Reduced injection volume applied to the quantitation of cylindrospermopsin and anatoxin-a in drinking water according to EPA Method 545

Authors
Ali Haghani,¹ Andy Eaton,¹ Neloni Wijeratne,² Claudia Martins²

¹Eurofins Eaton Analytical, Monrovia, CA
²Thermo Fisher Scientific, San Jose, CA

Keywords
Anatoxin-a, cylindrospermopsin, EPA Method 545, TSQ Quantis MS

Goal
To demonstrate a sensitive, accurate, and reliable LC-MS/MS workflow to quantify anatoxin-a and cylindrospermopsin in drinking water according to EPA Method 545, performed at a reduced injection volume to increase method robustness while maintaining the desired levels of sensitivity.

Introduction
Cyanobacteria naturally occur in surface waters. Under certain conditions, such as in warm water containing an abundance of nutrients, they can rapidly form harmful algal blooms (HABs). HABs can produce toxins known as cyanotoxins, which can be harmful to humans and animals. Anatoxin-a (also known as Very Fast Death Factor) is a neurotoxin with acute toxic effects and therefore subjected to monitoring and regulation efforts in several countries, including the United States. Cylindrospermopsin is toxic to liver and kidney tissues. As a result, the United States Environmental Protection Agency (EPA) has developed EPA Method 545 for the Unregulated Contaminant
Monitoring Rule 4 (UCMR 4) program, which collects data for contaminants suspected to be present in drinking water but that do not have health-based standards set under the Safe Drinking Water Act (SDWA)\(^2\). This study demonstrates the performance of the new Thermo Scientific™ TSQ Quantis™ triple quadrupole MS platform performance using EPA Method 545: Determination of cylindrospermopsin and anatoxin-a in drinking water by liquid chromatography electrospray ionization tandem mass spectrometry (LC/ESI-MS/MS).

**Experimental**

**Sample preparation and LC/ESI-MS/MS conditions**

The sample preparation was based on EPA Method 545. A triple freeze and thaw process was used on 2 mL of sample preserved with 0.1% sodium bisulfate and 0.01% ascorbic acid. The sample was filtered through a 0.2 µm pore size PVDF disposable filter to address the potential presence of intact algal cells in finished water samples. Then, 1 mL of sample was mixed with the phenylalanine-d5 and uracil-d4 internal standards and measured by direct injection-LC/ESI-MS/MS. Mobile phases used were 100 mM acetic acid in water and methanol.

Compounds were detected using the Thermo Scientific™ UltiMate™ 3000 LC system in conjunction with a Thermo Scientific™ TSQ Quantis™ triple quadrupole mass spectrometer equipped with a heated electrospray ionization source. LC conditions were as stated in the EPA 545 method, and MS experimental conditions are listed in Tables 1 and 2.

**Requirements**

The EPA has strict requirements that should be met before the analysis of any sample, referred to as the Initial Demonstration of Capability (IDC). These requirements include the demonstration of low background noise, precision by analyzing four laboratory fortified reagent water blanks (LFB) at mid-level, the demonstration of accuracy and, finally, the demonstration of capability necessary to meet the minimum reporting limit (MRL). The percent relative standard deviation (%RSD) of the results of the replicate analyses must be ≤ 20%. The average percent recovery for each analyte must be within ± 30% of the true value. All IDC samples need to be treated as field samples going through the entire method process. For comparability of results using the TSQ Quantis triple quadrupole MS, 5 µL, 10 µL, and 25 µL injections are reported.

**Results and discussion**

**Calibration**

The initial calibration was validated by calculating the concentration of each analyte as an unknown against its regression equation. For calibration levels that are ≤ MRL, the result for each analyte should be within ± 50% of the true value. All other calibration points must calculate to be within ± 30% of their true value. Results for all three injections met the calibration criteria. Figure 1 shows representative calibration curves for all compounds using a 5 µL injection.

### Table 1. MS source conditions.

<table>
<thead>
<tr>
<th>Ion Source Parameter</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Spray Voltage</td>
<td>3500 V</td>
</tr>
<tr>
<td>Sheath Gas</td>
<td>45 Arb</td>
</tr>
<tr>
<td>Aux Gas</td>
<td>10 Arb</td>
</tr>
<tr>
<td>Sweep Gas</td>
<td>0 Arb</td>
</tr>
<tr>
<td>Ion Transfer Tube Temperature</td>
<td>325 °C</td>
</tr>
<tr>
<td>Vaporizer Temperature</td>
<td>275 °C</td>
</tr>
</tbody>
</table>

### Table 2. Optimized SRM conditions.

<table>
<thead>
<tr>
<th>Compound</th>
<th>Polarity</th>
<th>Precursor (m/z)</th>
<th>Product (m/z)</th>
<th>Collision Energy (V)</th>
<th>RF Lens (V)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Anatoxin-a</td>
<td>Positive</td>
<td>166</td>
<td>131</td>
<td>18</td>
<td>100</td>
</tr>
<tr>
<td>Cylindrospermopsin</td>
<td>Positive</td>
<td>416</td>
<td>176</td>
<td>35</td>
<td>120</td>
</tr>
<tr>
<td>Phenylalanine-d5 IS for Anatoxin-a</td>
<td>Positive</td>
<td>171</td>
<td>125</td>
<td>12</td>
<td>80</td>
</tr>
<tr>
<td>Uracil-d4 IS for Cylindrospermopsin</td>
<td>Positive</td>
<td>115</td>
<td>98</td>
<td>10</td>
<td>55</td>
</tr>
</tbody>
</table>
Figure 1A. Calibration curves for (top) anatoxin-a and (bottom) cylindrospermospin.

Figure 1B. Calibration curves for reproducibility (%RSD) of internal standards with a 5 µL injection.

System background
All method blanks exhibited less than 1/3 MRL contamination or carryover (Table 3, Figure 2).

Table 3. Low background noise for all EPA Method 545 analytes.

<table>
<thead>
<tr>
<th>Analyte</th>
<th>MRL (µg/L)</th>
<th>1/3 MRL (µg/L)</th>
<th>5 µL</th>
<th>10 µL</th>
<th>25 µL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Anatoxin-a</td>
<td>0.03</td>
<td>0.01</td>
<td>NF</td>
<td>NF</td>
<td>NF</td>
</tr>
<tr>
<td>Cylindrospermospin</td>
<td>0.09</td>
<td>0.03</td>
<td>NF</td>
<td>NF</td>
<td>NF</td>
</tr>
</tbody>
</table>

NF=Not found
**Precision and accuracy**

The initial demonstration of precision and accuracy was met by analyzing four extracted LFBs spiked at 10x MRL level, which showed less than 20% RSD and ± 30% difference (Table 4 and Figure 3).

**Table 4.** Precision and accuracy at 10x MRL for all EPA Method 545 analytes.

<table>
<thead>
<tr>
<th>Analyte</th>
<th>Actual (µg/L)</th>
<th>LFB1 (µg/L)</th>
<th>LFB2 (µg/L)</th>
<th>LFB3 (µg/L)</th>
<th>LFB4 (µg/L)</th>
<th>%Rec</th>
<th>%RSD</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>5 µL Injection</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anatoxin-a</td>
<td>0.3</td>
<td>0.315</td>
<td>0.326</td>
<td>0.286</td>
<td>0.29</td>
<td>101%</td>
<td>6%</td>
</tr>
<tr>
<td>Cylindrospermopsin</td>
<td>0.9</td>
<td>1.011</td>
<td>0.952</td>
<td>1.182</td>
<td>1.12</td>
<td>118%</td>
<td>10%</td>
</tr>
<tr>
<td>IS-Phenylalanine-d5</td>
<td>90%</td>
<td>96%</td>
<td>123%</td>
<td>117%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IS-Uracil-d4</td>
<td>122%</td>
<td>119%</td>
<td>127%</td>
<td>119%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>10 µL Injection</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anatoxin-a</td>
<td>0.3</td>
<td>0.297</td>
<td>0.306</td>
<td>0.309</td>
<td>0.309</td>
<td>102%</td>
<td>2%</td>
</tr>
<tr>
<td>Cylindrospermopsin</td>
<td>0.9</td>
<td>1.09</td>
<td>0.994</td>
<td>0.992</td>
<td>1.02</td>
<td>114%</td>
<td>4%</td>
</tr>
<tr>
<td>IS-Phenylalanine-d5</td>
<td>90%</td>
<td>99%</td>
<td>100%</td>
<td>98%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IS-Uracil-d4</td>
<td>89%</td>
<td>101%</td>
<td>93%</td>
<td>83%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>25 µL Injection</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anatoxin-a</td>
<td>0.3</td>
<td>0.347</td>
<td>0.319</td>
<td>0.324</td>
<td>0.315</td>
<td>109%</td>
<td>4%</td>
</tr>
<tr>
<td>Cylindrospermopsin</td>
<td>0.9</td>
<td>0.956</td>
<td>0.899</td>
<td>1.072</td>
<td>1.055</td>
<td>111%</td>
<td>8%</td>
</tr>
<tr>
<td>IS-Phenylalanine-d5</td>
<td>88%</td>
<td>94%</td>
<td>96%</td>
<td>94%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IS-Uracil-d4</td>
<td>93%</td>
<td>101%</td>
<td>80%</td>
<td>78%</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**IS criteria**

<table>
<thead>
<tr>
<th>%Recovery</th>
<th>50–150%</th>
</tr>
</thead>
<tbody>
<tr>
<td>%RSD</td>
<td>&lt;20</td>
</tr>
</tbody>
</table>

%Recovery: 70–130%

%RSD: <20
MRL confirmation

MRL confirmation was evaluated by fortifying, extracting, and analyzing seven replicate LFBs at the proposed MRL concentration. The mean and the half range (HR) was then calculated. The Prediction Interval of Results (PIR) is defined as

\[ PIR = \text{Mean} + HR_{PIR} \]

where \( HR_{PIR} = 3.963s; \) s is the standard deviation and 3.963 is a constant value for seven replicates.

The upper and lower limits for the PIR meet the recovery criteria (upper PIR - 150%; lower PIR - 50%) (Table 5).

Tap water analysis

A Monrovia, CA tap water sample (comprised of ground and surface water) was extracted and analyzed using the methodology developed. Results are shown in Table 6.

---

**Table 5. Minimum reporting limit confirmation for all EPA Method 545 analytes.**

<table>
<thead>
<tr>
<th>Analyte</th>
<th>Actual (µg/L)</th>
<th>LFB1 (µg/L)</th>
<th>LFB2 (µg/L)</th>
<th>LFB3 (µg/L)</th>
<th>LFB4 (µg/L)</th>
<th>LFB-5 (µg/L)</th>
<th>LFB-6 (µg/L)</th>
<th>LFB-7 (µg/L)</th>
<th>Lower PIR (%)</th>
<th>Upper PIR (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>5 µL Injection</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anatoxin-a</td>
<td>0.03</td>
<td>0.038</td>
<td>0.03</td>
<td>0.031</td>
<td>0.035</td>
<td>0.031</td>
<td>0.029</td>
<td>0.032</td>
<td>66%</td>
<td>149%</td>
</tr>
<tr>
<td>Cylindrospermopsin</td>
<td>0.09</td>
<td>0.087</td>
<td>0.104</td>
<td>0.1</td>
<td>0.099</td>
<td>0.106</td>
<td>0.098</td>
<td>0.113</td>
<td>77%</td>
<td>148%</td>
</tr>
<tr>
<td>IS-Phenylalanine-d5</td>
<td>134%</td>
<td>133%</td>
<td>129%</td>
<td>125%</td>
<td>127%</td>
<td>131%</td>
<td>127%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IS-Uracil-d4</td>
<td>122%</td>
<td>119%</td>
<td>127%</td>
<td>119%</td>
<td>126%</td>
<td>125%</td>
<td>118%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>10 µL Injection</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anatoxin-a</td>
<td>0.03</td>
<td>0.034</td>
<td>0.033</td>
<td>0.035</td>
<td>0.036</td>
<td>0.036</td>
<td>0.037</td>
<td>0.035</td>
<td>99%</td>
<td>135%</td>
</tr>
<tr>
<td>Cylindrospermopsin</td>
<td>0.09</td>
<td>0.076</td>
<td>0.091</td>
<td>0.09</td>
<td>0.093</td>
<td>0.096</td>
<td>0.084</td>
<td>0.08</td>
<td>65%</td>
<td>129%</td>
</tr>
<tr>
<td>IS-Phenylalanine-d5</td>
<td>111%</td>
<td>130%</td>
<td>129%</td>
<td>130%</td>
<td>129%</td>
<td>129%</td>
<td>129%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IS-Uracil-d4</td>
<td>105%</td>
<td>112%</td>
<td>111%</td>
<td>110%</td>
<td>115%</td>
<td>113%</td>
<td>116%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>25 µL Injection</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anatoxin-a</td>
<td>0.03</td>
<td>0.039</td>
<td>0.039</td>
<td>0.041</td>
<td>0.04</td>
<td>0.04</td>
<td>0.042</td>
<td>0.041</td>
<td>120%</td>
<td>149%</td>
</tr>
<tr>
<td>Cylindrospermopsin</td>
<td>0.09</td>
<td>0.091</td>
<td>0.1</td>
<td>0.099</td>
<td>0.106</td>
<td>0.102</td>
<td>0.096</td>
<td>0.1</td>
<td>89%</td>
<td>131%</td>
</tr>
<tr>
<td>IS-Phenylalanine-d5</td>
<td>105%</td>
<td>105%</td>
<td>105%</td>
<td>104%</td>
<td>101%</td>
<td>105%</td>
<td>101%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IS-Uracil-d4</td>
<td>103%</td>
<td>101%</td>
<td>102%</td>
<td>103%</td>
<td>106%</td>
<td>111%</td>
<td>110%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>IS criteria</strong></td>
<td>50–150%</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

LFB stands for Fortified Laboratory Blank. PIR stands for Prediction Interval of Results.
Table 6. Monrovia water sample analyzed using the TSQ Quantis triple quadrupole MS.

<table>
<thead>
<tr>
<th>Analyte</th>
<th>Actual (µg/L)</th>
<th>FS</th>
<th>LFSM</th>
<th>LFSMD</th>
<th>%Rec</th>
<th>%RSD</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>5 µL Injection</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anatoxin-a</td>
<td>0.3</td>
<td>0</td>
<td>0.337</td>
<td>0.367</td>
<td>117%</td>
<td>6%</td>
</tr>
<tr>
<td>Cylindrospermopsis</td>
<td>0.9</td>
<td>0</td>
<td>0.954</td>
<td>0.966</td>
<td>107%</td>
<td>1%</td>
</tr>
<tr>
<td>IS-Phenylalanine-d5</td>
<td>120%</td>
<td>111%</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IS-Uracil-d4</td>
<td>142%</td>
<td>130%</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>10 µL Injection</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anatoxin-a</td>
<td>0.3</td>
<td>0</td>
<td>0.347</td>
<td>0.352</td>
<td>117%</td>
<td>1%</td>
</tr>
<tr>
<td>Cylindrospermopsis</td>
<td>0.9</td>
<td>0</td>
<td>1.162</td>
<td>1.085</td>
<td>125%</td>
<td>5%</td>
</tr>
<tr>
<td>IS-Phenylalanine-d5</td>
<td>119%</td>
<td>109%</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IS-Uracil-d4</td>
<td>89%</td>
<td>113%</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>25 µL Injection</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Date Analyzed</td>
<td>4/7/2017</td>
<td>4/7/2017</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anatoxin-a</td>
<td>0.3</td>
<td>0</td>
<td>0.35</td>
<td>0.341</td>
<td>115%</td>
<td>2%</td>
</tr>
<tr>
<td>Cylindrospermopsis</td>
<td>0.9</td>
<td>0</td>
<td>0.788</td>
<td>0.843</td>
<td>91%</td>
<td>5%</td>
</tr>
<tr>
<td>IS-Phenylalanine-d5</td>
<td>96%</td>
<td>87%</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IS-Uracil-d4</td>
<td>122%</td>
<td>107%</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>IS criteria</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>%Recovery</td>
<td>70–130%</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>%RSD</td>
<td>&lt;30</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

FS stands for Field Sample. LFSM stands for Laboratory Fortified Sample Matrix. LFSMD stands for Laboratory Fortified Sample Matrix Duplicate.

Conclusion

- The TSQ Quantis triple quadrupole MS proved to be sensitive, accurate, reproducible, and reliable in the quantitation of cylindrospermopsis, and anatoxin-a in drinking water according to EPA Method 545.
- Adequate sensitivity was obtained with 5 µL, 10 µL, and 25 µL injection volumes for drinking water matrices. This represents an up to 10-fold reduction in the injection volume than what the EPA is recommending and results in less matrix injected, thereby reducing maintenance of the LC-MS system.

References


Find out more at thermofisher.com/Altis-Quantis