

# Simultaneous Phosphorus and Sulfur Speciation by HPLC Interfaced with High Resolution ICP-MS

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## Abstract

A highly sensitive method for the simultaneous speciation of phosphorus and sulfur is described. Phosphorus and sulfur containing molecules are separated by High Performance Liquid Chromatography (HPLC) and detected on-line by High Resolution Sector Field Inductively Coupled Plasma Mass Spectrometry (ICP-SFMS). With a resolution of 4000 ( $R = m/\Delta m$ ), phosphorus and sulfur are completely resolved from all polyatomic interferences, resulting in high selectivity. This allows the use of gradient organic mobile phases of up to 100% acetonitrile, which would normally lead to worsened analytical performance, due to the formation of spectral interferences. By using high mass resolution simple and clear spectra are obtained without creating new interferences. Detection limits of 0.06 to 0.19 ng g<sup>-1</sup> P and 1.3 to 1.9 ng g<sup>-1</sup> S are obtained, corresponding to absolute amounts of 0.6 to 1.9 pg P and 13 to 19 pg S. Sensitivities of up to 1700 cps / ng g<sup>-1</sup> P and up to 1100 cps / ng g<sup>-1</sup> S are achieved. The investigated compounds are deoxyribonucleotides, peptides, l-methionine and N-acetyl-dl-methionine. Analytical precisions are between 2.3 and 3.8% RSD. Phosphorus and sulfur can be simultaneously detected within one chromatographic run with a scan duty cycle of 99.8%. This work shows the potential of ICP-SFMS for the determination of phosphorylation states in proteins.

## Introduction

Phosphorus and sulfur containing molecules play an important role in biochemistry and proteomics. Phosphorus is present in the backbone of the DNA and RNA chain and in phospholipids. It is involved in energy storage processes and protein phosphorylation. The sulfur containing amino acids cysteine and methionine are found in proteins. Some other examples of sulfur containing biomolecules are: thiaminepyrophosphate, liponamide, acetyl coenzyme A, glutathione and S-adenosylmethionine.

The classical techniques for the investigation of phosphorus and sulfur in these biomolecules are ESI-MS (electrospray ionization mass spectrometry) and MALDI-MS (matrix assisted laser desorption/ionization mass spectrometry). Other techniques employed are based on the incorporation of radioisotopes (<sup>32</sup>P, <sup>33</sup>P or <sup>35</sup>S) and subsequent monitoring of the  $\beta$  radiation emitted. More recent developments for the determination of phosphorus and sulfur compounds are the hyphenation of HPLC with ICP-MS<sup>[1-15]</sup> and gel electrophoresis followed by laser ablation with ICP-MS detection<sup>[16-19]</sup>.

The fundamental limitation in the detection of phosphorus and sulfur by ICP-MS is the existence of polyatomic interferences formed in the ICP ion source; for example: <sup>15</sup>N<sup>16</sup>O<sup>+</sup>, <sup>14</sup>N<sup>17</sup>O<sup>+</sup>, <sup>14</sup>N<sup>16</sup>O<sup>1</sup>H<sup>+</sup>, <sup>12</sup>C<sup>18</sup>O<sup>1</sup>H<sup>+</sup> and <sup>1</sup>H<sub>3</sub><sup>12</sup>C<sup>16</sup>O<sup>+</sup> at mass <sup>31</sup>P (Figures 1 and 2), <sup>16</sup>O<sup>16</sup>O<sup>+</sup>, <sup>14</sup>N<sup>18</sup>O<sup>+</sup> and <sup>15</sup>N<sup>16</sup>O<sup>1</sup>H<sup>+</sup> on mass <sup>32</sup>S (Figure 3) and <sup>16</sup>O<sup>18</sup>O<sup>+</sup> and <sup>1</sup>H<sub>2</sub><sup>16</sup>O<sub>2</sub><sup>+</sup> on mass <sup>34</sup>S (Figure 4).

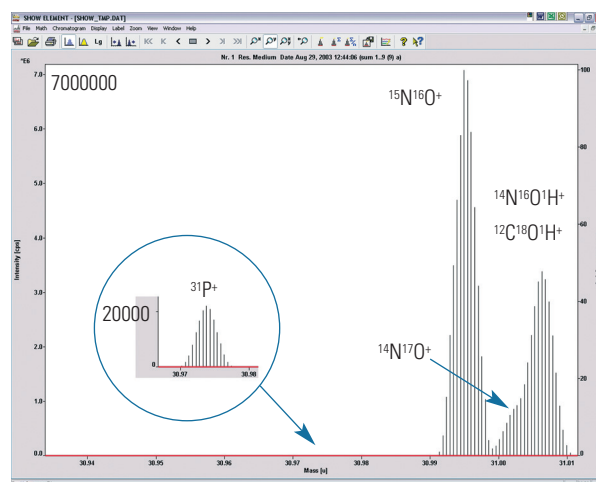


Figure 1: Phosphorus interferences in acetonitrile.

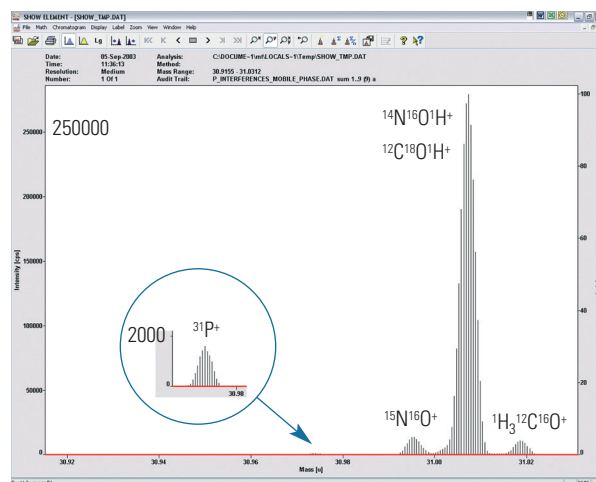


Figure 2: Phosphorus interferences in 4 mmol L<sup>-1</sup> ammonium acetate and 2% methanol.

## Key Words

- ELEMENT 2
- Chromatography
- High Resolution ICP-MS
- Phosphorus and Sulfur Speciation
- Phosphorylation
- Proteomics

	P SPECIATION OF DEOXYRIBONUCLEOTIDES	S SPECIATION OF SULFUR COMPOUNDS	P & S SPECIATION OF PHOSPHORYLATED PEPTIDES
Stationary phase	Discovery RP Amide C16	Discovery RP Amide C16	Discovery RP Amide C16
Mobile phase	4 mmol L <sup>-1</sup> ammonium acetate, pH 6.4, 2% methanol (v/v)	0.1% trifluoroacetic acid, 10% acetonitrile, 89.9% water (v/v/v)	A = 0.1% trifluoroacetic acid in water (v/v) B = 0.1% trifluoroacetic acid in acetonitrile (v/v)
Isocratic / Gradient	Isocratic	Isocratic	Gradient: 10% B to 100% B (3-15 min)
Sample loop	10 µL	10 µL	10 µL
Flow rate	200 µL/min	200 µL/min	200 µL/min
Nebulizer	PFA µ-Flow-LC	PFA µ-Flow-LC	PFA µ-Flow-LC
Sample gas	1.09 L/min Ar	0.69 L/min Ar	0.69 L/min Ar
Additional Gas	---	0.04 L/min O <sub>2</sub>	0.04 L/min O <sub>2</sub>
Spray chamber	Quartz, double pass, room temperature	Quartz, double pass, peltier-cooled to -2 °C	Quartz, double pass, peltier-cooled to -5 °C
Injector	Quartz, 1.75 mm ID	Quartz, 1mm ID	Quartz, 1 mm ID
Cones	Ni sampler and Ni-X skimmer	Pt sampler and Pt-X skimmer	Pt sampler and Pt-X skimmer
RF power	1450 W	1450 W	1450 W
Isotopes monitored	<sup>31</sup> P	<sup>32</sup> S	<sup>31</sup> P and <sup>32</sup> S
Resolution	Medium (4000)	Medium (4000)	Medium (4000)

Table 1: Instrument settings for the analysis of deoxyribonucleotides, sulfur compounds and phosphorylated peptides.

One approach used to reduce these interferences is the application of collision/reaction cell ICP-MS. Helium has been reported as collision gas for reducing interferences on <sup>31</sup>P<sup>[6]</sup>. But due to the stability of the O<sub>2</sub><sup>+</sup> interference, helium is not an effective collision gas for the determination of <sup>32</sup>S or <sup>34</sup>S. While xenon is useful for reducing interferences at sulfur, it has the disadvantage of reducing sensitivities for other elements. Therefore, either a single, non-ideal collision gas has to be used for all elements or ideal collision gases have to be changed repeatedly in a single chromatographic run, leading to long delays.

A simpler approach is the application of High Resolution Sector Field ICP-MS (ICP-SFMS). This technique avoids the use of any collision gas. The principle of ICP-SFMS is to physically resolve the analyte ions from interferences by their small difference in mass. A resolution of 4000 is sufficient to completely resolve phosphorus and sulfur from all interferences even in 100% organic solvents. Other advantages of ICP-SFMS are the outstanding sensitivity and a very high signal-to-noise ratio, which result in extremely low detection limits.

ICP-SFMS has been widely used for the speciation of metals, e.g. aluminum<sup>[20]</sup>, calcium<sup>[21-23]</sup>, chromium<sup>[23-26]</sup>, manganese<sup>[23]</sup>, vanadium<sup>[25,27]</sup>, iron<sup>[19,23,25,27-30]</sup>, nickel<sup>[25,26,31]</sup>, copper<sup>[19,23,25,26,29,32-35]</sup>, zinc<sup>[19,23,25,26,29,33,34-36]</sup>, arsenic<sup>[37-41]</sup>, selenium<sup>[23,38,42-45]</sup>, rhodium<sup>[46]</sup>, palladium<sup>[46]</sup>, cadmium<sup>[25,26,33-35,47]</sup>, tin<sup>[23,48]</sup>, antimony<sup>[26]</sup>, tellurium<sup>[38]</sup>, neodymium<sup>[49]</sup>, gadolinium<sup>[15]</sup>, platinum<sup>[22,46,50]</sup>, mercury<sup>[51]</sup>, lead<sup>[23,26]</sup>, thorium<sup>[25]</sup> and uranium<sup>[25]</sup>. It has also been used for the speciation of silicon<sup>[52]</sup> and non-metals like iodine<sup>[5,26,53]</sup>, phosphorus<sup>[1-5,12,13,16-19,52]</sup> and sulfur<sup>[2,8-11,14-16,18,19,22,27,33-35]</sup>.

This application note shows the potential of HPLC interfaced to ICP-SFMS for the investigation of phosphorus and sulfur containing biomolecules and evaluates this technique through the analysis of deoxyribonucleotides, sulfur-containing molecules and peptides.

## Experimental

The Thermo Scientific ELEMENT 2 High Resolution Sector Field ICP-MS was used with the Organic Matrix Kit (part number: 1067531; including an oxygen mass flow controller, platinum-tipped sampler and skimmer cones and a 1.0 mm quartz injector). A new design of Peltier cooled spray chamber with software based temperature control was used in order to reduce the solvent load in the plasma by decreasing the spray chamber temperature.

For acetonitrile containing solvents, oxygen was added in the spray chamber in order to maintain a stable plasma and to avoid carbon deposition on the cones. The highly increased oxides (e.g. NO, NOH, COH, O<sub>2</sub>, H<sub>2</sub>O<sub>2</sub>) resulting are completely resolved from phosphorus and sulfur with a mass resolution of 4000.

The HPLC system consisted of the Thermo Scientific Surveyor™ LC Pump, containing a 4-channel in-line degasser and a six-port injection valve (Model 9125 from Rheodyne, Rohnert Park, CA) with a 100 µL PEEK™ sample loop. Separation was achieved using a Discovery™ RP Amide C16 column (150 x 2.1 mm, 5 µm, from Supelco®, Bellefonte, PA).

The HPLC and ICP-MS were connected with a PEEK capillary (0.25 mm ID), a PFA µ-flow nebulizer and a trigger cable (all contained in the HPLC Connection Kit, part number: 1159340). Data acquisition by the Element Software can be triggered by either contact closure or a 5 volts TTL signal from the HPLC system. In this work contact closure of the injection valve was used. Chromatograms can be automatically exported on-line in ASCII, ANDI (AIA netCDF), GRAMS™, Xcalibur™ and Spectacle compatible file formats. In this work, data evaluation was performed in GRAMS/AI v.7 software using peak heights. The instrument settings for different separation tasks are summarized in Table 1. Sample gas and torch position were optimized by tuning on the <sup>74</sup>Ge signal of the mobile phase containing 50 ng g<sup>-1</sup> germanium.

## Results and Discussion

With a mass resolution of 4000, all polyatomic interferences on phosphorus and sulfur are eliminated. This is demonstrated for phosphorus in Figures 1 and 2, which show that the  $^{31}\text{P}$  peak at  $m/z$  30.974 is completely resolved from interferences originating from different mobile phases. In acetonitrile, the most abundant interferences are  $^{15}\text{N}^{16}\text{O}^+$ ,  $^{14}\text{N}^{17}\text{O}^+$ ,  $^{14}\text{N}^{16}\text{O}^+\text{H}^+$  and  $^{12}\text{C}^{18}\text{O}^+\text{H}^+$  (Figure 1).

Even in this solvent, consisting largely of just carbon and nitrogen, phosphorus can be resolved from the carbon and nitrogen based interferences. In a 2% methanol solution containing 4 mmol  $\text{L}^{-1}$  ammonium acetate, an additional interference from  $^1\text{H}_3^{12}\text{C}^{16}\text{O}^+$  can be detected (Figure 2).

Figure 3 shows that  $^{32}\text{S}$  is resolved from all interferences with a resolution of 4000. The most abundant interferences on  $^{32}\text{S}$  in acetonitrile are  $^{16}\text{O}^{16}\text{O}^+$ ,  $^{14}\text{N}^{18}\text{O}^+$  and  $^{15}\text{N}^{16}\text{O}^+\text{H}^+$ . Also  $^{34}\text{S}$  is resolved from all interferences, mainly  $^{16}\text{O}^{18}\text{O}^+$  and  $^1\text{H}_2^{16}\text{O}_2^+$  (Figure 4). This enables the analysis of isotopically enriched sulfur compounds, which is necessary for sulfur tracer studies.

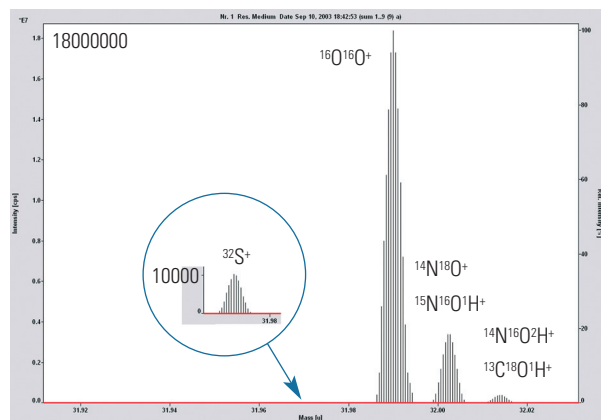


Figure 3:  $^{32}\text{S}$  interferences in acetonitrile.

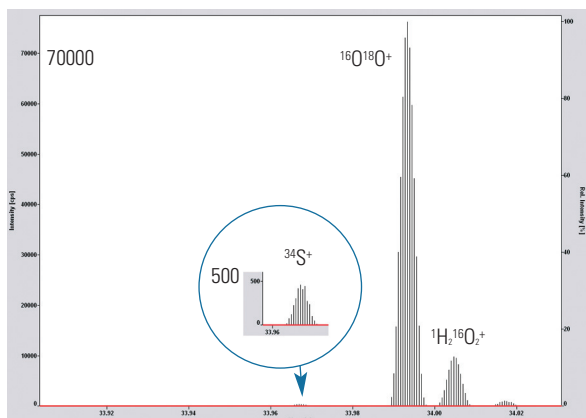


Figure 4:  $^{34}\text{S}$  interferences in acetonitrile.

By using Medium Resolution ( $m/\Delta m = 4000$ ),  $^{31}\text{P}$ ,  $^{32}\text{S}$  and  $^{34}\text{S}$  are completely resolved from all carbon and nitrogen based polyatomic interferences, resulting in extraordinary specificity. Simple and clear spectra are obtained without creating new interferences and without changing any ICP-MS conditions.

## Phosphorus Speciation in a Mixture of Deoxyribonucleotides

In order to show the potential of HPLC interfaced to Sector Field ICP-MS for phosphorus speciation, a mixture of four deoxyribonucleotides (dAMP, dTMP, dGMP and dCMP) has been analyzed with this technique, using the parameters listed in Table 1. A chromatogram obtained after the injection of a mixture containing 50 ng  $\text{g}^{-1}$  P of each deoxyribonucleotide is shown in Figure 5.

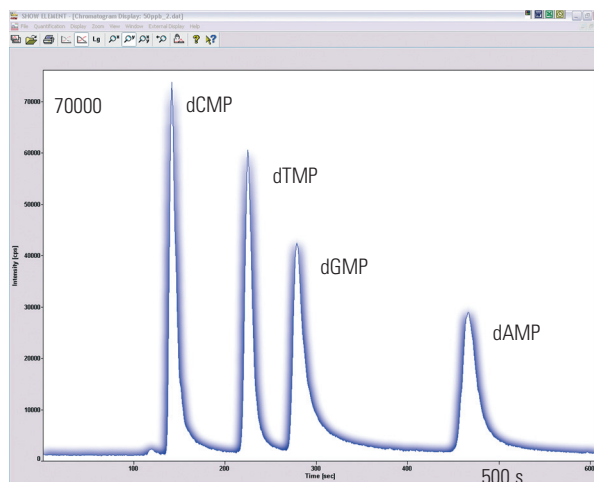


Figure 5: Online P chromatogram of a mixture containing 50 ng  $\text{g}^{-1}$  P of each deoxyribonucleotide.

The figures of merit, obtained by evaluation of peak heights, are shown in Table 2.

RETENTION TIME [min]	CALIBRATED RANGE [ng $\text{g}^{-1}$ P]	RSD [%] height*	RSD [%] time**	SENSITIVITY [cps/ng $\text{g}^{-1}$ P]	LOD [ng $\text{g}^{-1}$ P]	LOD [pg P]
dCMP 2.4	1 - 200	2.6	0.6	1681	0.11	1.1
dTMP 3.7	1 - 200	3.8	0.4	1276	0.07	0.7
dGMP 4.6	1 - 200	3.4	0.8	1013	0.19	1.9
dAMP 7.7	1 - 200	2.9	0.7	581	0.06	0.6

\*  $n = 5$  (10 ng  $\text{g}^{-1}$  P)

\*\*  $n = 19$

Table 2: Figures of merit for the determination of deoxyribonucleotides.

Limits of detection were calculated by dividing three times the standard deviation from 8 measurements of a low concentration standard by the slope of the calibration curve. The limits of detection obtained range between 0.6 to 1.9 pg P (0.06 to 0.19 ng  $\text{g}^{-1}$ ) and are limited by effects from the chromatographic system, e.g. P contamination of the mobile phase and column washout effects after the injection of high concentration samples etc. With HPLC interfaced to a collision cell ICP-MS, detection limits of 30 to 60 pg P (3 - 6 ng  $\text{g}^{-1}$ ) have been reported for the speciation of the same deoxyribonucleotides by Proefrock et al.<sup>[6]</sup>. For the speciation of phospholipids by HPLC interfaced to collision cell ICP-MS, detection limits of 210 to 1200 pg P have been reported by Kovacevic et al.<sup>[7]</sup>. The low limits of detection achievable with High Resolution Sector Field ICP-MS are due to the high sensitivity (600 to 1700 cps/ng  $\text{g}^{-1}$  P), high ion transmission and the specificity, caused by the unambiguous mass resolution of the analyte from polyatomic interferences. The different sensitivities obtained for different species are due to the different peak heights in the chromatogram.

A wide linear dynamic range (1 to 200 ng g<sup>-1</sup> P) and good stability and precision (2 to 4% RSD) are also achieved. As an example, the calibration curve for dAMP is shown in Figure 6.

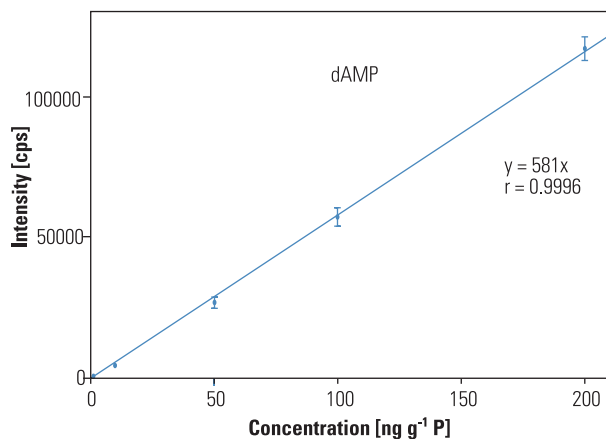


Figure 6: Calibration curve for dAMP.

### Sulfur Speciation

L-Methionine and N-acetyl-dl-methionine were chosen as sulfur containing model compounds in order to evaluate the performance of HPLC interfaced to Sector Field ICP-MS for sulfur speciation. The chromatogram obtained after injection of a mixture containing 250 ng g<sup>-1</sup> S of each species is shown in Figure 7. Figures of merit are listed in Table 3. Limits of detection range between 1.3 to 1.9 ng g<sup>-1</sup> S, equivalent to 13 to 19 pg S and are limited by the sulfur content of the mobile phase. The sensitivity is approximately 1000 cps / ng g<sup>-1</sup> S.

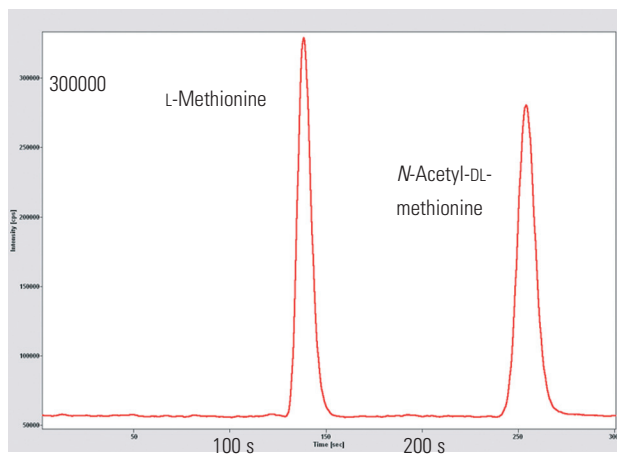


Figure 7: Online S chromatogram of L-Methionine and N-Acetyl-DL-methionine (each 250 ng g<sup>-1</sup> S).

	RETENTION TIME [min]	CALIBRATED RANGE [ng g <sup>-1</sup> S]	RSD [%]* height	RSD [%]* time	SENSITIVITY [cps/ng g <sup>-1</sup> S]	LOD [ng g <sup>-1</sup> S]	LOD [pg S]
L-M.	2.4	10 - 2000	2.3	0.6	1095	1.3	13
N-A.	4.3	10 - 2000	2.3	0.3	917	1.9	19

Table 3: Figures of merit for sulfur speciation.

### Simultaneous Phosphorus and Sulfur Speciation of Phosphorylated Peptides

Unlike any other technique, both phosphorus and sulfur can be detected simultaneously in organic mobile phases by High Resolution Sector Field ICP-MS using identical instrument conditions. With a resolution of 4000, P and S analyte ions are resolved from all polyatomic interferences originating from the mobile phase. Therefore this technique, when interfaced to HPLC, is ideally suited for the simultaneous speciation of phosphorus and sulfur, e.g. for the investigation of phosphorylated peptides. In order to demonstrate this, a tryptic digest of casein was separated by HPLC with on-line phosphorus and sulfur detection by Sector Field ICP-MS. The chromatogram obtained is shown in Figure 8.

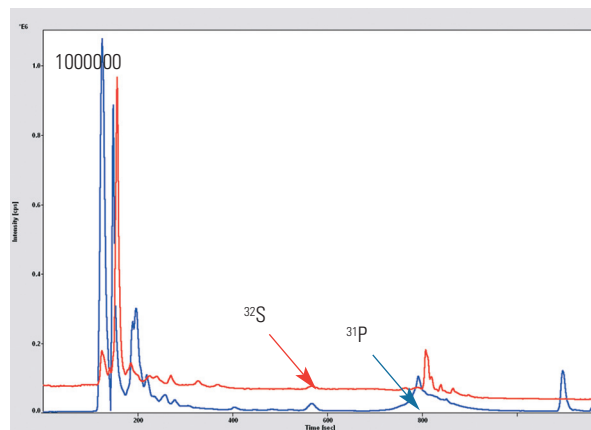


Figure 8: Online P and S chromatogram of a tryptic digest of casein, containing phosphorylated peptides.

The peaks of the <sup>31</sup>P trace correspond to phosphorylated peptides and the peaks of the <sup>32</sup>S trace correspond to peptides with sulfur containing amino acids. The chromatogram shows that, even with gradients from 10 to 100% acetonitrile (which are frequently used in proteomics), no drift of the baseline is observed as has been seen with other ICP techniques due to changing levels of interferences. The duty cycle (time spent actually acquiring data divided by the total analysis time) of the method used was 99.8%. Advantages of High Resolution ICP-MS for this application are:

- All polyatomic interferences are completely physically resolved from P and S with a single set of ICP-MS parameters
- Phosphorus and sulfur can be detected simultaneously without any collision gas
- Chromatographic gradients can be used and the variable organic content of the mobile phase does not lead to an increased baseline
- High sensitivity (up to 1700 cps / ng g<sup>-1</sup> P and 1100 cps / ng g<sup>-1</sup> S)
- The <sup>31</sup>P signal intensity is independent of the chemical form of phosphorus and proportional to the molar amount of phosphorus in the HPLC eluate

## Conclusions

- With on-line HPLC/ICP-SFMS, the simultaneous speciation of phosphorus and sulfur has been demonstrated. The determination of phosphorus and sulfur (as a measure of the protein or peptide content) enables the determination of phosphorylation states.
- By using Sector Field ICP-MS, phosphorus and sulfur are completely resolved from all interferences, resulting in high specificity.
- Simple and clear spectra are obtained without creating new interferences.
- Very low limits of detection for phosphorus and sulfur speciation are possible with this technique. These low detection limits are due to the high sensitivity and the low background that is not influenced by polyatomic interferences.
- Gradients can be used, because the chromatographic phosphorus and sulfur baselines are not affected by interferences from different solvents.
- Organic solvents like acetonitrile are frequently used as a mobile phase in proteomics for the separation of peptides. Therefore the complete removal of interferences caused by organic solvents is important for biochemical applications.
- Since the analytes are completely resolved from all interferences by the small difference in mass, Sector Field ICP-MS is matrix independent, making it an ideal tool for speciation analysis.

## References

- [1] Wind, M., Edler, M., Jakubowski, N., Linscheid, M., Wesch, H., Lehmann, W.D., 2001. Analysis of protein phosphorylation by capillary liquid chromatography coupled to element mass spectrometry with 31P detection and to electrospray mass spectrometry. *Anal. Chem.*, 73, 29-35.
- [2] Wind, M., Wesch, H., Lehmann, W.D., 2001. Protein phosphorylation degree: determination by capillary liquid chromatography and inductively coupled plasma mass spectrometry. *Anal. Chem.*, 73, 3006-3010.
- [3] Wind, M., Gosenca, D., Kuebler, D., Lehmann, W.D., 2003. Stable isotope phospho-profiling of fibrinogen and fetuin subunits by element mass spectrometry coupled to capillary liquid chromatography. *Anal. Biochem.*, 317, 26-33.
- [4] Wind, M., Kelm, O., Nigg, E.A., Lehmann, W.D., 2002. Identification of phosphorylation sites in the polo-like kinase Plx1 and Plk1 by a novel strategy based on element and electrospray high resolution mass spectrometry. *Proteomics*, 2, 1516-1523.
- [5] Wind, M., Eisenmenger, A., Lehmann, W.D., 2002. Modified direct injection high efficiency nebulizer with minimized dead volume for the analysis of biological samples by micro- and nano LC-ICP-MS. *J. Anal. At. Spectrom.*, 17, 21-26.
- [6] Proefrock, D., Leonhard, P., Prange, A., 2003. Determination of phosphorus in phosphorylated deoxyribonucleotides using capillary electrophoresis and high performance liquid chromatography hyphenated to inductively coupled plasma mass spectrometry with an octopole reaction cell. *J. Anal. At. Spectrom.*, 18, 708-713.
- [7] Kovacevic, M., Leber, R., Kohlwein, S.D., Goessler, W., 2004. Application of inductively coupled plasma mass spectrometry to phospholipid analysis. *J. Anal. At. Spectrom.*, 19, 80-84.
- [8] Wind, M., Wegener, A., Eisenmenger, R., Kellner, R., Lehmann, W.D., 2003. Sulfur as the key element for quantitative protein analysis by capillary liquid chromatography coupled to element mass spectrometry. *Angew. Chem.*, 42, 3425-3427.
- [9] de la Flor St. Remy, R.R., Montes-Bayon, M., Sanz-Medel, A., 2003. Determination of total homocysteine in human serum by capillary gas chromatography with sulfur-specific detection by double focusing ICP-MS. *Anal. Bioanal. Chem.*, 377, 299-305.
- [10] Rodriguez-Fernandez, J., Montes-Bayon, M., Pereiro, R., Sanz-Medel, A., 2001. Gas chromatography double focusing sector-field ICP-MS as an innovative tool for bad breath research. *J. Anal. At. Spectrom.*, 16, 1051-1056.
- [11] Montes-Bayon, M., Meija, J., LeDuc, D.L., Terry, N., Caruso, J.A., Sanz-Medel, A., 2004. HPLC-ICP-MS and ESI-Q-TOF analysis of biomolecules induced in *Brassica juncea* during arsenic accumulation. *J. Anal. At. Spectrom.*, 19, 153-158.
- [12] Helferich, A., Bettmer, J., 2004. Determination of phytic acid and its degradation products by ion-pair chromatography (IPC) coupled to inductively coupled plasma-sector field-mass spectrometry (ICP-SF-MS). *J. Anal. At. Spectrom.*, 19, 1330-1334.
- [13] Cartwright, A.J., Jones, P., Wolff, J.C., Evans, E.H., 2005. Detection of phosphorus tagged carboxylic acids using HPLC-SF-ICP-MS. *J. Anal. At. Spectrom.*, 20, 75-80.
- [14] Evans, E.H., Wolff, J.C., Eckers, C., 2001. Sulfur-specific detection of impurities in Cimetidine drug substance using liquid chromatography coupled to high resolution inductively coupled plasma mass spectrometry and electrospray mass spectrometry. *Anal. Chem.*, 73, 4722-4728.
- [15] Krueger, R., Braun, K., Pipkorn, R., Lehmann, W.D., 2004. Characterization of a gadolinium-tagged molecular contrast agent by element and molecular mass spectrometry. *J. Anal. At. Spectrom.*, 19, 852-857.
- [16] Becker, J.S., Zoriy, M., Becker, J.S., Pickhardt, C., Przybylski, M., 2004. Determination of phosphorus and metals in human brain proteins after isolation by gel electrophoresis by laser ablation inductively coupled plasma source mass spectrometry. *J. Anal. At. Spectrom.*, 19, 149-152.
- [17] Wind, M., Feldmann, I., Jakubowski, N., Lehmann, W.D., 2003. Spotting and quantification of phosphoproteins purified by gel electrophoresis and laser ablation-element mass spectrometry with phosphorus-31 detection. *Electrophoresis*, 24, 1276-1280.
- [18] Becker, J.S., Boulyga, S.F., Becker, J.S., Pickhardt, C., Damoc, E., Przybylski, M., 2003. Structural identification and quantification of protein phosphorylations after gel electrophoretic separation using fourier transform ion cyclotron resonance mass spectrometry and laser ablation inductively coupled plasma mass spectrometry. *Int. J. Mass Spectrom.*, 228, 985-997.
- [19] Becker, J.S., Zoriy, M., Krause-Buchholz, U., Becker, J.S., Pickhardt, C., Przybylski, M., Pompe, W., Rödel, G., 2004. In-gel screening of phosphorus and copper, zinc and iron in proteins of yeast mitochondria by LA-ICP-MS and identification of phosphorylated protein structures by MALDI-FT-ICR-MS after separation with two-dimensional gel electrophoresis. *J. Anal. At. Spectrom.*, 19, 1236-1243.
- [20] Tsunoda, K., Umemura, T., Ohshima, K., Aizawa, S., Yoshimura, E., Satake, K., 2001. Determination and speciation of aluminum in environmental samples by cation exchange high-performance liquid chromatography with high resolution ICP-MS detection. *Water, Air, Soil Poll.*, 130, 1589-1594.
- [21] Dahlqvist, R., Benedetti, M.F., Andersson, K., Turner, D., Larsson, T., Stolpe, B., Ingri, J., 2004. Association of calcium with colloidal particles and speciation of calcium in the Kalix and Amazon rivers. *Geochim. Cosmochim. Acta*, 68, 4059-4075.
- [22] Klueppel, D., Jakubowski, N., Messerschmidt, J., Stuewer, D., Klockow, D., 1998. Speciation of platinum metabolites in plants by size-exclusion chromatography and inductively coupled plasma mass spectrometry. *J. Anal. At. Spectrom.*, 13, 255-262.
- [23] Montes Bayon, M., Soldado Cabezero, A.B., Blanco Gonzalez, E., Garcia Alonso, J.I., Sanz-Medel, A., 1999. Capabilities of fast protein liquid chromatography coupled to a double focusing inductively coupled plasma mass spectrometer for trace metal speciation in human serum. *J. Anal. At. Spectrom.*, 14, 947-951.
- [24] Vanhaecke, F., Saverwyns, S., De Wannemacker, G., Moens, L., Dams, R., 2000. Comparison of the application of higher mass resolution and cool plasma conditions to avoid spectral interferences in Cr(III)/Cr(VI) speciation by means of high-performance liquid chromatography - inductively coupled plasma mass spectrometry. *Anal. Chim. Acta*, 419, 55-64.

[25] Wu, F., Evans, D., Dillon, P., Schiff, S., 2004. Molecular size distribution characteristics of the metal-DOM complexes in stream waters by high-performance size-exclusion chromatography (HPSEC) and high-resolution inductively coupled plasma mass spectrometry (ICP-MS). *J. Anal. At. Spectrom.*, 19, 979-983.

[26] Boulyga, S.F., Loreti, V., Bettmer, J., Heumann, K.G., 2004. Application of SEC-ICP-MS for comparative analyses of metal-containing species in cancerous and healthy human thyroid samples. *Anal. Bioanal. Chem.*, 380, 198-203.

[27] Nagaoka, M.H., Akiyama, H., Maitani, T., 2004. Binding patterns of vanadium to transferrin in healthy serum studied with HPLC/high resolution ICP-MS. *Analyst*, 129, 51-54.

[28] Rottmann, L., Heumann, K.G., 1994. Determination of heavy metal interactions with dissolved organic materials in natural aquatic systems by coupling a high-performance liquid chromatography system with an inductively coupled plasma mass spectrometer. *Anal. Chem.*, 66, 3709-3715.

[29] Sariago Muniz, C., Marchante Gayon, J.M., Garcia Alonso, J.I., Sanz-Medel, A., 2001. Speciation of essential elements in human serum using anion-exchange chromatography coupled to post-column isotope dilution analysis with double focusing ICP-MS. *J. Anal. At. Spectrom.*, 16, 587-592.

[30] Harrington, C.F., Elahi, S., Merson, S.A., Ponnampalavanar, P., 2001. A method for the quantitative analysis of iron speciation in meat by using a combination of spectrophotometric methods and high-performance liquid chromatography coupled to sector field inductively coupled plasma mass spectrometry. *Anal. Chem.*, 71, 4422-4427.

[31] Bouysiere, B., Knispel, T., Ruhnau, C., Denkhau, E., Prange, A., 2004. Analysis of nickel species in cytosols of normal and malignant human colonic tissues using two dimensional liquid chromatography with ICP-sector field MS detection. *J. Anal. At. Spectrom.*, 19, 196-200.

[32] Wolf, C., Schaumloeffel, D., Richarz, A.N., Prange, A., Braetter, P., 2003. CZE-ICP-MS separation of metallothioneins in human brain cytosols: comparability of electropherograms obtained from different sample matrices. *Analyst*, 128, 576-580.

[33] Polec-Pawlak, K., Schaumloeffel, D., Szpunar, J., Prange, A., Lobinski, R., 2002. Analysis for metal complexes with metallothionein in rat liver by capillary zone electrophoresis using ICP double-focusing sector-field isotope dilution MS and electrospray MS detection. *J. Anal. At. Spectrom.*, 17, 908-912.

[34] Schaumloeffel, D., Prange, A., Marx, G., Heumann, K.G., Braetter, P., 2002. Characterization and quantification of metallothionein isoforms by capillary electrophoresis - inductively coupled plasma-isotope-dilution mass spectrometry. *Anal. Bioanal. Chem.*, 372, 155-163.

[35] Prange, A., Schaumloeffel, D., Braetter, P., Richarz, A.N., Wolf, C., 2001. Species analysis of metallothionein isoforms in human brain cytosols by use of capillary electrophoresis hyphenated to inductively coupled plasma-sector field mass spectrometry. *Fres. J. Anal. Chem.*, 371, 764-774.

[36] Van Lierde, V., Chery, C.C., Strijckmans, K., Galleni, M., Devreese, B., Van Beuemen, J., Moens, L., Vanhaecke, F., 2004. Capillary electrophoresis hyphenated to inductively coupled plasma-sector field-mass spectrometry for the stoichiometric determination of Zn bound to *Aeromonas hydrophila* Zn beta-lactamase. *J. Anal. At. Spectrom.*, 19, 888-893.

[37] Thermo Fisher Scientific, Application Note AN30012, High sensitivity arsenic speciation: HPLC sector field ICP-MS, Thermo Scientific ELEMENT 2.

[38] Prange, A., Schaumloeffel, D., 1999. Determination of element species at trace levels using capillary electrophoresis-inductively coupled plasma sector field mass spectrometry. *J. Anal. At. Spectrom.*, 14, 1329-1332.

[39] Zheng, J., Hintelmann, H., Dimock, B., Dzurko, M.S., 2003. Speciation of arsenic in water, sediment, and plants of the Moira watershed, Canada, using HPLC coupled to high resolution ICP-MS. *Anal. Bioanal. Chem.*, 377, 14-24.

[40] Zheng, J., Hintelmann, H., 2004. Hyphenation of high performance liquid chromatography with sector field inductively coupled plasma mass spectrometry for the determination of ultra-trace level anionic and cationic arsenic compounds in freshwater fish. *J. Anal. At. Spectrom.*, 19, 191-195.

[41] Koellensperger, G., Nurmi, J., Hann, S., Stingeder, G., Fitz, W.J., Wenzel, W.W., 2002. CE-ICP-SFMS and HPIC-ICP-SFMS for arsenic speciation in soil solution and soil water extracts. *J. Anal. At. Spectrom.*, 17, 1042-1047.

[42] Jakubowski, N., Stuewer, D., Klockow, D., Thomas, C., Emons, H., 2001. Speciation of organic selenium compounds using reversed-phase liquid chromatography and inductively coupled mass spectrometry. Part III. Application of a sector field instrument with low and high mass resolution for selenium speciation in herring gull eggs. *J. Anal. At. Spectrom.*, 16, 135-139.

[43] Thomas, C., Jakubowski, N., Stuewer, D., Klockow, D., Emons, H., 1998. Speciation of organic selenium compounds using reversed-phase liquid chromatography and inductively coupled mass spectrometry. Part I. Sector field instrument with low mass resolution. *J. Anal. At. Spectrom.*, 13, 1221-1226.

[44] Feldmann, I., Jakubowski, N., Stuewer, D., Thomas, C., 2000. Speciation of organic selenium compounds using reversed-phase liquid chromatography and inductively coupled mass spectrometry. Part II. Sector field instrument with high mass resolution. *J. Anal. At. Spectrom.*, 15, 371-376.

[45] Yang, L., Sturgeon, R.E., Wolf, W.R., Goldschmidt, R.J., Mester, Z., 2004. Determination of selenomethionine in yeast using CNBr derivatization and species specific isotope dilution GC ICP-MS and GC-MS. *J. Anal. At. Spectrom.*, 19, 1448-1453.

[46] Lesniewska, B.A., Messerschmidt, J., Jakubowski, N., Hulanicki, A., 2004. Bioaccumulation of platinum group elements and characterization of their species in *Lolium multiflorum* by size-exclusion chromatography coupled with ICP-MS. *Sci. Tot. Environ.*, 322, 95-108.

[47] Alvarez-Llamas, M.R., de la Campa, F., Fernandez Sanchez, M.L., Sanz-Medel, A., 2002. Comparison of two CE-ICP-MS interfaces based on microflow nebulizers: application to cadmium speciation in metallothioneins using quadrupole and double focusing mass analyzers. *J. Anal. At. Spectrom.*, 17, 655-661.

[48] Yang, L., Mester, Z., Sturgeon, R.E., 2003. Comparison of sector field- and quadrupole-ICP-MS for the determination of DBT and TBT in sediment following GC separation. *J. Anal. At. Spectrom.*, 18, 1365-1370.

[49] Sonke, J.E., Salters, V.J.M., 2004. Determination of neodymium-fulvic acid binding constants by capillary electrophoresis inductively coupled plasma mass spectrometry (CE-ICP-MS). *J. Anal. At. Spectrom.*, 19, 235-240.

[50] Hann, S., Koellensperger, G., Stefanka, Z., Stingeder, G., Fühacker, M., Buchberger, W., Mader, R.M., 2003. Application of HPLC-ICP-MS to speciation of cisplatin and its degradation products in water containing different chloride concentrations and in human urine. *J. Anal. At. Spectrom.*, 18, 1391-1395.

[51] Silva da Rocha, M., Soldado, A.B., Blanco-Gonzalez, E., Sanz-Medel, A., 2000. Speciation of mercury compounds by capillary electrophoresis coupled on-line with quadrupole and double-focusing inductively coupled plasma mass spectrometry. *J. Anal. At. Spectrom.*, 15, 513-518.

[52] Carter, J., Ebdon, L., Evans, E.H., 2004. Speciation of silicon and phosphorous using liquid chromatography coupled with sector field high resolution ICP-MS. *Microchem. J.*, 76, 35-41.

[53] Buchberger, W., Czizsek, B., Hann, S., Stingeder, G., 2003. Preliminary comparison of inductively coupled plasma mass spectrometry and electrospray mass spectrometry hyphenated with ion chromatography for trace analysis of iodide. *J. Anal. At. Spectrom.*, 18, 512-514.

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