

#### **ThermoFisher** SCIENTIFIC

# HRAM iQuan 2016: Thermo Scientific<sup>™</sup> Q Exactive<sup>™</sup> Mass Spectrometer Series Calibration & Maintenance

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### HRAM iQuan Agenda

#### **Tuning and Calibration**

- Tune Software Interface (v.2.6)
- Preparation
- Tuning for Calibration Solvents
- Calibration Procedure

#### Maintenance

- Considerations
- Schedules
- How-to's



#### Thermo Scientific<sup>™</sup> Q Exactive<sup>™</sup> Mass Spectrometer Calibration & Maintenance



#### Calibration

#### Preparation, Tuning, Calibration





#### Q Exactive MS Series Tune Interface (Tune v2.6): Overview





#### Q Exactive MS Series Tune Interface (Tune v2.6): Display Panel Options

Thermo Q Exactive Focus — operational	
File Windows Reports Help	Role: Adva
Interventions   Instrument Control     Scan parameters   Ititory   Scan type   Full MS   Scan range   150.0 to 2.000.0 m/z   Fragmentation   None   Resolution   17.500   Polarity   Positive   Microscans   1   Lock masses   Off   AGC target   1e6   Maximum inject time   50   Apply   Help   Help   Hot link   Source Auto-Defaults   Apply   Help   Hot link	Spectrum       Spectrum         Spect
Mass Traces     Calibrate     Surfaces	<ul> <li>14:54:10.37: Document C\Xcalibur\system\Exactive\instrument\reports\90180E665870_Spectral Mass Calibration (pos)_2016-06-06_14-53-52.pdf generated.</li> <li>15:00:56.37. Set user role to Standard.</li> <li>15:00:56.37. Set user role to Standard.</li> <li>15:00:14:146. Set user role to Advanced.</li> </ul>
Vacuum / Bakeout	Messages Analysis Graphs



#### Q Exactive MS Series Tune Interface (Tune v2.6): Display Panel Options





## Q Exactive MS Series Tune Interface (Tune v2.6): Display Panel Options

e Windows Reports Help		Kole: Aava
Instrument Control	Spectrum	🥸 Instrument Status
Mass Traces	🚃 📶 🖕 🔄 🔄 🔄 🔄 💽 🔍 🐝 🎰 🔆 🗡	🤣 🗆 Instrument
Calibrata		Current Scan
Calibrate	Type: FTMS + p ESIFull ms [150.00-2000.00]	Total Ion Current 2035.07 E6 ions/
		TIC Variation 4 %
Cambrate	100	Inject time 0.64 ms
0.0 %		AGC larget reached 100 %
		Prescan Mode -1
	80	k marrer
Calmix Calibration	Tune v2.8SP1 available now	k masses
Base Calibration OK (2016-05-11 14:03)		
Electronics		sis Graph
Quadrupole RF Frequency 2016-05-11 12:27		⊕ Performance Ok
Flatapole RF 2016-05-11 12:27		Electronics
HCD RF 2016-05-11 12:28		ESI Ion Source
Basics - Positive Ion OK (2016-05-11 14:03)		Peripheral Devices
Ouadrupole Basic (pos)		Vacuum System
Quad Iso Mass/Res Cal (rough) 2016-05-11 14:03	$20 - \frac{1}{262,636}$	
Quad RF freq. fine adjustment 2016-05-11 14:03	360.238 524.265 1221.991 1321.984 1421.977 1521.972 17	21.960
eFT Parameter (pos) OK (2016-05-11 14:03)	196.091 338.341 1121.997	21.967
Analyzer Accuracy (pos) OK (2016-05-11 14:04)		Vacuum System
Basics - Negative Ion OK (2016-05-12 09:46) Ion transfer (neg)	500 1000 1500	2000
Quadrupole Basic (neg)	m/z	
eFT Parameter (neg) OK (2016-05-12 09:46)	Analysis Graphs	
Analyzer Accuracy (neg) OK (2016-05-12 09:47)		
□ Isolation Mass and Res. (pos) OK (2016-05-11 14:36)		
Iso Mass/Res. Cal. (wide iso) 2016-05-11 14:25	Mass accuracy BMS deviation mit 198.1 mit 195.1 mit 572.3 mit 197.0 mit 197.0	
Iso Mass / Kes. Cal. (narrow) OK (2010-05-11 14:36) □ Isolation Mass and Res. (neg) OK (2016-05-12 10:04)		
Iso Mass/Res. Cal. (neg)(wide iso) 2016-05-12 09:54		
Iso Mass/Res. Cal. (neg)(narrow) OK (2016-05-12 10:04)		
MS Mass Calibration (pos) OK (due in 7 days)		
MS Mass Calibration (neg) Overdue (2016-05-12 10:17)		
Customized Calibration	Tune v2.8SP1 available now!	
	* E.2	
		* * * *
Evaluate	5 IV 15	20



#### Instrument Control Software Update

- Q Exactive MS Series 2.8 SP1 is released for Windows 7 32-bit and 64-bit
- Recommended to upgrade all instruments running Q Exactive MS Series Tune 2.7 or older to this release
  - BRE0008597 Exactive MS Series 2.8SP1 / Xcalibur 3.1 (for Windows 7 32-bit)
  - BRE0008596 Exactive MS Series 2.8SP1 / Xcalibur 4.0 (for Windows 7 64-bit).
- Full list of improvements and defect fixes can be found in release notes included with the software
  - Main changes with the release of 2.8 SP1 are:
    - Modified bakeout procedure to improve lifetime of turbo pumps
    - Defect fix for some calibration procedures (Isolation Transmission Endurance Test, normal mode and HMR mode eFT)
    - TIC variation readback in v 2.7 did not generate reliable output





### Calibration is Qualitative, Tuning is Quantitative

- Calibration ... Mass spectrometer optimization
  - Calibration is independent of source type
    - Exactive MS series calibrations must be performed using the ESI or HESI-II probe
  - Successful calibrations are saved automatically
    - Only one calibration file exists

#### • Tuning ... Compound optimization

- Efficiently generating ions from the source into the mass spectrometer
- Tune files must be manually saved after the tuning process
- Compound/Application/Method dependent
  - Multiple tune files can be saved to a method



#### Q Exactive MS Series: What is **Tuned** and What is **Calibrated**?





- Calibrations can be performed immediately if the instrument switched from "STANDBY" to "ON" mode
- If the instrument switched from "OFF" to "ON" mode, scan in FTMS mode for 80 minutes prior to calibration
  - Does not require spray voltage or liquid flow
- Instrument reset, instrument shut down, bakeout and leaving the source open for more than 60 min, will place the instrument's electronics in "OFF" mode
  - Make sure your instrument has warmed up (scanned) for 80 minutes if the electronics have been off
- Use the correct calibration solutions for your instrument (store in the dark at room temp.)
  - Pierce<sup>™</sup> LTQ Velos ESI Positive Ion Calibration Solution (*Catalog no. 88323*)
  - Pierce<sup>™</sup> Negative Ion Calibration Solution (*Catalog no. 88324*)





 Electronics service switch is in the Operating Mode position





- Electronics service switch is in the Operating Mode position
- Check that the gas pressure is within the operational limit

Nitrogen: 800  $\pm$  30 kPa (8  $\pm$  0.3 bar, 116  $\pm$  4 psi)





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Nitrogen: 800  $\pm$  30 kPa (8  $\pm$  0.3 bar, 116  $\pm$  4 psi)

 Check that the vacuum levels are sufficient for operating the instrument

_		
<b>V</b> 91	Instrument Status	×
0	Instrument	
	🗆 Current Scan	
	Total Ion Current	2049.90 E6 ions/sec
	TIC Variation	5 %
	Inject time	0.66 ms
0	AGC Target reached	100 %
	AGC Prescan Mode	-1
	Scan Rate	13.3 scans/sec
	Lock masses	
	Control	
	🕀 System	
	Analysis Graph	
	Performance	Ok
0	Electronics	
0	ESI Ion Source	
0	Peripheral Devices	
0	🖃 Vacuum System	
0	Status	Ok - HV enabled
0	Fore Vacuum	1.43e+00 mbar
	High Vacuum	Turned off
0	Ultra High Vacuum	1.87e-10 mbar
	Source TMP Speed	100 %
	UHV TMP Speed	100 %



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- Check that the gas pressure is within the operational limit

Nitrogen: 800  $\pm$  30 kPa (8  $\pm$  0.3 bar, 116  $\pm$  4 psi)

- Check that the vacuum levels are sufficient for operating the instrument
  - Fore Vacuum < 2 mbar
  - High Vacuum "Turned off"
  - Ultra High < 5E-10 mbar
  - Source and UHV TMP Speed 100%

<b>V</b> 91	instrument Status		×
0	🗉 Instrument		
	Current Scan		
	Total Ion Current	2049.90 E6 ions/sec	:
0	TIC Variation	5 %	
	Inject time	0.66 ms	
0	AGC Target reached	100 %	
	AGC Prescan Mode	-1	
	Scan Rate	13.3 scans/sec	
	Control		
	표 Analysis Graph		
	Performance	Ok	
0	Electronics		
0	ESI Ion Source		
0			
0	🖃 Vacuum System		1
0	Status	Ok - HV enabled	Т
0	Fore Vacuum	1.43e+00 mbar	
	High Vacuum	Turned off	
0	Ultra High Vacuum	1.87e-10 mbar	
	Source TMP Speed	100 %	
	UHV TMP Speed	100 %	



- An infusion line should be installed between the syringe pump and the grounding union that is held by the grounding bar of the lon Max API source
  - Clean 500 µL Hamilton (Thermo) syringe
  - Pierce LTQ Velos ESI Positive/Negative Ion Calibration Solution
  - Tubing, PEEK<sup>™</sup> (0.005-in ID × 1/16-in. OD (red))
  - Fitting, fingertight, Upchurch Scientific (natural) (used with (red) PEEK<sup>™</sup> tubing)





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  - Fitting, fingertight, Upchurch Scientific (natural) (used with (red) PEEK<sup>™</sup> tubing)
- Ion Max API source with HESI II probe are installed on the mass spectrometer and proper position of the HESI II probe is established





### Establishing a Tune: Conditions for Calibration Solutions

- Remember to create a tune under the conditions used for calibration
  - Fill syringe with calmix solution
    - Use 500 µL syringe
    - Start with positive calmix first
  - Set flow rate to 5 µL/min
  - Set up scan parameters appropriately
    - Mass range 150-2000 m/z
    - AGC target 1e6
    - 17,500 resolution



- Optimize source conditions for best signal stability
  - Source conditions include: gas flows, voltages, temperatures, source probe adjustments
    - TIC Variation <10% RSD</li>
    - NL signal (min 1e8)
    - IT < 2 ms

HESI source		\$
		actual
Sheath gas flow rate	5	6
Aux gas flow rate	1	1
Sweep gas flow rate 0		0
Spray voltage (  kV  )	3.50	3.50
Spray current (µA)		0.50
Capillary temp. (°C)	300	300
S-lens RF level	50.0	
Aux gas heater temp (°C)	SI source         acts           ss flow rate         5         6           low rate         1         1           s flow rate         0         0           use (lkV)         3.50         350           temp. (*C.)         300         300           level         50.0         41           Auto-Defaults         W Hot link         1	41
Source Auto-Defaults		
Apply Help	↓ Int link	

Save Tune File (pos/neg)



### Typical Source Settings for Calibration Solutions

Example for HESI-II probe and syringe pump infus Syringe Pump Flowrate: 5 µL/min	Sion Aux gas flow is either not required or set lower than Sheath
Instrument Control  Scan parameters HESI source	Spray voltage for positive and negative mode is between 3 and 4.5 kV
Sheath gas flow rate     5       Aux gas flow rate     1       Sweep gas flow rate     0       Spray voltage ( kV )     3.50	actual 6 1 Capillary temp (set at 320 °C for calibration) 3.50
Spray current (μA)         Capillary temp. (°C)       300         S-lens RF level       50.0         Aux gas heater temp (°C)       0	<ul> <li>0.50</li> <li>300</li> <li>S-lens level may have to be adjusted manually (typical range is 50-60)</li> </ul>
Source Auto-Defaults Apply Help V Hot link	HESI-II Probe Heater temp is NOT used and can be set to 0 °C



### Establishing a Tune: Requirements

- Optimize source settings for calmix solution using 5 µL/min flow (see next slides)
- Check spray stability: monitor the **TIC Variation** (RSD < 10%)
- Monitor injection time (IT): < 2 ms and establish strong signal (NL) (≥ 1e8)





#### Establishing a Tune: What to Know About the Tune File

- Unlike calibration files, results of tuning procedure are **not** saved automatically
- Tune files only save the source parameters (e.g. gases, voltages, temps)
- The source parameters for both positive and negative polarity modes can be saved in the same tune file
- To save optimized parameters, select File > Save Tune to overwrite the previous Tune file or File > Save Tune as to capture the conditions in a new file

Next order of operation ... calibration!

×		
HESI source		actual
Sheath gas flow rate	5	6
Aux gas flow rate	1	1
weep gas flow rate	0	0
opray voltage (  kV  )	3.50	3.50
Spray current (µA)		0.50
Capillary temp. (°C)	300	300
S-lens RF level	50.0	
Aux gas heater temp (°C)	0	41
Source Auto-Defaults		



#### Q Exactive MS Series: A Good Positive Calibration Spectrum





### Q Exactive MS Series: A Good Negative Calibration Spectrum





### Calibration: Tips for Success

- Make sure calibration solutions are fresh
- Ensure infused calibration solution contains all calibrant masses (pos/neg)
- Maintain stable spray
  - Focus on the injection time (IT, < 2 ms) and normalized level (NL, minimum 1e8) in the Tune Page scan header
  - In the instrument status pane, monitor the TIC Variation, RSD < 10%</li>
- All successful calibration procedures are updated and saved automatically in the "master.cal" file

















- NOTE: Always **check/evaluate**, if a procedure fails then calibrate it
- Positive Ion Evaluation: evaluate every month; if it fails, calibrate Isolation Mass and Resolution

Evaluate		Role: Standard
<b>A</b>		
Evaluate		
0.0×		
0.0 % valuate Stop Help Calmix Evaluation Positive Ion Evaluation Base Evaluation (pos) Isolation Evaluation (pos)	🤣 Instrument Status	×
	🕑 🗆 Instrument	
Calmix Evaluation	Current Scan	
Positive Ion Evaluation		
Base Evaluation (pos)		
<ul> <li>Isolation Evaluation (pos)</li> </ul>		
MS Mass Check (pos)		
📃 😑 Negative Ion Evaluation		
Base Evaluation (neg)		
Isolation Evaluation (neg)		
MS Mary Chark (mag)		
ma mass check (neg)		
Extra Evaluation		



- NOTE: Always check/evaluate, if a procedure fails then calibrate it
- Positive Ion Evaluation: evaluate every month; if it fails, calibrate Isolation Mass and Resolution
- **MS Mass Calibration** (positive/negative): evaluate daily/weekly; if it fails, calibrate
- Calibrate unused polarity modes (e.g. negative mode) at least every 3 months





#### Electronics:

Calibrated in factory, calibrate only when hardware is replaced

#### Basic Positive or Negative:

- Ion transfer: calibrated in the factory
- Quadrupole Basic: evaluate monthly, calibrate only when Isolation Mass and Resolution calibration fails
- **eFT**: evaluate monthly, calibrate if it fails. Always perform a MS Mass Calibration after eFT calibration
- Analyzer Accuracy: evaluate monthly, if it fails calibrate (C-trap Charge Detection)
- Isolation Mass and Resolution: evaluate monthly, if it fails calibrate. If calibration fails, check quadrupole Basic calibration
- MS Mass Calibration (positive/negative): evaluate daily/weekly; if it fails, calibrate





#### Extra Evaluation – Calmix Evaluation (Introduced in ICSW v2.5)



Done: Isolation Transmission Endurance Test: Transmission score at 0.98

Electronics evaluations are NOT recommended on a regular basis

These evaluations should be performed upon request with guidance from tech support

The evaluation REQUIRES a stable spray of calmix (Intensity > 1e8; TIC Variation < 10%; IT < 2 ms) Calmix infusion starts automatically



2

OK

#### Calibration: Regular Backup of the Calibration File





#### Calibration: Reports (Tune v2.6)

🗊 Thermo Q Exactive I	Focus — operational			
File Windows Repo	ts Help Calibration  Spectral Mass Calibration (neg)  Spectral Mass Calibration (pos)  C		10.	Role: Advanced
🕤 Instrume		Image: State in the state		
al .		📲 🍻 🔍 🔍 🔍 🍳 🔍 👾 🎬 🎇 俊	Current Soon	
MU Scan parame	eters 🎗	Scan: #109850 µS: 1 IT: 0.65 NL: 5.17E8	Total Ion Current	1005 84 E6 ions/sec
History		Type: FTMS + p ESI Full ms [150.00-2000.00]	TIC Variation	2 %
Scan type	Full MS		Inject time	0.66 ms
Scan range	150.0 to 2,000.0 m/z	100 195.088	AGC Target reached	100 %
Fragmentation	None		AGC Prescan Mode	-1
Resolution	17,500		Scan Bate	13.1 scans/sec
Polarity	Positive		+ Lock masses	
Microscans	1	80 -	Control	
Lock masses	Off		+ System	
AGC target	1e6	<b>8</b>	Analysis Graph	
Maximum inject time	50		Performance	Ok
Annha			Electronics	
	The link		ESI Ion Source	
	<u>^</u>		Peripheral Devices	
HEST SOURCE	actual		Vacuum System	
Sheath gas flow rate	5 5		Status	Ok - HV enabled
Aux gas flow rate	1 1		Fore Vacuum	1.43e+00 mbar
Sweep gas flow rate	0 0		High Vacuum	Turned off
Spray voltage (  kV  )	3.50 3.50		Ultra High Vacuum	1.87e-10 mbar
Spray current (µA)	0.40	262,636	Source TMP Speed	100 %
Capillary temp. (°C)	300 300	20 -	UHV TMP Speed	100 %
S-lens RF level	50.0	300/238 524/265 1221/991 1321/985 1421/9/8 1521/972 1621/969		
Aux gas heater temp	(°C) 0 41	121.197 121.298 122.298		
Source Auto-Defau	ilts		Vacuum System	
Apply Help	V Hot link	500 1000 1500 2000 m/z		
	*	Analysis Graphs		×
Acquisition state	adv.	III 😹 😣		
Progress 0.0	0 min	Mate accuracy		
File in use	known			
Destination file C4	Xcalibur\Data\Bus Nol C2			
Method file by	time 1			
Mass Traces				
			· · ·	
Galibrate				
🌂 Evaluate		0 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	20	25
Vacuum / Bak	eout	Messages 🔐 Analysis Graphs		



### Calibration: Reports (Tune v2.6)

📁 Thermo Q Exactive	Focus — operational					
File Windows Rep	orts Help					Role: Advanced
	Calibration  Latest					
	Spectral Mass Calibration (neg)	Sole Edit View Window Help	- Adobe Reader	_		
	Spectral Mass Calibration (pos)					Tools Sign Comment
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History	$\rightarrow$	Ca Ca	alibration Report	Thermo	19 138 06619 195 08765 524 26497 1221 99037 1421 97738 1621 96584	
Scan type	Full MS			SCIENTIFIC	20 138.06618 195.08765 524.26493 1221.99038 1421.97730 1621.96576	
Scan range	150.0 to 2,000.0 m/z	Date	e of calibration: 2016-06-06 14.53.52 dows user name: FDKW252/Thermol)		mean value 138.06619 195.08765 524.26493 1221.99046 1421.97731 1621.96581	
Fragmentation	None	QB	active Focus 2.6-26401/2.6.0.2640		std. dev.[ppm] 0.023 0.000 0.045 0.065 0.058 0.055	
Resolution	17,500	inst	rument identification. Exactive Series slot 4150 (S01606065870)		Mass accuracy	
Polarity	Positive	Type	e of calibration: Spectral Mass Calibration (pos)			
Microscans	1	Scar scar	n settings: n range: 150.0 - 2000.0			
Lock masses	Off	reso AGC	Nution: 17500 : target : 1000000			
AGC target	1e6	mice	ro scans : 1			
Maximum inject tim	50 S0					
Apply	Help 🛛 Hot link	Che	ck ok: ion signal (ion s/n 2683, TIC 1.59e+09)		• •	
P-d		Mes	is accuracy without lock mass			
HESI source			Scan 138.06619 195.08765 524.26496 1221.99064 1421.97786 1621.96509		0 5 10 15 20 25	
	-		1 138.06621 195.08769 524.26505 1221.99070 1421.97754 1621.96612		Mass Calibration Trend / History	
Sheath gas flow rate	. 5		2 138.06620 195.08767 524.26500 1221.99054 1421.97731 1621.96606 8 198.06510 195.08767 524.26500 1221.99056 1421.97738 1521.96512		Hos Calculation ( Here / Hosey)	
Aux gas flow rate	1		4 138.06622 195.08769 524.26510 1221.99081 1421.97756 1621.96616		10	
Sprawyoltage (Jk//	350		5 138.06621 195.08769 524.26503 1221.99065 1421.97758 1621.96608			
Spray current (uA)	, 5.50		6 138.06618 195.08764 524.26492 1221.99037 1421.97721 1621.96553 7 138.06618 195.08764 524.26495 1221.99042 1421.97725 1621.96574			
Capillary temp. (°C)	300		8 138.06620 195.08767 524.26498 1221.99064 1421.97748 1621.96578			
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		std	dev.[ppm] 0.093 0.097 0.101 0.106 0.096 0.131		-15	
Source Auto-Defa	ults				dayn	
Apply Hel	p V Hot link	Mas	s accuracy with lock mass 195.08765		Procedure result: passed (rms = 0.25/0.27 ppm)	
Acquisition			Scan 138.06619 195.08765 524.26496 1221.99064 1421.97786 1621.96509		Calibration Reagent:	
Acquisition state	adv		12 138.06619 195.08765 524.26491 1221.99042 1421.97727 1621.96564		Lot Number:	
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#### Report Files

 Saved in data system's local C:Xcalibur\system\Exactive\instrument \reports

Organize 🔻 🛛 Include in	library ▼ Share with ▼ Burn New fo	older				
Favorites	Name	Date modified	Туре	Size		
🧮 Desktop	📔 temp	5/11/2016 1:06 PM	File folder			
Downloads		5/10/2016 12:11 PM	Adobe Acrobat D	82 KB		
🗐 Recent Places	901B0E665B70_Calibration_2016-05-10_19	5/10/2016 12:16 PM	Adobe Acrobat D	78 KB		
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	12_901B0E665B70_Calibration_2016-05-12_09	5/12/2016 9:48 AM	Adobe Acrobat D	300 KB		
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	12.10 12.10 12.10 12.10	5/12/2016 10:18 AM	Adobe Acrobat D	91 KB		
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	🔁 901B0E665B70_Spectral Mass Calibration	5/11/2016 2:36 PM	Adobe Acrobat D	100 KB		



#### Calibration: Reports (Tune v2.6)

 File
 Windows
 Reports
 Help

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🕁 Calibrate

#### Tune v 2.8 SP1

- Evaluation reports are now available
- Structure of drop-down menu under reports sorted and cleaned up to enable quick finding of reports

#### **Evaluation Report**

 Date of evaluation:
 2016-08-24 11:01:50

 Windows user name:
 WINDOWS-3FU3DB0\Thermo(Thermo)

 Q. Exactive HF
 2.8-268801/2.8.0.2688

 Instrument identification:
 Exactive Series slot #1 (901B0E49DA3D)

#### Type of evaluation: C-Trap Charge Detection Scaling

Scan settings: scan range: 150.0 - 2000.0 resolution: 60000 AGC target : 1000000.0 micro scans : 1




• To avoid persistent contamination with Ultramark or Caffeine in the Ion Max API source



#### **Reminder:**

- Use dedicated syringe and infusion line for calibration
- Use dedicated ion transfer capillary for calibration
- After calibrations, flush lines with LC/MS grade methanol



## Tuning and Calibration: Take Home Message

## • Key Considerations:

- Successful evaluation/calibration must be performed under stable spray conditions using HESI-II probe
  - Ensure <10% TIC Variation for >100scans located in instrument status panel via tune page
- Use fresh calmix for all calibrations
  - Pierce LTQ Velos ESI Positive Ion Calibration Solution
    - Catalog no. 88323
  - Pierce Negative Ion Calibration Solution
    - Catalog no. 88324



### Calibration Schedule:

- Weekly
  - Perform MS Mass Calibration
- Monthly



- Run Ion Transfer calibration
- Run Base Evaluation, if failed run Analyzer Accuracy calibration and eFT calibration
- Run MS Mass Calibration
- Run Isolation Evaluation, if failed, calibrate Wide Isolation and then Narrow Isolation
- Following vent/clean → perform monthly procedure



#### Maintenance





### Maintenance Considerations



#### **Maintenance Considerations**

- Always wear clean, talc-free and lint free gloves when handling instrument components
- Always place the components on a clean, lint-free surface, such as Aluminum foil
- Never overtighten a screw or use excessive force
- Ensure tools are clean; dirty tools can contaminate your system
- Never insert a test probe into the sockets of female cable connectors on Printed Circuit Boards (PCBs)



## **Recommended Maintenance Schedule**

## Daily

- Check the vacuum gauge pressures and make sure the vacuum system is operational. Typical values are:
  - Fore Vacuum: <2 mbar
  - High Vacuum: Turned off
  - Ultra High Vacuum : < 5e-10 mbar
  - Source and UHV TMP Speed: 100%
- If bottled nitrogen gas is being used, check nitrogen gas pressure. Recommended value for the Q Exactive MS series is 800  $\pm$  30 kPa (8  $\pm$  0.3 bar, 116  $\pm$  4 psi)
- Make sure the solvent waste bottle for drain tube is empty
- Flush (clean) sample transfer line
- Perform MS Mass Calibration (optional)



## **Recommended Maintenance Schedule**

## • Weekly

• Check the forepump oil level and check for condensation in the pump

NOTE: An increase of the pump oil level is a sign of vapor condensation in the pump

- Fill the forepump with oil as needed
- Ballast the forepump to drain oil back into the pump (~30 min)
- Clean the ion transfer capillary
- Perform MS Mass Calibration
- Monthly
  - Evaluate Q Exactive MS series calibrations and recalibrate only if the evaluation(s) fails



## **Recommended Maintenance Schedule**

## Quarterly

- Back up the data folder and master calibration file
- Check relevant websites for available software updates
  - Thermo Fisher Scientific BRIMS portal for Omics software <a href="https://portal.thermo-brims.com/">https://portal.thermo-brims.com/</a>
  - Thermo Fisher Scientific Customer Download Site -<u>https://thermo.flexnetoperations.com/control/thmo/login</u>

#### Yearly

- Schedule Preventative Maintenance (PM)
- Leak Check Gas Lines
- Defragment computer hard drive
- Replace the forepump oil



### Removing the Sweep Cone



CAUTION: During operation of the mass spectrometer, sweep cone might reach temperatures of up to 450° <sup>C</sup>.

Let the spray cone cool down for at least 60 minutes before removing.

Remove the ion sweep cone by grasping its outer ridges and pulling the cone straight off of the API cone seal.

Clean the ion sweep cone by wiping the inside and outside a lint-free tissue soaked in LC/MS grade methanol.

Remove and inspect the O-ring that is seated in the spray cone under the entrance end of the ion transfer capillary and clean with LC/MS grade methanol or replace if necessary.



### Removing the Sweep Cone



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Remove and inspect the O-ring that is seated in the spray cone under the entrance end of the ion transfer capillary and clean with LC/MS grade methanol or replace if necessary.



## Removing Ion Transfer Capillary



#### Turn off flow from LC.

Turn off Capillary temperature and allow Ion Transfer capillary to cool (temperature needs to be below 200°C).

Remove source housing from the front of the mass spectrometer.

Remove the Ion Sweep Cone.

Remove Ion Transfer Capillary by turning it counterclockwise with the custom removal tool.



• Click here to see <u>video</u>



# Cleaning Ion Transfer Capillary





# Cleaning Ion Transfer Capillary





## **Cleaning Ion Transfer Capillary**



Sonicate the Ion Transfer Capillary for 30 minutes in 50:50 solution of methanol/water containing 20% formic acid.

Rinse the Ion Transfer Capillary thoroughly with LC/MS grade water.

Sonicate the Ion Transfer Capillary in LC/MS grade water for 15 minutes.

Rinse the Ion Transfer Capillary with LC/MS grade methanol.

Sonicate the Ion Transfer Capillary in LC/MS grade methanol for 15 minutes.

Blow the Ion Transfer Capillary dry with a stream of nitrogen gas.



## In Case of Extreme Contamination



Add a cleaning step that uses Liquinox<sup>™</sup> to the standard procedure as follows:

Sonicate the Ion Transfer Capillary overnight in a 10% Liquinox<sup>™</sup> solution in LC/MS grade water.

Rinse the Ion Transfer Capillary, forcing a strong stream of LC/MS grade water through the orifice for 2 minutes.

Sonicate the Ion Transfer Capillary for 30 minutes in LC/MS grade water.

Continue with standard procedure (see previous slide).



## Reinstalling Ion Transfer Capillary



NOTICE: Use caution when reinstalling the Ion Transfer Capillary.

Ensure that everything is properly aligned to prevent stripping the threads on the Ion Transfer Capillary.

Insert the Ion Transfer Capillary into the heater block and rotate the Ion Transfer Capillary as you insert it.

After it is inserted, turn the Ion Transfer Capillary clockwise until it is finger tight.



## Reinstalling Ion Transfer Capillary



NOTICE: Use caution when reinstalling the Ion Transfer Capillary.

Ensure that everything is properly aligned to prevent stripping the threads on the Ion Transfer Capillary.

Insert the Ion Transfer Capillary into the heater block and rotate the Ion Transfer Capillary as you insert it.

After it is inserted, turn the Ion Transfer Capillary clockwise until it is finger tight.

Align the gas inlet on the Ion Sweep Cone with the sweep gas supply port on the ion source mount. Firmly press the Ion Sweep Cone onto the ion source mount.



#### Instrument Shutdown Procedure



In the Tune software window, click the On/Standby button to put the instrument in the Off condition.

Place the electronics service switch (located on the power panel) in the Service Mode position.

Put the main power circuit breaker switch of the mass spectrometer in the Off position.

NOTICE: An instrument that is shut down still consumes nitrogen because the vent valve is connected to the nitrogen supply of the laboratory. Keeping on the nitrogen flow prevents humidity from contaminating the vacuum system of the mass spectrometer.



## Removing the Ion Source Interface



- **CAUTION:** The ion source interface can become hot enough to cause severe burns when the mass spectrometer is in operation. Do not touch the ion source interface immediately after removing the ion source housing. Allow the ion source interface to cool to room temperature before removing it from the mass spectrometer.
- Use the tool to lever out the interface assembly.
- Grasp the ridges on either side of the ion source interface and firmly pull the assembly straight out.



• Click here to see <u>video</u>



## **Removing S-Lens**



Prepare a clean work surface by covering the area with lint-free paper or Aluminum foil.

Put on a new pair of lint- and powder-free gloves.

Loosen and extend the two thumbscrews that secure the S-lens to the ion source interface cage and the exit lens to the S-lens.

Remove the exit lens from the S-lens and place it on a clean, lint-free surface.

Grasp the two thumbscrews and carefully pull the S-lens straight out of the ion source interface cage and place it on a clean, lint-free surface.



## **Cleaning the S-Lens**



#### DO NOT DISASSEMBLE THE S-LENS!

NOTICE: Do not clean the exit lens or S-lens with abrasives, acidic or caustic substances, or detergents not stated.

Clean the S-lens by sonicating the unit for 10– 15 minutes in a 50:50 solution of LCMS-grade methanol and water or a 1% solution of Liquinox in LCMS-grade water.

Rinse the components thoroughly with LCMSgrade water, then rinse the exit lens and Slens with fresh LCMS-grade methanol.

Blow dry the exit lens and S-lens with oil-free nitrogen gas.



## Cleaning the S-Lens



Ensure that all solvent has evaporated from the components before reassembly.

Inspect the components under magnification for any lint or particulates.

NOTICE: Inspect the orifices to confirm that no lint or particulates are present in the bore of the orifices.

Use tweezers or a similar tool to remove the lint or particulate.



## Removing and Cleaning the S-Lens

• Click here to see <u>video</u>



## Reinstalling Ion Source Interface

• Click here to see <u>video</u>



## Forepump Maintenance

Maintenance job	Frequency
Check oil level	Daily
Check oil condition	Depends on process
Check gas ballast valve	Monthly
Change oil	Every 8000 h (~ one year) of operation
Replace exhaust filter	If oil mist appears at exhaust or annually
Check anti-suckback valve	Annually
01 0	

Clean fan cover

Annually



Oil level should be close to the MAX marks. If below the MIN mark, add the required amount.

Make certain the oil is not turbid.

Open the ballast valve and operate the pump for 30 minutes.

Oil change must be done with a switched off and still warm pump.

Pump switched off, open the exhaust hood, take out the filter and replace it.

If dirty, clean with an appropriate solvent.

Remove the filter and clean with stream of air.



## Purging the Oil of the Forepump



Turn off the flow of liquid from the LC (or other sample introduction device to the API source).

#### Put the MS into Standby mode.

Open the gas ballast valve on the forepump.

Allow the pump to run for 30 minutes with the gas ballast valve open.

After 30 minutes, close the gas ballast valve.



### Instrument Startup Procedure After a Shutdown



Switch on the computer and wait until the operating system is completely loaded.

Start the Tune software. NOTICE: data system must be running before start up of instrument, otherwise the instrument will not operate.

Turn on nitrogen flow at the tank, if it is off. NOTICE: Make sure the main power circuit breaker switch is in the off position (O) and the electronics service switch is in the Service Mode position.

Place the main power circuit breaker switch in the On (|) position to start up the forepump and turbomolecular pumps (TMPs); all LEDs on the front panel will remain off.



#### Instrument Startup Procedure After a Shutdown



Within a short time, a significant pressure decrease must be observed. The quality of the vacuum can be estimated by the rotation speed of the TMPs.

# Allow the mass spectrometer to pump down for 5 minutes.

Place electronics service switch into Operating Mode position and continue to monitor the vacuum readings in the Tune software.

Proceed to the bake out procedure to bake out the system before data acquisition occurs.





In the Q Exactive MS Series Tune software window, click the On/Standby button to put the instrument in Off condition (*see image*).



🕺 Vacuum / Bakeout	
Vacuum / Bakeout	
Fore vacuum (mbar) 1.46E+00 O UHV (mbar) 1.77E-10 O	
Bakeout time (h) 12.0 - Enter standby after Bakeout 🗹	
0.0 % Bake out Stop Help	

In the Q Exactive MS Series Tune software window, click the On/Standby button to put the instrument in Off condition (*see image*).

In the Tasks panel of the Q Exactive MS Series Tune software window, click to display the Vacuum / Bakeout.



Vacuum / Bakeout Vacuum / Bakeout Fore vacuum (mbar) 1.46E+00 UHV (mbar) 1.77E-10 Bakeout time (h) 12.0 Enter standby after Bakeout 0.0 %	
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Bake out Stop Help	Bake out Stop Help

In the Q Exactive MS Series Tune software window, click the On/Standby button to put the instrument in Off condition (*see image*).

In the Tasks panel of the Q Exactive MS Series Tune software window, click to display the Vacuum / Bakeout.

Enter the baking duration (in hours) into the spin box. The available range is 4 to 30 hours (v 2.7 SP1), 12 hours is recommended as the maximum.



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Click Bakeout.



Vacuum / Ba	ikeout
Fore vacuum (mb UHV (mbar)	Bakeout ar) 1.46E+00 1.77E-10 Baka aut
Bakeout time (h) Enter standby afte Bake out Sto	Bake out Bake out Bakeout will last 12.0 hours and will be followed by a cooling and stabilization time of about 3 hours. The instrument will be inaccessible for approximately 15 hours. Continue?
	Yes No

In the Q Exactive MS Series Tune software window, click the On/Standby button to put the instrument in Off condition (*see image*).

In the Tasks panel of the Q Exactive MS Series Tune software window, click to display the Vacuum / Bakeout.

Enter the baking duration (in hours) into the spin box. The available range is 4 to 30 hours (v 2.7 SP1), 12 hours is recommended as the maximum.

Click Bakeout.

A dialog box shows the duration of the baking procedure. Click Yes to confirm the message.



## Fan Filters Maintenance





## Fan Filters Maintenance



Each fan filter bracket is mounted on hinges. Insert a finger into the recess in the instrument frame and pull at the fan filter bracket to open it.

Remove each fan filter from the rear of the mass spectrometer by pulling it out of the filter bracket.


## Fan Filters Maintenance



Each fan filter bracket is mounted on hinges. Insert a finger into the recess in the instrument frame and pull at the fan filter bracket to open it.

Remove each fan filter from the rear of the mass spectrometer by pulling it out of the filter bracket.

Wash the fan filters in a solution of soap and water and rinse the fan filters with tap water.

Squeeze the water from the fan filters and allow them to air dry, then reinstall the fan filters in the fan filter brackets.





Unscrew the finger tight fitting from the sample inlet port.





# Remove the HESI-II probe from the Ion Max API source.

Unscrew the finger tight fitting from the sample inlet port.

Remove the metal needle insert from the probe using a 3 mm (7/64 in.) hex wrench or ball driver, by removing the two M4×35 mm length, socket head cap screws.





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Unscrew the finger tight fitting from the sample inlet port.

Remove the metal needle insert from the probe using a 3 mm (7/64 in.) hex wrench or ball driver, by removing the two M4×35 mm length, socket head cap screws.

Pull off the end cover of the probe and unscrew the metal needle insert, and then pull it out of the probe body.





# Remove the HESI-II probe from the Ion Max API source.

Unscrew the finger tight fitting from the sample inlet port.

Remove the metal needle insert from the probe using a 3 mm (7/64 in.) hex wrench or ball driver, by removing the two M4×35 mm length, socket head cap screws.

Pull off the end cover of the probe and unscrew the metal needle insert, and then pull it out of the probe body.

Insert a new metal needle insert into the probe body and hand tighten the adjustable union fitting until the tip of the needle insert protrudes from the probe nozzle by 1.5 mm.

## • Key Considerations:

- Practice good lab habits
  - Work bench surfaces should be clean and lint-free
  - Wear clean, talc-free and lint free gloves when handling instrument components
  - Ensure tools are clean; dirty tools can contaminate your system

### Maintenance Schedule

- Weekly
  - Check vacuum and gas tank pressures
  - Ballast forepump
  - Clean ion transfer capillary
  - Perform MS Mass Calibration
- Monthly
  - Evaluate Q Exactive MS series calibrations, only calibrate procedures that fail
- Quarterly
  - Back up data and master calibration file
  - Check for software updates (see next slide)
- Yearly
  - Schedule Preventative Maintenance (PM)
  - Replace forepump oil



## **Useful Reminders**

## Technical Support-North America

- Priority issues:
  - Call 800-532-4752, Select option 2
- Non-urgent issues:
  - Webform:
    <u>http://www.unitylabservices.com/contact.php</u>
  - Email:

US.Techsupport.Analyze@ThermoFisher.com

- Enter serial number in subject line
- Provide brief Description of Issue
- Technical Support-Europe
  - Email:

EU.techsupport.CMS@thermofisher.com

### Websites

- planetorbitrap.com
- https://portal.thermo-brims.com
- <u>www.mytracefinder.com</u>
- <u>https://appslab.thermofisher.com</u>
- Thermo Fisher Scientific Customer
  Download Site:
  <u>https://thermo.flexnetoperations.com/control/</u>
  <u>thmo/login</u>



## Thank You for Your Attention!

### HRAM iQuan 2016



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