

# Quantitative Bioanalysis of Oligonucleotides

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Thermo BioSeparations Workshop



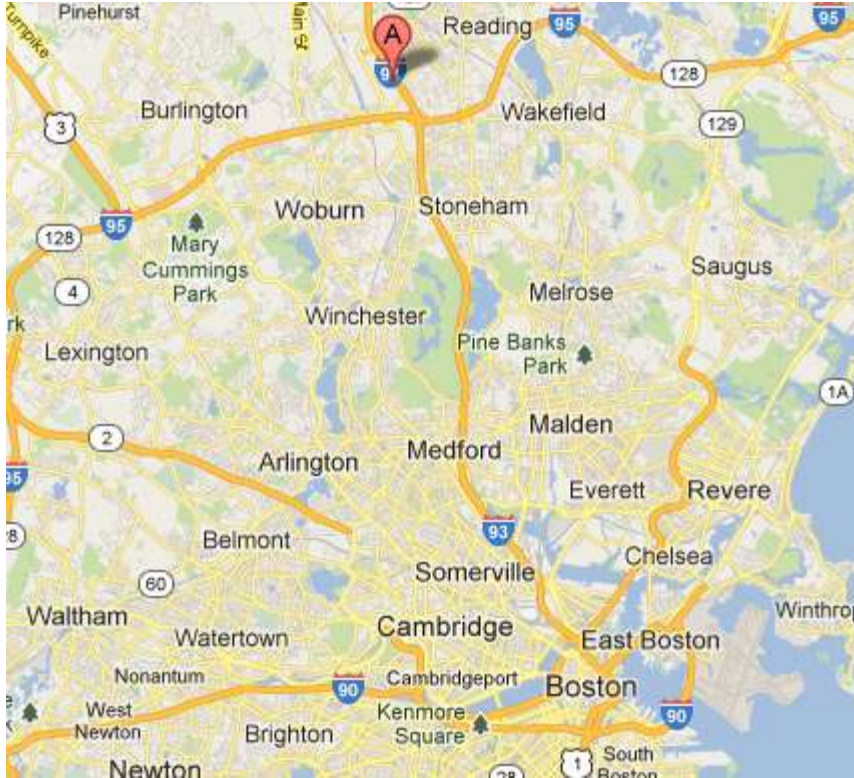
# Outline

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- Overview
  - Thermo instruments at NovaBioAssays
  - Bioanalysis of oligonucleotide therapeutics
- DNAPac PA200 columns for hybridization based LC-fluorescence assays
- DNAPac RP columns for ion-pair LC-MS/MS and LC-HRAM analysis
  - Full Scan, target-SIM and PRM/DIA analysis on Q-Exactive
- Q & A



# Where Are We



**52 Dragon Court, Suite 3B  
Woburn, MA 01801, USA  
Phone: (781)933-3480**



# What Do We Have (1/3)



AB SCIEX 5000 LC/MS/MS  
(x2)



Thermo TSQ Vantage  
(nano and ESI source)



Thermo Q Exactive MS (Plus)  
(x2)



Bruker MicroTOFQ II  
(nano and ESI source)  
(x2)



Waters Acquity UPLC  
Waters nanoAcquity UPLC  
(x7)



Accela 1250 and CTC PAL



Thermo Easy nLC



## What Do We Have (2/3)

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Thermo Vanquish Flex UHPLC System  
With Vanquish™ Fluorescence  
Detector F (4 Excitation/Emission)



Waters Acquity UPLC with PDA  
And Fluorescence Detectors



# What Do We Have (3/3)



TomTec Quadra 3 SPE  
Automated Liquid Handler  
(x2)

PPE, LLE, SPE



Thermo Versette™  
Automated Liquid Handler



Thermo KingFiser Flex 96  
Magnetic Particle Processor

Affinity/Immunoaffinity Purification  
Mass Spectrometric Immunoassay (MSIA)





# What Can We Do?

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## **Quantitative Bioanalysis (API5000 or Q-Exactive or FD/PDA)**

- Protein bioanalysis using LC-MS/MS or LC-HRAM
- Lipids/Polymers using LC-MS/MS or LC-HRAM
- Small molecule analysis using LC-MS or LC-HRAM
- Oligonucleotides bioanalysis using
  - LC-MS/MS or LC-HRAM
  - Hybridization UPLC fluorescence
  - UPLC-UV

## **Qualitative analysis (Q-Exactive and Q-tof)**

- Primary structural characterization using LC-HRAM and LC-MS/MS
- Tertiary structural characterization using H/D exchange LC-HRAM and LC-MS/MS
- Quant-Qual analysis using LC-Q Exactive

## ***In Vitro* Protein Binding and Stability/Metabolite ID**

- Rapid Equilibrium Dialysis(RED) and Ultracentrifugation
- Microsomal/Plasma/Whole blood stability
- Hepatocyte incubation



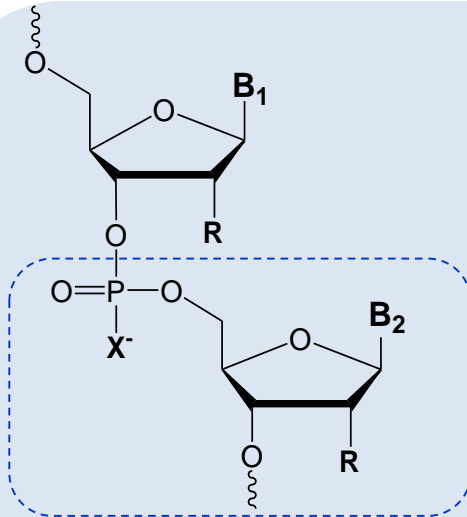
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# Quantitative Bioanalysis of Oligonucleotides



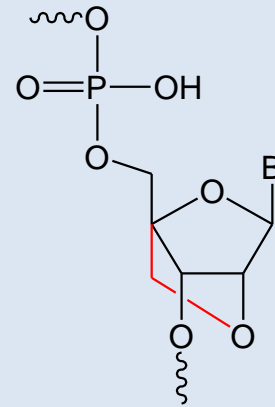


# Chemical Structures of Oligonucleotides

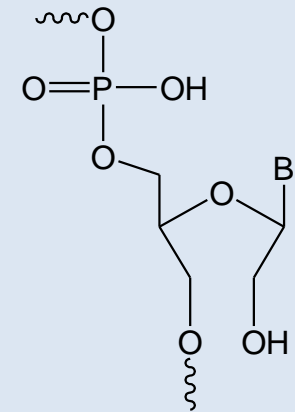


X = O (Phosphodiester)  
S (Phosphorothioate)

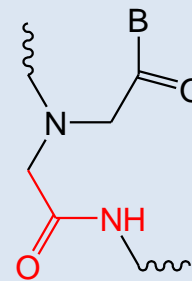
R = -H (DNA)  
-OH (RNA)  
-OCH<sub>3</sub> (2'-Methyl)  
-F (2'-Fluoro)  
-OCH<sub>2</sub>CH<sub>2</sub>OCH<sub>3</sub> (2'-MOE)



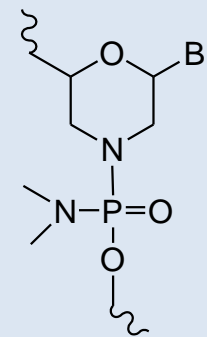
LNA



UNA



PNA

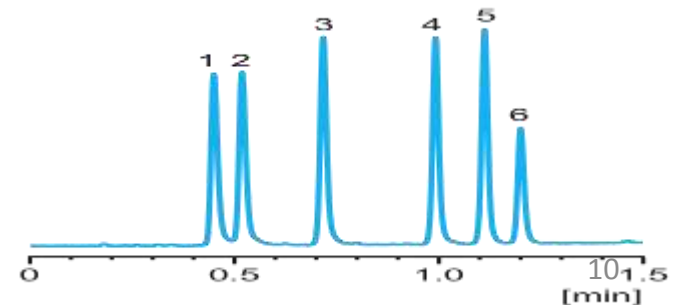
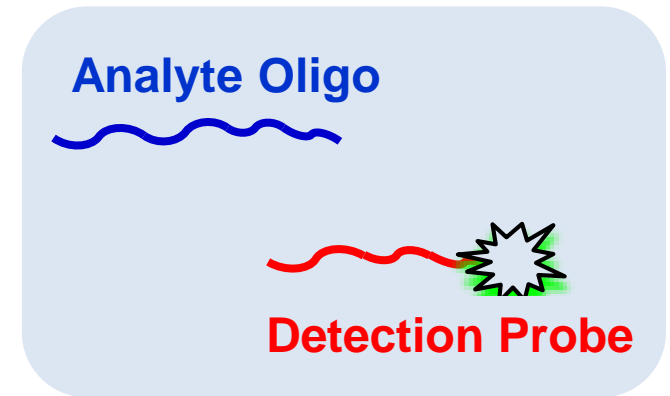


PMO



# Quantitative Bioanalysis of Oligonucleotides/mRNA

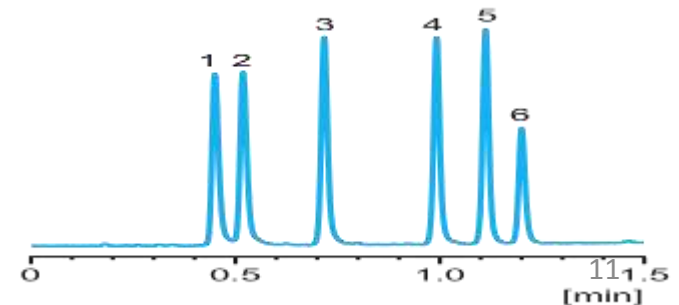
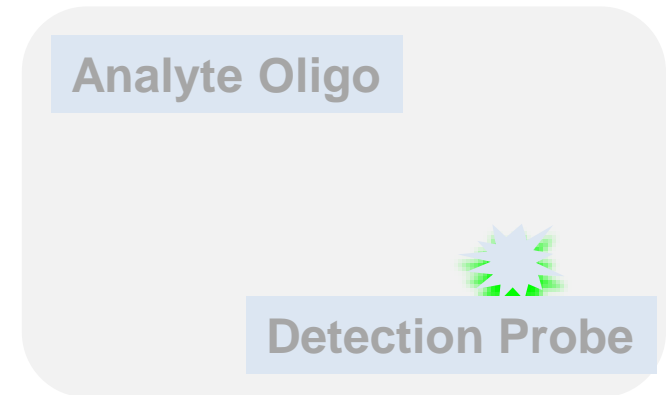
- Hybridization-based assay (indirect analysis)
  - Northern blot/*in situ* hybridization
  - Microarray
  - Deep sequencing (NGS)
  - RT-qPCR
  - Hybridization ELISA
    - Direct ELISA, Sandwich ELISA, etc
    - Branched DNA (bDNA) signal amplification technology (QuantiGene)
- Chromatographic-based assay (direction analysis)
  - HPLC-UV/Fluorescence
  - Hybridization-based LC-fluorescence
  - LC-MS/MS and LC-HRAM





# Quantitative Bioanalysis of Oligonucleotides/mRNA

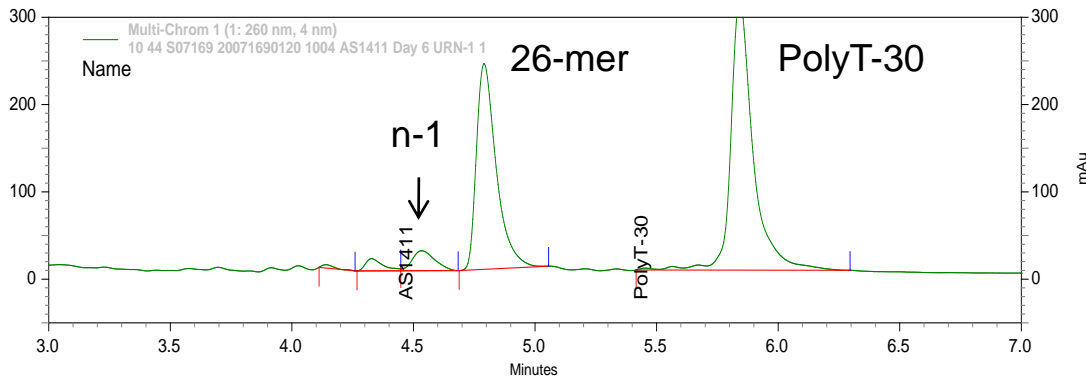
- Hybridization-based assay (indirect analysis)
  - Northern blot/*in situ* hybridization
  - Microarray
  - Deep sequencing (NGS)
  - RT-qPCR
  - Hybridization ELISA
    - Direct ELISA, Sandwich ELISA, etc
    - Branched DNA (bDNA) signal amplification technology (QuantiGene)
- Chromatographic-based assay (direction analysis)
  - HPLC-UV/Fluorescence
  - Hybridization-based LC-fluorescence
  - LC-MS/MS and LC-HRAM





# Ion-Pair (IP)-UPLC-UV Assays

- Simple sample preparation (single LLE or SPE)
- High reproducibility (15/20% acceptance criteria)
- Relatively selective (primarily by length/charge)
- Estimation of metabolite concentrations
- Reasonable run time (<10 minutes/sample)
- **Low sensitivity** (~100 ng/mL LLOQ)

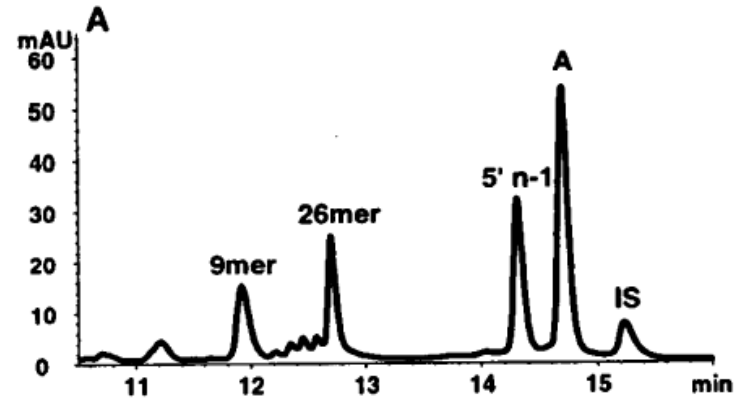


Mobile Phase:  
20 mM tetrabutylammonium  
sulfate in water/MeCN  
pH=9



# Strong Anion Exchange (SAX)-HPLC-UV

- Relatively selective (primarily by length/charge)
- High reproducibility (15/20% acceptance criteria)
- Simple sample preparation (single LLE or protease k digest)
- Estimation of metabolite concentrations
- Low sensitivity (~200 ng/mL LLOQ)
- Only works for charged oligos
- Long run time (20-35 min)



**Analyte:** 35-mer modified ribozyme RNA

**IS:** 43-mer analog RNA

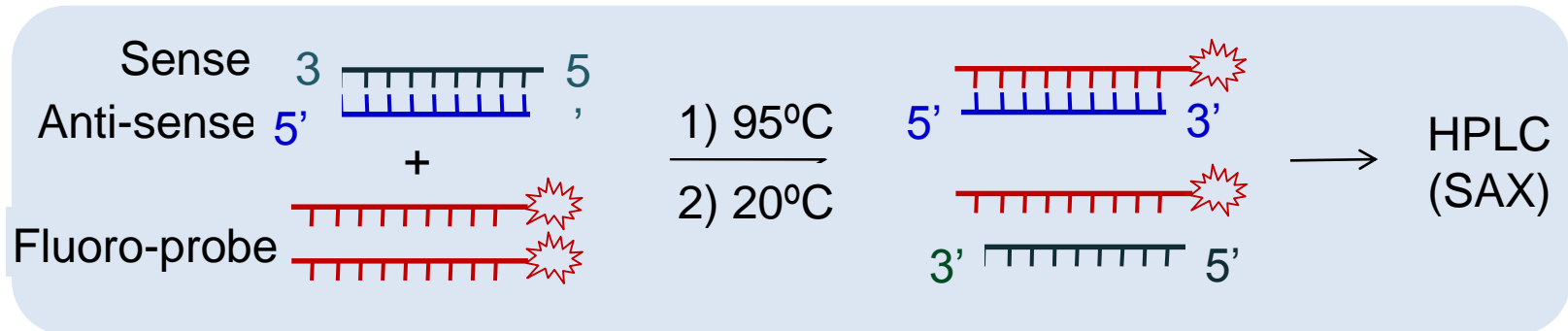
**LC column:** DNAPac® PA100

**MP:** (50 mM Tris pH=8.0)/Ethanol 80/20 (v/v) with 50-250 mM NaClO<sub>4</sub>



# Hybridization-based HPLC-Fluorescence Assays

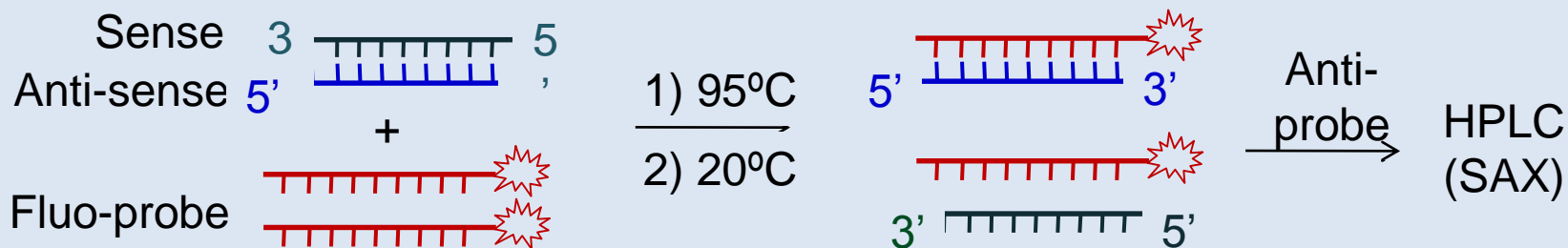
- High sensitivity ( $\sim 1$  ng/mL LLOQ )
- Good specificity (Hybridization and LC separation)
- Large dynamic range (1000x)
- Simple sample preparation (protein digestion or LLE)



- Good reproducibility ( $\pm 15\%$  accuracy/precision)
- Tolerance to structure modifications
  - Base, sugar and phosphate backbone modifications



# Quantify siRNA in HuPI Using Fluorescence Labeled DNA Probe (1.00 - 500 ng/mL)



Analyte: 21-mer siRNA

Probe: 21-mer Atto dye labeled DNA oligo

Anti-probe: Cholesterol conjugated DNA oligo

Sample Prep: Protease K digestion

Column: DNA Pac-PA-200, 4x250 mm

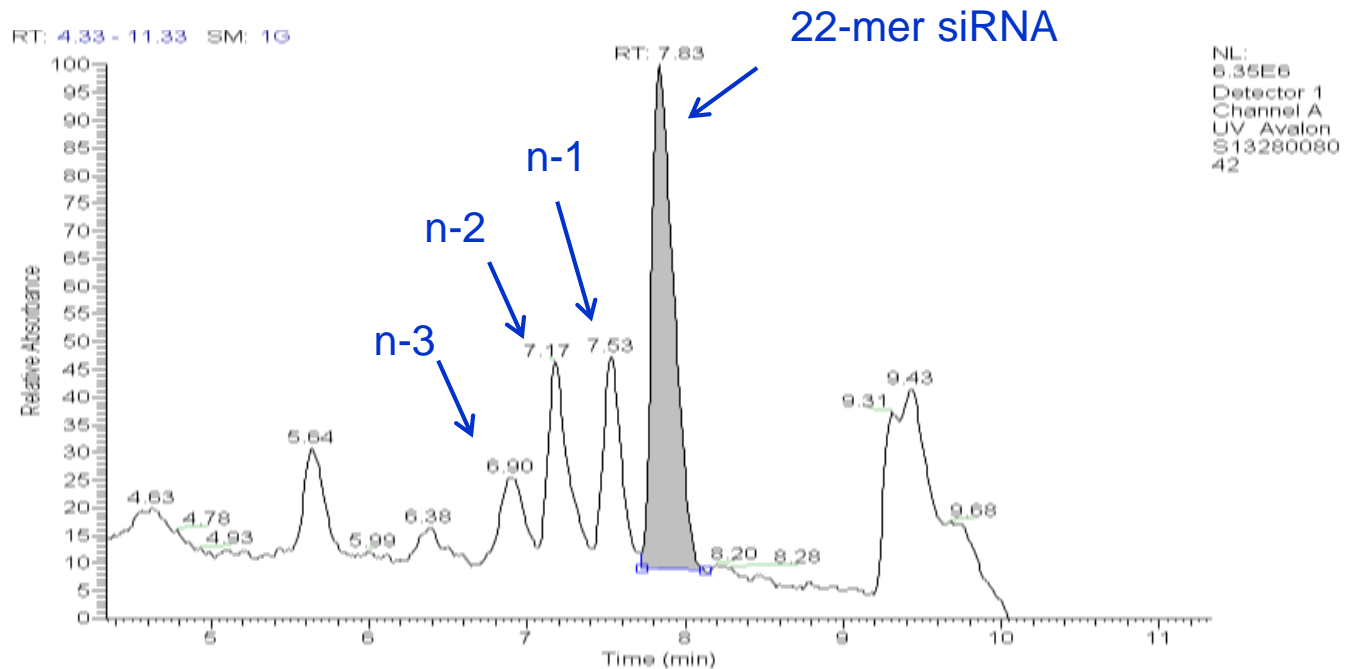
MPA: 1 mM EDTA in 25 mM Tris-HCl (pH 8.0) with 10% MeCN

MPB: 1 mM EDTA and with 800 mM NaClO<sub>4</sub> in 25 mM Tris-HCl with 10% MeCN



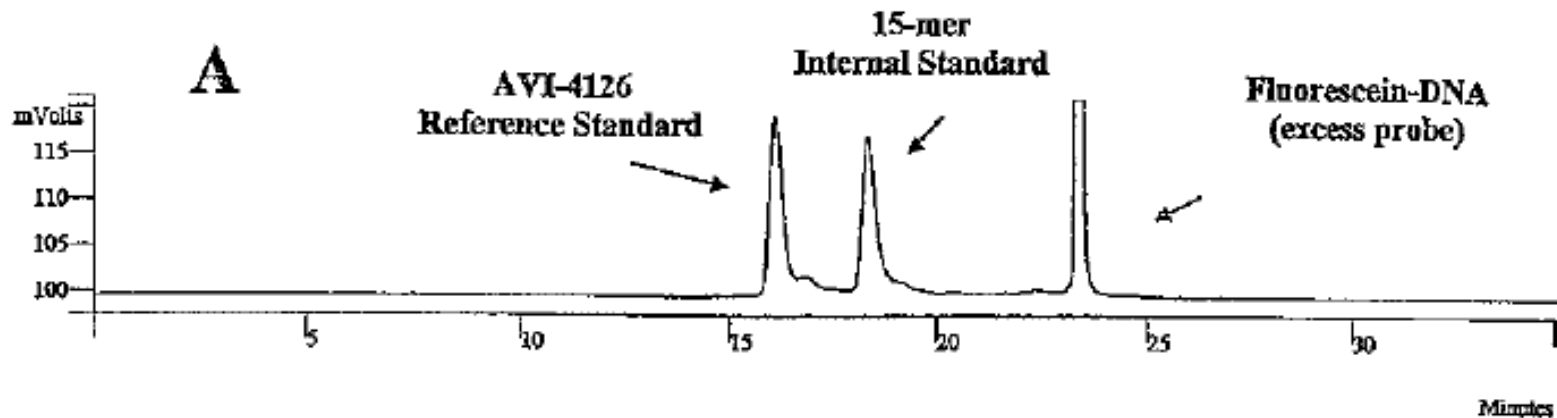


# Typical Chromatogram of 22-mer siRNA in HuPI Samples (>24 Hours)





# Quantification of a Neutral 20-mer PMO Using a 15-mer Fluorescein-DNA (40.0-4,000 ng/mL)



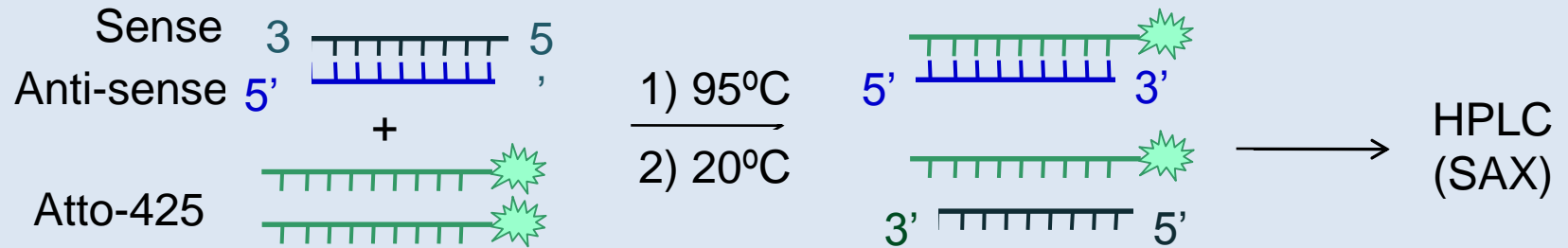
AVI-4126: 5'-ACGTTGAGGGGCATCGTCGC-3' PMO  
(Phosphorodiamidate Morpholino Oligomer)

LC column: Dionex DNA Pac PA-100 (4 x 250 mm)

Mobile Phase: A) 25 mM Tris pH=8.0;  
B) 25 mM Tris pH=8.0 with 1.0 M NaCl



# Quantification of siRNA in HuPI Using Fluorescence-labeled PNA probe (1.00-1000 ng/mL)



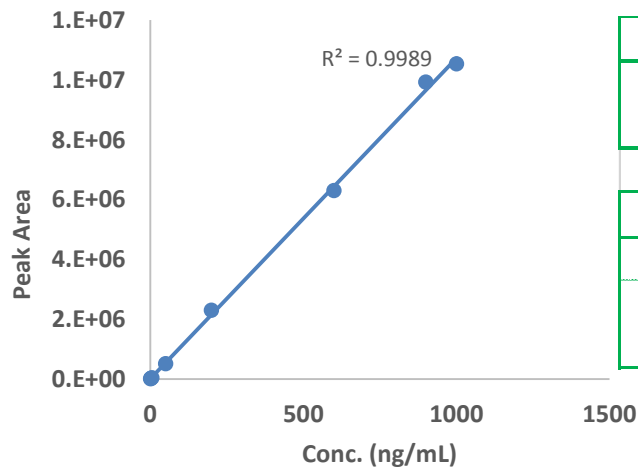
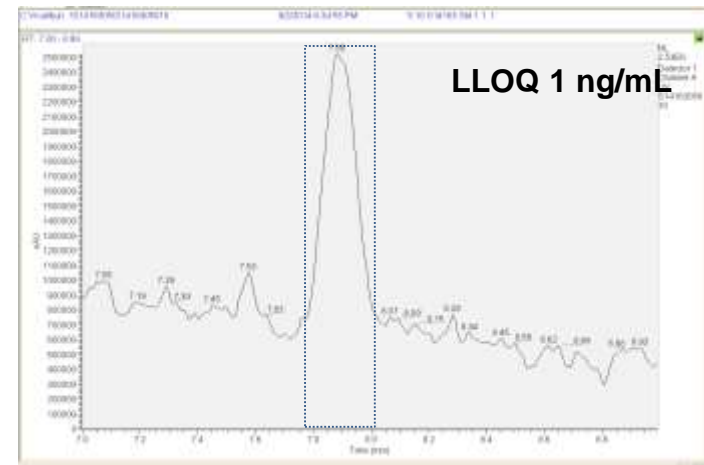
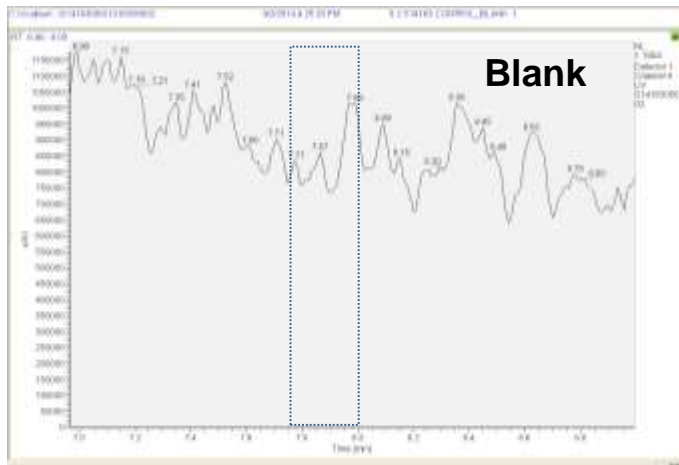
Analyte: 21-mer siRNA

Probe: 18-mer Atto dye labeled PNA oligo

Sample Prep: Protease K digestion

Column: DNA Pac-PA-200, 4x250 mm

# Method Performance of Quantitative Determination of a siRNA in Rat Plasma using PNA Assay



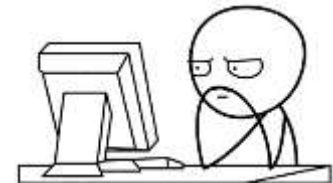
Quality Control Samples			
QC (10 Runs)	Precesion (CV%)	Accuracy (%Bias)	
	2.9%~11.2%	-0.7%~2.5%	
Incurred Sample Reanalysis			
ISR Samples	20%Difference from Original	%Difference from original	ISR samples
45	45 (100% pass)	< ±10%	43
		±10% ~ ±15%	2



# Pros and Cons of Hybridization-based HPLC-Fluorescence Assays

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- High sensitivity ( $\sim 1$  ng/mL LLOQ )
- Good specificity (Hybridization and LC separation)
- Large dynamic range (1000x)
- Simple sample preparation
- Internal standard is not necessary
- Good reproducibility ( $\pm 15\%$  accuracy/precision)
- Tolerance to structure modifications
  - Base, sugar and phosphate backbone modifications
- Long run time (15-30 minutes/sample)
- One analyte/fluorophore at a time





# Fast and Multiple Analyte Hybridization based LC-Fluorescence Assays



DNApac PA200:  
(PEEK) 8  $\mu$ m,  
4.6 x 250 mm

**DNApac PA200 RS:** 4  $\mu$ m,  
**(Pek -lined** 4.6 x 150 mm  
**Stainless Steel)** 4.6 x 50 m



Thermo Vanquish Focused UHPLC  
System  
With Vanquish™ Fluorescence  
Detector F (4 Excitation/Emission)  
Chromeleon 7.2 Chromatography  
Data System

*Data to be shared soon...*



# LC-MS/MS & LC-HRAM Assays

- Highest specificity
  - Molecular weight identification
  - Product ion characterization
  - LC Separation
- High throughput (~5 minutes/sample)
- Wide dynamic range (1000x)
- Good sensitivity (< 5 ng/mL LLOQ)
- No enzymes or special reagents needed
- Good reproducibility ( $\pm 15/20\%$  acceptance criteria)
- Need high performance mass spectrometer
- Potentially challenging assay development







# Challenges & Solutions of Developing LC-MS/MS Assays

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- Non-specific adsorption (stickiness)
  - Add disruption reagent(s) in extraction buffer
  - Avoid extensive drying down of samples
  - Minimize use of polypropylene container
  - Keep stock solution at high concentrations.
- Stability in matrix
  - Use  $K_2EDTA$  as anticoagulant
  - Add CHAPS or BSA to urine samples.
- Short column life time and carryover
  - Clean-up sample “appropriately”
  - Re-generate the column after each injection (back flush the column).
- Charge distribution and metal ion adducts in mass spectrum
  - HFIP and TEA/DIPEA buffered water and MeOH/MeCN as mobile phase.

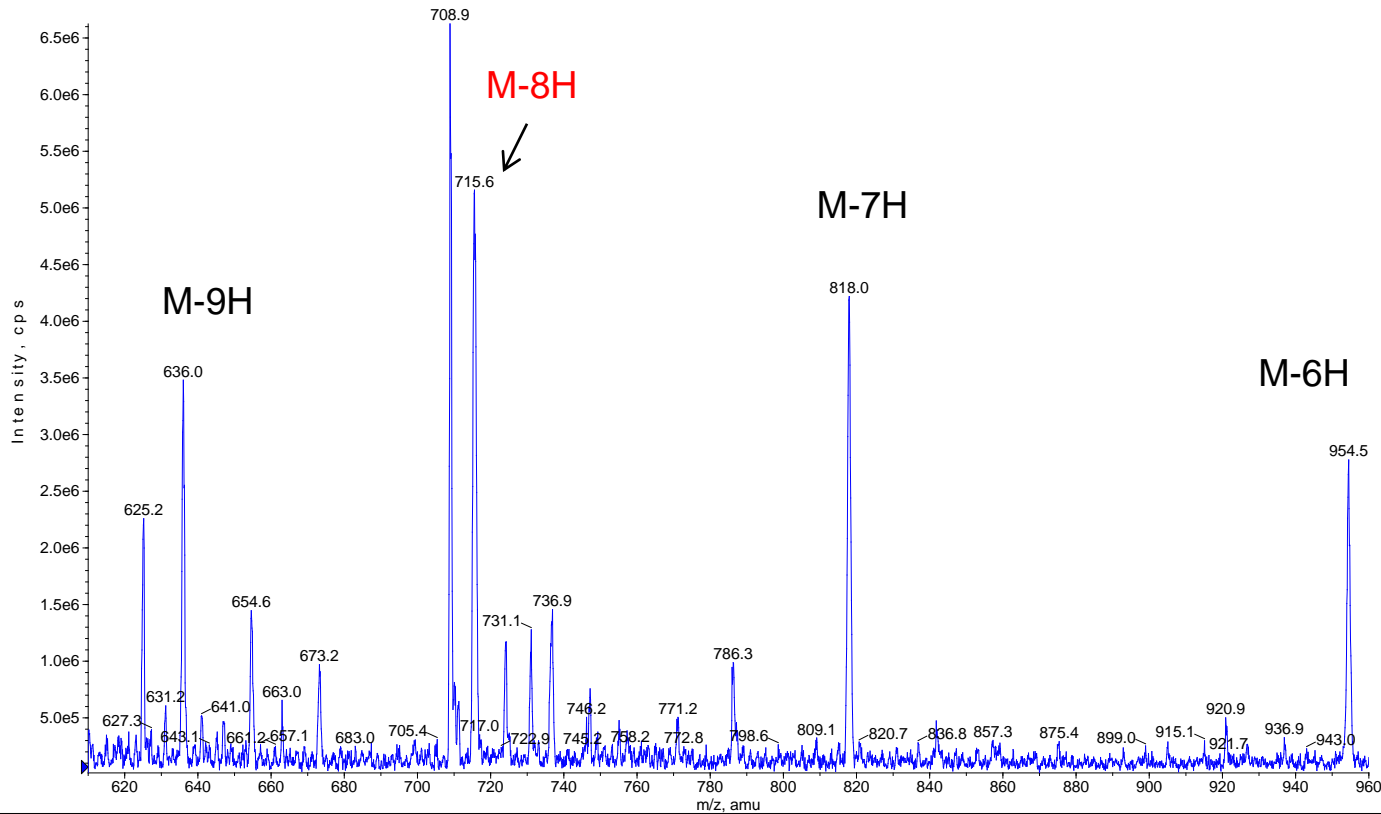


# Q1 MS Scan of a 18-mer PS-ODN on a Triple Quad

Operator: Laixin Wang  
Printing Date: Thursday, May 28, 2009  
Results Name: N/A

■ -Q1: 20 MCA scans from Sample 1 (TuneSampleID) of MT20090528154857.wiff (Turbo Spray)

Max. 6.6e6 cps.

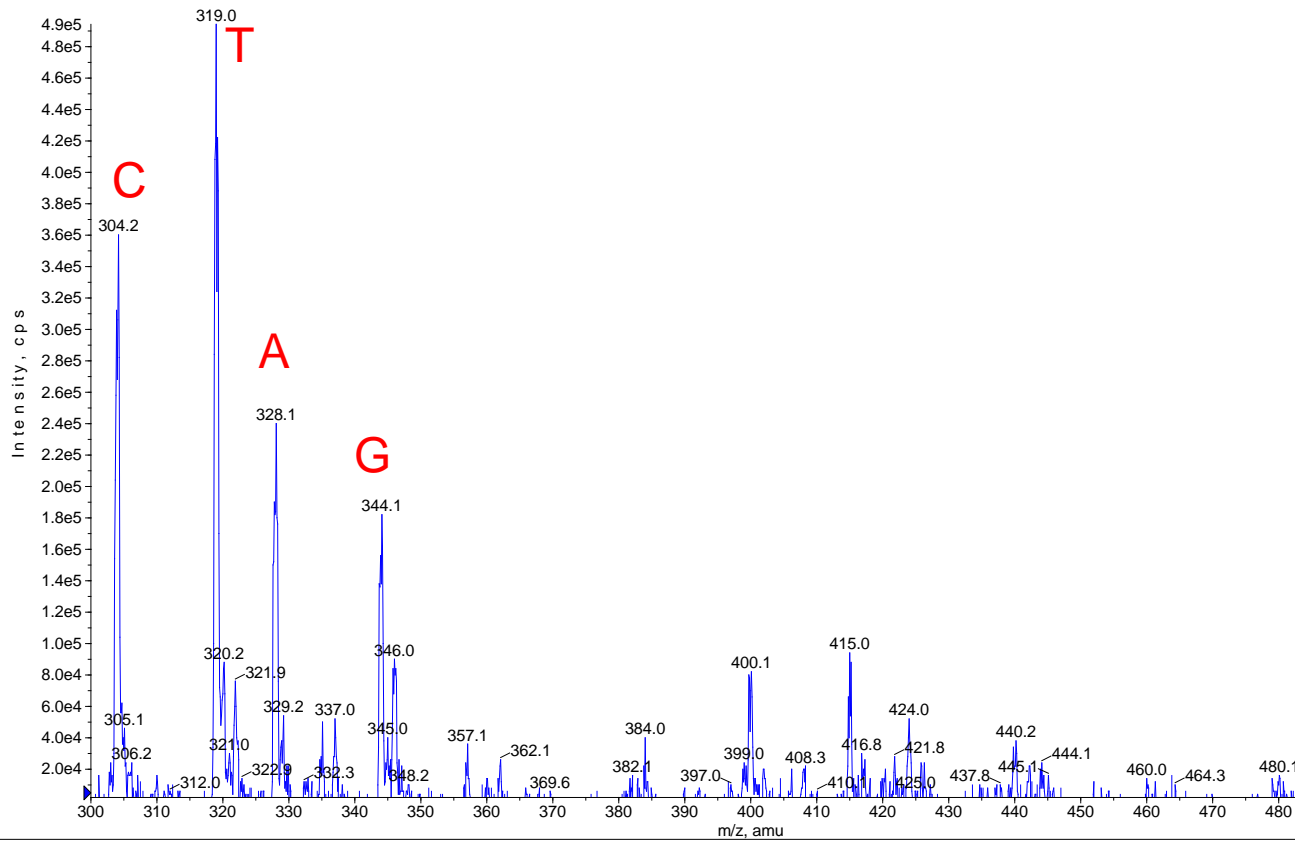


Calculated $m/z$	
M	5731.6
M-6H	954.3
M-7H	817.8
M-8H	715.5
M-9H	636.8



# Product Ion Scan of a 18-mer PS-ODN on a Triple Quad

■ -MS2 (715.40): 20 MCA scans from Sample 1 (TuneSampleID) of MT20090608140619.wiff (Turbo Spray)



SRM Transitions  
evaluated  
for TL0901

715.5 → 319.0

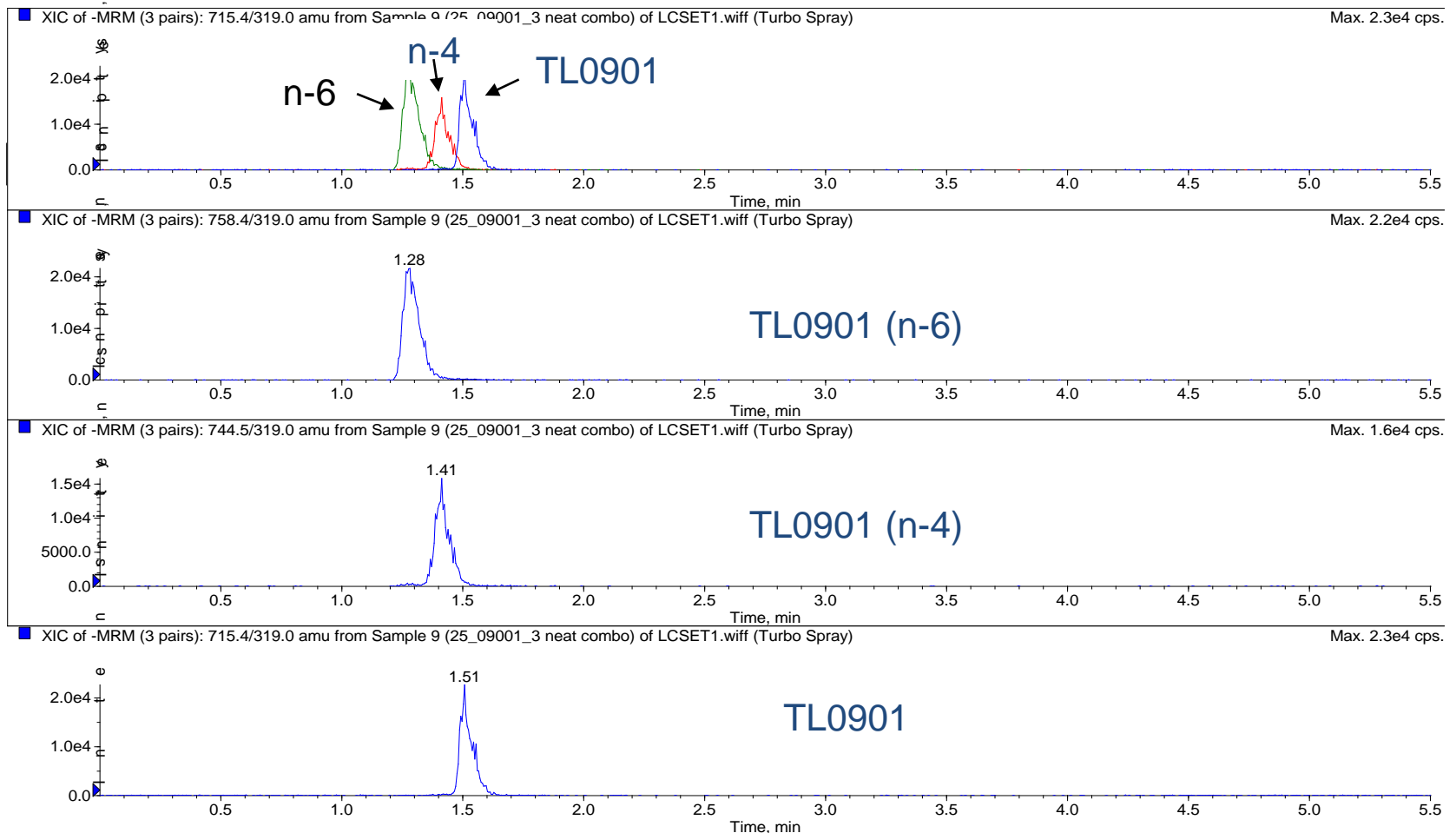
715.5 → 304.2

818.0 → 319.0

818.0 → 304.2

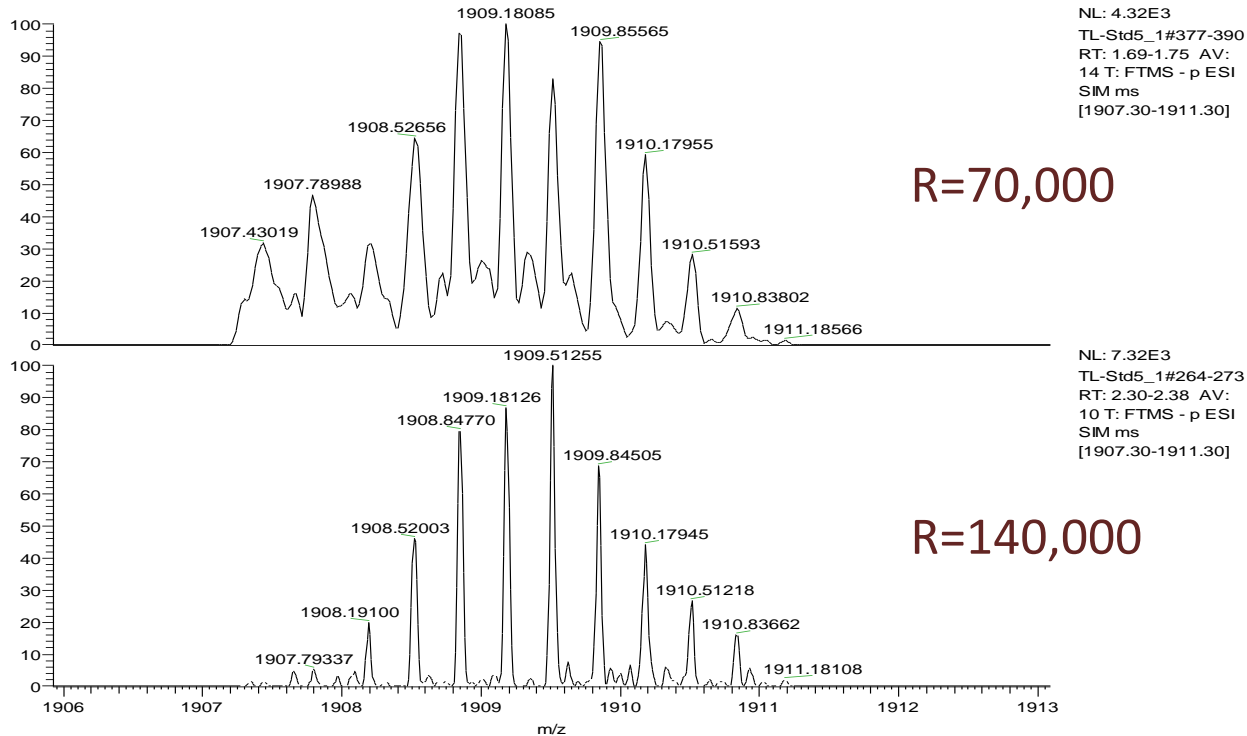


# LC Separation of a 18-mer PS-ODN and its Metabolites (Neat Mixture)





# HRAM Improves Selectivity by Using Higher Resolution (Q Exactive)

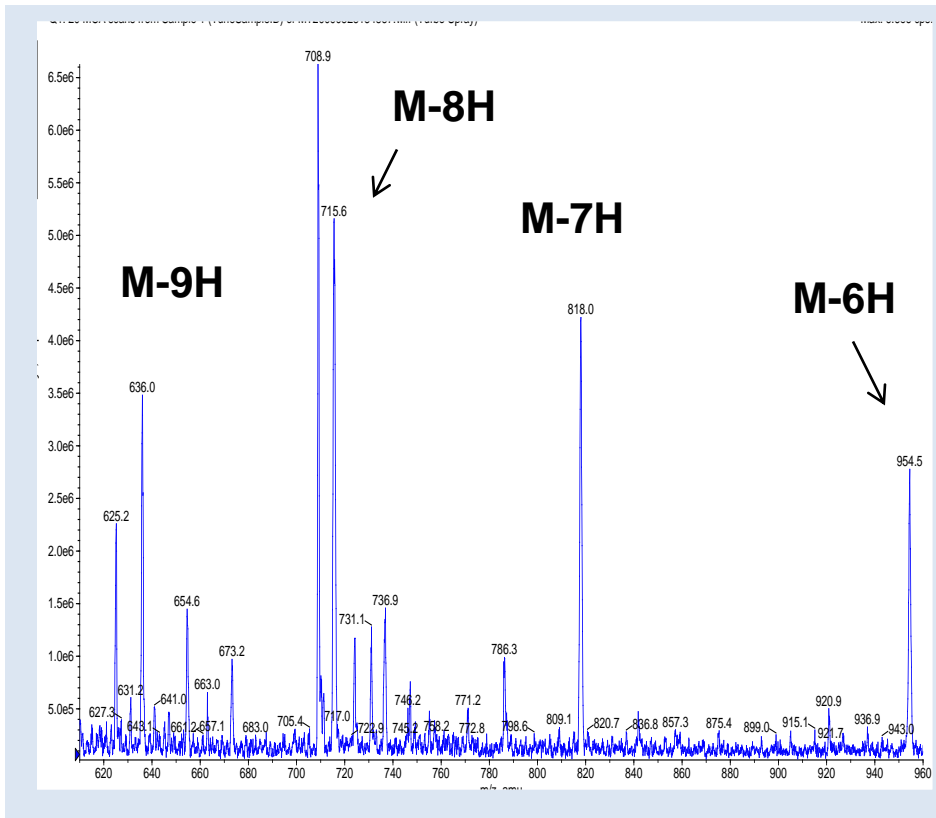


Isotope envelopes of the  $[M-3H]^{3-}$  ion of an 18-mer PS-ODN on a high resolution Q-Exactive Orbitrap<sup>®</sup> (Thermo Finnigan, LLC)

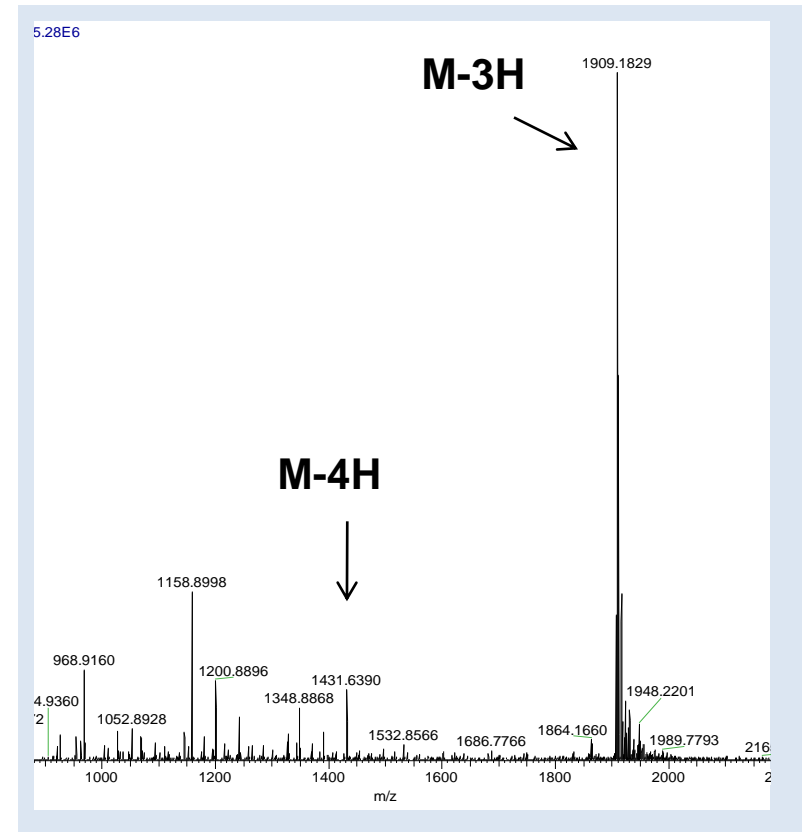


# Different Charge Distribution of a 18-mer PS-ODN on Different Platforms

API-5000

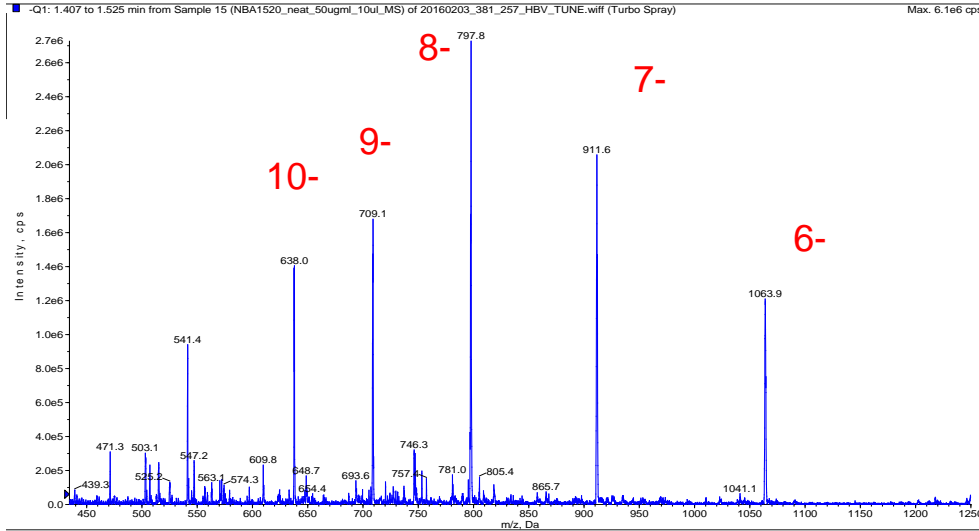


Q-Exactive



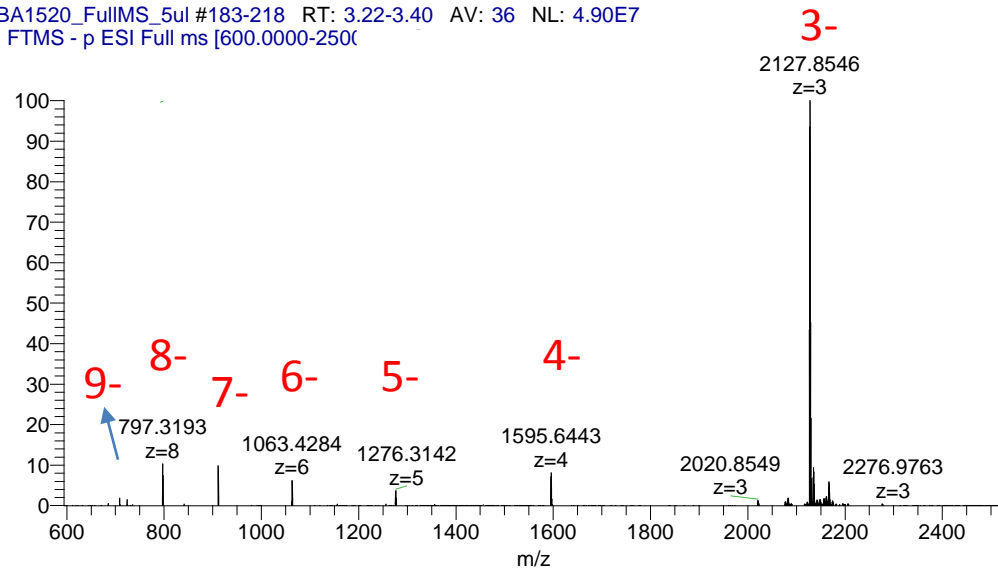


# Different Charge Distribution of a 20-mer PS-ODN on Different Platforms



API5000 Triple Quads

NBA1520\_FullMS\_5ul #183-218 RT: 3.22-3.40 AV: 36 NL: 4.90E7  
T: FTMS - p ESI Full ms [600.0000-2500]

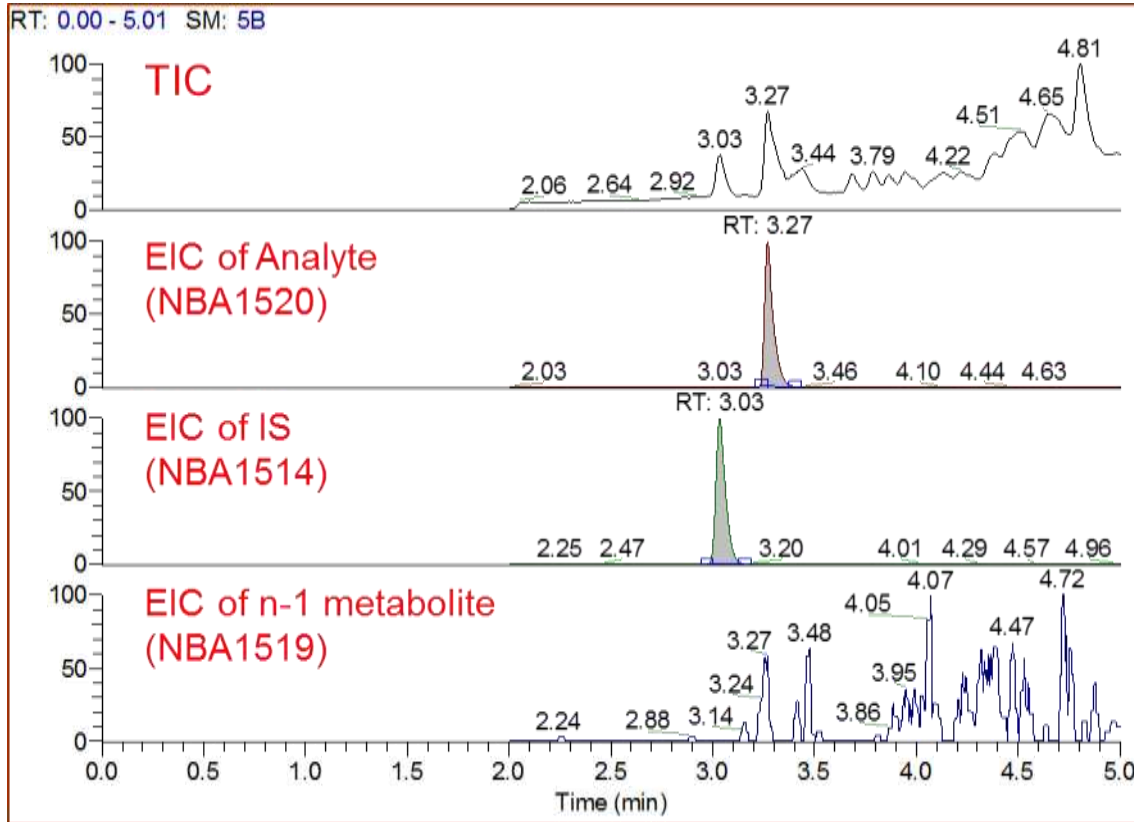


Q Exactive Plus





# Full Scan Analysis of Extracted 20-mer PS-ODN (HuPL) on Q-Exactive Plus (1/3)



Extraction:

*SPE from 200  $\mu$ L HuPI*

LC Column:

*Thermo DNAPac RP  
4  $\mu$ m, 2x50 mm*

Column Temp: 60  $^{\circ}$ C

Mobile Phase:

*HFIP/TEA buffered water/  
MeOH Gradient*

Run Time: 8 minutes

LC System:

*Thermo Vanquish UPLC*

MS: Q-Exactive Plus

Ionization Mode: Negative

Resolution: 70K

Scan Range 600-2500  $m/z$

**NBA1520:** 5'--TAG TCA ATC TGC TTA TGT CA--3'

**NBA1519:** 5'--TAG TCA ATC TGC TTA TGT C--3'

**NBA1514:** 5'--TAG TCA ATC TGC TT--3'

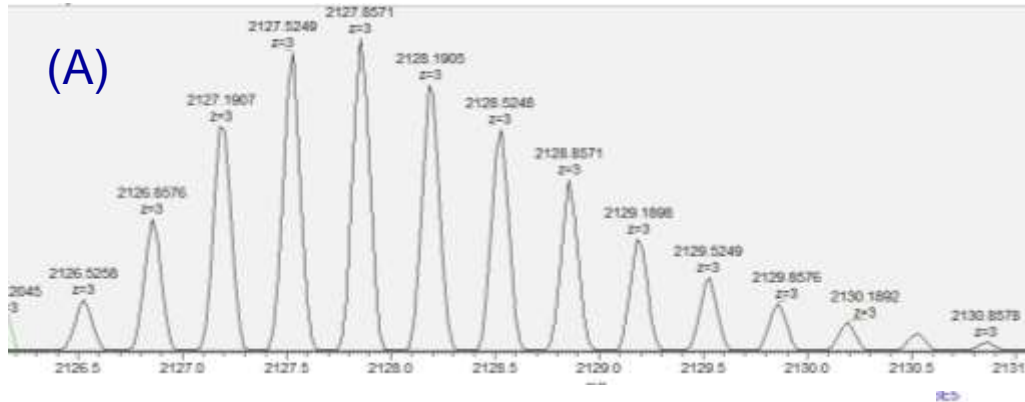
**C196H249N68O102P19S19**

**C186H237N63O98P18S18**

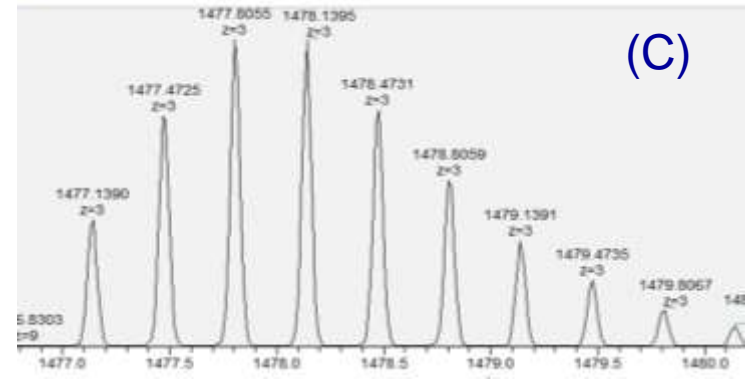
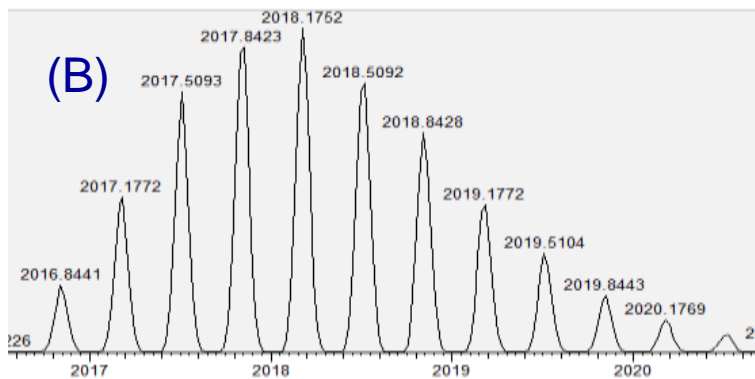
**C137H175N46O72P13S13**



# Full Scan Analysis of Extracted 20-mer PS-ODN (HuPL) on Q-Exactive Plus (2/3)



(A): NBA1520  
(B): NBA1519  
(C): NBA1514



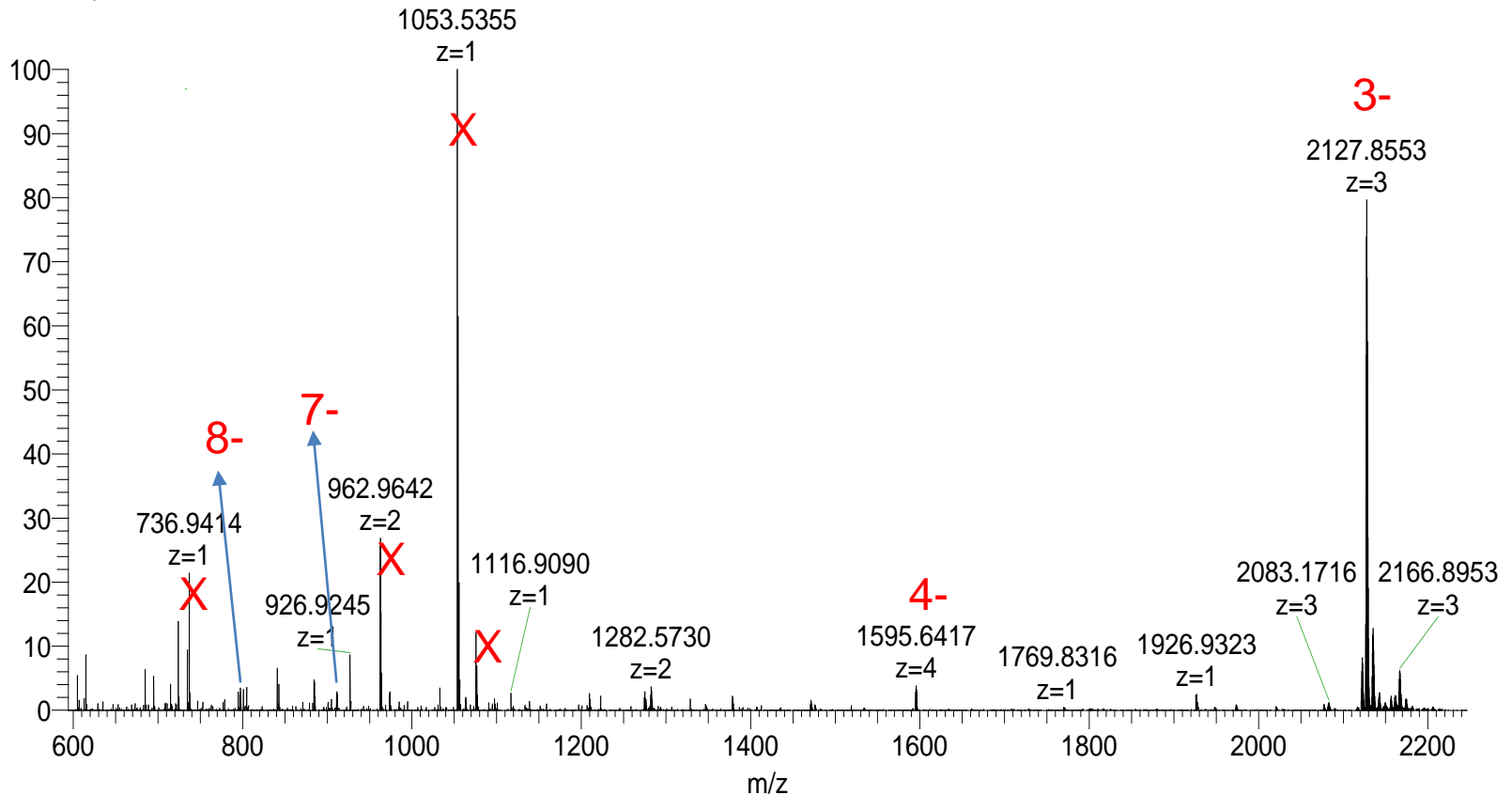
**NBA1520:** 5'--TAG TCA ATC TGC TTA TGT CA--3'  
**NBA1519:** 5'--TAG TCA ATC TGC TTA TGT C--3'  
**NBA1514:** 5'--TAG TCA ATC TGC TT--3'

**C196H249N68O102P19S19**  
**C186H237N63O98P18S18**  
**C137H175N46O72P13S13**



# Full Scan Analysis of Extracted 20-mer PS-ODN (HuPL) on Q-Exactive Plus (3/3)

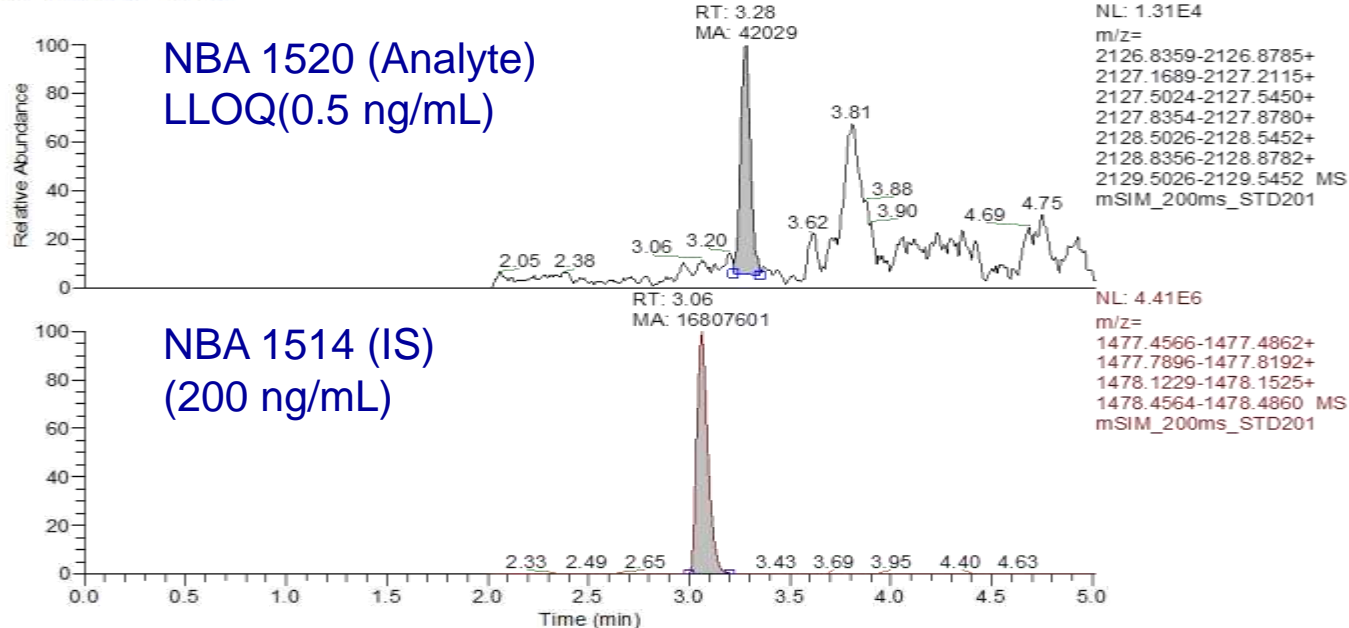
Full-MS\_STD210 #192-207 RT: 3.25-3.33 AV: 16 NL: 2.09E6  
T: FTMS - p ESI Full ms [600.0000-2500.00]





# Target-SIM Analysis of Extracted 20-mer PS-ODN (HuPL) on Q-Exactive Plus (1/3)

RT: 0.00 - 5.02 SM: 5B



**Extraction:** SPE from 200  $\mu$ L HuPL

**LC Column:** Thermo DNAPac RP

4  $\mu$ m, 2x50 mm

**Mobile Phase:**

HFIP/TEA buffered water/MeOH

**LC System:**

Thermo Vanquish Flux UPLC

**MS:** Q-Exactive Plus

**Ionization Mode:** Negative

**Resolution:** 70K

**t-SIM:**

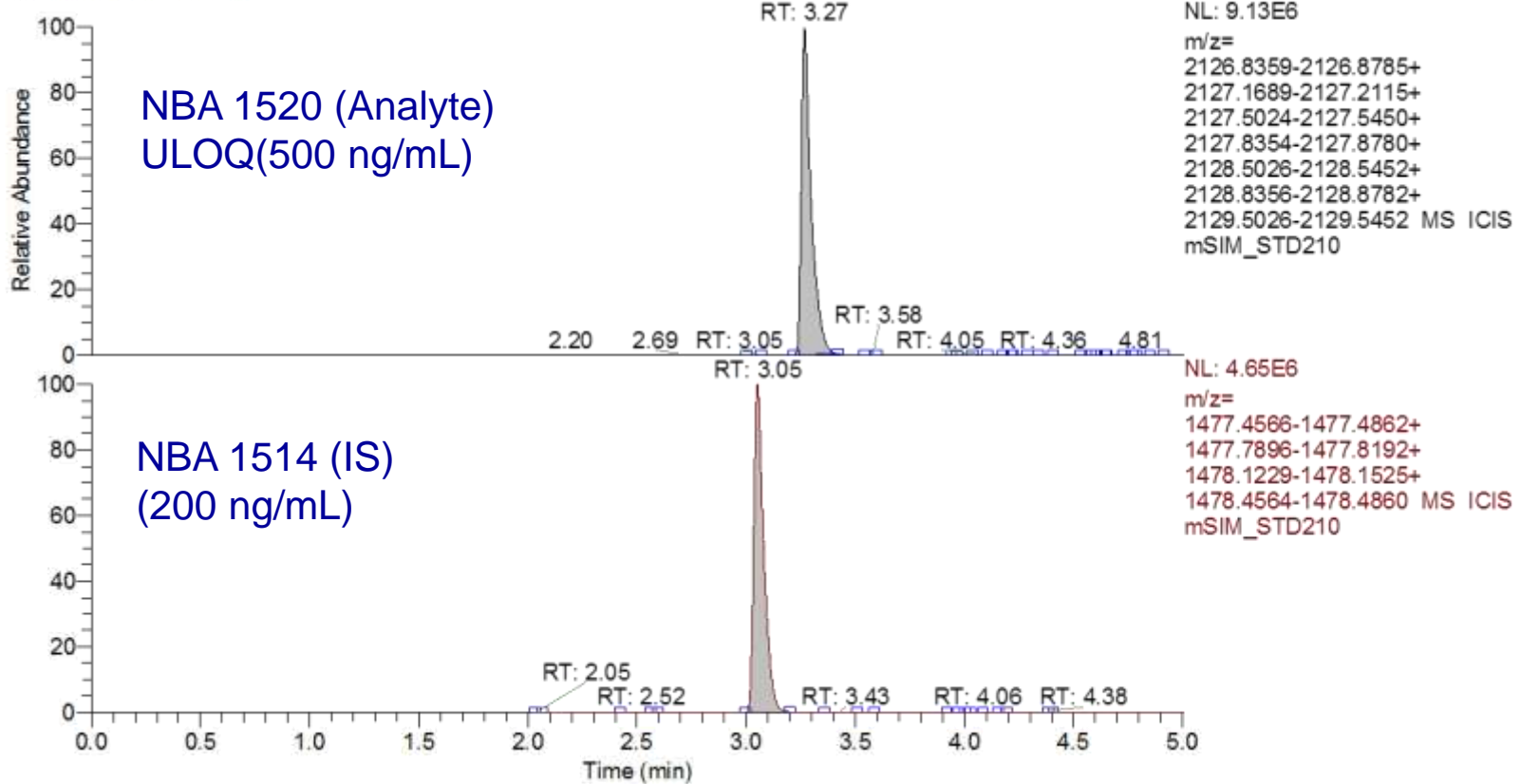
**Analyte:** 2123.8610-2131.8610

**IS:** 1473.8065-1481.8065



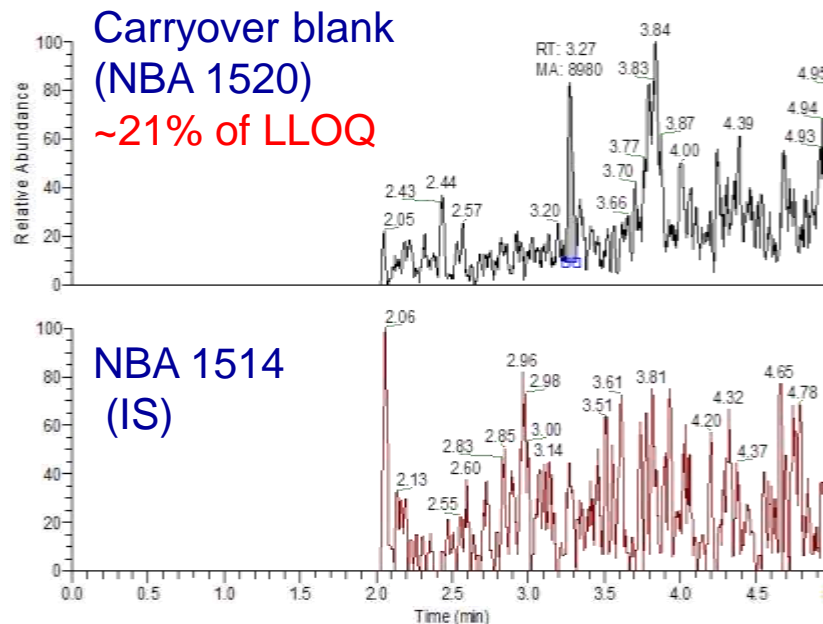
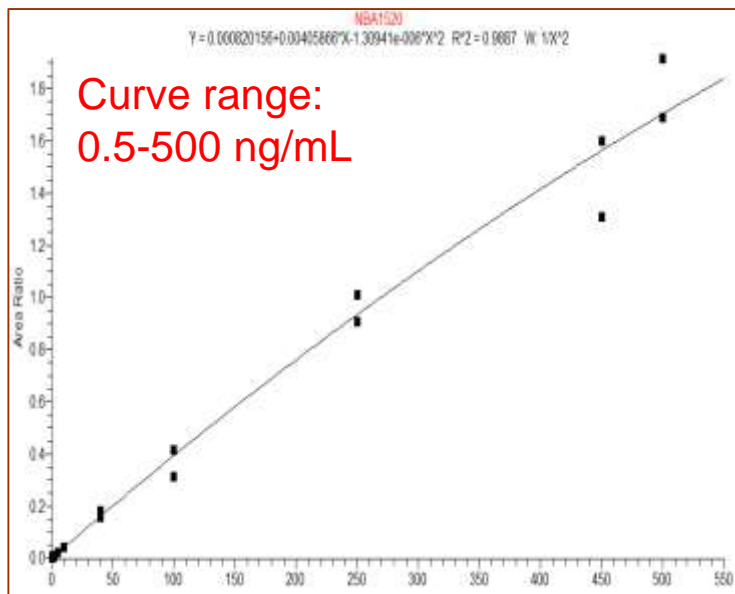
# Target-SIM Analysis of Extracted 20-mer PS-ODN (HuPL) on Q-Exactive Plus (2/3)

RT: 0.00 - 5.01 SM: 5B





# Target-SIM Analysis of Extracted 20-mer PS-ODN (HuPL) on Q-Exactive Plus (3/3)

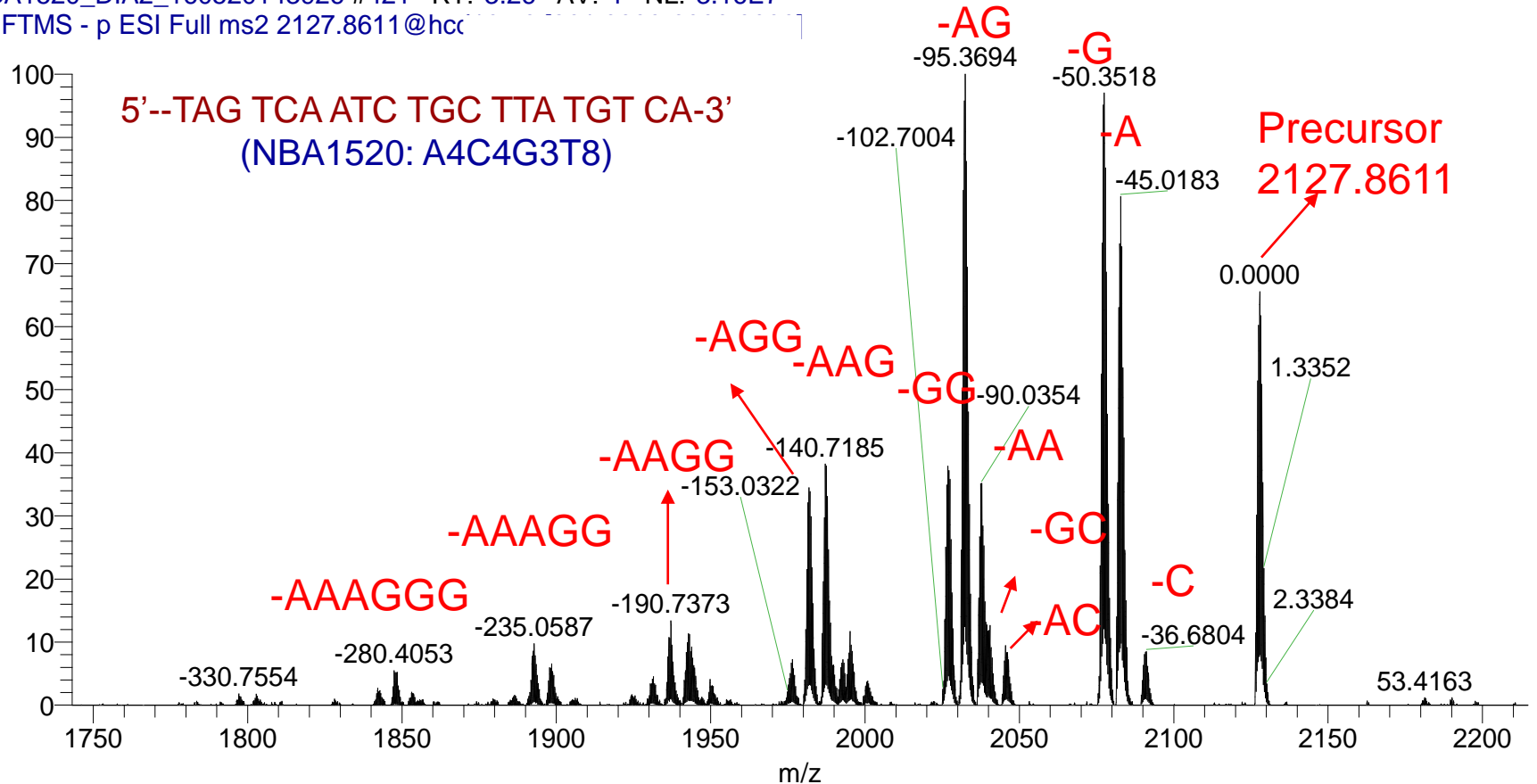


	Nominal C. (ng/mL)	Average C. (ng/mL)	% Bias	CV%
High QC	400.	395	-1.25%	4.51%
Medium QC	200.	193.	-3.50%	5.29%
Low-Medium	15.0	14.2	-5.33%	7.10%
Low QC	1.50	1.41	-6.00%	4.48%



# Fragmentations of PS-ODN on Q-Exactive for PRM/DIA Analysis Plus (1/2)

NBA1520\_DIA2\_160520143926 #421 RT: 3.26 AV: 1 NL: 3.16E7  
T: FTMS - p ESI Full ms2 2127.8611@hcc

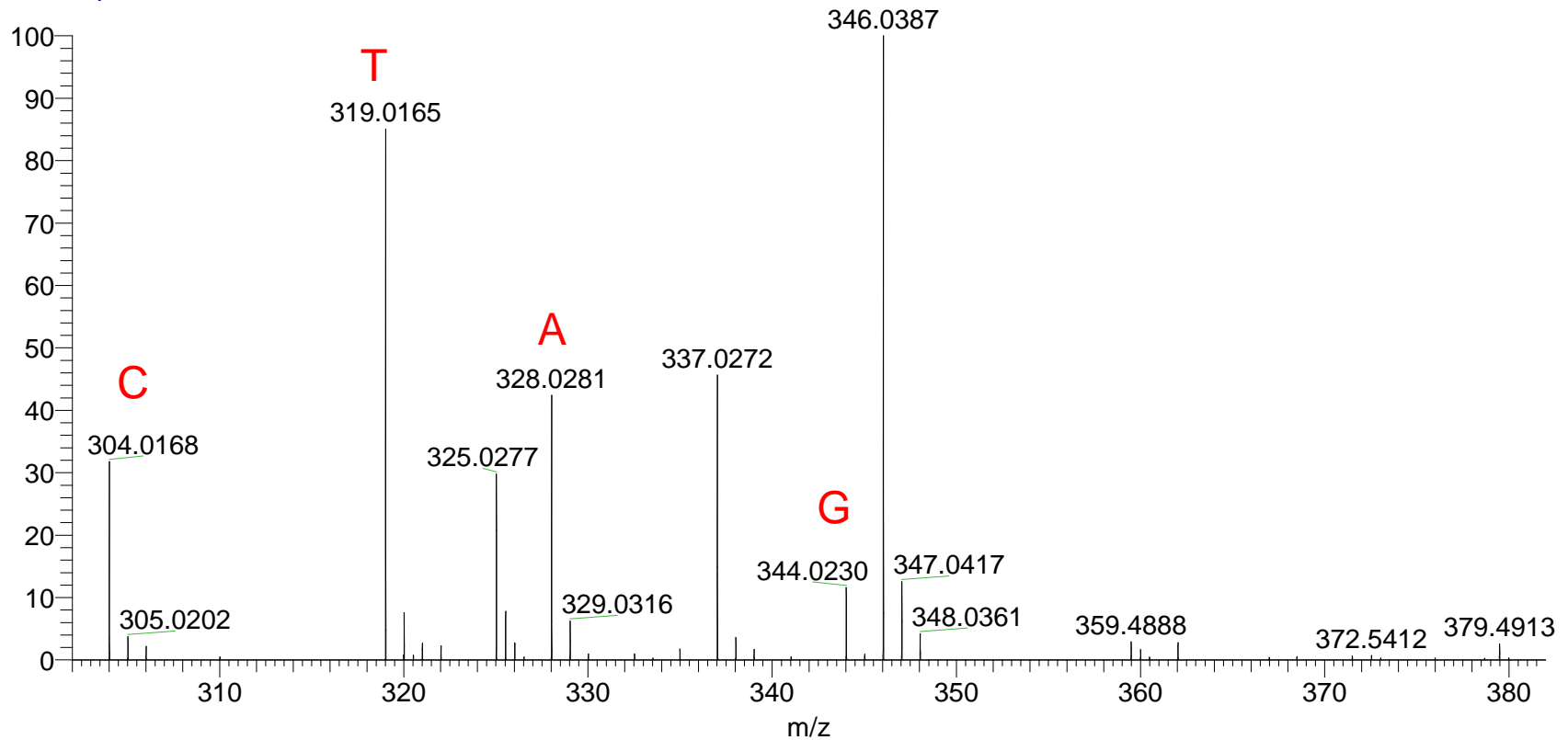






# Fragmentations of PS-ODN on Q-Exactive for PRM/DIA Analysis Plus (2/2)

NBA1520\_DIA2\_160520143926 #425 RT: 3.27 AV: 1 NL: 6.11E5  
T: FTMS - p ESI Full ms2 797.3195@hcd'





# SPE Extraction of the 20-mer PS-ODN From Human Plasma

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**Extraction SPE Plate:** 96-Well WAX SPE plate, 30 mg,

**Sample Aliquot:** *200  $\mu$ L Human Plasma*

**Extraction Procedure:**

- 1) Aliquot 200  $\mu$ l sample
  - a) Add 50  $\mu$ l IS to each well
  - b) Add 500  $\mu$ l loading buffer
  - c) Vertex briefly and spin down
- 2) Load the samples onto the SPE plate  
(Pre-equilibrated with 1 mL MeOH and 1 mL loading buffer)
- 3) Wash the plate with 1 ml loading buffer and 1 mL of washing buffer
- 4) Heat sample plate for 1 hr incubation at 70°C
- 5) Elute the sample with 250-500  $\mu$ l elution buffer  
(contains 10% methanol)
- 6) Inject 10  $\mu$ l for LC-MS analysis



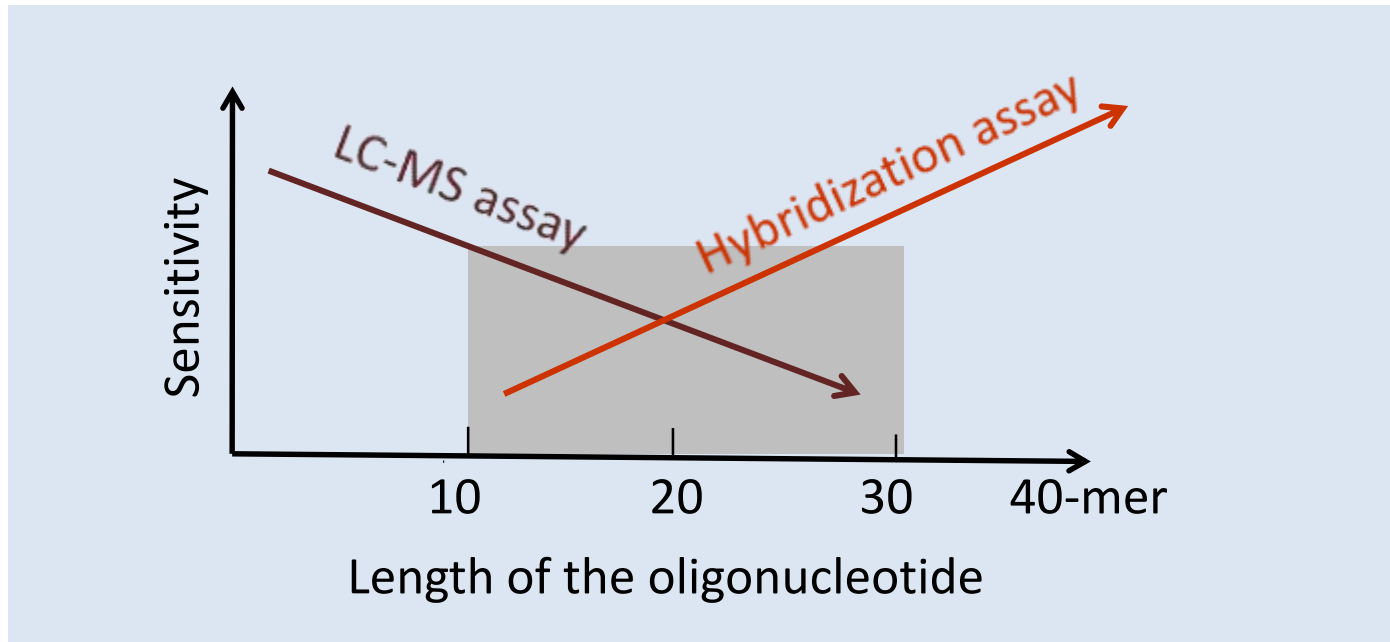
## LC-MS/MS vs LC-HRAM

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- Q1 mass spectra of oligos have different charge distributions on different instrument platforms
- LC-HRAM on Q-Exactive can achieve comparable sensitivity as LC-MS/MS on triple quads for some analytes.
- LC-HRAM can be performed without any prior knowledge of the compound, thus avoiding the need for compound-specific tuning (setting up the SRM transition). Therefore, it is very convenient for monitoring unknown metabolite information or estimate the known metabolite concentration(s) without standard reference material while quantifying the parent compounds.



# Size Matters for The Assay Sensitivities



- LC-MS/MS (HRAM) assays are a good choice for small oligonucleotides;
- Hybridization-based assays are advantageous for large oligonucleotides.



# THANK YOU

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