Simplified routine maintenance with front access to pre-aligned cells and lamps
- ID chips on flow cells and lamps for identification and life-span monitoring
- Chromeleon CDS software for full control and flexible data handling
- Front-panel display for easy monitoring of detector status to maximize uptime
- Flow cells for semi-micro, semi-analytical, analytical, and semi-preparative applications
- Flow cells available in stainless steel and biocompatible versions



UltiMate 3000 DAD-3000 Diode Array Detector

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## RefractoMax 521 Refractive Index Detector

The Thermo Scientific RefractoMax 521 Refractive Index Detector from ERC Inc. This detector, in combination with the UltiMate 3000 system, is the right choice for the isocratic analysis of sugars, polymers, and fatty acids. It features fast baseline stabilization and excellent reproducibility, combined with high sensitivity. The RefractoMax 521 is fully controlled by the Chromeleon CDS, and can also operate in stand-alone mode.

- The detector is highly sensitive and applicable universally. It provides very stable baselines with a drift of 0.2 µRIU/h and a noise specification of 2.5 nRIU or less
- The optical bench, thermostatically regulated from 30 °C to 55 °C, and the superior signal-to-noise ratio ensure highly precise measurement results

## Standard HPLC Detectors

- The extended flow rate range from 1 mL/min up to 10 mL/min and the operating range of 1.00 to 1.75 RIU enable the use of this detector for a wide range of applications
- Applications include the analysis of all compounds with low UV-Vis activity, such as alcohols, mono- and polysaccharides, esters, fatty acids, or polymers
- An Auto Set-up function automates purging, equilibration, autozero, and the control baseline stability and noise
- Operation with Chromeleon CDS makes the detector easy to use and ensures maximum productivity in instrument control, data processing, and reporting of results



RefractoMax 521 Refractive Index Detector

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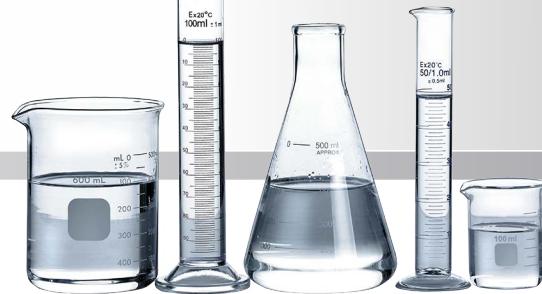
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## Corona Veo Charged Aerosol Detector

Charged Aerosol Detection provides near universal detection independent of chemical structure for non- or semi-volatile analytes with HPLC and UHPLC. A Thermo Scientific™ Dionex™ Corona™ Veo™ Charged Aerosol detector is ideally suited as a primary detector for any laboratory, while providing complementary data to UV or MS methods. No other LC detector available today can match the performance of a Corona Veo detector.

- High sensitivity – single-digit nanogram on column
- Consistent response – independent of chemical structure
- Wide dynamic range – to four orders of magnitude or greater
- Simple to use – easy to integrate with any HPLC/UHPLC system

The Corona Veo detector gives the simplicity, reproducibility and performance required for a full range of applications from basic research to manufacturing QC/QA. With charged aerosol detection you get predictable responses to measure analytes in direct proportion to their relative amounts for quantitation without actual standards.

This detector offers the flexibility to use reversed-phase gradients, as well as normal phase and HILIC modes of separation on any LC system. And, in many cases eliminates the need for derivatization or sample pre-treatment to provide real dilute-and-shoot simplicity.

## Specialty HPLC Detectors



Corona Veo Charged Aerosol Detector

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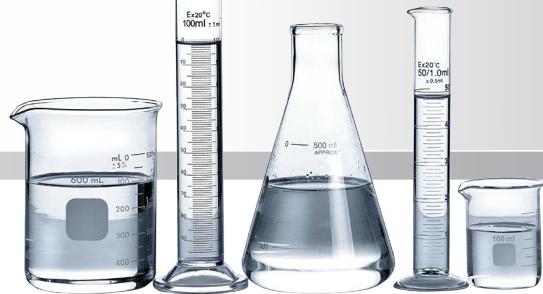
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## Ultimate 3000 Electrochemical Detector

Electrochemical detection delivers high sensitivity for neurotransmitter analysis, simplicity and robustness for pharmaceutical or clinical diagnostics, and the selectivity for the characterization of complex samples such as natural products, biological tissues and fluids. For today's researcher, there is a continuing need for detecting vanishingly small quantities of analyte and often in complex samples. Because electrochemical detection measures only compounds that can undergo oxidation or reduction it is both highly sensitive and very selective.

The Thermo Scientific Dionex UltiMate 3000 Electrochemical Detector, designed by the pioneers of coulometric electrochemical detection, delivers state-of-the-art sensor technologies complete with an entire range of high performance and ultra-high performance LC systems optimized for electrochemical detection. The UltiMate 3000 ECD-3000RS takes electrochemical detection to the next level with UHPLC compatibility, total system integration, and selection of detection mode, all with unprecedented operational simplicity.

## Specialty HPLC Detectors

### Features include:

- Detection Modes – choose from DC and PAD for optimum analyte response
- Choice of sensors – both coulometric and amperometric sensors to meet the demands of any application
- UHPLC compatibility – ultralow peak dispersion and high data acquisition rates for conventional or fast, high resolution chromatography
- Modularity – easily expandable to multiple independent sensors for unrivaled flexibility
- Autoranging – simultaneously measure both low and high levels of analytes without losing data
- SmartChip™ technology – easy operation with automatic sensor recognition, event logging and electrode protection



UltiMate 3000 Electrochemical Detector

Learn more at [www.thermoscientific.com/ECDetection](http://www.thermoscientific.com/ECDetection)

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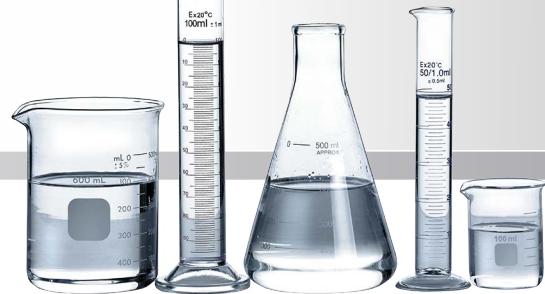
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## CoulArray Multi-electrode Array Detector

The Thermo Scientific™ Dionex™ CoulArray™ Multi-electrode Array detector is the only practical multi-channel electrochemical detection system that allows you to measure multiple analytes simultaneously, including those that are chromatographically unresolved. The CoulArray detector delivers the widest dynamic range of any available electrochemical detector with unmatched selectivity for detection of trace components in complex matrixes, even when used with aggressive gradients.

- Measures analytes from femtomole to micromole levels
- Greatly simplify sample preparation and eliminate interferences
- Simultaneously analyze multiple analytes in very complex samples
- Easily produce qualitative information for compound identification

Multiple system configurations offer 4, 8, 12, or 16 channels that can be upgraded anytime. The unique data acquisition and processing software uses automatic signal ranging and a unique patented baseline correction algorithms to provide identification and quantitation of single or multiple analytes and powerful 3D data for quick sample fingerprint confirmation with integration to pattern recognition platforms.

With the power of coulometric array technology, the CoulArray detector can give you the qualitative data of a optical PDA with 1,000 fold greater sensitivity to profile the characteristic qualities of products, determine integrity, identify adulteration and even evaluate competitors' products.

## Specialty HPLC Detectors



CoulArray Multi-electrode Array Detector

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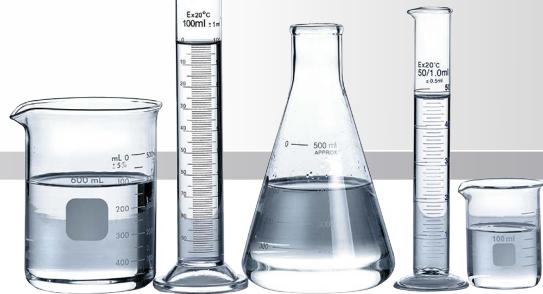
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## Ultimate 3000 Fluorescence Detector

The Thermo Scientific Dionex UltiMate 3000 FLD-3000 is a high-sensitivity fluorescence detector series for UltiMate 3000 HPLC systems. It is available in Rapid Separation (RS) and Standard (SD) versions. The optics of the FLD-3000 series provide maximum stray-light suppression for best detection sensitivity. Operated with the Chromeleon CDS software, the detector provides automated qualification, various tools for method development, and instrument wellness monitoring for ease of use, maximum uptime, and the highest degree of regulatory compliance.

- Data collection at up to 200 Hz for optimal support of even the fastest UHPLC separations (FLD-3400RS)
- Standard detectors operate at up to 100 Hz data rate for optimum support of 62 MPa (9,000 psi) UltiMate 3000 standard systems
- Lowest limits of detection with a Raman signal-to-noise ratio (S/N): > 550 ASTM (> 2100 using dark signal as noise reference)

## Specialty HPLC Detectors

- Unsurpassed reproducibility with active flow cell temperature control for stable fluorophore activity independent of changes in ambient temperature
- Long-life xenon flash lamp for highest sensitivity and long-term operation without the need for frequent lamp changing
- Optional second photomultiplier (PMT) for unique Dual-PMT operation, offering an extended wavelength range up to 900 nm without sacrificing sensitivity in the standard wavelength range
- Two-dimensional (2D) or three dimensional (3D) excitation, emission, or synchro scans to provide the highest degree of flexibility for method development or routine sample characterization
- Innovative Variable Emission Filter for real-time compound-related sensitivity optimization (FLD-3400RS only)
- Large front-panel display for easy monitoring of the detector status
- Two flow-cell sizes for easy optimization to application requirements: the 8 µL flow cell is ideal for trace analysis, and the 2 µL flow cell offers best peak resolution with narrow-bore HPLC and UHPLC columns



Ultimate 3000 Fluorescence Detector

Learn more at [www.thermoscientific.com/liquidchromatography](http://www.thermoscientific.com/liquidchromatography)

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# Food Compendium: Analytical Technologies



## Ion Chromatography

Thermo Scientific Dionex IC systems have led the analytical instrument industry for over 30 years with solutions that represent state-of-the art technological advancements and patented technologies.

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## Innovative Ion Chromatography Solutions

Our High-Pressure™ Ion Chromatography (HPIC™) systems include the Thermo Scientific Dionex ICS-5000+ HPIC system, which is optimized for flexibility, modularity, and ease-of-use, combining the highest chromatographic resolution with convenience. In addition, the Thermo Scientific Dionex ICS-4000 Capillary HPIC system is the world's first commercially available dedicated capillary high-pressure Reagent-Free™ (RFIC™) IC system. The Dionex ICS-4000 system is always ready for the next analysis, delivering high-pressure IC on demand.

Reagent-Free IC systems eliminate daily tasks of eluent and regenerant preparation in turn saving time, preventing errors, and increasing convenience. RFIC-EG systems use electrolytic technologies to generate eluent on demand from deionized water, and to suppress the eluent back to

pure water to deliver unmatched sensitivity. RFIC-ER systems are designed to use carbonate, carbonate/ bicarbonate, or MSA eluents for isocratic separations.

At the heart of our ion chromatography portfolio is a unique set of column chemistries that provide high selectivities and efficiencies with excellent peak shape and resolution. Thermo Scientific™ Dionex™ IonPac™ chromatography columns address a variety of chromatographic separation modes including ion exchange, ion exclusion, reversed-phase ion pairing, and ion suppression. Our column chemistries are designed to solve specific applications, and we offer a variety of selectivities and capacities for simple and complex samples. Additionally, our Dionex IonPac column line is available in standard bore, microbore and capillary formats for the ultimate application flexibility.



Thermo Scientific Dionex IC instrument family

Learn more at [www.thermoscientific.com/IC](http://www.thermoscientific.com/IC)

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# Food Compendium: Analytical Technologies



## Mass Spectrometry

Thermo Fisher Scientific provides advanced integrated IC/MS and LC/MS solutions with superior ease-of-use and modest price and space requirements. UltiMate 3000 System Wellness technology and automatic MS calibration allow continuous operation with minimal maintenance. The Dionex ion chromatography family automatically removes mobile phase ions for effort-free transition to MS detection.

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## Single-Point Control and Automation

Thermo Fisher Scientific provides advanced integrated IC/MS and LC/MS solutions with superior ease-of-use and modest price and space requirements. UltiMate 3000 System Wellness technology and automatic MS calibration allow continuous operation with minimal maintenance. The Dionex ion chromatography family automatically remove mobile phase ions for effort-free transition to MS detection.

- Thermo Scientific™ MSQ Plus™ mass spectrometer, the smallest and most sensitive single quadrupole on the market for LC and IC
- Self-cleaning ion source for low maintenance operation

## Mass Spectrometry Instruments

- Chromeleon CDS software for single-point method setup, instrument control, and data management compatible with existing IC and LC methods
- The complete system includes the MSQ Plus mass spectrometer, PC data system, electrospray ionization (ESI) and atmospheric pressure chemical ionization (APCI) probe inlets, and vacuum system

Now, you no longer need two software packages to operate your LC/MS system. Chromeleon CDS software provides single-software method setup and instrument control; powerful UV, conductivity, and MS data analysis; and fully integrated reporting.



MSQ Plus Mass Spectrometer

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# Food Compendium: Analytical Technologies



## Chromatography Data Systems

Tackle chromatography management challenges with the world's most complete chromatography software. Whether your needs are simple or complex or your scope is a single instrument, a global enterprise, or anything in between – the combination of Chromleon CDS' scalable architecture and unparalleled ease-of use, makes your job easy and enjoyable with one Chromatography Data System for the entire lab.

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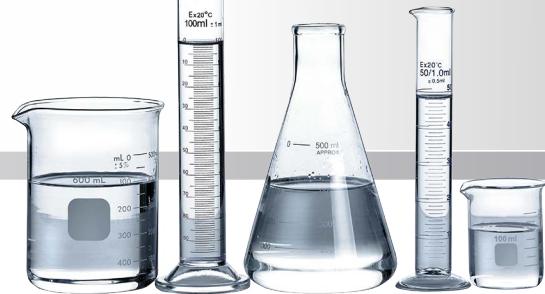
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## The Fastest Way from Samples to Results

The 7.2 release of Chromeleon Chromatography Data System software is the first CDS that combines separation (GC/IC/LC) and Mass Spectrometry (MS) in an enterprise (client/server) environment. By extending Chromeleon 7.2 CDS beyond chromatography into MS, lab technicians can now streamline their chromatography and MS quantitation workflows with a single software package. MS support in Chromeleon 7.2 CDS is focused on routine and quantitative workflows, which provides access to rich quantitative data processing and automation capabilities — ultimately boosting your overall lab productivity and increasing the quality of your analytical results.

## Chromeleon CDS Software

- Enjoy a modern, intuitive user interface designed around the principle of operational simplicity
- Streamline laboratory processes and eliminate errors with eWorkflows™, which enable anyone to perform a complete analysis perfectly with just a few clicks
- Access your instruments, data, and eWorkflows instantly in the Chromeleon Console
- Locate and collate results quickly and easily using powerful built-in database query features
- Interpret multiple chromatograms at a glance using MiniPlots
- Find everything you need to view, analyze, and report data in the Chromatography Studio
- Accelerate analyses and learn more from your data through dynamic, interactive displays
- Deliver customized reports using the built-in Excel compatible spreadsheet



**CHROMELEON 7.2**  
*Simply Intelligent*

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# Food Compendium: Analytical Technologies



## Process Analytical Systems

Thermo Scientific Dionex process analytical systems provide timely results by moving chromatography-based measurements on-line.

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## Process Analytical Systems and Software

### Improved Process Monitoring with On-line Chromatography IC and LC Systems

Information from the Thermo Scientific Dionex Integral process analyzer can help reduce process variability, improve efficiency, and reduce downtime. These systems provide comprehensive, precise, accurate information faster than is possible with laboratory-based results. From the lab to the factory floor, your plant's performance will benefit from the information provided by on-line LC.

- Characterize your samples completely with multicomponent analysis
- Reduce sample collection time and resources with automated multipoint sampling
- Improve your process control with more timely results
- See more analytes with unique detection capabilities
- The Thermo Scientific Integral Migration Path approach lets you choose the systems that best meets your needs



Integral process analyzer

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# Food Compendium: Analytical Technologies



## Automated Sample Preparation

Solvent extractions that normally require labor-intensive steps are automated or performed in minutes, with reduced solvent consumption and reduced sample handling using the Thermo Scientific™ Dionex™ ASE™ Accelerated Solvent Extractor system or Thermo Scientific™ Dionex™ AutoTrace™ 280 Solid-Phase Extraction instrument.

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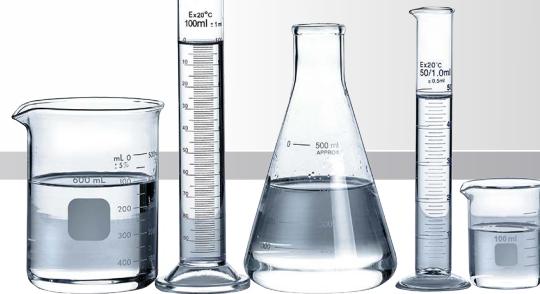
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## Accelerated Solvent Extractor System

### Complete Extractions in Less Time Using Less Solvent

Thermo Scientific Dionex ASE systems extract of solid and semisolid samples using common solvents at elevated temperature and pressure. The Dionex ASE 150 and 350 systems feature pH-hardened pathways with Dionium™ components to support extraction of acidic or alkaline matrices, and combine pretreatment, solvent extraction, and cleanup into one step. Dionium is zirconium that has undergone a proprietary

hardening process that makes it inert to chemical attack by acids and bases at elevated temperatures.

Dionex ASE systems are dramatically faster than Soxhlet, sonication, and other extraction methods, and require significantly less solvent and labor. Accelerated solvent extraction methods are accepted and established in the environmental, pharmaceutical, foods, polymers and consumer product industries. Accelerated solvent extraction methods are accepted and used by government agencies worldwide.



Dionex ASE 150/350 and Dionex AutoTrace 280 SPE instruments

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## Chapter 2: Sugar Substitutes

### Types of Sugar Substitutes

The majority of sugar substitutes approved for food use by the FDA are artificial. However, a growing number of natural sugar substitutes are now commercially available. Stevia components and mogrosides are obtained from plants. Although sorbitol and xylitol are found in berries, fruits, vegetables, and mushrooms, their extraction is not feasible so these are produced industrially by the catalytical hydrogenation of appropriate sugar starting materials.

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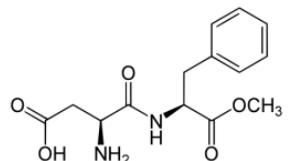
Sorbitol and Xylitol

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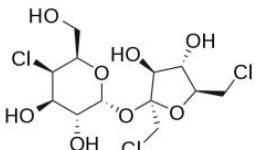
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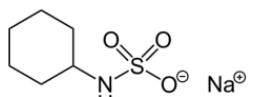
## Artificial Sweeteners



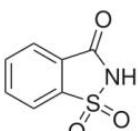
Aspartame



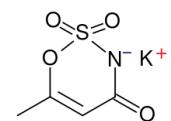
Sucratose



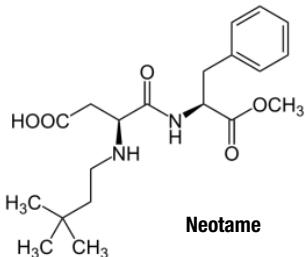
Sodium Cyclamate  
(Banned)



Saccharin



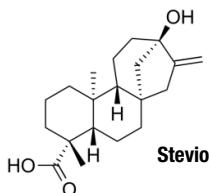
Acesulfam Potassium



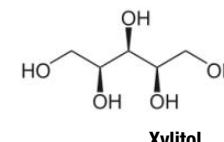
Neotame

## Structures of Sugar Substitutes

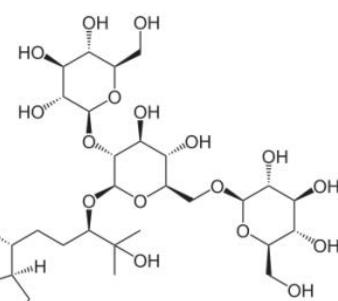
## Natural Sweeteners



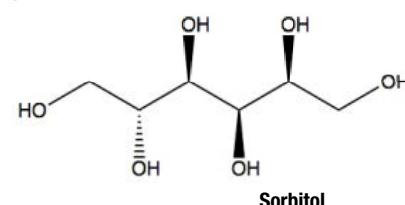
Steviol



Xylitol



Mogroside V



Sorbitol

Figure 2-1. Chemical structures of various artificial sweeteners.

Figure 2-2. Chemical structures of various natural sweeteners.

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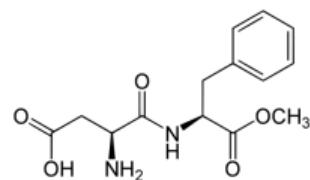
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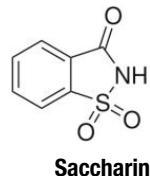


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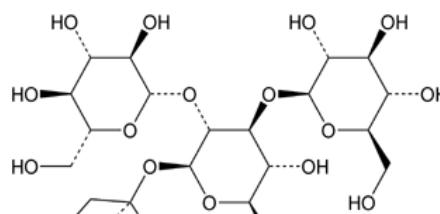
### Absorbance Properties of Common Sweeteners



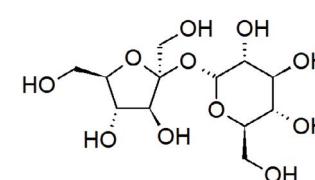
Aspartame



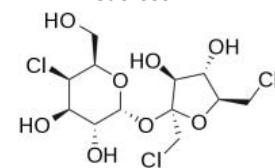
Saccharin



Rebaudioside A



Sucrose



Sucratose

Strong UV

Weak

None

Figure 2-3. Absorbance properties of common sweeteners. Weak and no UV absorbance compounds need an alternative detection technique.

### Did You Know?

Lead acetate (sometimes called *sugar of lead*) is an artificial sugar substitute made from lead that was widely used by ancient Romans. Lead acetate was abandoned as a food additive throughout most of the world after the high toxicity of lead compounds became apparent.

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### Stevia

In December 2008, the U.S. FDA recognized rebaudioside A purified from *Stevia rebaudiana* (Bertoni) as “Generally Recognized as Safe” (GRAS) for use as a sugar substitute in foods. Since this recognition, stevia products have become popular as table-top and beverage sweeteners.

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## Stevia

Although the stevia plant and extracts from stevia leaves have long been used as sweeteners in Asia and Latin America, the terpene glycosides have different flavor profiles with both sweet and unpleasant bitter flavors. Two steviol glycosides, stevioside and rebaudioside A, are largely responsible for the desired sweet flavor of the leaves, with rebaudioside A preferred for sweeteners.

Steviol glycoside determination is challenging for multiple reasons. The structures of the steviol glycosides are quite similar, differing in small changes in glycosylation. For example, rebaudioside B, an impurity

that can be formed during processing of the leaves, differs in structure from rebaudioside A primarily by the presence or absence of a glucose residue at the R1 position on the terpene. These structural similarities make chromatographic separation difficult. In addition to the separation challenges, sensitive detection of these compounds also can be difficult. They do not absorb strongly in the UV, and typical detection wavelengths for steviol glycosides, such as 210 nm, are nonspecific.

Application Note 293 describes an HPLC-charged aerosol detection method capable of measuring all relevant analytes in Stevia.

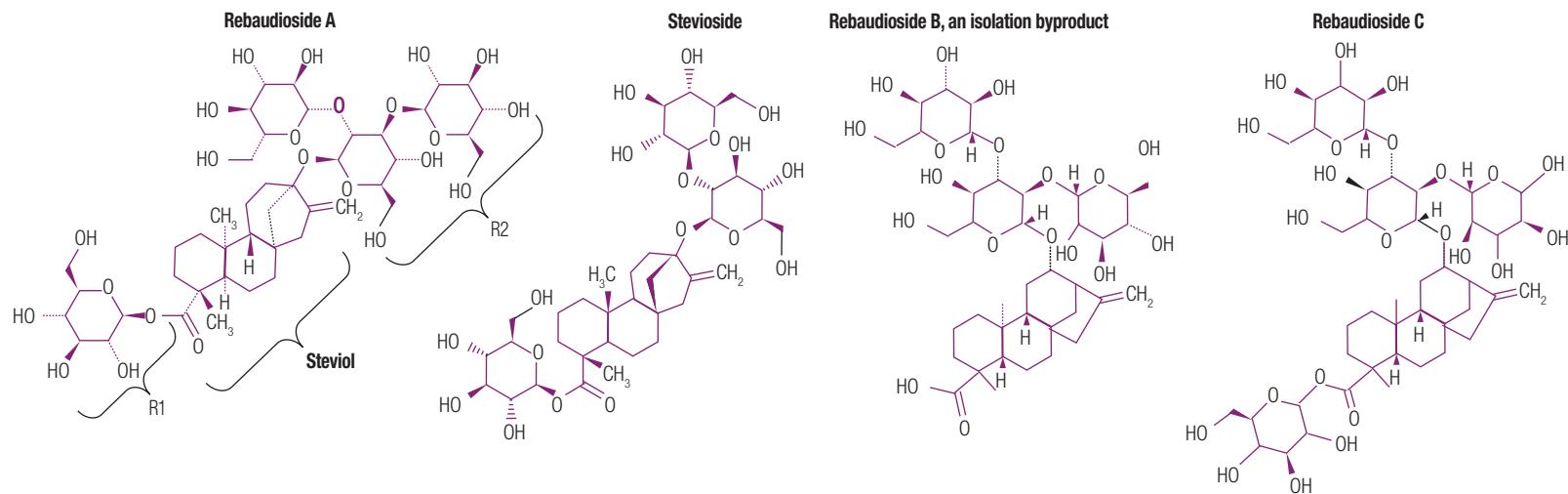


Figure 2-4. Chemical structures of the steviol glycosides stevioside and rebaudioside A, rebaudioside B, and rebaudioside C.

**Download Application Note 293: Steviol Glycoside Determination by HPLC with Charged Aerosol and UV Detections Using the Acclaim Trinity P1 Column**

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Column: Acclaim Trinity P1, 2.1 × 100 mm and guard  
Flow Rate: 0.3 mL/min  
Temperature: 20 °C  
Injection Volume: 5 µL  
Mobile Phase: 81/19 acetonitrile/10 mM ammonium formate, pH = 3.00  
Detection: A) UV detection, 210 nm  
B) Charged aerosol detection, nebulizer temp. 10 °C  
Sample: Brand A extracted sweetener

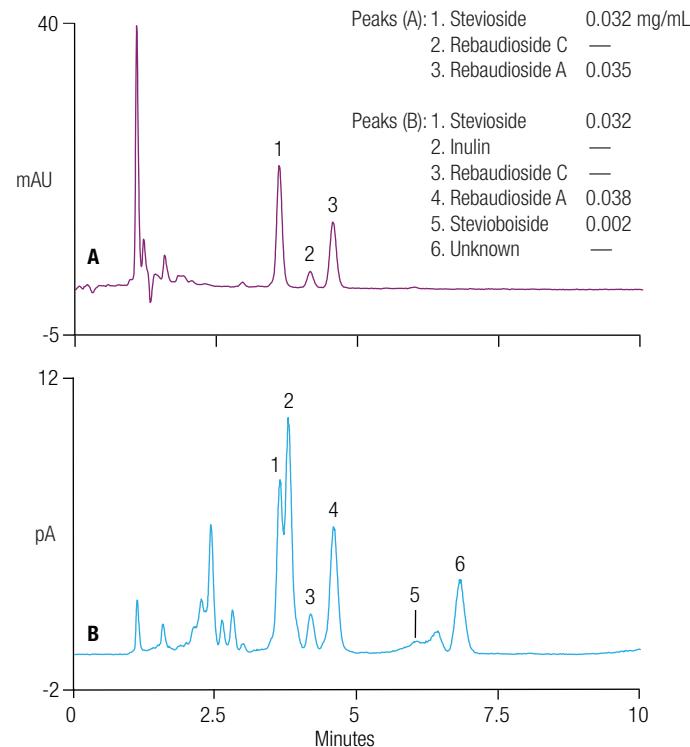


Figure 2-6. Separation of Brand A sweetener on the Acclaim Trinity P1 column and detected by A) UV and B) charged aerosol detections.

Column: Acclaim Trinity P1, 3 µm analytical, 2.1 × 100 mm and guard  
Mobile Phase: 81/19 acetonitrile/10 mM ammonium formate, pH = 3.00  
Flow Rate: 0.3 mL/min  
Temperature: 20 °C  
Injection Volume: 5 µL  
Detection: Charged aerosol detection, nebulizer temp. 10 °C  
Sample: A) Brand A extracted sweetener  
B) Steviol glycoside standards

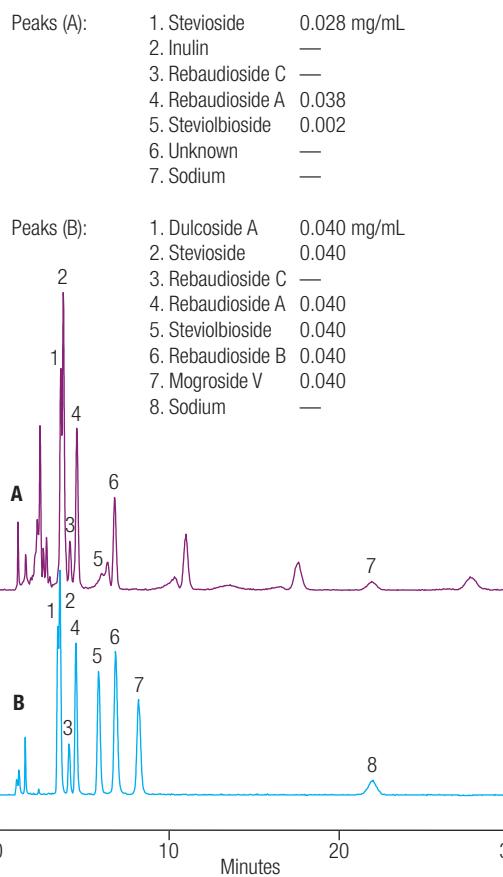


Figure 2-7. Separation of A) stevia sweetener Brand A in comparison to B) standards on the Acclaim Trinity P1 column detected by charged aerosol detection.

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Column: Acclaim Trinity P1,  
3  $\mu$ m analytical,  
2.1  $\times$  100 mm and guard  
Flow Rate: 0.3 mL/min  
Temperature: 20 °C  
Injection Volume: 5  $\mu$ L  
Mobile Phase: 81/19 acetonitrile/  
10 mM ammonium formate,  
pH = 3.00  
Peaks (A): 1. Rebaudioside A 0.044 mg/mL  
(Inset): 2. Rebaudioside B <LOD

Peaks (B): 1. Erythritol —  
2. Rebaudioside A 0.044  
3. Rebaudioside B 0.003

Detection: A) UV detection, 210 nm  
B) Charged aerosol detection

Sample: Brand B extracted sweetener

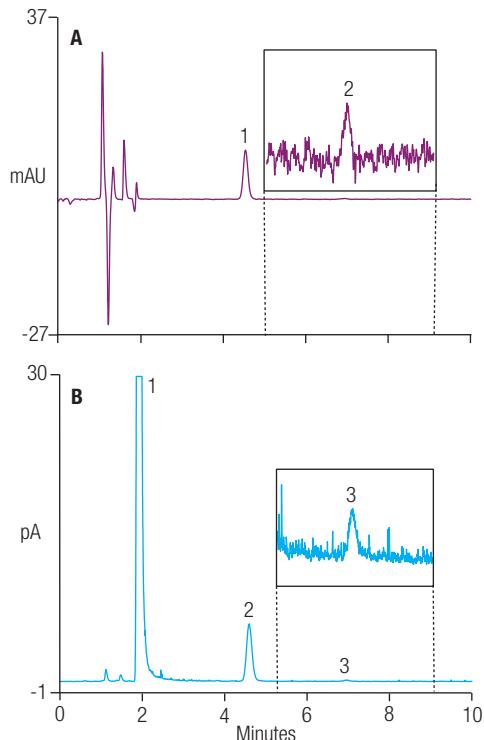


Figure 2-8. Rebaudioside A determination in Brand B, commercial sweetener, by A) UV and B) charged aerosol detection.

## Stevia

### Alternate HPLC-Charged Aerosol Detection Approach

Column: C18, 4.6  $\times$  250 mm, 5  $\mu$ m  
Flow: 1.0 mL/min  
Column Temp.: 50 °C  
Injection Volume: 10  $\mu$ L  
Mobile Phase: A. Deionized Water (D), acetonitrile, trifluoroacetic acid (TFA) (95:5:0.1)  
B. D:acetonitrile (5:95)  
Gradient: 0–90% B in 30 min; hold for 5 min and return to starting conditions  
Sample Prep.: Commercially available stevia extract powder was dissolved in deionized water (0.9 mg/mL)

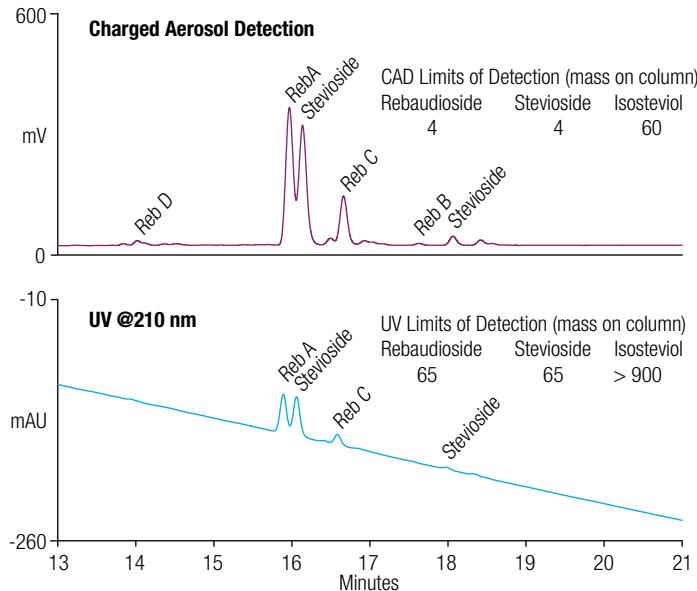


Figure 2-9. Alternate HPLC-charged aerosol detection method for Stevia analysis.

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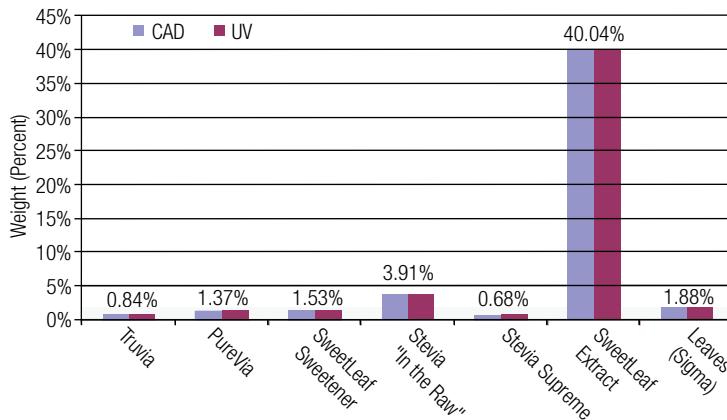


Figure 2-10. Percentage of Reb A in stevia-containing products.



## Stevia

### Comparison of HPLC and RSLC for the Analysis of a Stevia Extract

#### HPLC (A)

Column: C18, 4.6 × 250 mm, 5 µm

Flow Rate: 1.0 mL/min

Column Temp.: 50 °C

Injection Volume: 10 µL

Mobile Phase: A. Deionized Water (DI), acetonitrile, trifluoroacetic acid (TFA) (95:5:0.1)

B. DI:acetonitrile (5:95)  
5–90% B in 30 min;  
hold for 5 min and return to starting conditions

#### UHPLC (B)

Column: Acclaim RSLC Polar Advantage II 2.1 × 250 mm, 2.2 µm

Flow Rate: 0.7 mL/min

Column Temp.: 40 °C

Injection Volume: 5 µL

Mobile Phase: A. Deionized Water (DI) + 0.1% formic acid

B. Acetonitrile + 0.1% formic acid  
5% to 60% B in 9 min;  
hold 1 min and return to 5% B

Gradient:  
Sample Prep.: Commercially available stevia extract powder was dissolved in deionized water (0.9 mg/mL)

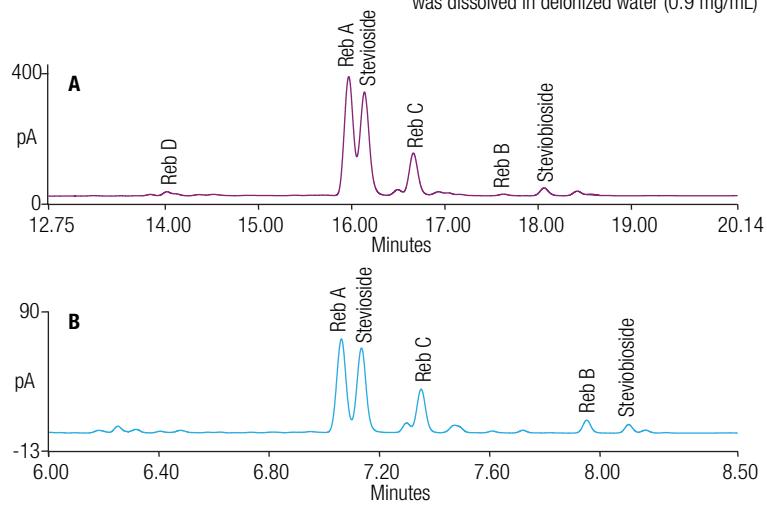


Figure 2-11. Using UHPLC conditions not only halves the time to perform analysis but improves resolution and also saves on mobile phase costs.

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### Stevia Degradation

Column: Acclaim RSLC Polar Advantage II,  
2.1 x 250 mm, 2.2  $\mu$ m  
Flow: 0.7 mL/min  
Column Temp.: 40 °C  
Injection Volume: 5  $\mu$ L  
Mobile Phase: A. Deionized Water (DI) + 0.1% formic acid  
B. Acetonitrile + 0.1% formic acid  
Gradient: 5% to 60% B in 9 min; hold 1 min and return to 5% B

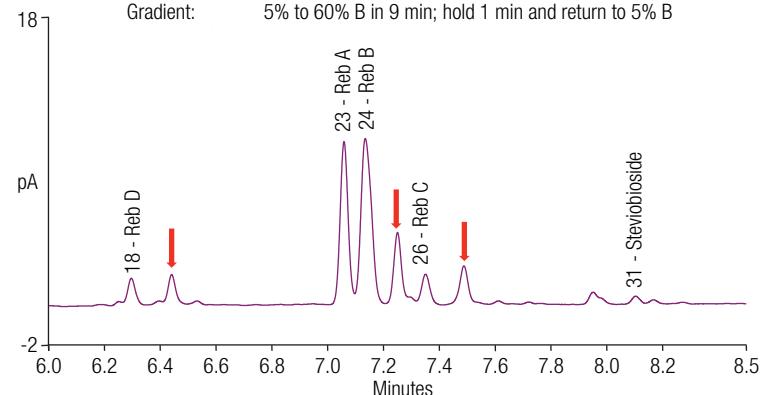


Figure 2-12. The UHPLC charged aerosol detection method could readily distinguish degradation products formed in a stevia-containing beverage left unopened at room temperature for over a year.

### Trivia Question

Q: Do you know who discovered the first artificial sweetener?

A: Constantine Fahlberg, a German scientist, discovered Saccharin—the first artificial sweetener—in 1879, completely on accident! He was doing research entirely unrelated to sweeteners and was instead conducting research on coal tar derivative products.



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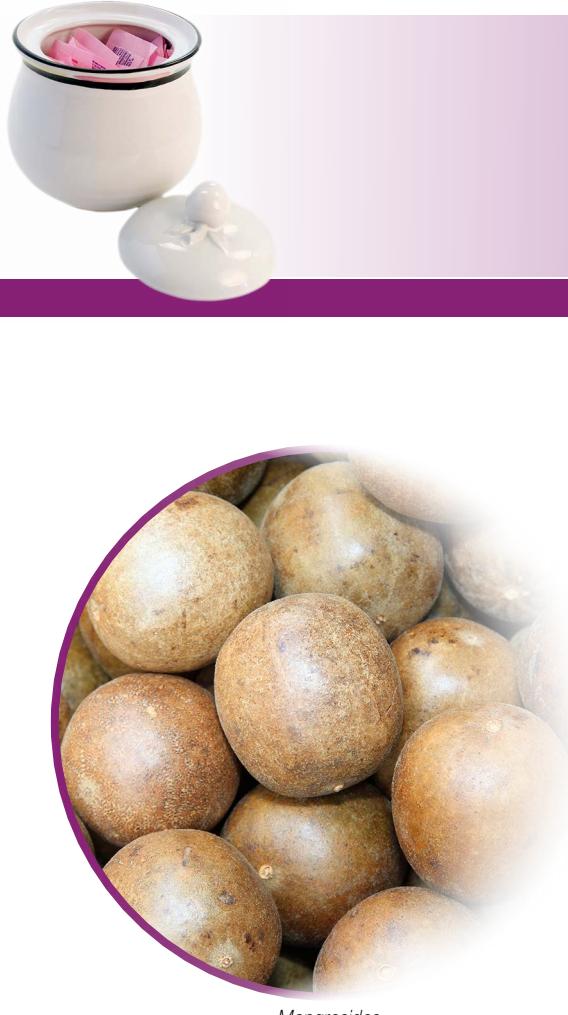
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### Mongrosides

Luo han kuo fruit (*Siraitia grosvenori* Swingle) has long been used in traditional Asian medicine. Recently cucurbitane-type and other triterpene glycosides have been isolated from the fruit and investigated for numerous potential health benefits such as antioxidant activity, anticancer effects, and antihyperglycemic effects. Many of these compounds are intensely sweet and therefore have also been investigated as sugar substitutes and flavor enhancers. Extracts of luo han kuo fruit used as sweeteners were acknowledged as "Generally Recognized as Safe" (GRAS) based on a GRAS submission to the U.S. FDA in January of 2010.

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Typical reversed-phase high-performance liquid chromatography methods to determine these glycosides are challenging due to the lack of a strong, specific chromophore in the compound. Other detection methods, such as charged aerosol detection, can be used to improve triterpene glycoside quantification.

In Application Update 184: Mogroside V determination by HPLC with Charged Aerosol and UV Detections, mogroside V is determined in a luo han kuo beverage by both charged aerosol and UV detections. The volatile mobile phase makes charged aerosol detection possible, which adds further flexibility to the method for detection of such glycosides.



Column: Acclaim Trinity P1, 3  $\mu$ m Analytical, 2.1  $\times$  100 mm and guard

Flow: 0.3 mL/min

Temperature: 20 °C

Injection Volume: 5  $\mu$ L

Mobile Phase: 81/19 acetonitrile/10 mM ammonium formate, pH 3.00

Detection: Charged aerosol detection, nebulizer temp. 10 °C

Sample: Steviol glycoside and mogroside V standards

Peaks: 1. Dulcoside A 0.190 mg/mL

2. Stevioside 0.190

3. Rebaudioside C —

4. Rebaudioside A 0.190

5. Steviolbioside 0.190

6. Rebaudioside B 0.190

7. Mogroside V 0.190

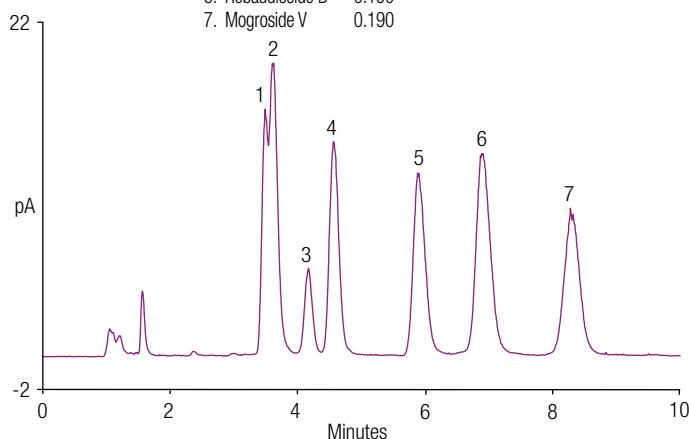


Figure 2-13. Detection of mogroside standards by charged aerosol detection.

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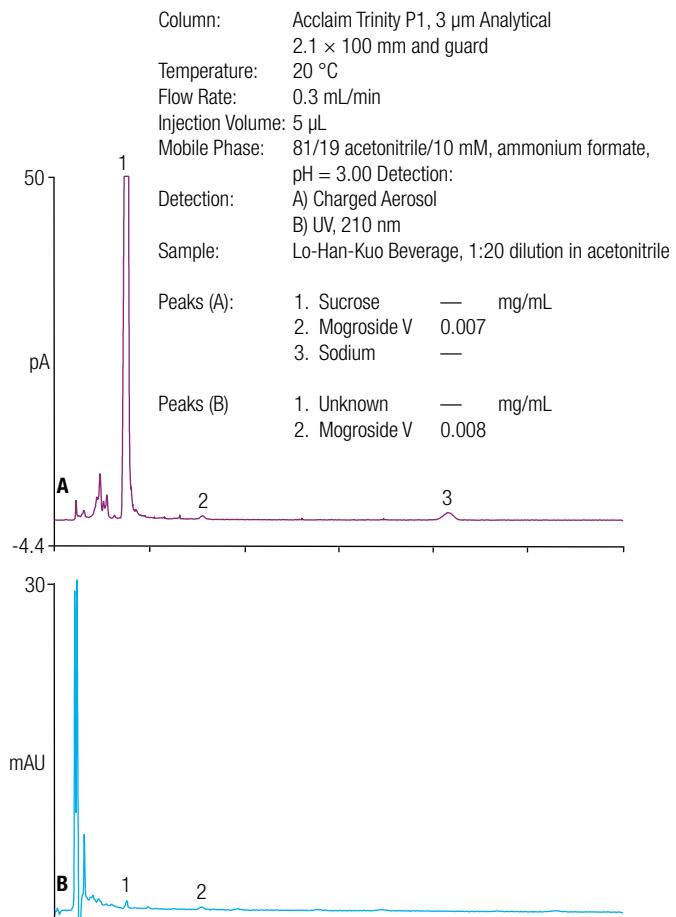


Figure 2-14. Superior performance of charged aerosol detection over UV detection for the detection of key analytes.

## Mogrosides

### Did You Know?

The pure mogroside mix extracted from the luo han guo fruit is approximately 300 times sweeter than sugar!



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### Sweetener Brand Ingredients

- Equal®: Aspartame, dextrose, and maltodextrin
- Splenda®: Sucratose, dextrose, and maltodextrin
- Sweet'n Low®: Saccharin, dextrose, and cream of tartar

Equal is a registered trademark of the Merisant Company, Splenda is a registered trademark of McNeil Nutritionals, LLC and Sweet'n Low is a registered trademark of Cumberland Packing Corporation.

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Aspartame was discovered by James M. Schlatter in 1965. Schlatter was a chemist working for G.D. Searle & Company. He had synthesized aspartame as an intermediate step in the process of making a tetrapeptide of the hormone gastrin, which was to be used in assessing the effectiveness of an anti-ulcer drug candidate. He accidentally discovered aspartame's sweet taste when he licked his finger, which had become contaminated with the compound.



## Aspartame

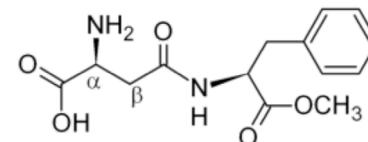


Figure 2-15. Aspartame, the methyl ester of the dipeptide of the natural amino acids L-aspartic acid and L-phenylalanine, is an artificial sweetener, that is approximately 200 times sweeter than sucrose, or table sugar.

### Did You Know?

A study in the journal *Physiology and Behavior* found that habitual diet soda sippers have more widespread activity in the reward processing regions of the brain when they consume other sweet foods and drinks than those who don't regularly opt for these beverages. That means they're more likely to overindulge in treats, which can pose a threat to maintaining a healthy weight.

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Equal contains multiple components, including the active ingredient, aspartame, along with the fillers dextrose and maltodextrin.

In this example, all components were separated using reversed phase chromatography and detected by the Corona charged aerosol detector. During method development a trace impurity/contaminant was found. Although several potential degradants (e.g. phenylalanine, aspartic acid) were analyzed, none corresponded to the impurity.

## Did You Know?

Equal was the first aspartame-based sweetener to be sold to the public in the United States. Up until its debut in the early 1980's the only other sweetener products contained only saccharin

## Equal and Impurity Method

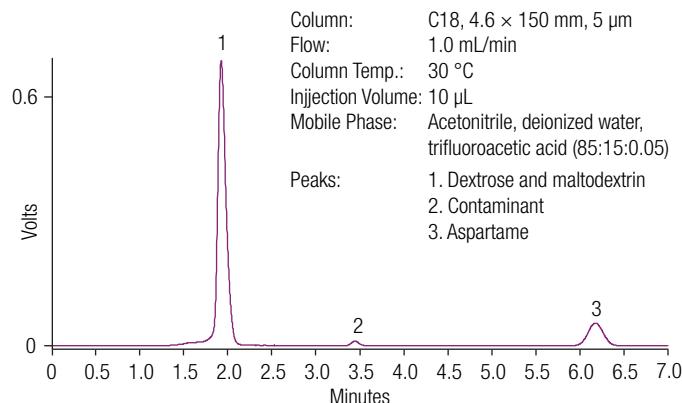


Figure 2-16. Chromatogram of Equal® sweeteners.



Download Poster Note: Sensitive Analysis of Commonly Used Artificial and Natural Sweeteners Including Stevia and Their Impurities and Degradation Products

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Sucralose (trichlorogalactosucrose or 1,6-dichloro- 1,6-dideoxy- $\beta$ -D-fructofuranosyl-4-deoxy- $\alpha$ -D-galacto- pyranoside) is a non-nutritive sweetener used to manufacture diabetic and dietetic foods and beverages. Detection of sucralose and other carbohydrates is challenging because they lack a strong chromophore and, therefore, cannot be detected at low concentrations with UV detection. Furthermore, sucralose would typically be present in foods containing compounds with strong UV chromophores. Refractive index detection can be used, but the sensitivity is poor. Charged aerosol detection is a viable approach to determination of sucralose (see above).

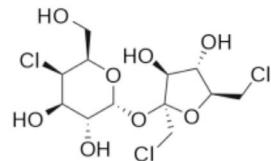


Figure 2-19. Chemical structure of sucralose.

Application Update 151 discusses the use of an alternate technique, High-Performance Anion-Exchange Chromatography with Pulsed Amperometric Detection (HPAE-PAD), to determine sucralose in reduced-carbohydrate colas. HPAE-PAD has been used to determine sucralose in other sugar-free beverages, after dilution, and foods. Reduced-carbohydrate cola samples have high concentrations of fructose and sucrose relative to sucralose, making these samples challenging for chromatographic analysis.

Column: Dionex CarboPac PA20 with guard  
Flow Rate: 0.5 mL/min  
Injection Volume: 25  $\mu$ L  
Mobile Phase: 100 mM sodium hydroxide/90 mM sodium acetate  
Detection: Pulsed amperometric detection (PAD), disposable Au working electrode, Carbohydrate waveform (Waveform A, TN 21)  
Sample:  
A. 10  $\mu$ M Sucralose  
B. Brand A  
C. Brand A with 10  $\mu$ M Sucralose  
D. Brand B  
E. Brand B with 10  $\mu$ M Sucralose  
Peaks:  
1. Fructose and Sucrose  
2. Unknown  
3. Sucralose

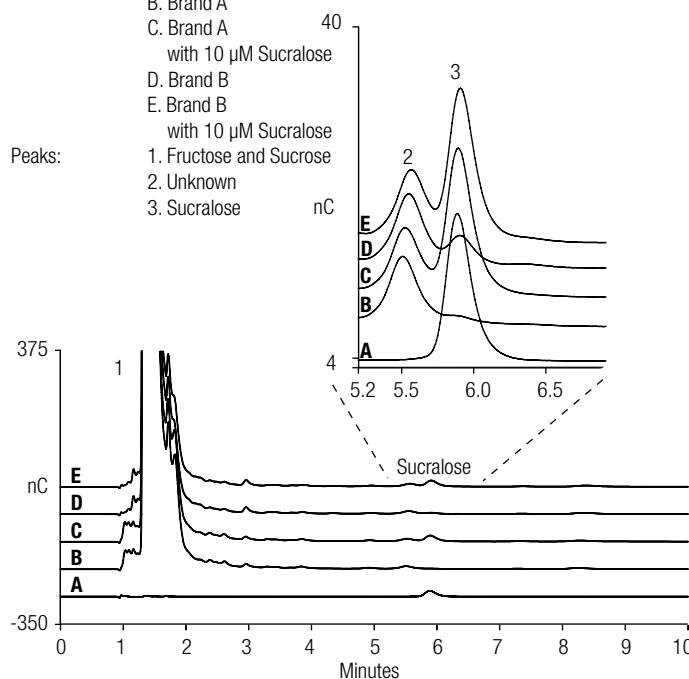


Figure 2-17. HPAE-PAD analysis of sucralose in a 100-fold dilution of the reduced-carbohydrate colas.

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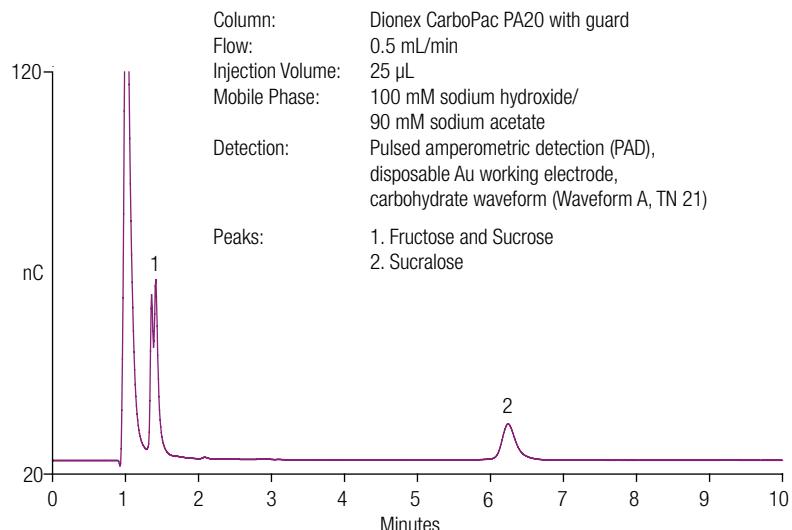


Figure 2-18. Determination of sucralose in a 50-fold dilution of Brand C peach citrus low-carbohydrate beverage.

## Sucralose

### Did You Know?

Sucralose is twice as sweet as saccharin, three times as sweet as aspartame and over 300 hundred times sweeter than table sugar!



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## Splenda

### HPLC-Charged Aerosol Detection Parameters:

Column: Amino, 4.6 x 250 mm, 5 µm  
Flow Rate: 1.0 mL/min  
Column Temp.: 30 °C  
Injection Volume: 10 µL  
Mobile Phase: A. Acetonitrile  
B. Deionized water  
Gradient: 30% to 70% B in 40 minutes

### Peaks:

1. Sucralose
2. Dextrose
3. Maltose
4. Maltotriose
5. Maltotetraose
6. Maltopentaose
7. Maltohexaose
8. Maltoheptaose
9. Maltooctaose
10. Maltonanase
11. Maltodecaose

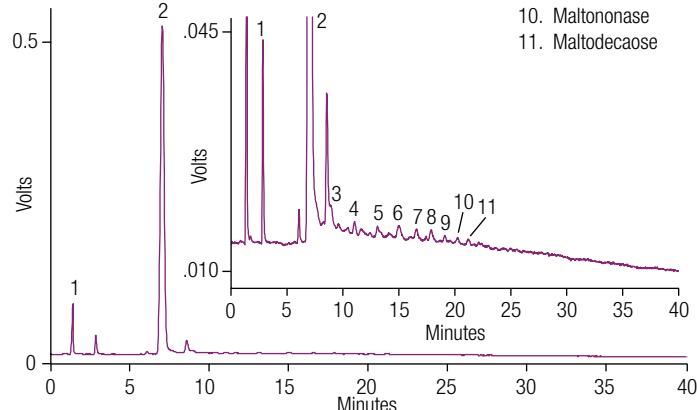


Figure 2-20. Analysis of Splenda. 10 µg on column. Inset presented at 10x sensitivity.

## Splenda and Sweet'N Low

## Sweet'N Low

### HPLC-Charged Aerosol Detection Parameters:

Column: C18, 4.6 mm x 150 mm, 5 µm  
Flow Rate: 1.0mL/min  
Column Temp.: Ambient  
Injection Volume: 10 µL  
Mobile Phase: 0.1% TFA in 20% MeOH(aq)  
Sample: Sweet and Low sachet dissolved in water at 1mg/mL

Inset at 10x sensitivity

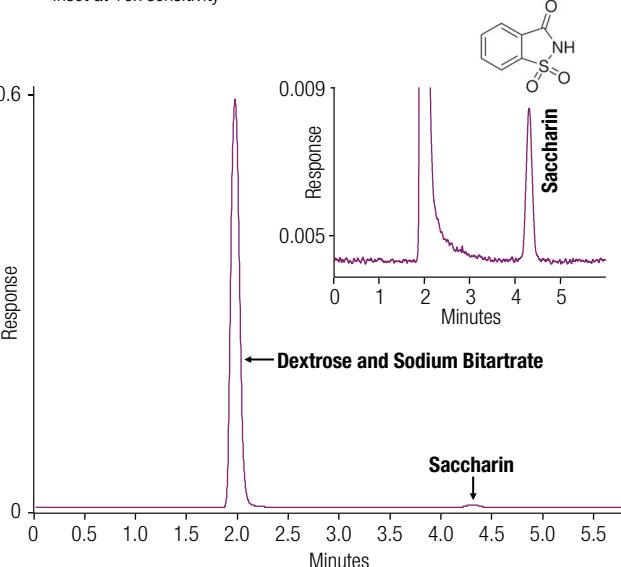


Figure 2-22. Analysis of Sweet'N Low. Saccharin, benzoic sulfilimine, is an artificial sweetener. It is much sweeter than sucrose, but has a bitter or metallic aftertaste, especially at high concentrations

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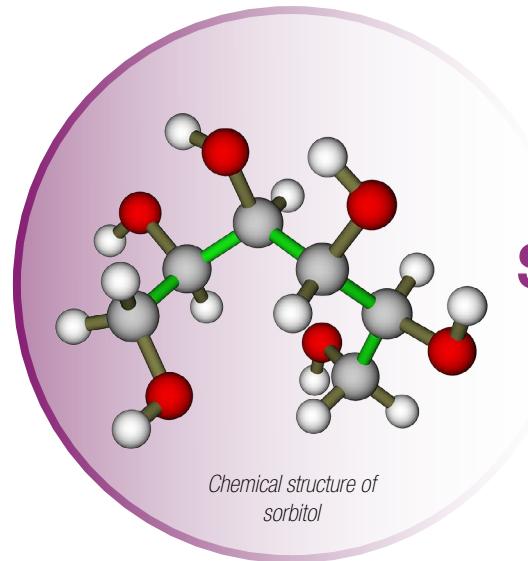
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## Chapter 2: Sugar Substitutes



### Sugar Alcohols

Sugar alcohols, such as sorbitol and mannitol, are used in confectionary products because they impart a sweet taste without the calories associated with sugars.

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Sorbitol (60% as sweet as sucrose) and mannitol are sugar alcohols commonly used as replacements for sucrose in dietetic candy. However, their use in foods is regulated because they exhibit laxative and diuretic properties.

Application Note 87 explores the use by pulsed amperometric detection for the measurement of sugar alcohols.

Column: Dionex CarboPac MA1, 4 × 250 mm  
Expected  
Operating Pressure: 5.5–7.6 MPa (800–1100 psi)  
Flow Rate: 0.4 mL/min  
Injection Volume: 10 µL  
Mobile Phase:  
A. Deionized water, 52%  
B. 1.0 M Sodium hydroxide, 48%  
Detection: Pulsed amperometry, gold working electrode  
ED40 Settings:

Time (min)	E (volts)	Integration (s)
400	+0.05	0.2–0.4
200	+0.75	
400	-0.15	

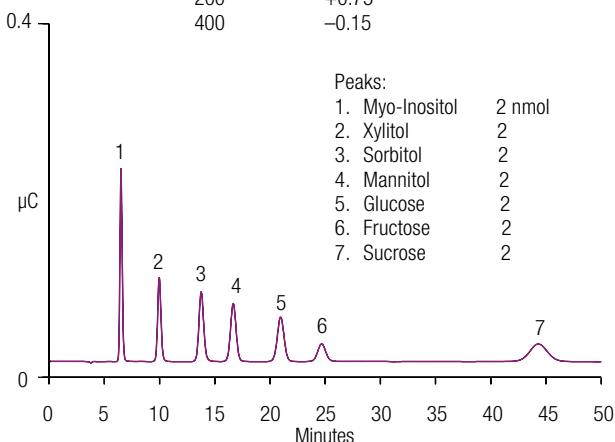


Figure 2-23. Sugar alcohols with high pKa values elute first.

## Sugar Alcohols

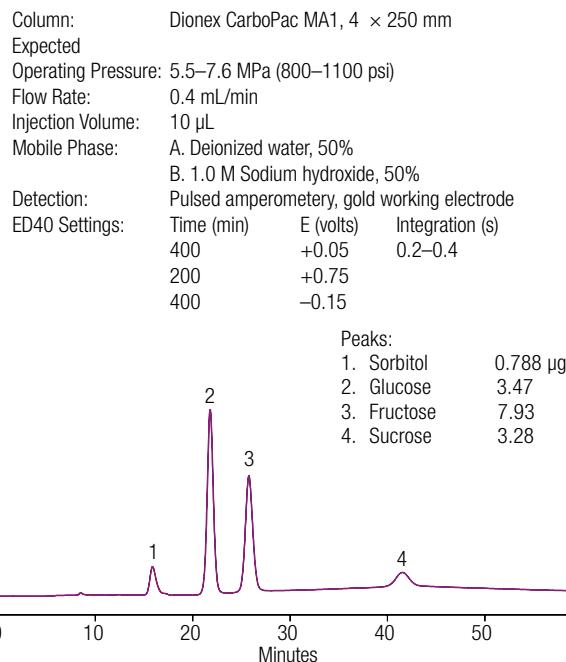


Figure 2-24. Diluted apple juice containing sorbitol.

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Column: Dionex CarboPac MA1, 4 × 250 mm  
Expected  
Operating Pressure: 5.5–7.6 MPa (800–1100 psi)  
Flow Rate: 0.4 mL/min  
Injection Volume: 10 µL  
Mobile Phase: A. Deionized water, 50%  
B. 1.0 M Sodium hydroxide, 50%  
Detection: Pulsed amperometry, gold working electrode

ED40 Settings:  
Time (min)      E (volts)      Integration (s)  
400               +0.05            0.2–0.4  
200               +0.75  
400               -0.15

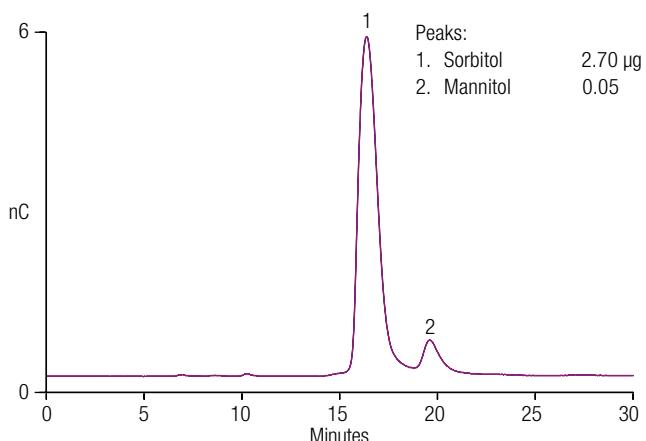


Figure 2-25. "Sugarless" hard candy containing sorbitol and mannitol.

Column: Dionex CarboPac MA1, 4 × 250 mm  
Expected  
Operating Pressure: 5.5–7.6 MPa (800–1100 psi)  
Flow Rate: 0.4 mL/min  
Injection Volume: 10 µL  
Mobile Phase: A. Deionized water, 50%  
B. 1.0 M Sodium hydroxide, 50%  
Detection: Pulsed amperometry, gold working electrode

ED40 Settings:  
Time (min)      E (volts)      Integration (s)  
400               +0.05            0.2–0.4  
200               +0.75  
400               -0.15

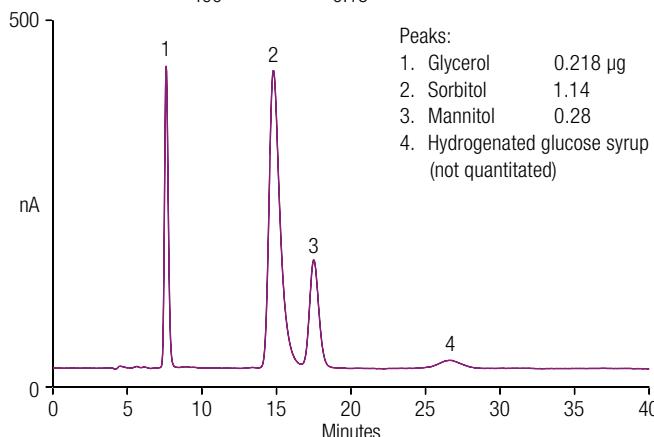
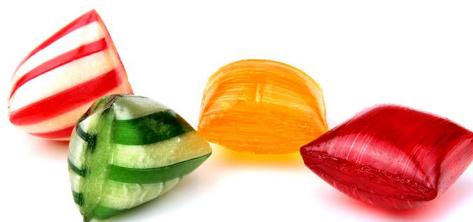


Figure 2-26. Chewing gum extract containing glycerol, sorbitol, and mannitol.



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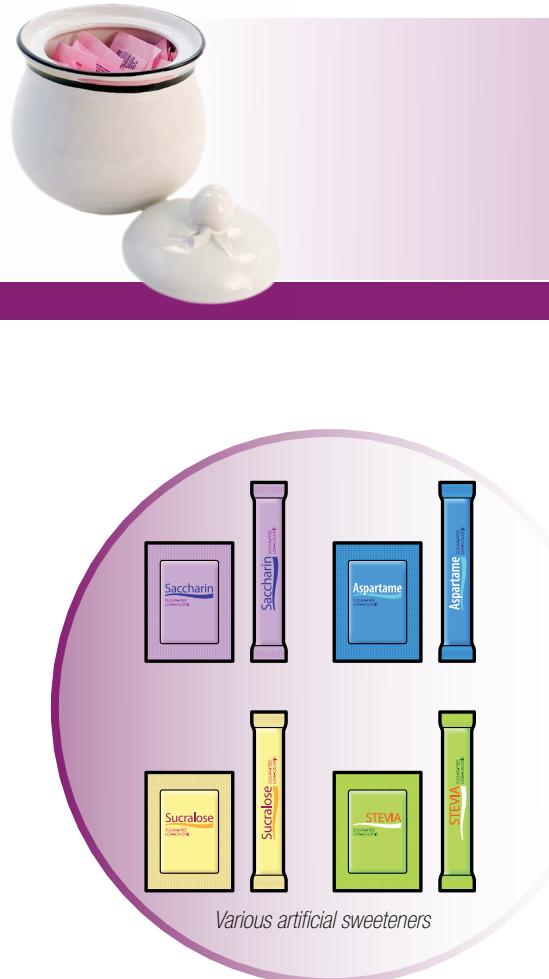
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## Chapter 2: Sugar Substitutes

### Global Artificial Sweetener Method

Many recently commercialized sweeteners tend to have increased potency, reducing the amount of active ingredient added to beverages and other food products and often providing cost savings to the manufacturer.

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## Global Artificial Sweetener Method

Using sugar substitutes of increased potency has also contributed to a need for sensitive analytical methods to quantify the active product and to detect low levels of breakdown products and impurities, which are required for quality and safety issues. Because compounds typically do not possess a chromophore, traditional HPLC-UV approaches are inappropriate.

The Poster Note "Sensitive Analysis of Commonly Used Artificial and Natural Sweeteners Including Stevia and Their Impurities and Degradation Products" describes a global gradient HPLC method with charged aerosol detection for the simultaneous measurement of several artificial sweeteners. This method is sensitive (low ng levels), with good reproducibility and accuracy.

Column :	C18, 5 µm, 4.6 mm x 250 mm	Peaks:	1. Acesulfame K
Flow:	1.00 mL/min		2. Cyclamate
Temperature:	30 °C		3. Saccharin
Injection Volume:	50 µL		4. Sucratose
Mobile Phase:	A: Deionized Water B: Acetonitrile and 0.1% Trifluoroacetic Acid		5. Aspartame
Gradient:	Isocratic: 2–40% B over 25 min; 40–60% from 25–30 min		6. Neotame
Samples	1.2 to 20 µg on column		7. Alitame
			8. NHDC
			9. Dulcin

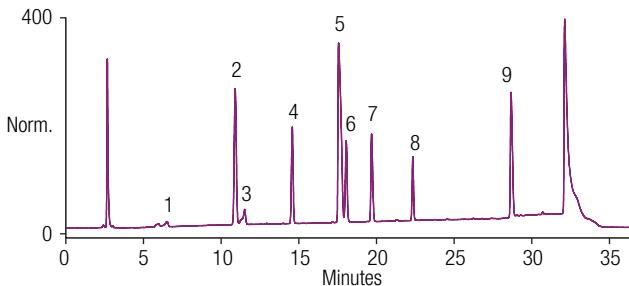


Figure 2-27. Chromatogram of artificial sweeteners.

### Did You Know?

Sugar substitutes are often cheaper to use than sugar due to their longer shelf-life and higher sweetening intensity, meaning a smaller amount can be used to achieve an equivalent level of sweetness.

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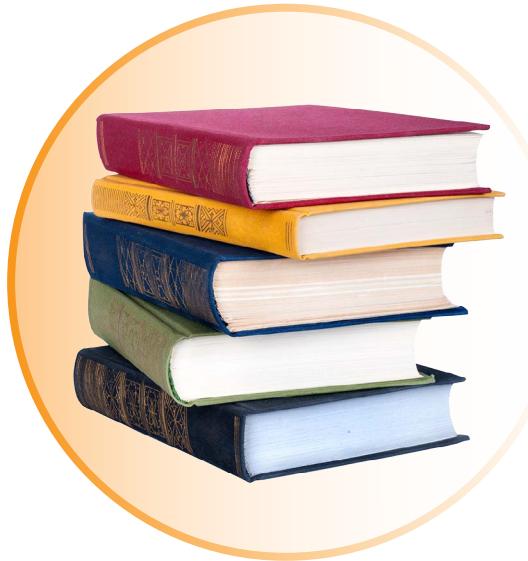
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## Technical Collateral and Peer Reviewed Journals

Here you'll find a multitude of references using our HPLC, ion chromatography and sample preparation solutions.

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<a href="#">Determination of levoglucosan in atmospheric aerosols using high performance liquid chromatography with aerosol charge detection.</a>	Dixon, R. W.; Baltzell, G.	<i>J. Chromatogr. A.</i> 1109 (2), 214–221	2006 Mar 24
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<a href="#">Characterization of an endoglucanase belonging to a new subfamily of glycoside hydrolase family 45 of the basidiomycete Phanerochaete chrysosporium.</a>	Igarashi, K.; Ishida, T.; Hori, C.; Samejima, M.	<i>Appl. Environ. Microbiol.</i> 74 (18), 5628–5634	2008 Sep
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### Food, Nutrition, Natural Products, and Supplements

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<a href="#">Analysis of lycopene geometrical isomers in biological microsamples by liquid chromatography with coulometric array detection</a>	Ferruzzi, M. G.; Nguyen, M. L.; Sander, L. C.; Rock, C. L.; Schwartz, S. J.	<i>J. Chromatogr., B: Biomed. Sci. Appl.</i> 760 (2), 289–299	2001 Sep 5
<a href="#">Charged aerosol detection to characterize components of dispersed-phase formulations</a>	Fox, C. B.; Sivananthan, S. J.; Mikasa, T. J.; Lin, S.; Parker, S. C.	<i>Adv. Colloid Interface Sci.</i> 199–200, 59–65	2013 Nov
<a href="#">HPLC with charged aerosol detection for the measurement of natural products</a>	Fukushima, K.; Kanedai, Y.; Hirose, K.; Matsumoto, T.; Hashiguchi, K.; Senda, M.; et al.	<i>Chromatography 27 (Suppl. 1)</i> , 83–86	2006
<a href="#">Determination of heterocyclic aromatic amines in beef extract, cooked meat and rat urine by liquid chromatography with coulometric electrode array detection</a>	Gerbl, U.; Cichna, M.; Zsivkovits, M.; Knasmüller, S.; Sontag, G.	<i>J. Chromatogr., B: Anal. Technol. Biomed. Life Sci.</i> 802 (1), 107–113	2004 Mar 25
<a href="#">Determination of macrolide antibiotics in porcine and bovine urine by high-performance liquid chromatography coupled to coulometric detection</a>	González de la Huebra, M. J.; Vincent, U.; Bordin, G.; Rodríguez, A. R.	<i>Anal. Bioanal. Chem.</i> 382 (2), 433–439	2005 May
<a href="#">Development and validation of HPLC-DAD-CAD-MS3 method for qualitative and quantitative standardization of polyphenols in <i>Agrimoniae eupatoriaie herba</i> (Ph. Eur.)</a>	Granica, S.; Krupa, K.; Klebowska, A.; Kiss, A. K.	<i>J. Pharm. Biomed. Anal.</i> 86, 112–122	2013 Dec
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<a href="#">Urinary 3-(3,5-dihydroxyphenyl)-1-propanoic acid, an alkylresorcinol metabolite, is a potential biomarker of whole-grain intake in a U.S. population</a>	Guyman, L. A.; Adlercreutz, H.; Koskela, A.; Li, L.; Beresford, S. A.; Lampe, J. W.	<i>J. Nutr.</i> 138 (10), 1957–1962	2008 Oct
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<a href="#">Determination of the urinary aglycone metabolites of vitamin K by HPLC with redox-mode electrochemical detection</a>	Harrington, D. J.; Soper, R.; Edwards, C.; Savidge, G. F.; Hodges, S. J.; Shearer, M. J.	<i>J. Lipid Res.</i> 46 (5), 1053–1060	2005 May

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Title	Authors	Publication	Publication Date
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<a href="#">RP-HPLC analysis of phenolic compounds and flavonoids in beverages and plant extracts using a CoulArray detector</a>	Jandera, P.; Skeifíková, V.; Rehová, L.; Hájek, T.; Baldriánová, L.; Skopová, G.; Kellner, V.; Horna, A.	<i>J. Sep. Sci.</i> 28 (9–10), 1005–1022	2005 Jun
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<a href="#">A combination of aspirin and <math>\gamma</math>-tocopherol is superior to that of aspirin and <math>\alpha</math>-tocopherol in anti-inflammatory action and attenuation of aspirin-induced adverse effects</a>	Jiang, Q.; Moreland, M.; Ames, B. N.; Yin, X.	<i>J. Nutr. Biochem.</i> 20 (11), 894–900	2009 Nov
<a href="#">HPLC analysis of rosmarinic acid in feed enriched with aerial parts of <i>Prunella vulgaris</i> and its metabolites in pig plasma using dual-channel coulometric detection</a>	Jirovský, D.; Kosina, P.; Myslínová, M.; Stýskala, J.; Ulrichová, J.; Simánek V.	<i>J. Agric. Food Chem.</i> 55 (19), 7631–7637	2007 Sep 19
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<a href="#">Sensitive electrochemical detection method for alpha-acids, beta-acids and xanthohumol in hops (<i>Humulus lupulus L.</i>)</a>	Kac, J.; Vovk, T.	<i>J. Chromatogr., B: Anal. Technol. Biomed. Life Sci.</i> 850 (1–2), 531–537	2007 May 1
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<a href="#">Optimization of pressurized liquid extraction for spicatoside A in <i>Liriope platyphylla</i></a>	Kim, S. H.; Kim, H. K.; Yang, E. S.; Lee, K. Y.; Kim, S. D.; Kim, Y. C.; Sung, S. H.	<i>Sep. Purif. Technol.</i> 71 (2), 168–172	2010
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<a href="#">Determination of 4-ethylcatechol in wine by high-performance liquid chromatography-coulometric electrochemical array detection</a>	Larcher, R.; Nicolini, G.; Bertoldi, D.; Nardin, T.	<i>Anal. Chim. Acta</i> 609 (2), 235–240	2008 Feb 25
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<a href="#">Determination of water-soluble vitamins in infant milk and dietary supplement using a liquid chromatography on-line coupled to a corona-charged aerosol detector</a>	Márquez-Sillero, I.; Cárdenas, S.; Valcárcel, M.	<i>J. Chromatogr., A.</i> 1313C, 253–258	2013 Oct 25
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<a href="#">High-performance liquid chromatography analysis of plant saponins: An update 2005–2010</a>	Negi, J. S.; Singh, P.; Pant, G. J.; Rawat, M. S.	<i>Pharmacogn. Rev.</i> 5 (10), 155–158	2011 Jul
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<a href="#">Practical preparation of lacto-<i>N</i>-biose I, a candidate for the bifidus factor in human milk</a>	Nishimoto, M.; Kitaoka, M.	<i>Biosci., Biotechnol., Biochem.</i> 71 (8), 2101–2104	2007 Aug
<a href="#">Hydrophilic interaction liquid chromatography—charged aerosol detection as a straightforward solution for simultaneous analysis of ascorbic acid and dehydroascorbic acid</a>	Nováková, L.; Solichová, D.; Solich, P.	<i>J. Chromatogr., A.</i> 1216 (21), 4574–4581	2009 May 22

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<a href="#">Assessment of probiotic strains ability to reduce the bioaccessibility of aflatoxin M 1 in artificially contaminated milk using an in vitro digestive model</a>	Serrano-Niño, J. C.; Cavazos-Garduño, A.; Hernandez-Mendoza, A.; Applegate, B.; Ferruzzi, M. G.; San Martin-González, M. F.; García, H. S.	<i>Food Control</i> 31 (1), 202–207	2013 May
<a href="#">Intestinal uptake of quercetin-3-glucoside in rats involves hydrolysis by lactase phlorizin hydrolase</a>	Sesink, A. L.; Arts, I. C.; Faassen-Peters, M.; Hollman, P. C.	<i>J. Nutr.</i> 133 (3), 773–776	2003 Mar
<a href="#">Quercetin glucuronides but not glucosides are present in human plasma after consumption of quercetin-3-glucoside or quercetin-4'-glucoside</a>	Sesink, A. L.; O'Leary, K. A.; Hollman, P. C.	<i>J. Nutr.</i> 131 (7), 1938–1941	2001 Jul
<a href="#">Co-administration of quercetin and catechin in rats alters their absorption but not their metabolism</a>	Silberberg, M.; Morand, C.; Manach, C.; Scalbert, A.; Remesy, C.	<i>Life Sci.</i> 77 (25), 3156–3167	2005 Nov 4
<a href="#">Nutritional status is altered in the self-neglecting elderly</a>	Smith, S. M.; Mathews Oliver, S. A.; Zwart, S. R.; Kala, G.; Kelly, P. A.; Goodwin, J. S.; Dyer, C. B.	<i>J. Nutr.</i> 136 (10), 2534–2541	2006 Oct

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### Food, Nutrition, Natural Products, and Supplements

Title	Authors	Publication	Publication Date
<a href="#">Binding of heterocyclic aromatic amines by lactic acid bacteria: results of a comprehensive screening trial</a>	Stidl, R.; Sontag, G.; Koller, V.; Knasmüller, S.	<i>Mol. Nutr. Food Res.</i> 52 (3), 322–329	2008 Mar
<a href="#">Direct separation and detection of biogenic amines by ion-pair liquid chromatography with chemiluminescent nitrogen detector</a>	Sun, J.; Guo, H. X.; Semin, D.; Cheetham, J.	<i>J. Chromatogr. A.</i> 1218 (29), 4689–4697	2011 Jul 22
<a href="#">Rapid purification method for fumonisin B1 using centrifugal partition chromatography</a>	Szekeres, A.; Lorántfy, L.; Bencsik, O.; Kecskeméti, A.; Szécsi, Á.; Mesterházy, Á.; Vágvölgyi, C.	<i>Food Addit. Contam.</i> 30 (1), 147–155	2013
<a href="#">Determination of coenzyme Q10 in over-the-counter dietary supplements by high-performance liquid chromatography with coulometric detection</a>	Tang, P. H.	<i>J. AOAC Int.</i> 89 (1), 35–39	2006 Jan–Feb
<a href="#">α-Tocopherol supplementation restores the reduction of erythrocyte glucose-6-phosphate dehydrogenase activity induced by forced training</a>	Tsakiris, S.; Reclos, G. J.; Parthimos, T.; Tsakiris, T.; Parthimos, N.; Schulpis, K. H.	<i>Pharmacol. Res.</i> 54 (5), 373–379	2006 Nov
<a href="#">Tissue distribution of isoflavones in ewes after consumption of red clover silage</a>	Urpi-Sarda, M.; Morand, C.; Besson, C.; Kraft, G.; Viala, D.; Scalbert, A.; Besle, J. M.; Manach, C.	<i>Arch. Biochem. Biophys.</i> 476 (2), 205–210	2008 Aug 15
<a href="#">Performance evaluation of charged aerosol and evaporative light scattering detection for the determination of ginsenosides by LC</a>	Wang, L.; He, W. S.; Yan, H. X.; Jiang, Y.; Bi, K. S.; Tu, P. F.	<i>Chromatographia</i> 70 (3–4), 603–608	2009 Aug
<a href="#">Catechins are bioavailable in men and women drinking black tea throughout the day</a>	Warden, B. A.; Smith, L. S.; Beecher, G. R.; Balentine, D. A.; Clevidence, B. A.	<i>J. Nutr.</i> 131 (6), 1731–1737	2001 Jun
<a href="#">Identification and quantification of polyphenol phytoestrogens in foods and human biological fluids</a>	Wilkinson, A. P.; Wähälä, K.; Williamson, G.	<i>J. Chromatogr., B: Anal. Technol. Biomed. Life Sci.</i> 777 (1–2), 93–109	2002 Sep 25
<a href="#">Bioavailability and pharmacokinetics of caffeoylquinic acids and flavonoids after oral administration of Artichoke leaf extracts in humans</a>	Wittemer, S. M.; Ploch, M.; Windeck, T.; Müller, S. C.; Drewelow, B.; Derendorf, H.; Veit, M.	<i>Phytomedicine</i> 12 (1–2), 28–38	2005 Jan
<a href="#">Validated method for the determination of six metabolites derived from artichoke leaf extract in human plasma by high-performance liquid chromatography-coulometric-array detection</a>	Wittemer, S. M.; Veit, M.	<i>J. Chromatogr., B: Anal. Technol. Biomed. Life Sci.</i> 793 (2), 367–375	2003 Aug 15
<a href="#">HPLC in natural product analysis: The detection issue</a>	Wolfender, J. L.	<i>Planta Med.</i> 75 (07), 719–734	2009 Jun
<a href="#">Simultaneous determination of isoflavones and bisphenol A in rat serum by high-performance liquid chromatography coupled with coulometric array detection</a>	Yasuda, S.; Wu, P. S.; Hattori, E.; Tachibana, H.; Yamada, K.	<i>Biosci., Biotechnol., Biochem.</i> 68 (1), 51–58	2004 Jan
<a href="#">Impurities from polypropylene microcentrifuge tubes as a potential source of interference in simultaneous analysis of multiple lipid-soluble antioxidants by HPLC with electrochemical detection</a>	Yen, H. C.; Hsu, Y. T.	<i>Clin. Chem. Lab. Med.</i> 42 (4), 390–395	2004 Apr

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Title	Authors	Publication	Publication Date
<a href="#">Simultaneous determination of triterpenoid saponins from <i>pulsatilla koreana</i> using high performance liquid chromatography coupled with a charged aerosol detector (HPLC-CAD)</a>	Yeom, H.; Suh, J. H.; Youm, J. R.; Han, S. B.	Bull. Korean Chem. Soc. 31 (5), 1159–1164	2010
<a href="#">DPPH radical scavenging activities of 31 flavonoids and phenolic acids and 10 extracts of Chinese <i>materia medica</i></a>	Yuan, Y.; Chen, C.; Yang, B.; Kusu, F.; Kotani, A.	Zhongguo Zhongyao Zazhi 34 (13), 1695–1700	2009 Jul
<a href="#">Determination of residual clenbuterol in pork meat and liver by HPLC with electrochemical detection</a>	Zhang, X. Z.; Gan, Y. R.; Zhao, F. N.	Yaoxue Xuebao 39 (4), 276–280	2004 Apr
<a href="#">Identification of equol producers in a Japanese population by high-performance liquid chromatography with coulometric array for determining serum isoflavones</a>	Zhao, J. H.; Sun, S. J.; Arao, Y.; Oguma, E.; Yamada, K.; Horiguchi, H.; Kayama, F.	Phytomedicine 13 (5), 304–309	2006 May
<a href="#">Simultaneous sampling of volatile and non-volatile analytes in beer for fast fingerprinting by extractive electrospray ionization mass spectrometry</a>	Zhu, L.; Hu, Z.; Gamez, G.; Law, W. S.; Chen, H.; Yang, S.; Chingin, K.; Balabin, R. M.; Wang, R.; Zhang, T.; Zenobi, R.	Anal. Bioanal. Chem. 398 (1), 405–413	2010 Sep
<a href="#">Comparison of various easy-to-use procedures for extraction of phenols from apricot fruits</a>	Zitka, O.; Sochor, J.; Rop, O.; Skalickova, S.; Sobrova, P.; Zehnalek, J.; Beklova, M.; Krská, B.; Adam, V.; Kizek, R.	Molecules 16 (4), 2914–2936	2011 Apr 4



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Title	Authors	Publication	Publication Date
<a href="#">Development of analytical procedures to study changes in the composition of meat phospholipids caused by induced oxidation</a>	Cascone, A.; Eerola, S.; Ritieni, A.; Rizzo, A.	<i>J. Chromatogr. A</i> 1120 (1–2), 211–220	2006 Jul 7
<a href="#">Evaporative light scattering and charged aerosol detector.</a>	Chaminade, P.	Chapter 5. In <i>Hyphenated and Alternative Methods of Detection in Chromatography, Chromatographic Science Series</i> ; Shalliker, A., Ed.; Taylor & Francis Group, LLC: Boca Raton, FL; 145–160	2012
<a href="#">Simple and efficient profiling of phospholipids in phospholipase D-modified soy lecithin by HPLC with charged aerosol detection</a>	Damjanovic', J.; Nakano, H.; Iwasaki, Y.	<i>J. Am. Oil Chem. Soc.</i> 90 (7), 951–957	2013 Jul
<a href="#">Discriminating olive and non-olive oils using HPLC-CAD and chemometrics</a>	de la Mata-Espinosa, P.; Bosque-Sendra, J. M.; Bro, R.; Cuadros-Rodríguez, L.	<i>Anal. Bioanal. Chem.</i> 399 (6), 2083–2092	2011 Feb
<a href="#">Olive oil quantification of edible vegetable oil blends using triacylglycerols chromatographic fingerprints and chemometric tools</a>	de la Mata-Espinosa, P.; Bosque-Sendra, J. M.; Bro, R.; Cuadros-Rodríguez, L.	<i>Talanta</i> 85 (1), 177–182	2011 Jul 15
<a href="#">Quantification of triacylglycerols in olive oils using HPLC-CAD</a>	de la Mata-Espinosa, P.; Bosque-Sendra, J.; Cuadros-Rodríguez, L.	<i>Food Analytical Methods</i> 4 (4), 574–581	2011 Dec
<a href="#">Quantification of pegylated phospholipids decorating polymeric microcapsules of perfluoroctyl bromide by reverse phase HPLC with a charged aerosol detector</a>	Díaz-López, R.; Libong, D.; Tsapis, N.; Fattal, E.; Chaminade, P.	<i>J. Pharm. Biomed. Anal.</i> 48 (3), 702–707	2008 Nov 4
<a href="#">Squalene emulsions for parenteral vaccine and drug delivery</a>	Fox, C. B.	<i>Molecules</i> 14 (9), 3286–3312	2009 Sep 1
<a href="#">Interactions between parenteral lipid emulsions and container surfaces</a>	Gonyon, T.; Tomaso, A.; Kotha, P.; Owen, H.; Patel, D.; Carter, P.; Cronin, J.; Green, J.	<i>PDA J. Pharm. Sci. and Tech.</i> 67 (3), 247–254	2013 May–Jun
<a href="#">Composition analysis of positional isomers of phosphatidylinositol by high-performance liquid chromatography</a>	Iwasaki, Y.; Masayama, A.; Mori, A.; Ikeda, C.; Nakano, H.	<i>J. Chromatogr. A</i> 1216 (32), 6077–6080	2009 Aug 7
<a href="#">Determination of phospholipid and its degradation products in liposomes for injection by HPLC-charged aerosol detection (CAD)</a>	Jiang, Q.; Yang, R.; Mei, X.	<i>Chinese Pharmaceutical Journal (Zhongguo Yaoxue Zazhi, Beijing, China)</i> 42 (23), 1794–1796	2007

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Title	Authors	Publication	Publication Date
<a href="#">Rapid quantification of yeast lipid using microwave-assisted total lipid extraction and HPLC-CAD</a>	Khoomrung, S.; Chumnanpuen, P.; Jansa-Ard, S.; Ståhlman, M.; Nookaew, I.; Borén, J.; Nielsen, J.	<i>Anal. Chem.</i> 85 (10), 4912–4919	2013 May 21
<a href="#">A new liquid chromatography method with charge aerosol detector (CAD) for the determination of phospholipid classes. Application to milk phospholipids</a>	Kiebowicz, G.; Micek, P.; Wawrzenczyk, C.	<i>Talanta</i> 105, 28–33	2013 Feb 15
<a href="#">An LC method for the analysis of phosphatidylcholine hydrolysis products and its application to the monitoring of the acyl migration process</a>	Kiebowicz, G.; Smuga, D.; Gladkowski, W.; Chojnacka, A.; Wawrzenczyk, C.	<i>Talanta</i> 94, 22–29	2012 May 30
<a href="#">Separation of acylglycerols, FAME and FFA in biodiesel by size exclusion chromatography</a>	Kittirattanapiboon, K.; Krisnangkurá, K.	<i>Eur. J. Lipid Sci. Technol.</i> 110 (5), 422–427	2008 Mar 17
<a href="#">Quantitation of triacylglycerols from plant oils using charged aerosol detection with gradient compensation</a>	Lísa, M.; Lynen, F.; Holčapek, M.; Sandra, P.	<i>J. Chromatogr. A.</i> 1176 (1–2), 135–142	2007 Dec 28
<a href="#">Quantitative study of the stratum corneum lipid classes by normal phase liquid chromatography: comparison between two universal detectors</a>	Merle, C.; Laugel, C.; Chaminade, P.; Baillet-Guffroy, A.	<i>J. Liq. Chromatogr. Relat. Technol.</i> 33, 629–644	2010 Mar
<a href="#">The analysis of lipids via HPLC with a charged aerosol detector</a>	Moreau, R. A.	<i>Lipids</i> 41 (7), 727–34	2006 Jul
<a href="#">Lipid analysis via HPLC with a charged aerosol detector</a>	Moreau, R. A.	<i>Lipid Technol.</i> 21 (8–9), 191–194	2009 Oct 23
<a href="#">Extraction and analysis of food lipids</a>	Moreau, R. A.; Winkler-Moser, J. K.	Chapter 6 in <i>Methods of Analysis of Food Components and Additives</i> , Second Edition; Ötles, S., Ed.; Taylor & Francis Group, LLC: Boca Raton, FL.; 115–134	2011 Nov
<a href="#">Aerosol based detectors for the investigation of phospholipid hydrolysis in a pharmaceutical suspension formulation</a>	Nair, L.; Werling, J.	<i>J. Pharm. Biomed. Anal.</i> 49 (1), 95–99	2009 Jan 15
<a href="#">Structure/function relationships of adipose phospholipase A2 containing a cys-his-his catalytic triad</a>	Pang, X. Y.; Cao, J.; Addington, L.; Lovell, S.; Battaile, K. P.; Zhang, Rao, J. L.; Dennis, E. A.; Moise, A. R.	<i>J. Biol. Chem.</i> 287 (42), 35260–35274	2012 Oct 12
<a href="#">Simultaneous assessment of lipid classes and bile acids in human intestinal fluid by solid-phase extraction and HPLC methods</a>	Persson, E.; Löfgren, L.; Hansson, G.; Abrahamsson, B.; Lennernäs, H.; Nilsson, R.	<i>J. Lipid Res.</i> 48 (1), 242–251	2007 Jan

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Title	Authors	Publication	Publication Date
<a href="#">The use of charged aerosol detection with HPLC for the measurement of lipids</a>	Plante, M.; Bailey, B.; Acworth, I.	<i>Methods Mol. Biol.</i> (Totowa, NJ, U.S.) 579, 469–482	2009
<a href="#">Comparison between charged aerosol detection and light scattering detection for the analysis of Leishmania membrane phospholipids</a>	Ramos, R. G.; Libong, D.; Rakotomanga, M.; Gaudin, K.; Loiseau, P. M.; Chaminade, P.	<i>J. Chromatogr. A.</i> 1209 (1–2), 88–94	2008 Oct 31
<a href="#">Authentication of geographical origin of palm oil by chromatographic fingerprinting of triacylglycerols and partial least square-discriminant analysis</a>	Ruiz-Samblás, C.; Arrebola-Pascual, C.; Tres, A.; van Ruth, S.; Cuadros-Rodríguez, L.	<i>Talanta.</i> 116, 788–793	2013 Nov 15
<a href="#">Simple and precise detection of lipid compounds present within liposomal formulations using a charged aerosol detector</a>	Schönherr, C.; Touchene, S.; Wilser, G.; Peschka-Süss, R.; Francese, G.	<i>J. Chromatogr. A.</i> 1216 (5), 781–786	2009 Jan 30
<a href="#">Determination of intraluminal individual bile acids by HPLC with charged aerosol detection</a>	Vertzoni, M.; Archontaki, H.; Reppas, C.	<i>J. Lipid Res.</i> 49 (12), 2690–2695	2008 Dec
<a href="#">Neurolipids and the use of a charged aerosol detector</a>	Waraska, J.; Acworth, I.	<i>Am. Biotechnol. Lab.</i> 26 (1), 12–13	2008



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Product Number	Technique	Title
AB 119	UV	Rapid Separation of Paclitaxel and Related Compounds in Paclitaxel Injection
AB 134	MS	LC-MS Analysis of Anthocyanins in Bilberry Extract
AB 139	UV	Separation of Schizandrin, Schizandrin A, and Schizandrin B in a Tablet Sample
AB 153	UV	Save the Flavor – Robust Iso- $\alpha$ -Acids Assaying in Beer within Ten Minutes
AB 155	UV	Monitor the Brewing Process with LC-Transformation of Hop alpha-Acids into Beer Iso-alpha-Acids
AN 109	FLD	Determination of Glyphosate by Cation-Exchange Chromatography with Postcolumn Derivatization
AN 156	UV	The Everlasting Paradigm-Keep Beer Tradition or Prevent Beer from a Skunk Off-Flavor?
AN 196	FLD	Determination of Polycyclic Aromatic Hydrocarbons (PAHs) in Edible Oils by Donor-Acceptor Complex Chromatography (DACC)-HPLC with Fluorescent Detection
AN 207	UV	Chromatographic Fingerprinting of <i>Flos Chrysanthema indicum</i> Using HPLC
AN 213	UV/FLD	Determination of Polycyclic Aromatic Hydrocarbons (PAHs) in Tap Water Using on-Line Solid-Phase Extraction Followed by HPLC with UV and Fluorescence Detections
AN 216	UV	Determination of Water- and Fat-Soluble Vitamins in Functional Waters by HPLC with UV-PDA Detection
AN 224	UV	Determination of Melamine in Milk Powder by Reversed-Phase HPLC with UV Detection
AN 232	UV	Determination of Anthraquinones and Stilbenes in Giant Knotweed Rhizome by HPLC with UV Detection
AN 236	UV	Determination of Iodide and Iodate in Seawater and Iodized Table Salt by HPLC-UV Detection
AN 245	UV	Fast Analysis of Dyes in Foods and Beverages
AN 251	UV	Determination of Water- and Fat-Soluble Vitamins in Nutritional Supplements by HPLC with UV Detection
AN 252	UV	HPLC Assay of Water-Soluble Vitamins, Fat-Soluble Vitamins, and a Preservative in Dry Syrup Multivitamin Formulation
AN 261	UV	Sensitive Determination of Microcystins in Drinking and Environmental Waters
AN 264	UV	Fast Determination of Anthocyanins in Pomegranate Juice
AN 266	FLD	Determination of Sialic Acids Using UHPLC with Fluorescence Detection
AN 272	FLD	Faster Yet Sensitive Determination of N-Methylcarbamates in Rice, Potato, and Corn by HPLC
AN 275	UV	Sensitive Determination of Catechins in Tea by HPLC
AN 287	UV	Two-Dimensional HPLC Combined with On-Line SPE for Determination of Sudan Dyes I–IV in Chili Oil
AN 292	UV	Determination of Aniline and Nitroanilines in Environmental and Drinking Waters by On-Line SPE
AN 293	CAD and UV	Steviol Glycoside Determination by HPLC with Charged Aerosol and UV Detections Using the Acclaim Trinity P1 Column
AN 299	UV	HPLC Analysis of Six Active Components of <i>Caulis Ilicicerae</i> Using a Phenyl-1 Column
AN 1008	UV	Determination of Nitidine Chloride, Toddalolactone, and Chelerythrine Chloride by HPLC

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Product Number	Technique	Title
AN 1020	EC, UV	Chalcinoids and Bitter Acids in Beer by HPLC with UV and ECD
AN 1023	UV	Determination of Sudan Dyes I–IV in Curry Paste
AN 1026	CAD	Fatty Acid Esters at Low Nanogram Levels
AN 1027	CAD	Ginseng
AN 1028	CAD	Ginkgo biloba
AN 1029	CAD	Black Cohosh
AN 1030	CAD	Soy Saponins
AN 1032	CAD	Unsaturated Fatty Acid: Arachidonic, Linoleic, Linolenic and Oleic Acids
AN 1033	CAD	Corn Syrup
AN 1034	CAD	Honey Sugars
AN 1035	CAD	Phenolic Acids
AN 1036	CAD	Water-Soluble Antioxidants: Ascorbic Acid, Glutathione and Uric Acid
AN 1037	CAD	Artificial Sweeteners-Global Method
AN 1039	CAD	Simultaneous Measurement of Glycerides (Mono-, Di- and Triglycerides) and Free Fatty Acids in Palm Oil
AN 1040	CAD	Analysis of Commercially Available Products Containing Stevia
AN 1041	CAD	Phytosterols
AN 1042	UV	Rapid Separation of Anthocyanins in Cranberry and Bilberry Extracts Using a Core-Shell Particle Column
AN 1045	UV	Determination of Phthalates in Drinking Water by UHPLC with UV Detection
AN 1046	UV	Determination of Phenylurea Compounds in Tap Water and Bottled Green Tea
AN 1055	CAD	Determination of Virginiamycin, Erythromycin, and Penicillin in Dried Distillers Grains with Solubles
AN 1063	ECD	Targeted Analyses of Secondary Metabolites in Herbs, Spices, and Beverages Using a Novel Spectro-Electro Array Platform
AN 1064	ECD	Product Authentication and Adulteration Determination Using a Novel Spectro-Electro Array Platform
AN 1067	UV	Determination of Carbendazim in Orange Juice
AN 1069	UV	Two-Dimensional HPLC Determination of Water-Soluble Vitamins in a Nutritional Drink
AN 1070	UV	Determination of Inositol Phosphates in Dried Distillers Grains and Solubles
AN 20583	UV	Determination of Catechins and Phenolic Acids in Red Wine by Solid Phase Extraction and HPLC
AN 20610	UV	Fast Analysis of Coffee Bean Extracts Using a Solid Core HPLC Column
AN 20663	CAD	Comparative Analysis of Cooking Oils Using a Solid Core HPLC Column
AN 20847	CAD	Analysis of a Sports Beverage for Electrolytes and Sugars Using Multi-Mode Chromatography with Charged Aerosol Detection

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AN 70158	CAD	Novel Universal Approach for the Measurement of Natural Products in a Variety of Botanicals and Supplements
AN 70277	CAD	Simultaneous Analysis of Glycerides and Fatty Acids in Palm Oil
AU 144	UV	Determination of Hexavalent Chromium in Drinking Water Using Ion Chromatography
AU 170	UV	Fast Determination of Vanillin and its Synthesis Precursor by HPLC
AU 182	CAD	Measuring Lactose in Milk: A Validated Method
AU 184	CAD, UV	Mogroside V Determination by HPLC with Charged Aerosol and UV Detections
CAN 106	UV	Determination of the Punicagins Found in Pomegranate by High Performance Liquid Chromatography
CAN 111	CAD	Determination of Triterpenes in <i>Centella asiatica</i> (Gotu Kola) by HPLC-CAD
CAN 112	CAD	Determination of Ginsenosides in Panax ginseng by HPLC-CAD
CAN 115	FLD	Clean-Up and Analysis of Aflatoxins and Ochratoxin A in Herbs and Spices
LPN 2062	MS	Profiling Analysis of 15 Prominent Naturally Occurring Phenolic Acids by LC-MS
LPN 2069	FLD	Fast and Effective Determination of Aflatoxins in Grains or Food Using Accelerated Solvent Extraction followed by HPLC
LPN 2421	UV	Achieving Maximum Productivity by Combining UHPLC with Advanced Chromatographic Techniques
LPN 2818	CAD	Analysis of Fat-Soluble Vitamins and Antioxidants in Supplements by RP-HPLC
LPN 2870	FLD	Benefits of High-Speed Wavelength Switching in UHPLC Methods Using Fluorescence Detection
LPN 2930	CAD	Determination of the Composition of Natural Products by HPLC with Charged Aerosol Detection
LPN 2923	CAD	Simple and Direct Analysis of Falcarinol and Other Polyacetylenic Oxylipins in Carrots by Reversed-Phase HPLC and Charged Aerosol Detection
LPN 2931	CAD	Quantification of Underivatized Omega-3 and Omega-6 Fatty Acids in Foods by HPLC CAD
LPN 2932	ECD	A Versatile Detector for the Sensitive and Selective Measurement of Numerous Fat-Soluble Vitamins and Antioxidants in Human Plasma and Plant Extracts
LPN 2934	CAD	Sensitive Analysis of Commonly Used Artificial and Natural Sweeteners Including Stevia and Their Impurities and Degradation Products
LPN 2991	CAD	Evaluation of Methods for the Characterization and Quantification of Polysorbates and Impurities Along with Other Surfactants and Emulsifiers Used in the Food and Pharmaceutical Industries
PN 70026	CAD	Carbohydrate Analysis Using PAD, FLD, CAD and MS Detectors
PN 70037	CAD	Sensitive HPLC Method for Triterpenoid Analysis Using Charged Aerosol Detection with Improved Resolution
PN 70055	CAD	Direct Analysis of Surfactants using HPLC with Charged Aerosol Detection
PN 70138	UV	Rapid Determination of Polyphenol Antioxidants in Green Tea and Cranberry Extract Using Core Shell Columns
PN 70538	CAD	Analysis of Silicone Oils by HPLC-CAD
PN 70540	CAD, ECD	Profiling <i>Hoodia</i> Extracts by HPLC with CAD, ECD, Principal Component Analysis

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Product Number	Technique	Title
AB 127	IC-PAD	Determination of Carbohydrates in Fruit Juice Using Capillary High-Performance Anion-Exchange Chromatography
AB 135	IC-SC	Determination of Anions and Organic Acids in Brewed Coffee Samples Using Capillary IC
AB 137	IC-SC	Determination of Inorganic and Organic Acids in Apple and Orange Juice Samples Using Capillary IC
AN 25	IC-SC	Determination of Inorganic Ions and Organic Acids in Non-Alcoholic Carbonated Beverages
AN 37	IC-PAD	Determination of Iodide and Iodate in Soy- and Mil-Based Infant Formulas
AN 46	IC-PAD	Ion Chromatography: A Versatile Technique for the Analysis of Beer
AN 54	IC-PAD	Determination of Total and Free Sulfite in Foods and Beverages
AN 67	IC-PAD	Determination of Plant-Derived Neutral Oligo- and Polysaccharides
AN 81	IC-SC	Ion Chromatographic Determination of Oxyhalides and Bromide at Trace Level Concentrations in Drinking Water Using direct Injection
AN 82	IC-PAD	Analysis of Fruit Juice Adulterated with Medium Invert Sugar from Beets
AN 87	IC-PAD	Determination of Sugar Alcohols in Confections and Fruit Juices by High-Performance Anion-Exchange Chromatography with Pulsed Amperometric Detection
AN 101	IC-SC	Trace Level Determination of Bromate in Ozonated Drinking Water Using Ion Chromatography
AN 112	IC-UV	Determination of Nitrate and Nitrite in Meat Using High-Performance Anion-Exchange Chromatography
AN 121	IC-SC	Analysis of Low Concentrations of Perchlorate in Drinking Water and Ground Water by Ion Chromatography
AN 123	IC-SC	Determination of Inorganic Anions and Organic Acids in Fermentation Broths
AN 133	IC-SC	Determination of Inorganic Anions in Drinking Water by Ion Chromatography
AN 136	IC-SC and IC-UV	Determination of Inorganic Oxyhalide Disinfection Byproduct Anions and Bromide in Drinking Water Using Ion Chromatography with the Addition of a Postcolumn Reagent for Trace Bromate Analysis
AN 140	IC-SC	Fast Analysis of Anions in Drinking Water by Ion Chromatography
AN 143	IC-SC	Determination of Organic Acids in Fruit Juices
AN 149	IC-SC	Determination of Chlorite, Bromate, Bromide, and Chlorate in Drinking Water by Ion Chromatography with an On-Line-Generated Postcolumn Reagent for Sub- $\mu$ g/L Bromate Analysis
AN 150	IC-PAD	Determination of Amino Acids in Cell Cultures and Fermentation Broths
AN 154	IC-SC	Determination of Inorganic Anions in Environmental Waters Using a Hydroxide-Selective Column
AN 155	IC-PAD	Determination of Trans-Galactooligosaccharides in Foods by AOAC Method 2001.02

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# Technical Collateral: Ion Chromatography Methods

Product Number	Technique	Title
AN 165	IC-SC	Determination of Benzoate in Liquid Food Products by Reagent-Free Ion Chromatography
AN 167	IC-SC	Determination of Trace Concentrations of Oxyhalides and Bromide in Municipal and Bottled Waters Using a Hydroxide-Selective Column with a Reagent-Free Ion Chromatography System
AN 168	IC-UV	Determination of Trace Concentrations of Disinfection By-Product Anions and Bromide in Drinking Water Using Reagent-Free Ion Chromatography Followed by Postcolumn Addition of Iol-Dianisidine for Trace Bromate Analysis
AN 169	IC-SC	Rapid Determination of Phosphate and Citrate in Carbonated Soft Drinks Using a Reagent-Free Ion Chromatography System
AN 172	IC-SC	Determination of Azide in Aqueous Samples by Ion Chromatography with Suppressed Conductivity Detection
AN 173	IC-PAD	Direct Determination of Cyanide in Drinking Water by Ion Chromatography with Pulsed Amperometric Detection (PAD)
AN 178	IC-SC	Improved Determination of Trace Concentrations of Perchlorate in Drinking Water Using Preconcentration with Two-Dimensional Ion Chromatography and Suppressed Conductivity Detection
AN 182	IC-SC and IC-PAD	Determination of Biogenic Amines in Alcoholic Beverages by Ion Chromatography with Suppressed Conductivity and Integrated Pulsed Amperometric Detections
AN 183	IC-SC and IC-PAD	Determination of Biogenic Amines in Fermented and Non-Fermented Foods Using Ion Chromatography with Suppressed Conductivity and Integrated Pulsed Amperometric Detections
AN 187	IC-SC	Determination of sub- $\mu$ g/L Bromate in Municipal Waters Using Preconcentration with Two-Dimensional Ion Chromatography and Suppressed Conductivity Detection
AN 188	IC-PAD	Determination of Glycols and Alcohols in Fermentation Broths Using Ion-Exclusion Chromatography and Pulsed Amperometric Detection
AN 197	IC-PAD	Determination of Glucosamine in Dietary Supplements Using HPAE-PAD
AN 227	ICE-PAD	Determination of Total Cyanide in Municipal Wastewater and Drinking Water Using Ion-Exclusion Chromatography with Pulsed Amperometric Detection (ICE-PAD)
AN 248	IC-PAD	Determination of Lactose in Lactose-Free Milk Products by High-Performance Anion-Exchange Chromatography with Pulsed Amperometric Detection
AN 253	HPAE-PAD	Determination of Infant Formula Sialic Acids
AN 270	IC-PAD	Determination of Hydroxymethylfurfural in Honey and Biomass
AN 273	IC-SC	Determination of Organic Acids in Fruit Juices and Wines by High-Pressure IC
AN 279	IC-SC	Time Savings and Improved Reproducibility of Nitrate and Nitrite Ion Chromatography Determination in Milk Samples
AN 280	IC-PAD	Carbohydrates in Coffee: AOAC Method 995.13 vs a New Fast Ion Chromatography Method
AN 295	IC-SC	Determination of Phytic Acid in Soybeans and Black Sesame Seeds
AN 1007	IC-SC	Determination of Mono-, Di-, and Triphosphates and Citrate in Shrimp by Ion Chromatography

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Product Number	Technique	Title
AN 1044	IC-SC	Determination of Anions in Dried Distillers Grains with Solubles
AN 1068	IC-SC	Determination of Organic Acids in Fruit Juices and Wines by High-Pressure IC
AU 132	IC-UV	Determination of Nitrite and Nitrate in drinking Water by Ion Chromatography with Direct UV Detection
AU 144	IC-UV	Determination of Hexavalent Chromium in Drinking Water Using Ion Chromatography
AU 148	IC-SC	Determination of Perchlorate in Drinking Water Using Reagent-Free Ion Chromatography
AU 150	IC-PAD	Determination of Plant-Derived Neutral Oligo- and Polysaccharides Using the CarboPac PA200
AU 151	IC-PAD	Determination of Sucratose in Reduced- Carbohydrate Colas using High-Performance Anion-Exchange Chromatography with Pulsed Amperometric Detection
AU 189	IC-SC	Determination of Choline in Infant Formula and Other Food Samples by IC
LPN 2982	IC-SC	Determination of Inorganic Anions and Organic Acids in Beverages Using a Capillary IC on a Monolith Anion-Exchange Column
PN 70743	IC-SC	Determination of Perchlorate Levels in Food and Soil Samples Using Accelerated Solvent Extraction and Ion Chromatography
TN 20	IC-PAD	Analysis of Carbohydrates by High-Performance Anion-Exchange Chromatography with Pulsed Amperometric Detection (HPAE-PAD)
TN 126	IC-SC	Determination of Organic Acids in Beer Samples Using a High-Pressure Ion Chromatography System
TN 135	IC-PAD	Determinations of Monosaccharides and Disaccharides in Beverages by Capillary HPAE-PAD

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<a href="#">Multi-residue method for the analysis of pesticide residues in fruits and vegetables by accelerated solvent extraction and capillary gas chromatography</a>	Adou, K.; Bontoyan, W. R.; Sweeney, P. J.	<i>J. Agric. Food Chem.</i> 49 (9), 4153–4160	2001 Sep
<a href="#">The development of an optimized sample preparation for trace level detection of 17<math>\alpha</math>-ethinylestradiol and estrone in whole fish tissue</a>	Al-Ansari, A. M.; Saleem, A.; Kimpe, L. E.; Trudeau, V. L.; Blais, J. M.	<i>J. Chromatogr. B Analys. Technol. Biomed. Life Sci.</i> 879 (30), 3649–52	2011 Nov
<a href="#">Determination of polyphenolic profiles of basque cider apple varieties using accelerated solvent extraction</a>	Alonso-Salces, R. M.; Korta, E.; Barranco, A.; Berrueta, L.A.; Gallo, B.; Vicent, F.	<i>J. Agric. Food Chem.</i> 49 (8), 3761–376	2001
<a href="#">Pressurized liquid extraction for the determination of polyphenols in apple</a>	Alonso-Salces, R. M.; Korta, E.; Barranco, A.; Berrueta, L. A.; Gallo, B.; Vicente, F.	<i>J. Chromatogr. A.</i> 933 (1–2), 37–43	2001 Nov
<a href="#">Methods for extraction and determination of phenolic acids in medicinal plants: a review</a>	Arceusz, A.; Wesolowski, M.; Konieczynski, P.	<i>Nat. Prod. Commun.</i> 8 (12), 1821–9	2013 Dec
<a href="#">Study of an accelerated solvent extraction procedure for the determination of acaricide residues in honey by high-performance liquid chromatography-diode array detector</a>	Bakkali, A.; Korta, E.; Berrueta, L. A.	<i>J. Food Protection</i> 65 (1), 161–166	2002
<a href="#">Pressurized liquid extraction of medicinal plants</a>	Benthin, B.; Danz, H.; Hamburger, M.	<i>J. Chromatogr. A.</i> 837 (1-2), 211–9	1999 Apr
<a href="#">Comparison of the chemical composition of extracts from <i>Scutellaria lateriflora</i> using accelerated solvent extraction and supercritical fluid extraction versus standard hot water or 70% ethanol extraction</a>	Bergeron, C.; Gafner, S.; Clausen, E.; Carrier, D. J.	<i>J. Agric. Food Chem.</i> 53 (8), 3076–80	2005 Apr
<a href="#">Polybrominated diphenyl ethers (PBDEs) in Mediterranean mussels (<i>Mytilus gallo-provincialis</i>) from selected Apulia coastal sites evaluated by GC-HRMS</a>	Bianco, G.; Novario, G.; Anzilotta, G.; Palma, A.; Mangone, A.; Cataldi, T. R.	<i>J. Mass Spectrom.</i> 45 (9), 1046–55	2010 Sep
<a href="#">Free and bound phenolic compounds in barley (<i>Hordeum vulgare L.</i>) flours. Evaluation of the extraction capability of different solvent mixtures and pressurized liquid methods by micellar electrokinetic chromatography and spectrophotometry</a>	Bonoli, M.; Marconi, E.; Caboni, M. F.	<i>J. Chromatogr. A.</i> 19; 1057 (1-2), 1–12	2004 Nov
<a href="#">Pressurized liquid extraction of lipids for the determination of oxysterols in egg-containing food</a>	Boselli, E.; Velazco, V.; Caboni, M. F.; Lercker, G.	<i>J. Chromatogr. A.</i> 11; 917 (1-2), 239–44	2001 May
<a href="#">Optimisation of accelerated solvent extraction of cocaine and benzoylecgonine from coca leaves</a>	Brachet, A.; Rudaz, S.; Mateus, L.; Christen, P.; Veuthey, J-L.	<i>J. Sep. Sci.</i> 24 (10-11), 865–873	2001 Nov

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<a href="#">Influence of extraction methodologies on the analysis of five major volatile aromatic compounds of citronella grass (<i>Cymbopogon nardus</i>) and lemongrass (<i>Cymbopogon citratus</i>) grown in Thailand</a>	Chanthai, S.; Prachakoll, S.; Ruangviriyachai, C.; Luthria, D. L.	<i>J. AOAC Int.</i> 95 (3), 763–72	2012 May-Jun
<a href="#">Accelerated solvent extraction of vitamin K<sub>1</sub> in medical foods in conjunction with matrix solid-phase dispersion</a>	Chase, G. W.; Thompson, B.	<i>J. AOAC Int.</i> 83 (2), 407–10	2000
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<a href="#">Determination of 88 pesticide residues in tea using gas chromatography-tandem mass spectrometry</a>	Chen, H.; Liu, X.; Wang, Q.; Jiang, Y.	<i>Se Pu.</i> 29 (5), 409–16	2011 May
<a href="#">Optimization of accelerated solvent extraction for the determination of chlorinated pesticides from animal feed</a>	Chen, S.; Gfrerer, M.; Lankmayr, E.; Quan, X.; Yang, F.	<i>Chromatographia</i> 58, 631–636	2003
<a href="#">Uptake of oxytetracycline, sulfamethoxazole and ketoconazole from fertilised soils by plants</a>	Chitescu, C. L.; Nicolau, A. I.; Stolker, A. A.	<i>Food Addit. Contam. Part A Chem. Anal. Control Expo. Risk Assess.</i> 30 (6), 1138–46	2013
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<a href="#">Application of accelerated solvent extraction to the investigation of saikosaponins from the roots of <i>Bupleurum falcatum</i></a>	Li, W.; Liu, Z.; Wang, Z.; Chen, L.; Sun, Y.; Hou, J.; Zheng, Y.	<i>J. Sep. Sci.</i> 33 (12), 1870–6	2010 Jun
<a href="#">Applicability of accelerated solvent extraction for synthetic colorants analysis in meat products with ultrahigh performance liquid chromatography-photodiode array detection</a>	Liao, Q. G.; Li ,W. H.; Luo, LG.	<i>Anal. Chim. Acta.</i> 716, 128–32	2012 Feb
<a href="#">Extraction, isolation, and purification of analytes from samples of marine origin – a multivariate task</a>	Liguori, L.; Bjørsvik, H. R.	<i>J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.</i> 910, 46–53	2012 Dec
<a href="#">Investigation on levels of polybrominated diphenyl ethers in retail fish and egg products in Shenzhen</a>	Liu, B.; Zhang, L. S.; Zhang, J. Q.; Jiang, Y. S.; Zhou, J.; Huang, H. Y.	<i>Zhonghua Yu Fang Yi Xue Za Zhi.</i> 45 (12), 1068–72	2011 Dec
<a href="#">Characterization of secondary volatile profiles in <i>Nigella sativa</i> seeds from two different origins using accelerated solvent extraction and gas chromatography-mass spectrometry</a>	Liu, X.; Abd El-Aty, A. M.; Cho, S. K.; Yang, A.; Park, J. H.; Shim, J. H.	<i>Biomed. Chromatogr.</i> 26 (10), 1157–62	2012 Oct
<a href="#">Accelerated solvent extraction of monacolin K from red yeast rice and purification by high-speed counter-current chromatography</a>	Liu, Y.; Guo, X.; Duan, W.; Wang, X.; Du, J.	<i>J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.</i> 878 (28), 2881–5	2010 Oct
<a href="#">Multi-residue determination of organophosphorus pesticides in ginkgo leaves by accelerated solvent extraction and gas chromatography with flame photometric detection</a>	Lu, Y.; Yi, X.	<i>J. AOAC Int.</i> 88 (3), 729–735	2005

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<a href="#">Pressurised solvent extraction for organotin speciation in vegetable matrices</a>	Marcic, C.; Lespes G.; Potin-Gautier, M.	<i>Anal. Bioanal. Chem.</i> 382 (7), 1574–83	2005 Aug
<a href="#">Comparison of different methods for the determination of the oil content in oilseeds</a>	Matthäus, B.; Brühl, L.	<i>J. AOCS</i> 78 95–102.	2001 Jan
<a href="#">A comparison of automated and traditional methods for the extraction of arsenicals from fish</a>	McKiernan, J. W.; Creed, J. T.; Brockhoff, C. A.; Caruso, J. A.; Lorenzana, R. M.	<i>J. Anal. At. Spectrom.</i> 14, 607–613	1999
<a href="#">Subcritical solvent extraction of anthocyanins from dried red grape pomace</a>	Monrad, J. K.; Howard, L. R.; King, J.; Srinivas, K.; Mauromoustakos, A.	<i>J. Agric. Food Chem.</i> 58 (5), 2862–8	2010 Mar
<a href="#">Subcritical solvent extraction of procyanidins from dried red grape pomace</a>	Monrad, J. K.; Howard, L. R.; King, J. W.; Srinivas, K.; Mauromoustakos, A.	<i>J. Agric. Food Chem.</i> 58 (7), 4014–21	2010 Apr
<a href="#">Pressurized liquid extraction of polar and nonpolar lipids in corn and oats with hexane, methylene chloride, isopropanol, and ethanol</a>	Moreau, R. A.; Powell, M. J.; Singh, V.	<i>J. Oil Fat Industr.</i> 80 (11), 1063–1067	2003 Jan
<a href="#">Accelerated solvent extraction for natural products isolation</a>	Mottaleb, M. A.; Sarker, S. D.	<i>Methods Mol. Biol.</i> 864, 75–87	2012
<a href="#">Optimization of extraction process for phenolic acids from black cohosh (<i>Cimicifuga racemosa</i>) by pressurized liquid extraction</a>	Mukhopadhyay, S.; Luthria, D. L.; Robbins, R. J.	<i>J. Sci. Food Agric.</i> 86 (1), 156–162, 15	2006 Jan
<a href="#">Anxiolytic activity of a supercritical carbon dioxide extract of <i>Sououbea sympetala</i> (Marcgraviaceae)</a>	Mullally, M.; Kramp, K.; Cayer, C.; Saleem, A.; Ahmed, F.; McRae, C.; Baker, J.; Goulah, A.; Otorola, M.; Sanchez, P.; Garcia, M.; Poveda, L.; Merali, Z.; Durst, T.; Trudeau, V. L.; Arnason, J. T.	<i>Phytother. Res.</i> 25 (2), 264–70	2011 Feb
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<a href="#">Analysis of multiple herbicides in soybeans using pressurized liquid extraction and capillary electrophoresis</a>	Nemoto, S.; Lehota, S. J.	<i>J. Agric. Food Chem.</i> ; 46 (6), 2190–2199	1998
<a href="#">Comparison of sample preparation methods, validation of an UPLC-MS/MS procedure for the quantification of tetrodotoxin present in marine gastropods and analysis of pufferfish</a>	Nzoughet, J. K.; Campbell, K.; Barnes, P.; Cooper, K. M.; Chevallier, O. P.; Elliott, C. T.	<i>Food Chem.</i> 15; 136 (3–4), 1584–9	2013 Feb
<a href="#">Multiresidue analysis of pesticides in vegetables and fruits using two-layered column with graphitized carbon and water absorbent polymer</a>	Obana, H.; Akutsu, K.; Okihashi, M.; Hori, S.	<i>The Analyst</i> 123, 711–714	1998
<a href="#">Analysis of 2-alkylcyclobutanones with accelerated solvent extraction to detect irradiated meat and fish</a>	Obana, H.; Furuta, M.; Tanaka, Y.	<i>J. Agric. Food Chem.</i> 53 (17), 6603–8	2005 Aug

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<a href="#">Pressurized hot water extraction of berberine, baicalein and glycyrrhizin in medicinal plants</a>	Ong, E. S.; Shea Mei, L.	<i>Anal. Chim. Acta.</i> 482 (1), 81–89	2003 Apr
<a href="#">Pressurized liquid extraction of berberine and aristolochic acids in medicinal plants</a>	Ong E. S.; Woo S. O.; Yong, Y. K.	<i>J. Chromatogr., A.</i> 904 (1), 57–6422	2000 Dec
<a href="#">Rapid determination of pesticide multiresidues in vegetables and fruits by accelerated solvent extraction coupled with online gel permeation chromatography-gas chromatography-mass spectrometry</a>	Ouyang, Y.; Tang, H.; Wu, Y.; Li, G.	<i>Se Pu.</i> 30(7), 654–9	2012 Jul
<a href="#">Determination of zearalenone from wheat and corn by pressurized liquid extraction and liquid chromatography-electrospray mass spectrometry</a>	Pallaroni, L.; von Holst, C.	<i>J. Chromatogr., A.</i> 993, 39–45	2003
<a href="#">Development of an extraction method for the determination of zearalenone in corn using less organic solvents</a>	Pallaroni, L.; von Holst, C.	<i>J. Chromatogr., A.</i> 5 1055 (1-2), 247–9	2004 Nov
<a href="#">Stability of phenolic compounds during extraction with superheated solvents</a>	Palma, M.; Piñeiro, Z.; Barroso, C. G.	<i>J. Chromatogr., A.</i> 6 921 (2), 169–74	2001 Jul
<a href="#">Extraction and analysis of trace amounts of cyclonite (RDX) and its nitroso-metabolites in animal liver tissue using gas chromatography with electron capture detection (GC-ECD)</a>	Pan, X.; Zhang, B.; Cobb, G. P.	<i>Talanta</i> 67 (4), 816–23	2005 Oct
<a href="#">Simultaneous determination of 405 pesticide residues in grain by accelerated solvent extraction then gas chromatography-mass spectrometry or liquid chromatography-tandem mass spectrometry</a>	Pang, G.; Liu, Y.; Fan, C.; Zhang, J.; Cao, Y.; Li, X.; Li, Z.; Wu, Y.; Guo, T.	<i>Anal. Bioanal. Chem.</i> 384, 1366–1408	2006 Mar
<a href="#">Automated sample preparation by pressurized liquid extraction-solid-phase extraction for the liquid chromatographic-mass spectrometric investigation of polyphenols in the brewing process</a>	Papagiannopoulos, M.; Mellenthin, A.	<i>J. Chromatogr., A.</i> 8 976 (1-2), 345–8	2002 Nov
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<a href="#">Determination of catechins by means of extraction with pressurized liquids</a>	Piñeiro, Z.; Palma, M.; Barroso C. G.	<i>J. Chromatogr., A.</i> 13 1026 (1-2), 19–23.	2004 Feb

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<a href="#">Extraction of polar and hydrophobic pollutants using accelerated solvent extraction (ASE)</a>	Pörschmann, J.; Plugge, J.	<i>Fresen. J. Anal. Chem.</i> 364 (7), 643–645	1999
<a href="#">Quantification of the total amount of artemisinin in leaf samples by thin layer chromatography</a>	Quennoz, M.; Bastian, C.; Simonnet, X.; Grogg, A. F.	<i>Chimia (Aarau)</i> 64 (10), 755–7.	2010
<a href="#">Determination of fat in dairy products using pressurized solvent extraction</a>	Richardson, R. K.	<i>J. AOAC Int.</i> 84 (5), 1522–1533	2001
<a href="#">Influence of altitudinal variation on the content of phenolic compounds in wild populations of <i>Calluna vulgaris</i>, <i>Sambucus nigra</i>, and <i>Vaccinium myrtillus</i></a>	Rieger, G.; Müller, M.; Guttenberger, H.; Bucar, F.	<i>J. Agric. Food Chem.</i> 56 (19), 9080–6.	2008 Oct
<a href="#">Pressurized liquid extraction of isoflavones from soybeans</a>	Rostagno, M. A.; Palma, M.; Barroso, C. G.	<i>Anal. Chim. Acta</i> 522 (2), 169–177.	2004 Sep
<a href="#">A multi-residue method for the analysis of organophosphorus residues in cooked and polished rice using accelerated solvent extraction and dispersive-solid phase extraction (D-SPE) technique and uncertainty measurement</a>	Sanyal, D.; Rani, A.; Alam, S.	<i>J. Environ. Sci. Health, B</i> 44 (7), 706–16.	2009 Sep
<a href="#">Accelerated solvent extraction of lipids for determining the fatty acid composition of biological material</a>	Schäfer, K.	<i>Anal. Chim. Acta</i> 358 (1), 69–77	1998 Jan
<a href="#">HPLC analysis of kaempferol and quercetin derivatives isolated by different extraction techniques from plant matrix</a>	Skalicka-Wozniak, K.; Szypowski, J.; Głowniak, K.	<i>J. AOAC Int.</i> 94 (1), 17–21.	Jan-Feb 2011
<a href="#">Statistical evaluation of fatty acid profile and cholesterol content in fish (common carp) lipids obtained by different sample preparation procedures</a>	Spiric, A.; Trbovic, D.; Vranic, D.; Djinovic, J.; Petronijevic, R.; Matekalo-Sverak, V.	<i>Anal. Chim. Acta</i> 672 (1-2), 66–71.	2010 Jul
<a href="#">Application of accelerated solvent extraction in the analysis of organic contaminants, bioactive and nutritional compounds in food and feed</a>	Sun, H.; Ge, X.; Lv, Y.; Wang, A.	<i>J. Chromatogr., A</i> 1237, 1–23.	2012 May
<a href="#">Development of an accelerated solvent extraction, ultrasonic derivatization LC-MS/MS method for the determination of the marker residues of nitrofurans in freshwater fish</a>	Tao, Y.; Chen, D.; Wei, H.; Yuanhu, P.; Liu, Z.; Huang, L.; Wang, Y.; Xie, S.; Yuan, Z.	<i>Food Addit. Contam. Part A Chem. Anal. Control Expo. Risk Assess.</i> 29 (5), 736–45.	2012
<a href="#">Simultaneous determination of lincomycin and spectinomycin residues in animal tissues by gas chromatography-nitrogen phosphorus detection and gas chromatography-mass spectrometry with accelerated solvent extraction</a>	Tao, Y.; Chen, D.; Yu, G.; Yu, H.; Pan, Y.; Wang, Y.; Huang, L.; Yuan, Z.	<i>Food Addit. Contam. Part A Chem. Anal. Control Expo. Risk Assess.</i> 28 (2), 145–54.	2011 Feb
<a href="#">Determination of 17 macrolide antibiotics and avermectins residues in meat with accelerated solvent extraction by liquid chromatography-tandem mass spectrometry</a>	Tao, Y.; Yu, G.; Chen, D.; Pan, Y.; Liu, Z.; Wei, H.; Peng, D.; Huang, L.; Wang, Y.; Yuan, Z.	<i>J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.</i> 897, 64–71.	2012 May
<a href="#">Determination of seven toxaphene congeners in ginseng and milkvetch root by gas chromatography tandem mass spectrometry</a>	Tian, S.; Mao, X.; Miao, S.; Jia, Z.; Wang, K.; Ji, S.	<i>Se Pu</i> 30 (1), 14–20.	2012 Jan

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<a href="#">A consecutive preparation method based upon accelerated solvent extraction and high-speed counter-current chromatography for isolation of aesculin from <i>Cortex fraxinus</i></a>	Tong, X.; Zhou, T; Xiao, X.; Li, G.	<i>J. Sep. Sci.</i> 35 (24), 3609–14	2012 Dec
<a href="#">Characterization of anthocyanins and anthocyanidins in purple-fleshed sweetpotatoes by HPLC-DAD/ESI-MS/MS</a>	Truong, V. D.; Deighton, N.; Thompson, R. T.; McFeeters, R. F.; Dean, L. O.; Pecota, K. V.; Yencho, G. C.	<i>J. Agric. Food Chem.</i> 58 (1), 404–10	2010 Jan
<a href="#">Fat extraction from acid- and base-hydrolyzed food samples using accelerated solvent extraction</a>	Ullah, S. M.; Murphy, B.; Dorich, B.; Richter, B.; Srinivasan, K.	<i>J. Agric. Food Chem.</i> 59 (6), 2169–74.	2011 Mar
<a href="#">Analysis of zearalenone in cereal and swine feed samples using an automated flow-through immunosensor</a>	Urraca, J. L.; Benito-Peña, E.; Pérez-Conde, C.; Moreno-Bondi, M. C.; Pestka, J. J.	<i>J. Agric. Food Chem.</i> 53 (9), 3338–3344	2005
<a href="#">Accelerated solvent extraction and gas chromatography/mass spectrometry for determination of polycyclic aromatic hydrocarbons in smoked food samples</a>	Wang, G.; Lee, A. S.; Lewis, M.; Kamath, B.; Archer, R. K.	<i>J. Agric. Food Chem.</i> 47 (3), 1062–6.	1999 Mar
<a href="#">Subcritical water extraction of alkaloids in <i>Sophora flavescens</i> Ait. and determination by capillary electrophoresis with field-amplified sample stacking</a>	Wang, H.; Lu, Y.; Chen, J.; Li, J.; Liu, S.	<i>J. Pharm. Biomed. Anal.</i> 58, 146–51.	2012 Jan
<a href="#">Evaluation of Soxhlet extraction, accelerated solvent extraction and microwave-assisted extraction for the determination of polychlorinated biphenyls and polybrominated diphenyl ethers in soil and fish samples</a>	Wang, P.; Zhang, Q.; Wang, Y.; Wang, T.; Li X.; Ding, L.; Jiang, G.	<i>Anal. Chim. Acta.</i> 663 (1), 43–8.	2010 Mar
<a href="#">Determination of ten pesticides of pyrazoles and pyrroles in tea by accelerated solvent extraction coupled with gas chromatography-tandem mass spectrometry</a>	Xu, D.; Lu, S.; Chen, D.; Lan, J.; Zhang, Z.; Yang, F.; Zhou, Y.	<i>Se Pu.</i> 31 (3), 218–22.	2013 Mar
<a href="#">Online cleanup of accelerated solvent extractions for determination of adenosine 5'-triphosphate (ATP), adenosine 5'-diphosphate (ADP), and adenosine 5'-monophosphate (AMP) in royal jelly using high-performance liquid chromatography</a>	Xue, X.; Wang, F.; Zhou, J.; Chen, F.; Li, Y.; Zhao, J.	<i>J. Agric. Food Chem.</i> 57 (11), 4500–5.	2009 Jun
<a href="#">Identification and quantitation of eleven sesquiterpenes in three species of <i>Curcuma</i> rhizomes by pressurized liquid extraction and gas chromatography-mass spectrometry</a>	Yang, F. Q.; Li ,S.; Chen, Y.; Lao, S. C.; Wang, YT.; Dong, T. T. X.; Tsim, K. W. K.	<i>J. Pharm. Biomed. Anal.</i> 39 (3/4), 552–558	2005 Sep
<a href="#">Dispersive solid-phase extraction cleanup combined with accelerated solvent extraction for the determination of carbamate pesticide residues in <i>Radix glycyrrhizae</i> samples by UPLC-MS-MS</a>	Yang, R. Z.; Wang, J. H.; Wang, M. L.; Zhang, R.; Lu, X. Y.; Liu, W. H.	<i>J. Chromatogr. Sci.</i> 49 (9), 702–8.	2011 Oct
<a href="#">Simultaneous determination of amitraz and its metabolite residue in food animal tissues by gas chromatography-electron capture detector and gas chromatography-mass spectrometry with accelerated solvent extraction</a>	Yu, H.; Tao, Y.; Le, T.; Chen, D.; Ihsan, A.; Liu, Y.; Wang, Y.; Yuan, Z.	<i>J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.</i> 878 (21), 1746–52.	2010 Jul
<a href="#">Simultaneous determination of fluoroquinolones in foods of animal origin by a high performance liquid chromatography and a liquid chromatography tandem mass spectrometry with accelerated solvent extraction</a>	Yu, H.; Tao, Y.; Chen, D.; Pan, Y.; Liu, Z.; Wang, Y.; Huang, L.; Dai, M.; Peng, D.; Wang, X.; Yuan, Z.	<i>J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.</i> 885-886, 150–9.	2012 Feb

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<a href="#">Response surface modeling and optimization of accelerated solvent extraction of four lignans from <i>fructus schisandrae</i></a>	Zhao, L. C.; He, Y.; Deng, X.; Yang, G. L.; Li, W.; Liang, J.; Tang, Q. L.	<i>Molecules</i> . 17 (4), 3618–29	2012 Mar
<a href="#">Determination of acetanilide herbicides in cereal crops using accelerated solvent extraction, solid-phase extraction and gas chromatography-electron capture detector</a>	Zhang, Y.; Yang, J.; Shi, R.; Su, Q.; Yao, L.; Li, P.	<i>J. Sep. Sci.</i> 34 (14), 1675–82	2011 Jul
<a href="#">Application of accelerated solvent extraction coupled with high-performance counter-current chromatography to extraction and online isolation of chemical constituents from <i>Hypericum perforatum</i> L</a>	Zhang, Y.; Liu, C.; Yu, M.; Zhang, Z.; Qi, Y.; Wang, J.; Wu, G.; Li, S.; Yu, J.; Hu, Y.	<i>J. Chromatogr., A.</i> 1218 (20), 2827–34	2011 May
<a href="#">Analysis of volatile components in Qingshanlvshui tea using solid-phase microextraction/accelerated solvent extraction-gas chromatography-mass spectrometry</a>	Zhan, J.; Lu, S.; Meng, Z.; Xiang, N.; Cao, Q.; Miao, M.	<i>Se Pu</i> . 26 (3), 301–5.	2008 May



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AN 335	HPLC-UV	Accelerated Solvent Extraction (ASE) of Active Ingredients from Natural Products
AN 356	IC-conductivity	Determination of Perchlorate in Vegetation Samples Using Accelerated Solvent Extraction and Ion Chromatography
AN 357	HPLC	Extraction of Phenolic Acids from Plant Tissue Using Accelerated Solvent Extraction (ASE)
AN 363	HPLC	Extraction of Herbal Marker Compounds Using Accelerated Solvent Extraction Compared to Traditional Pharmacopoeia Protocols



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