



Improved metabolome coverage and increased confidence in unknown identification through novel automated acquisition strategy combining sequential injections and MSⁿ

Anas Kamleh, PhD

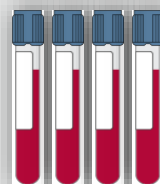
Nordic Metabolomics Society meeting, Örebro, 28/08/2018

Challenges



- Structural Diversity
- Background Interference
- Sample Limitations
 - Detection Limit
 - Dynamic Range
 - Spectral Density
- MS² is Insufficient
- MSⁿ is Difficult to Setup and Analyze

Applications



Metabolomics



Metabolite, Degradant and Impurity Identification



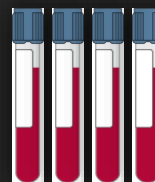
Extractables and Leachables Identification

Challenges



- Structural Diversity
- Background Interference
- Sample Limitations
 - Detection Limit
 - Dynamic Range
 - Spectral Density
- MS² is Insufficient
- MSⁿ is Difficult to Setup and Analyze

Applications



Metabolomics



Metabolite, Degradant and Impurity Identification



Extractables and Leachables Identification

Small Molecule Dedicated Mass Spectrometer on Thermo Scientific Tribrid Platform



**Thermo Scientific™ Orbitrap ID-X™
Tribid™ Mass Spectrometer System**

Improved
Instrumentation

Orbitrap
ID-X
MS

Advanced
Data Processing

Novel Data
Acquisition

Small Molecule Dedicated Mass Spectrometer on Thermo Scientific Tribrid Platform



**Thermo Scientific™ Orbitrap ID-X™
Tribrid™ Mass Spectrometer System**

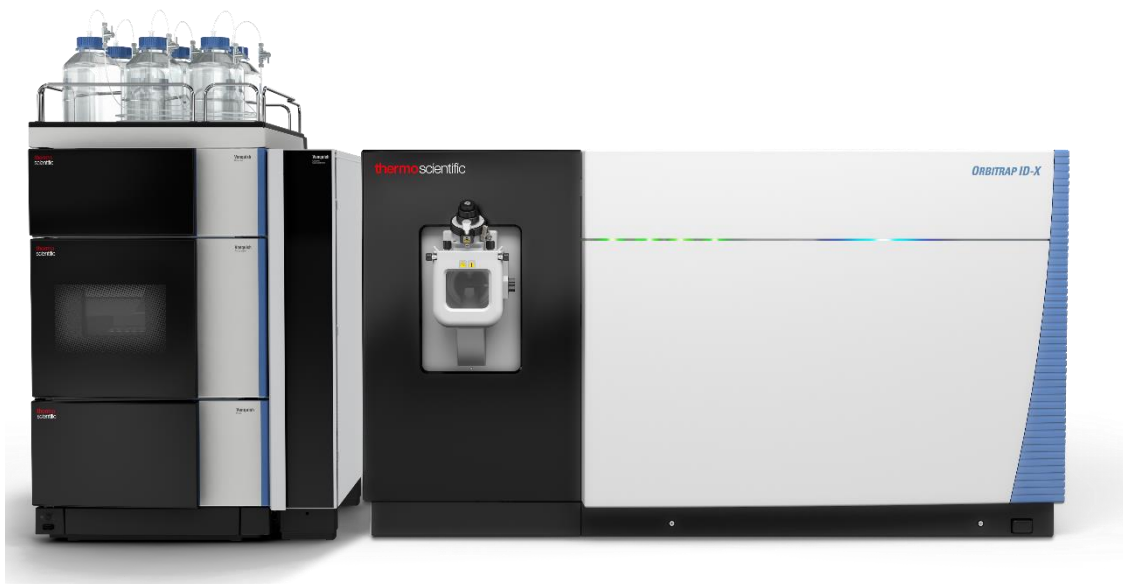
Improved
Instrumentation

Orbitrap
ID-X
MS

Advanced
Data Processing

Novel Data
Acquisition

Orbitrap ID-X Tribrid Mass Spectrometer Instrument Features



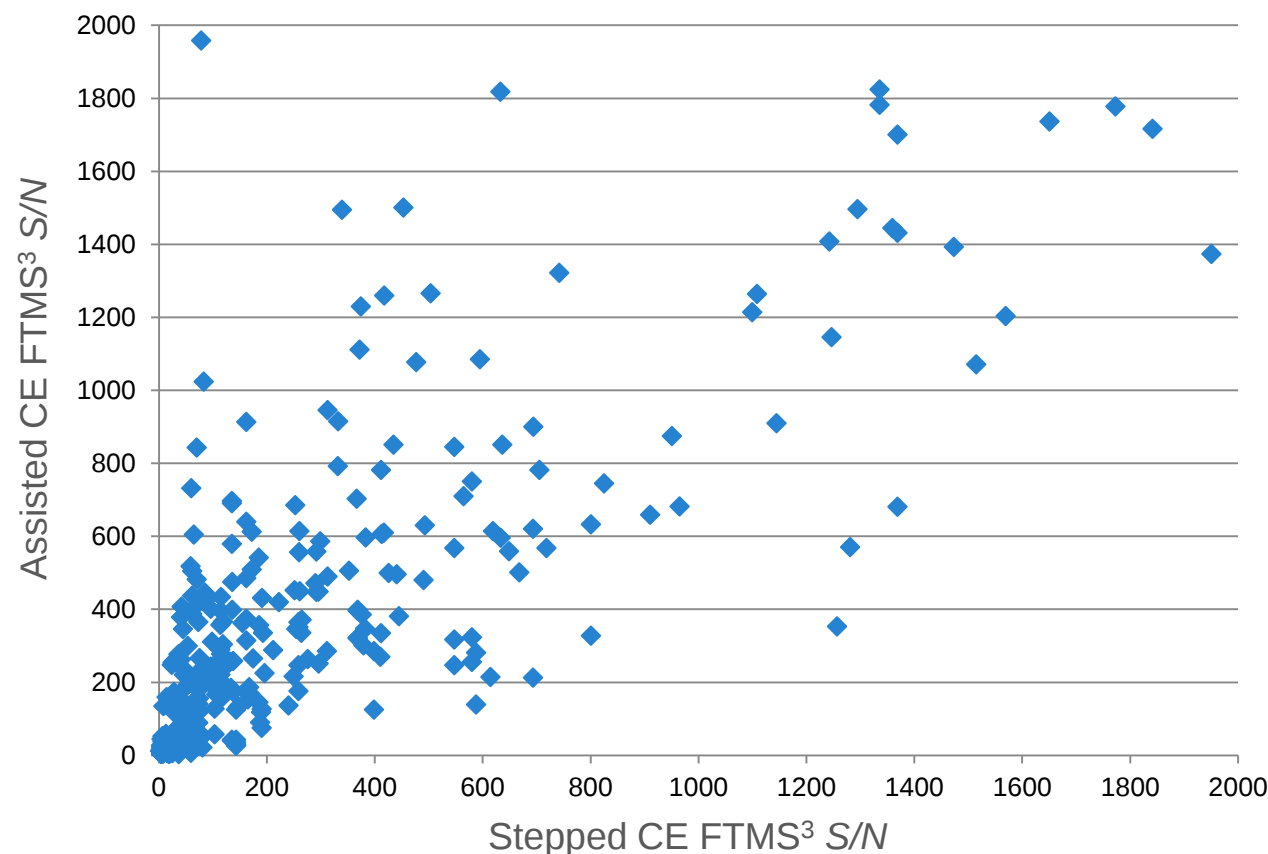
Max Resolution	500,000 at m/z 200
Scan Rate OTMS ²	30 Hz
Scan Rate ITMS ²	40 Hz
Quad Mass Selection	Precursor isolation to 0.4 amu
Ion Trap MS ⁿ	Up to MS ¹⁰
Mass Accuracy	3 ppm external, 1 ppm internal
Dissociation	CID, HCD

Instrument Improvements

- **Thermo Scientific™ OptaMax™ NG** ion source for enhanced usability and robustness
- **Streamlined calibrations** with improved mass calibration for ions with $m/z < 200$
- **User interface** and default parameters optimized for small molecule analysis
- **Expansive collection** of application specific small molecule methods
- **Assisted CE**, allowing for real-time collision energy optimization
- **Library Builder** method for the acquisition of high-quality MSⁿ spectral trees for local library generation

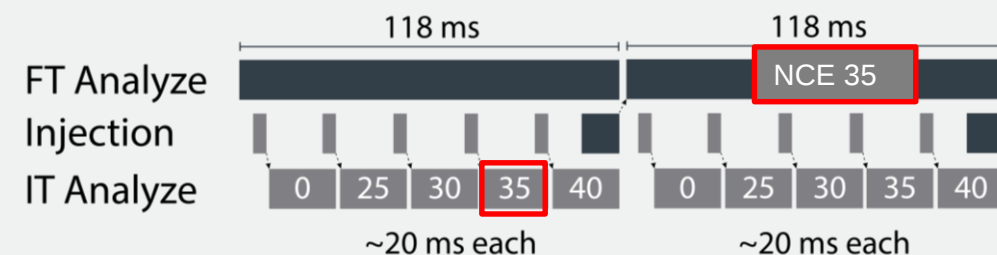
Assisted CE Allows Maximizing MSⁿ Spectral Quality

Assisted CE Improves MSⁿ S/N



Comparison of FTMS³ signal to noise for LC-MS runs with Assisted CE vs. Stepped CE shows higher number of fragment ions with increased S/N resulting from Assisted CE

Assisted CE Determination



- Instrument collects hidden ion trap scans to generate break-down curves per compound in parallel with acquisition of the preceding FTMS scan
- Optimal collision energy is ascertained based on specified precursor depletion threshold
- FTMS² analytical scan is collected using the optimal collision energy

Bailey et al. ThP 831

Small Molecule Dedicated Mass Spectrometer on Thermo Scientific Tribrid Platform



**Thermo Scientific™ Orbitrap ID-X™
Tribrid™ Mass Spectrometer System**

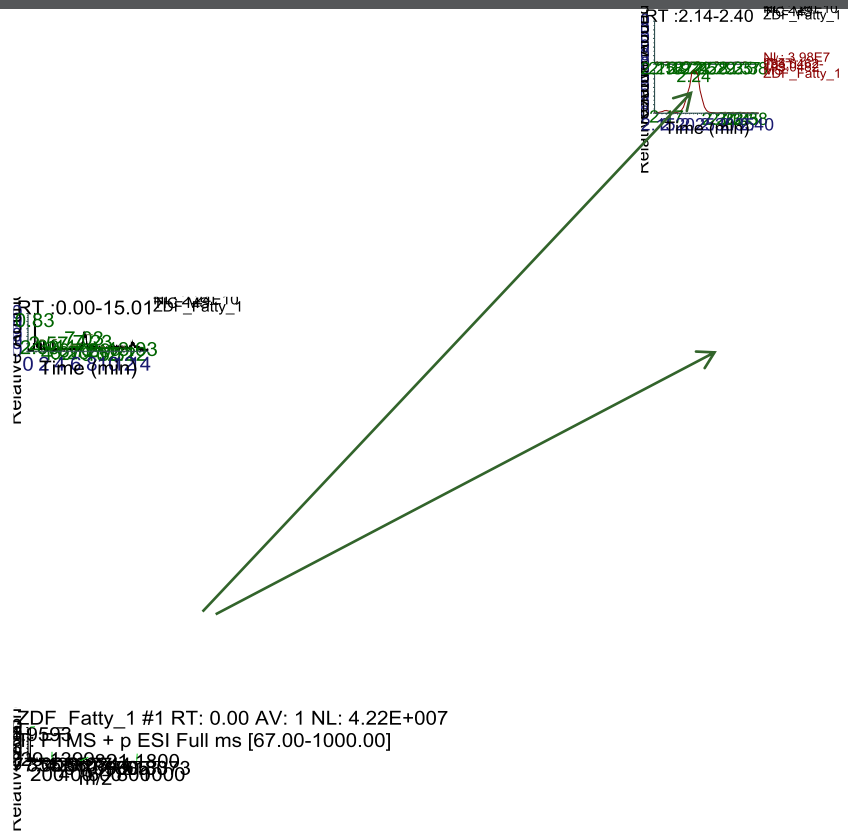
Improved
Instrumentation

Orbitrap
ID-X
MS

Advanced
Data Processing

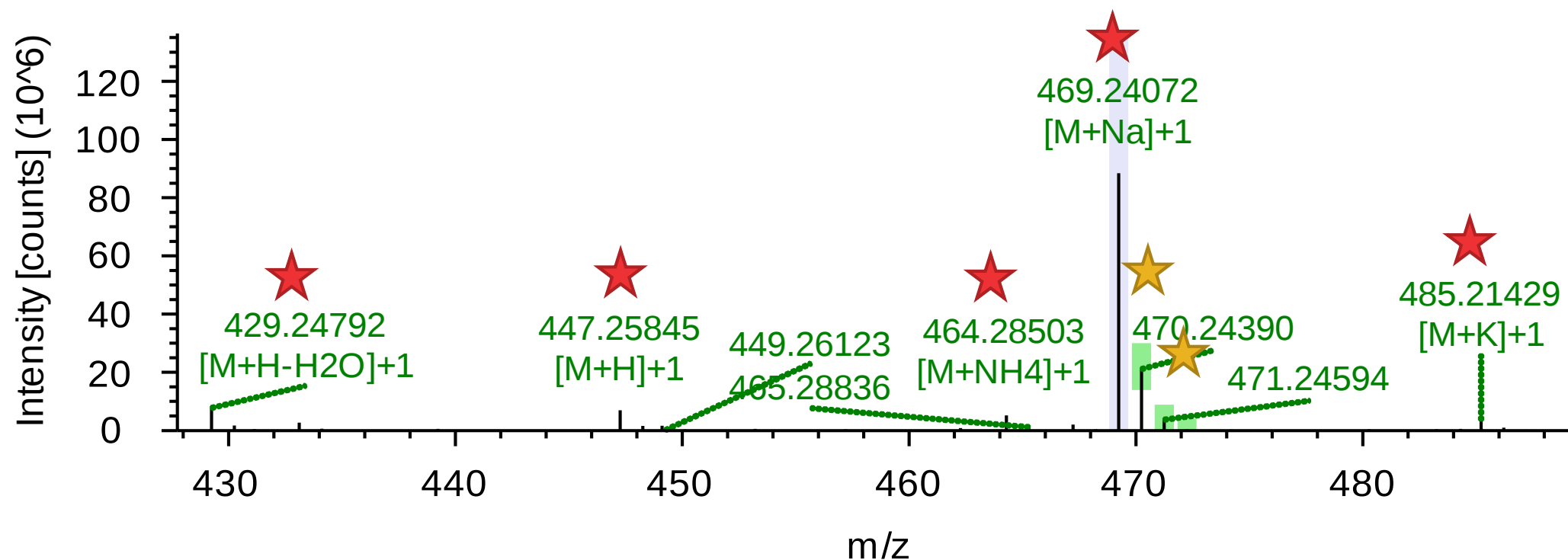
Novel Data
Acquisition

Unknown analysis of complex samples

[illegible]

Electrospray ionization produces multiple ions per compound

12162_serum_asthma_female_52_28, #1379, RT=7.317 min, FTMS (+)
C₂₂H₃₈O₉ as [M+Na]⁺1

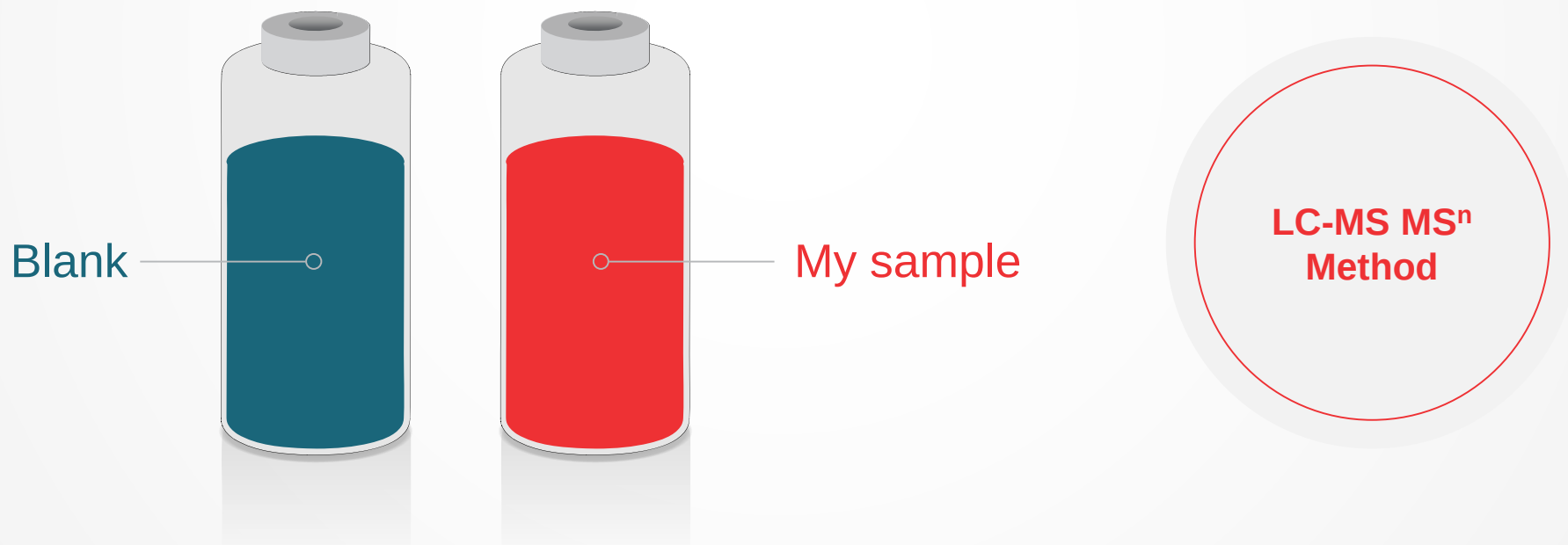


AcquireX

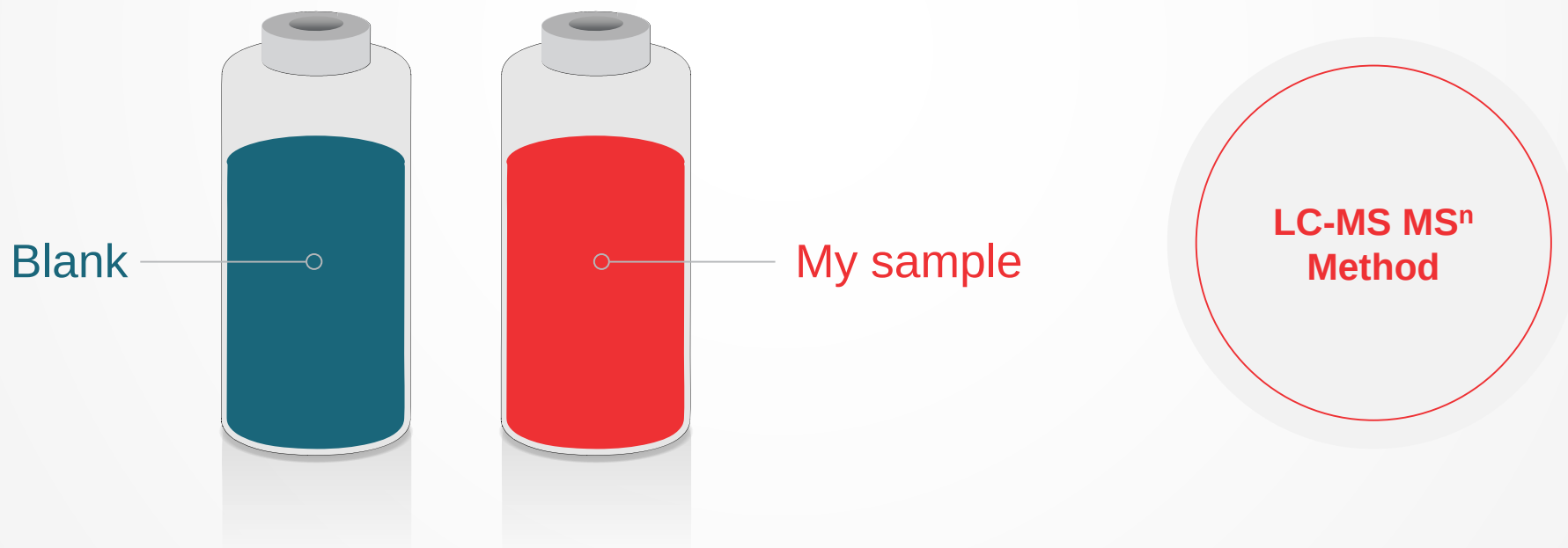
Automated acquisition workflow for comprehensive sample interrogation

This is my challenge!

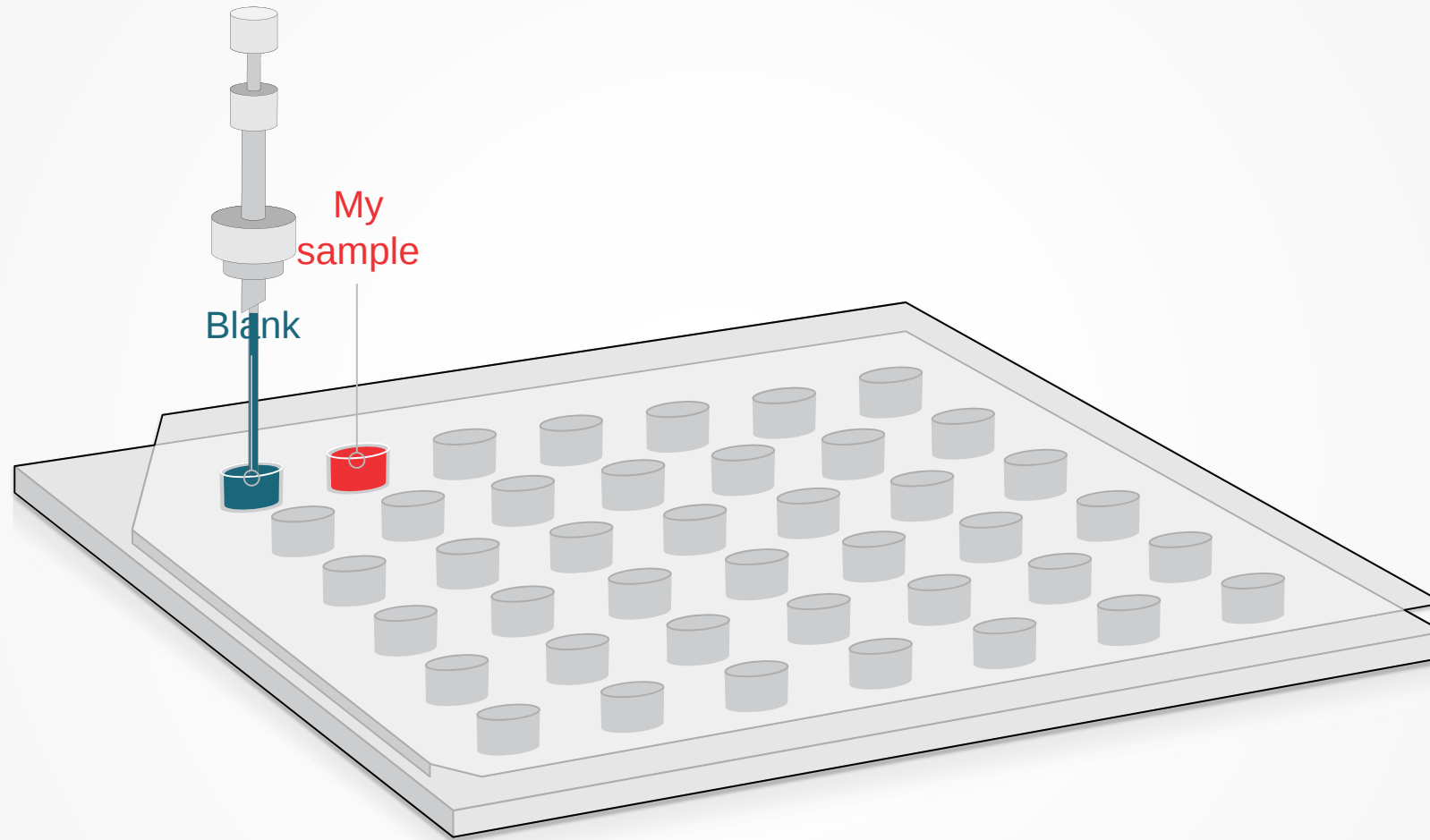
- I need to generate **high quality** MSⁿ spectra on **all** components in my sample
- My sample is too complex and not all features are fragmented
- Duty cycle is often wasted triggering on background features
- Manually excluding background and including relevant features is too laborious



**How do I analyze all of the features
in my sample using a data-dependent MSⁿ method?
In an automated manner?**



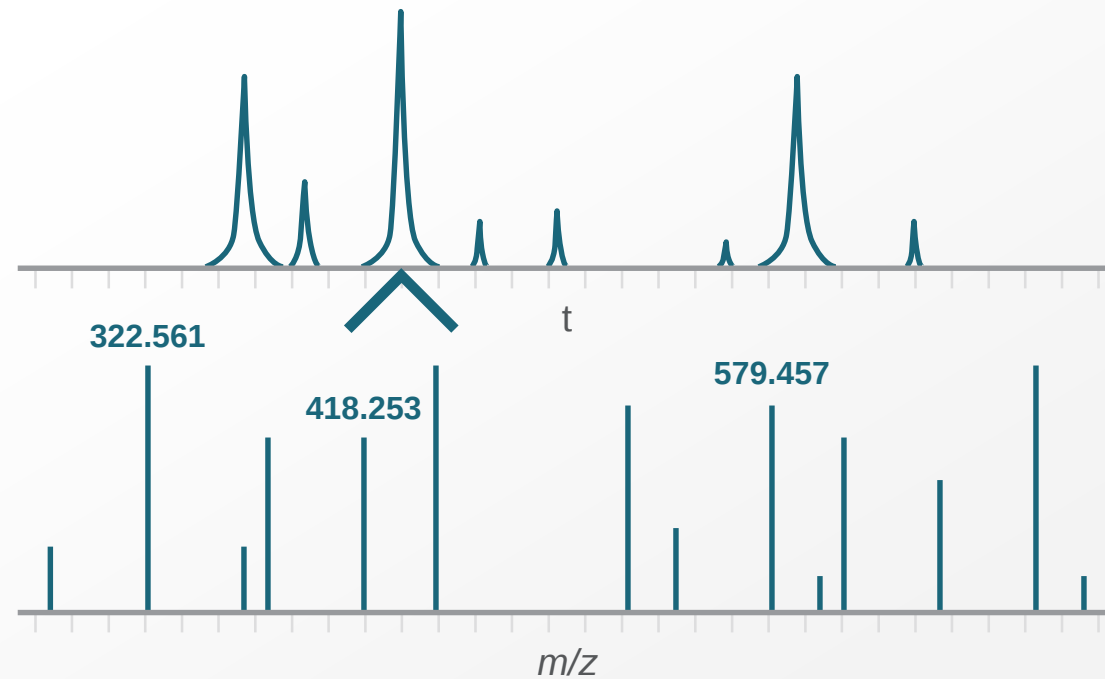
Step 1: LC-MS analysis of background (blank)



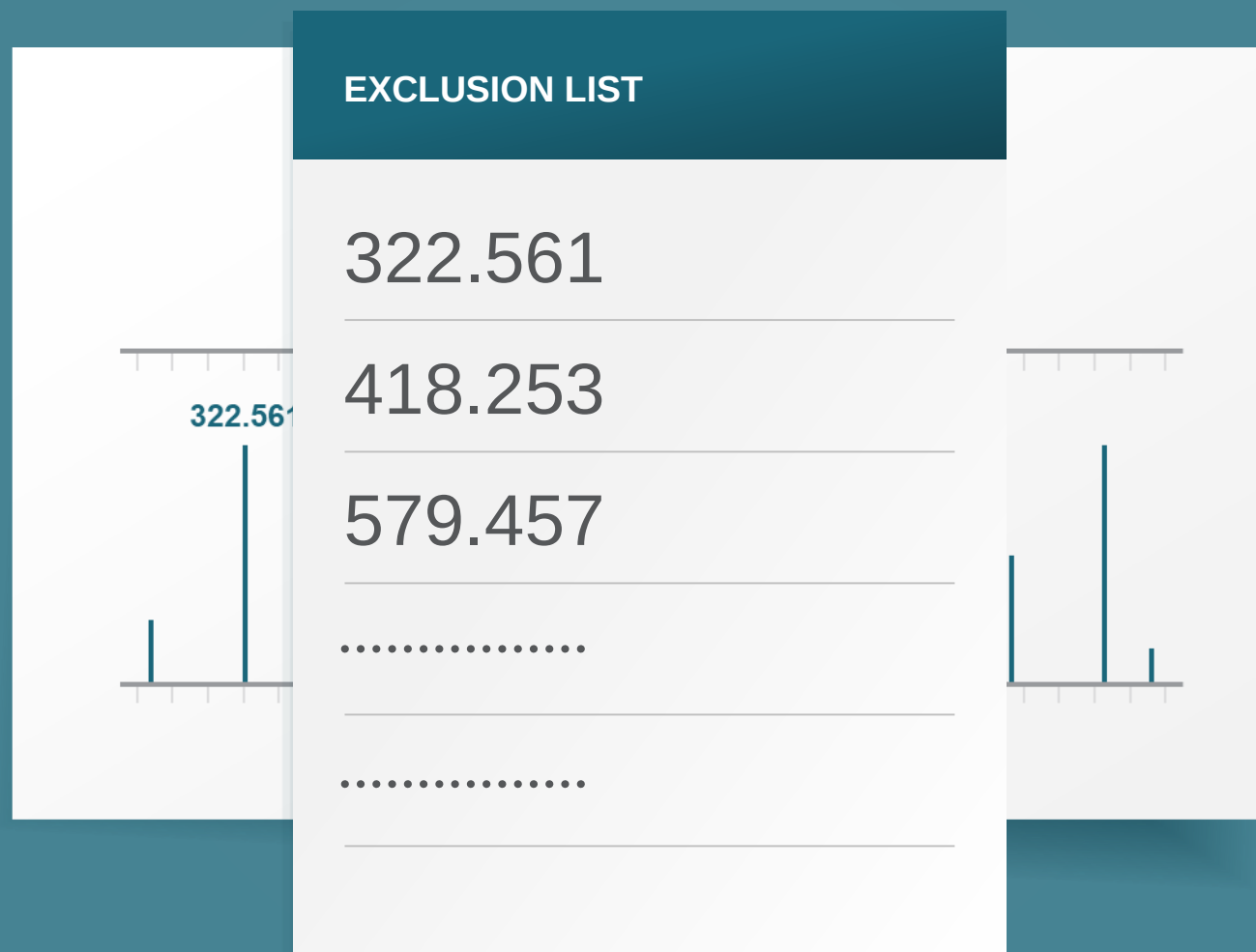
Step 1: LC-MS analysis of background (blank)



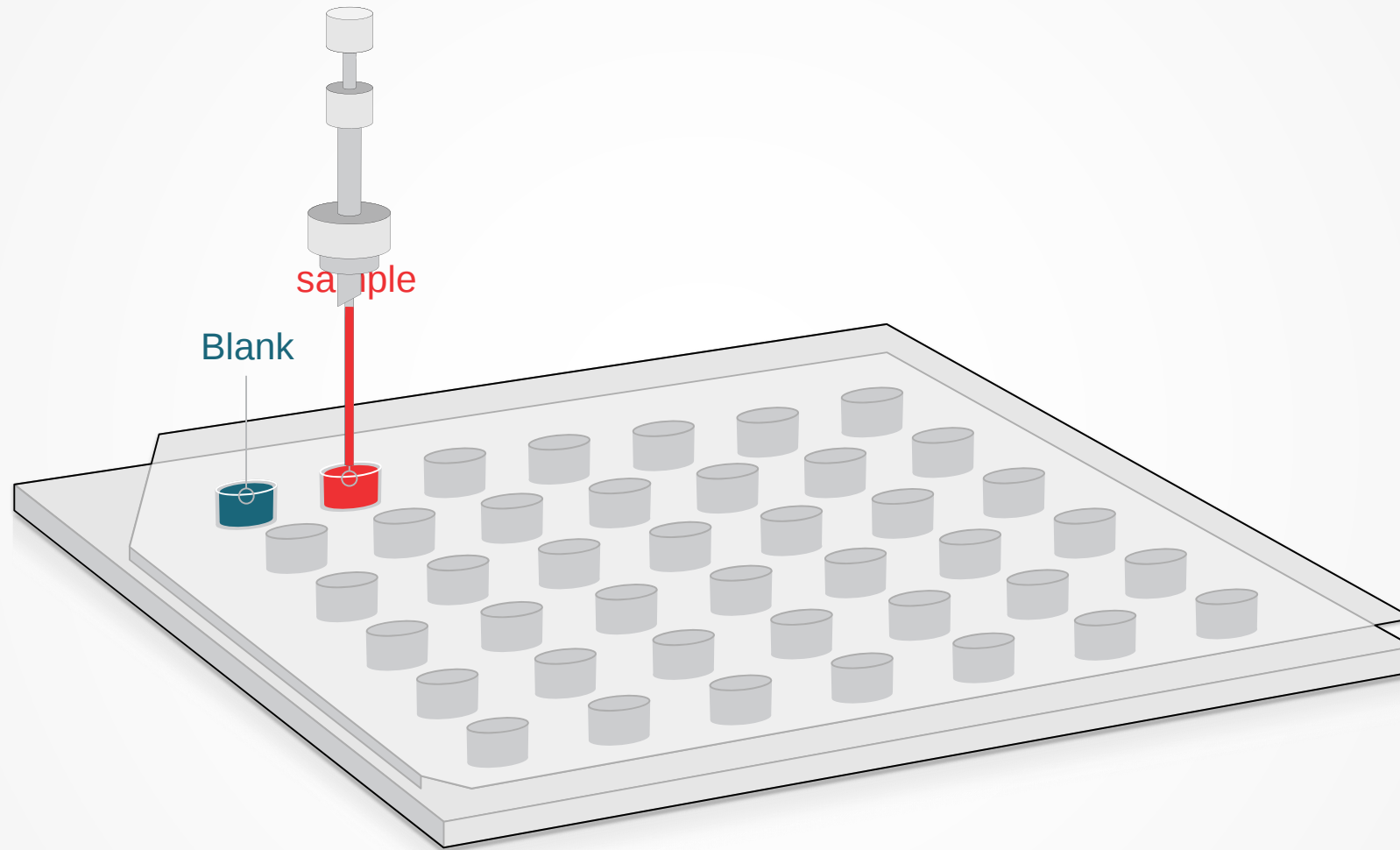
Step 1: LC-MS analysis of background (blank)



Step 2: Automatic exclusion list generation from the blank LC-MS



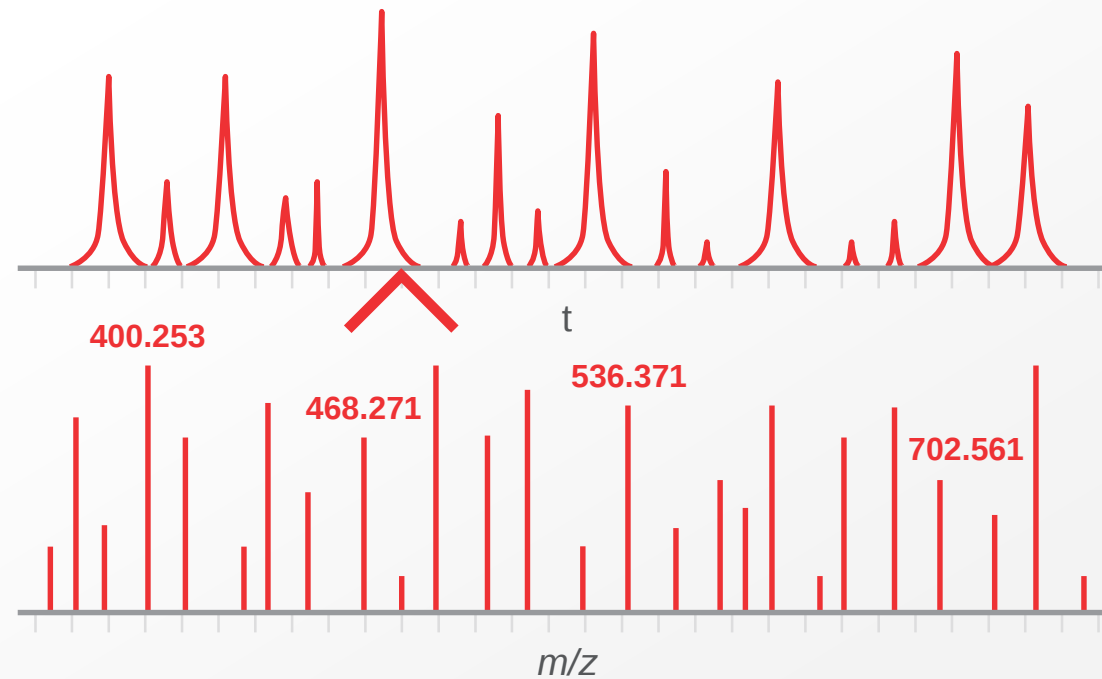
Step 3: Full scan LC-MS analysis of my sample



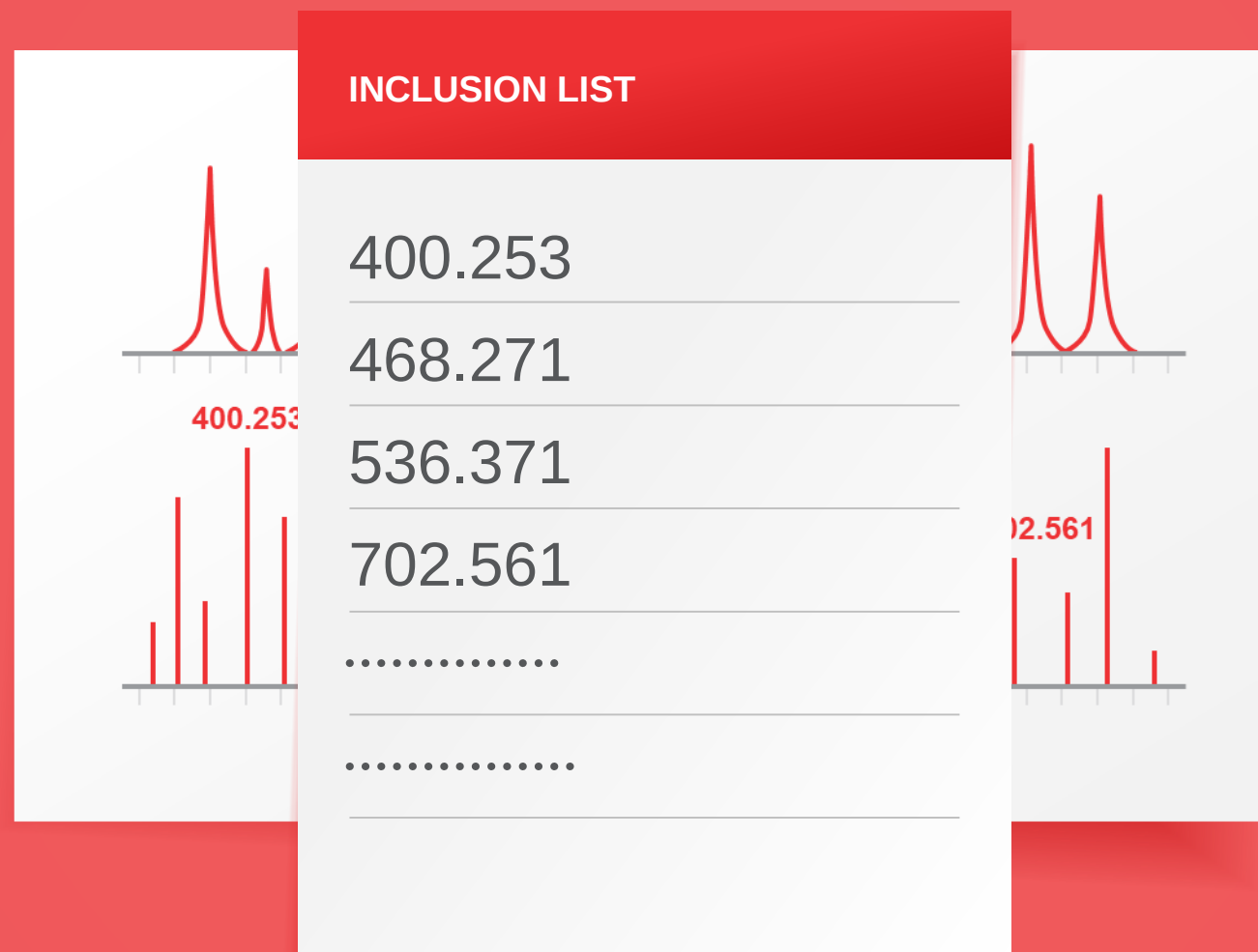
Step 3: Full scan LC-MS analysis of my sample



Step 3: Full scan LC-MS analysis of my sample



Step 4: Automatic inclusion list generation from my sample



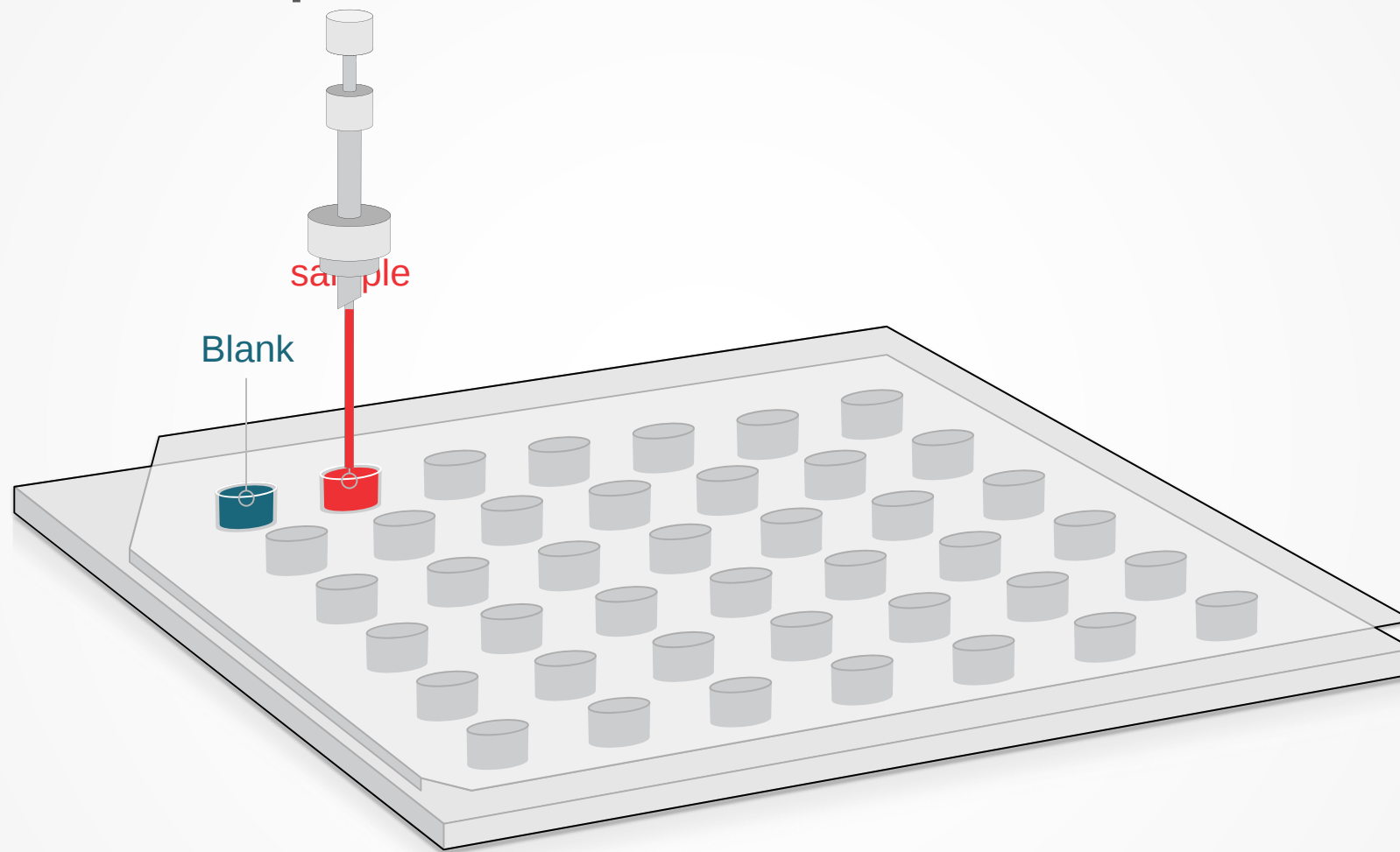
Step 5: Automatic method update with exclusion and inclusion lists

EXCLUSION LIST
322.561
418.253
579.457
.....
.....

INCLUSION LIST
400.253
468.271
536.371
702.561
.....
.....



Step 6: First automated LC-MSⁿ analysis using the updated inclusion and exclusion lists

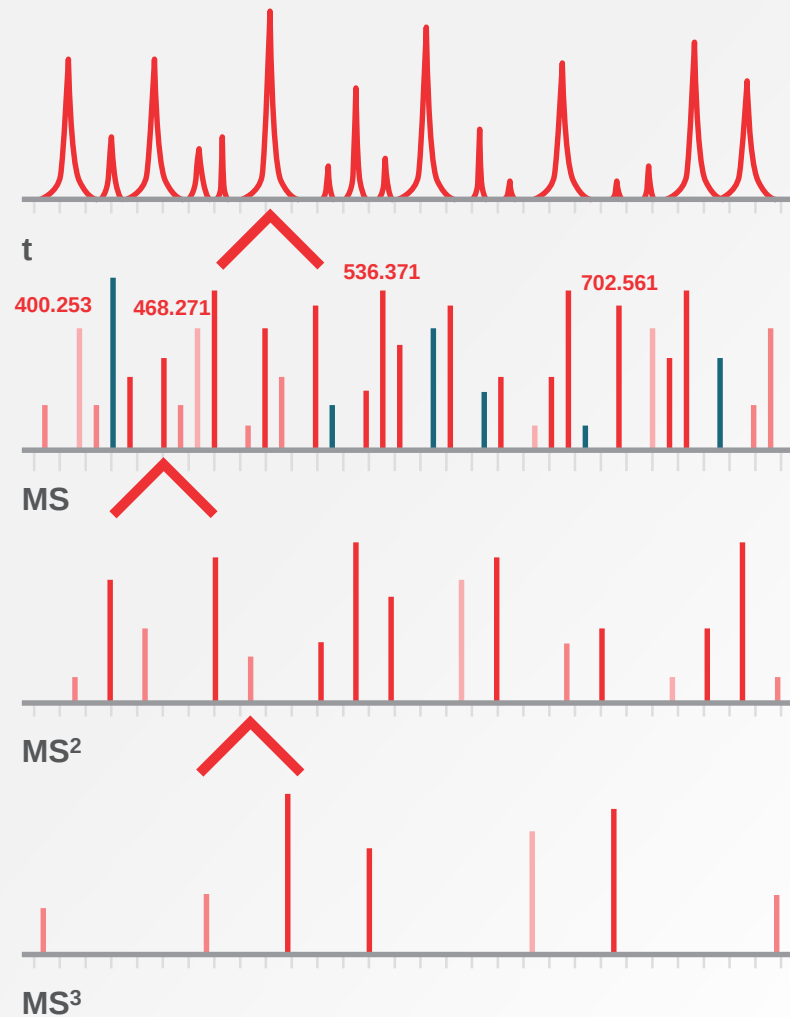


Step 6: First automated LC-MSⁿ analysis using the updated inclusion and exclusion lists



Step 7: Updating the lists automatically

Moving triggered features from inclusion to exclusion list



INCLUSION LIST

400.253

468.271

536.371

702.561

.....

EXCLUSION LIST

322.561

418.253

468.271

579.457

.....

Step 8: Updating the MS method automatically

with the current inclusion and exclusion lists

MSⁿ Method

INCLUSION LIST

400.253

.....

536.371

702.561

.....

EXCLUSION LIST

EXCLUSION LIST

322.561

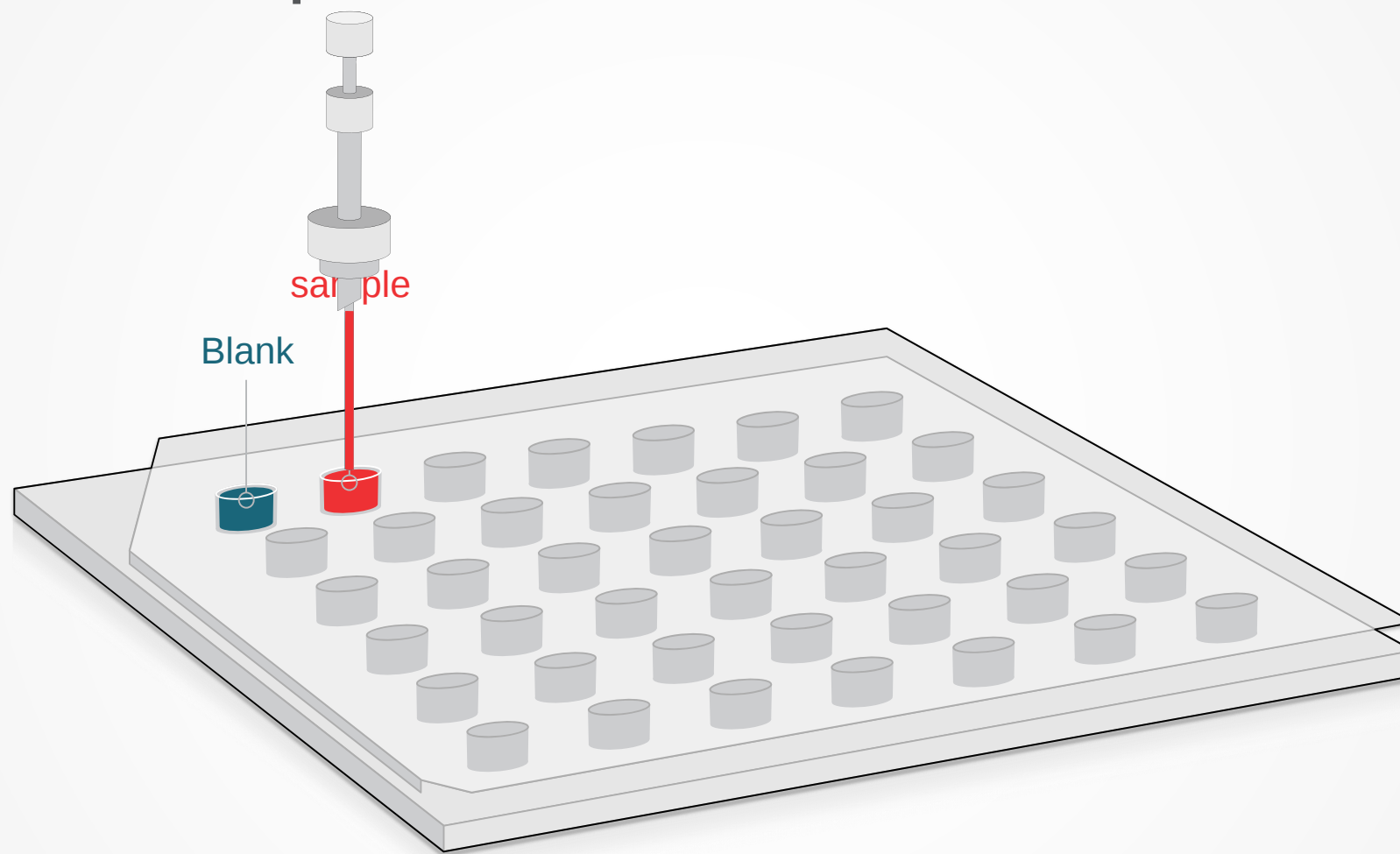
418.253

468.271

579.457

.....

Step 9: Second automated LC-MSⁿ analysis using the updated inclusion and exclusion lists

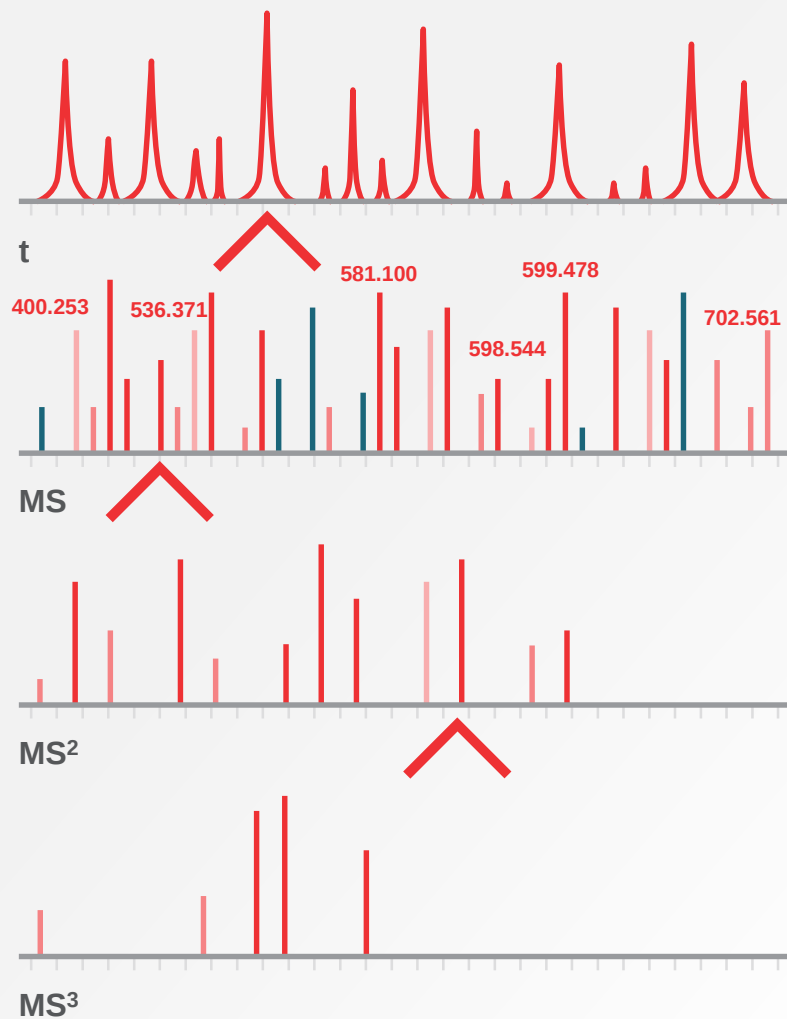


Step 9: Second automated LC-MSⁿ analysis using the updated inclusion and exclusion lists



Step 10: Updating the lists automatically

Moving newly triggered features from inclusion to exclusion list



INCLUSION LIST

400.253

536.371

581.100

598.544

599.478

702.561

EXCLUSION LIST

322.561

418.253

468.271

536.371

579.457

Repeat steps 9-10 until all of the features on the inclusion list are sampled

Injection 4

INCLUSION LIST

598.544

599.478

EXCLUSION LIST

322.561

400.253

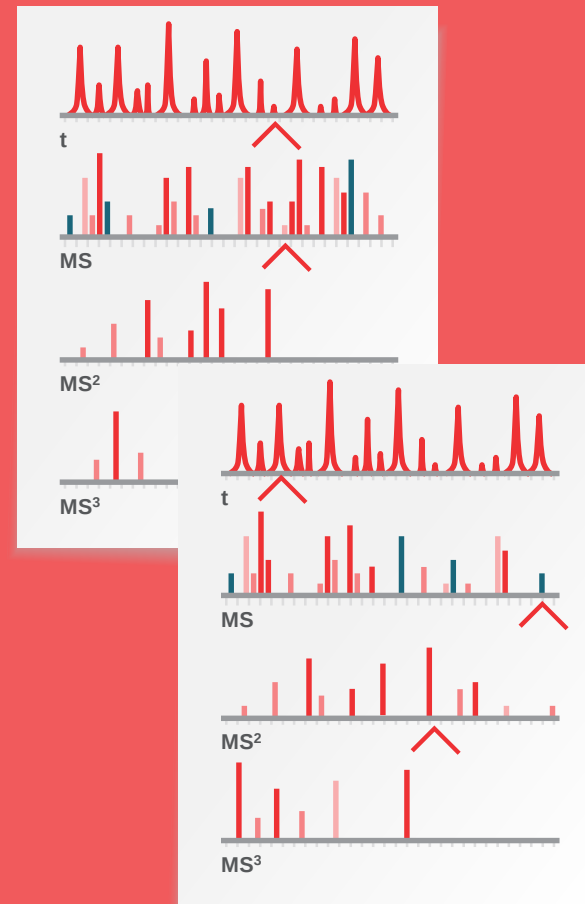
418.253

468.271

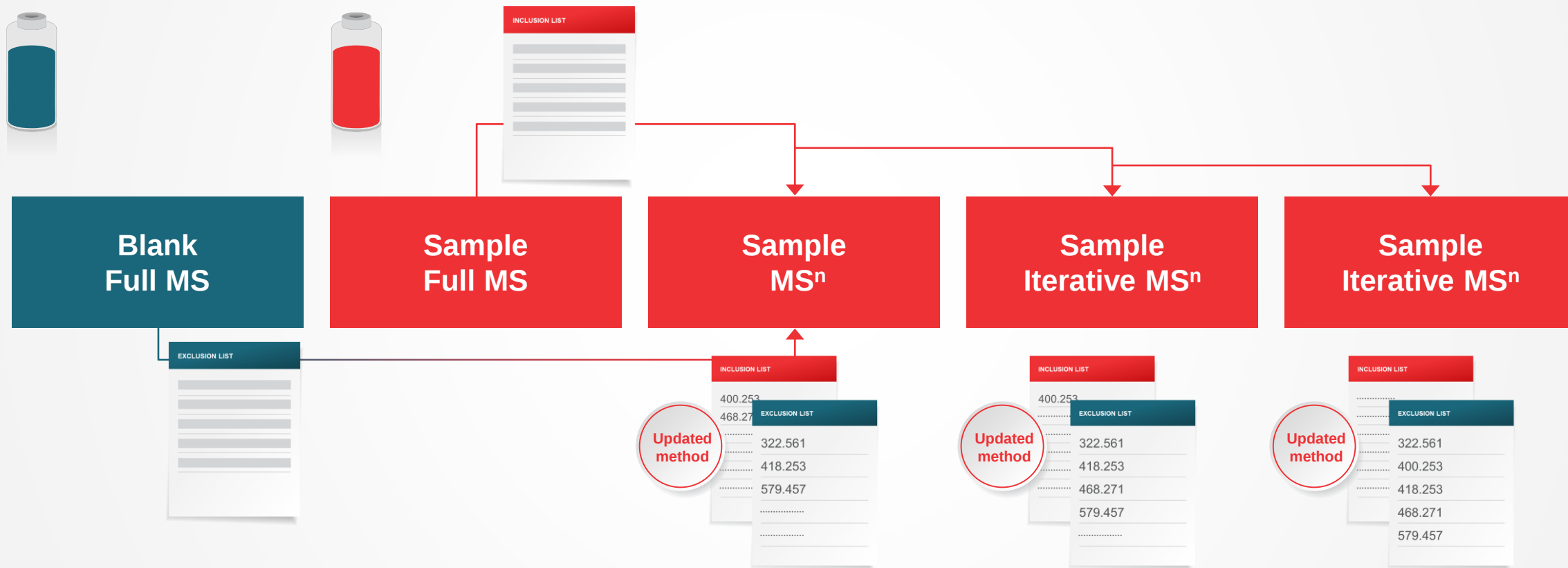
536.371

579.457

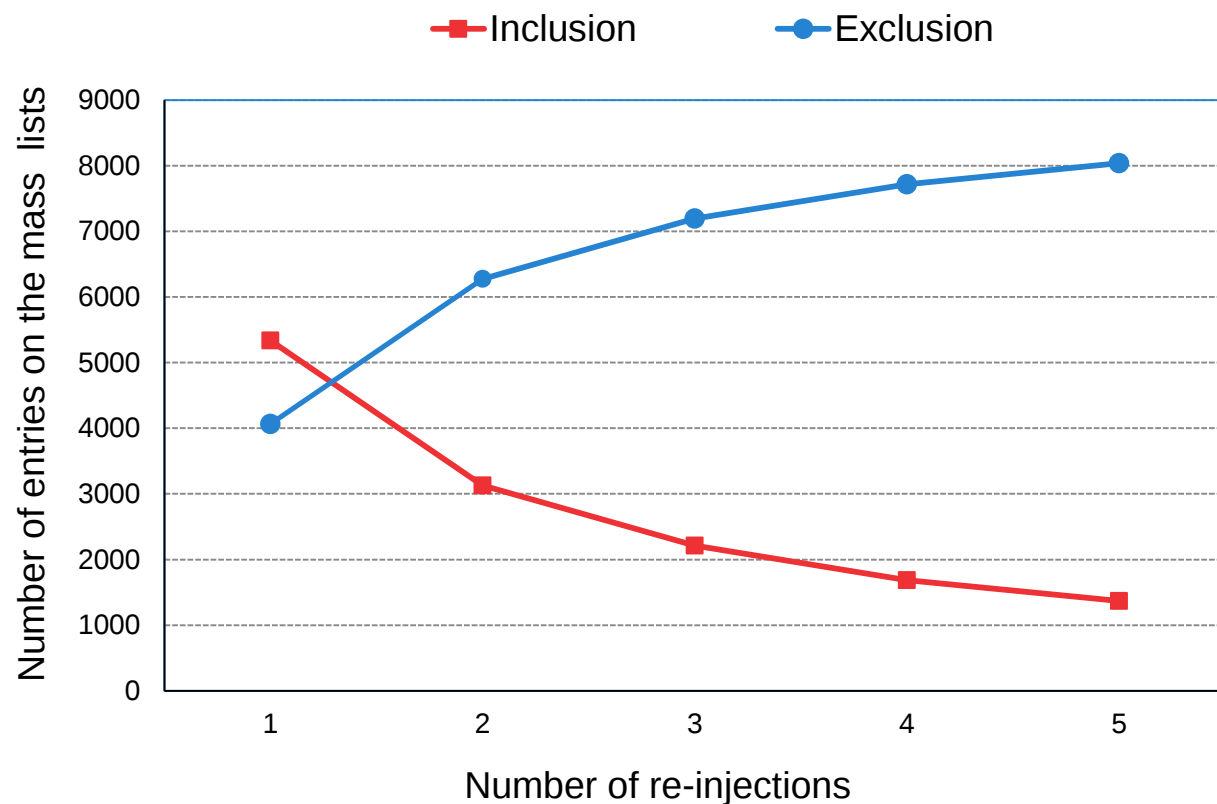
581.100



Acquire^X: Comprehensive MSⁿ analysis of all of the features in the sample



Automatically updated run-to-run inclusion/exclusion lists



LC-MS analysis of NIST SRM 1950 metabolites in frozen human plasma. The instrument automatically updates the inclusion and exclusion lists during successive iterative AcquireX re-injections, such that exclusion list continues to grow, while inclusion list declines with each additional re-injection

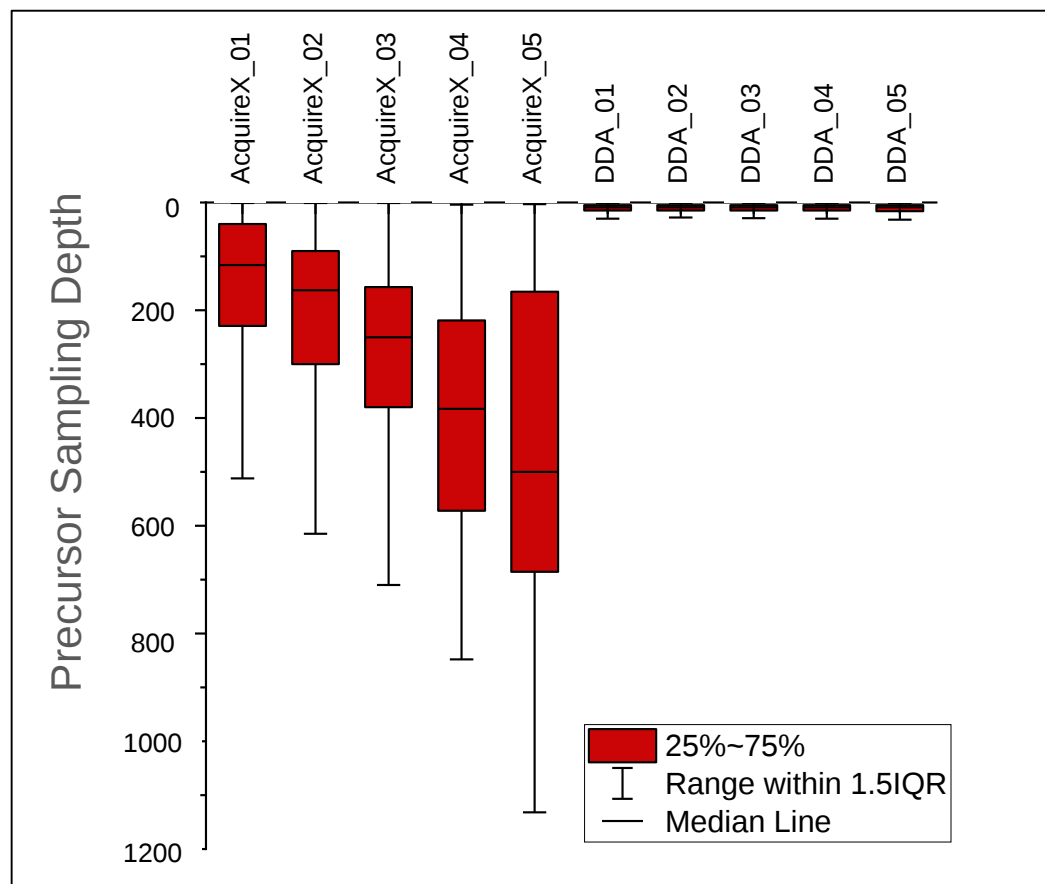


Deep Scan Analysis

- 2 µl of NIST SRM 1950 plasma extract was injected using 2.1mm x 150 mm Hypersil Gold column
- Automated generation of exclusion list >4000 features
- Automated generation of inclusion list >5000 features
- Inclusion list size decreases with each re-injection while exclusion list size increases

AcquireX: Deeper Interrogation of Sample

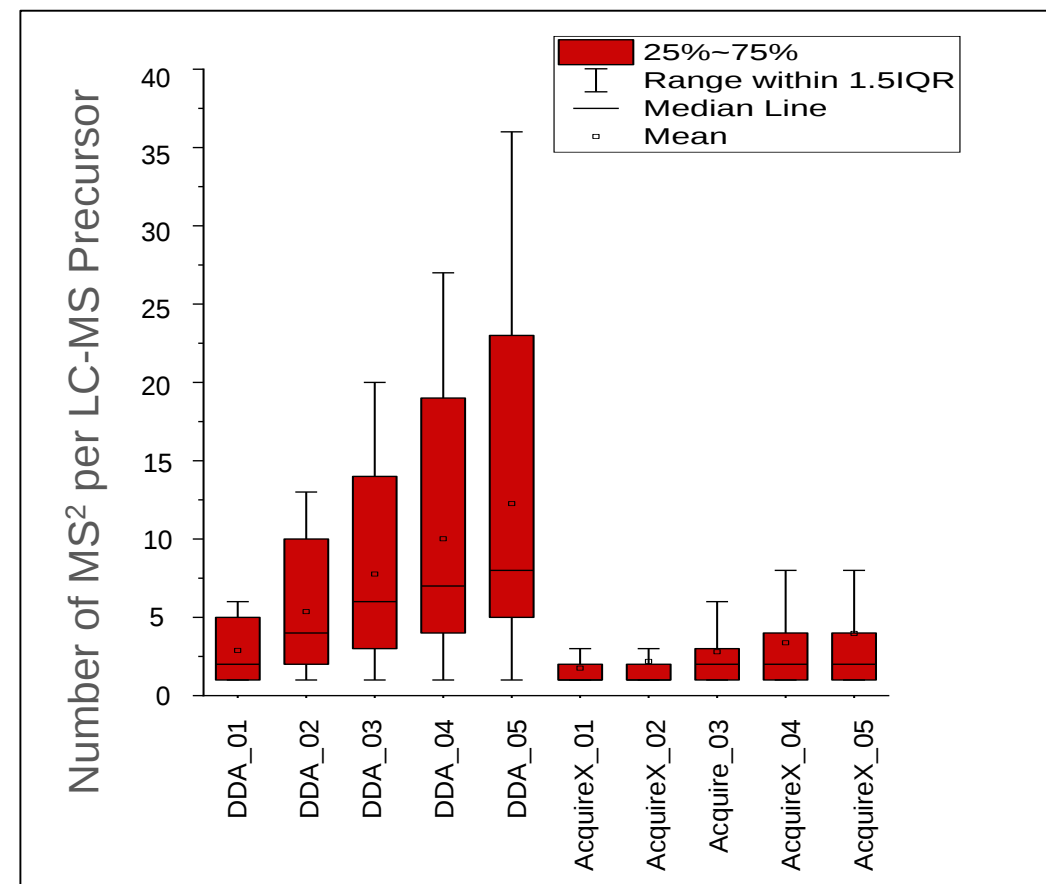
Increased Precursor Sampling Depth



Comparison of traditional DDA and AcquireX re-injections shows increased coverage with the AcquireX acquisitions

McAlister et al. MP 120

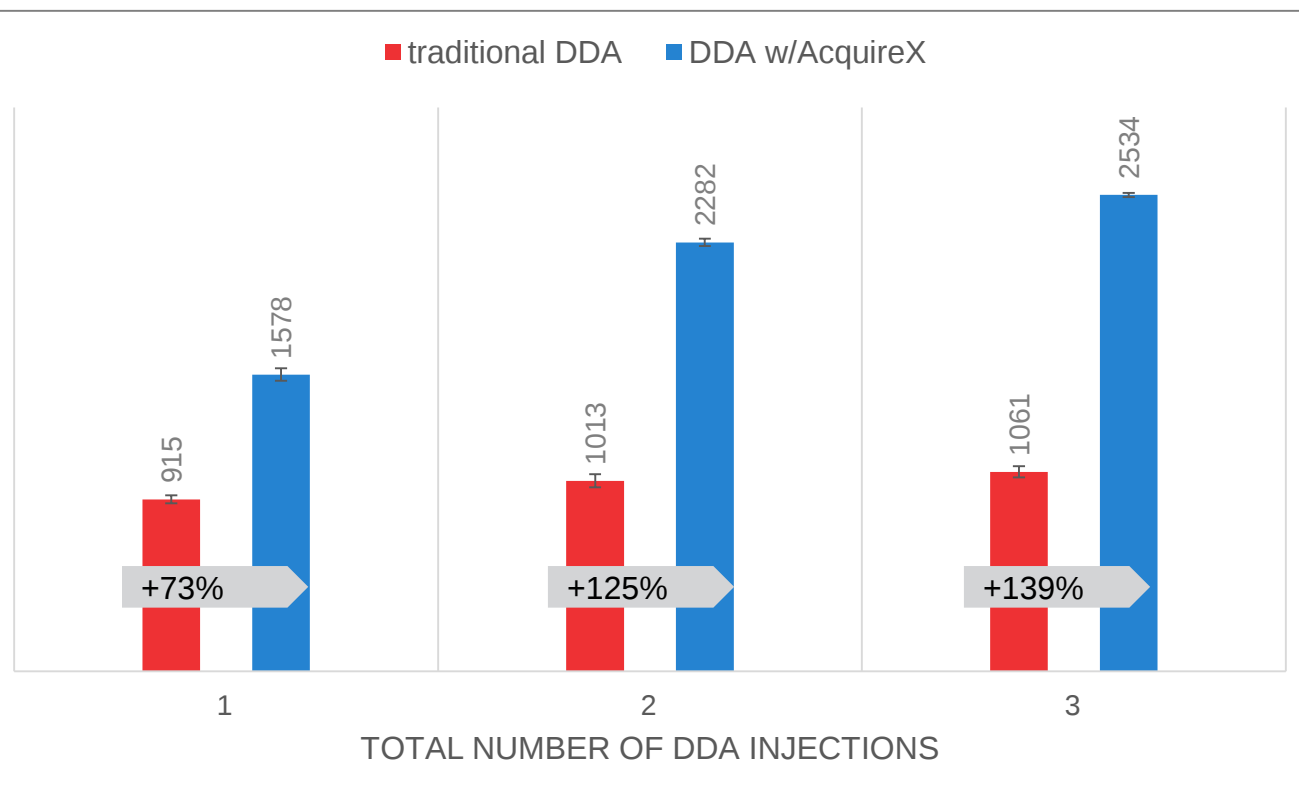
.....With Less Redundancy



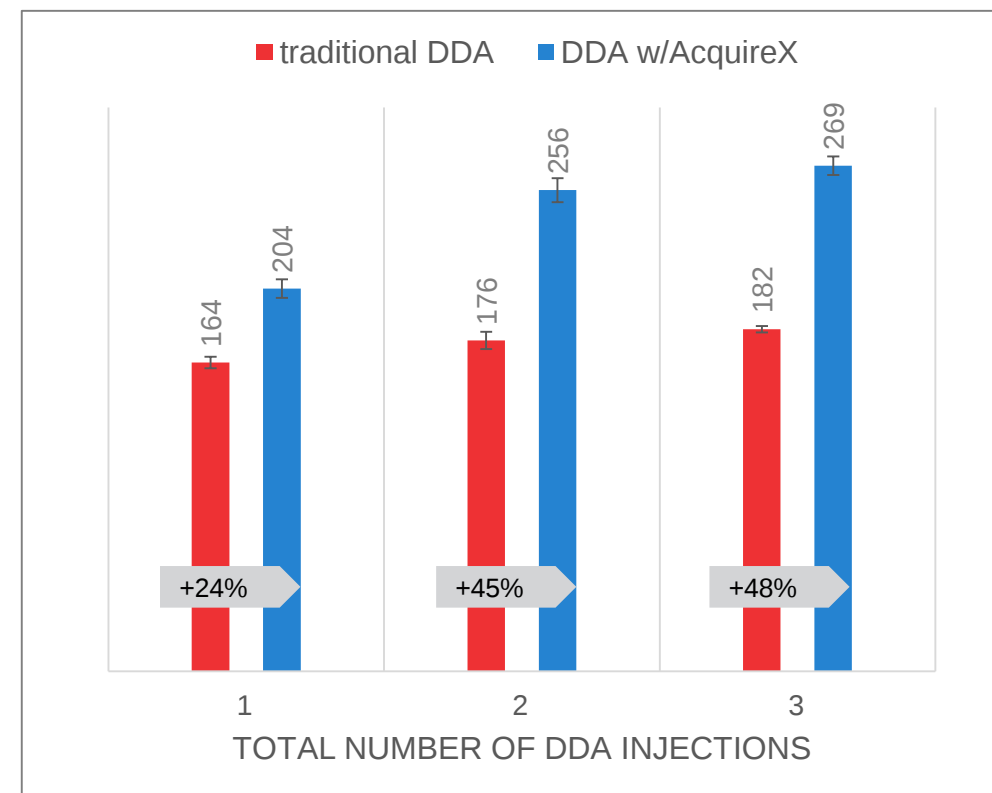
Comparison of traditional DDA and AcquireX re-injections shows less redundancy in total number of fragmentation spectra per compound during the AcquireX acquisitions

AcquireX: More Confident Identifications with Increased Productivity

Increased Number of MS features with Fragmentation Spectra



Increased Number of mzCloud Spectral Matches Using Compound Discoverer 3.0 software

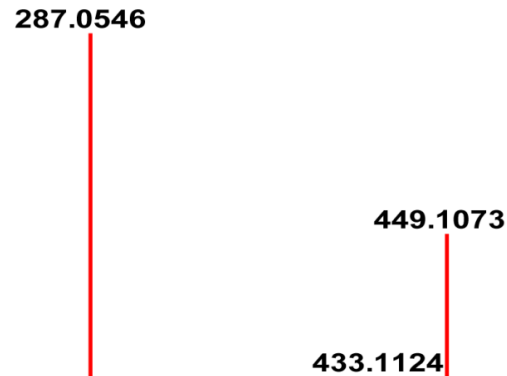


Comparison of three re-injections of the **extracted human plasma** using traditional DDA and DDA with AcquireX showing more than 2x increase in the number of compounds with fragmentation spectra (left) and almost 50% gain in confidently identified compounds (right) when DDA with AcquireX is used

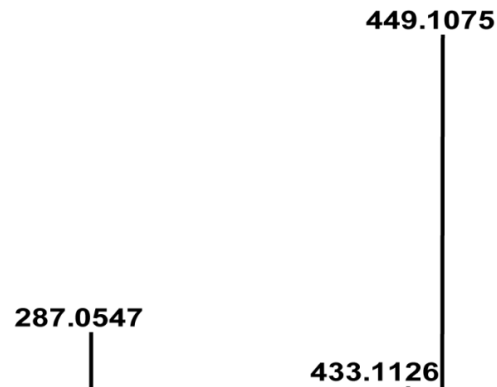
MSⁿ Based Structural Characterization of Flavonoid Isomers

Indistinguishable by MS²

MS/MS
595.1650

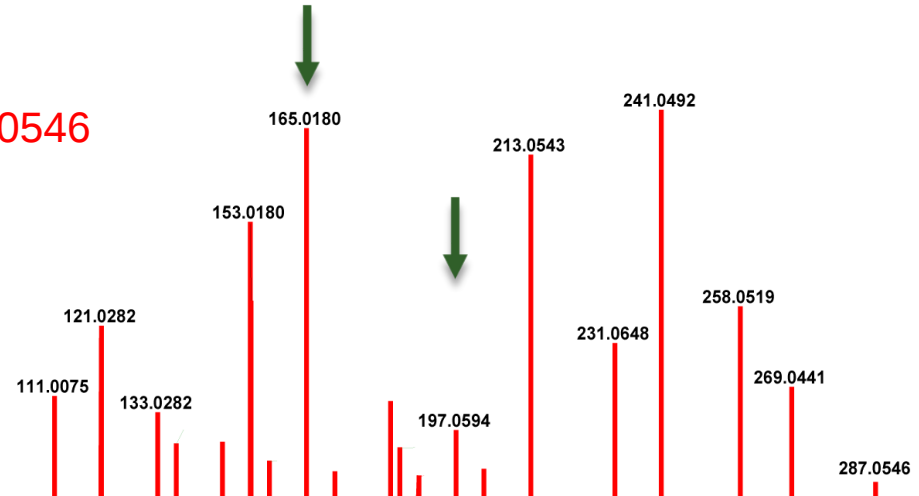


MS/MS
595.1650

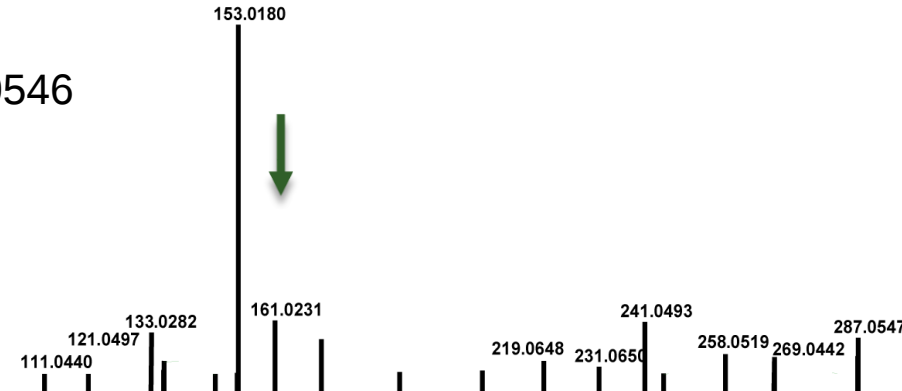


Distinguishable by MS³

MS³
287.0546

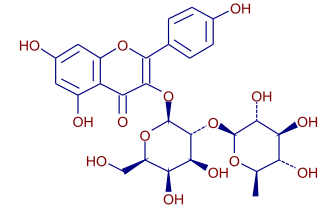


MS³
287.0546

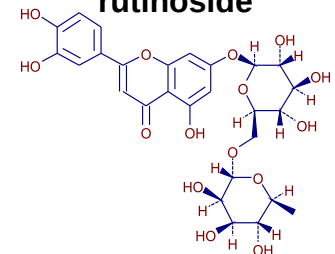


Isomers

Kaempferol 3-O-β-rutinoside

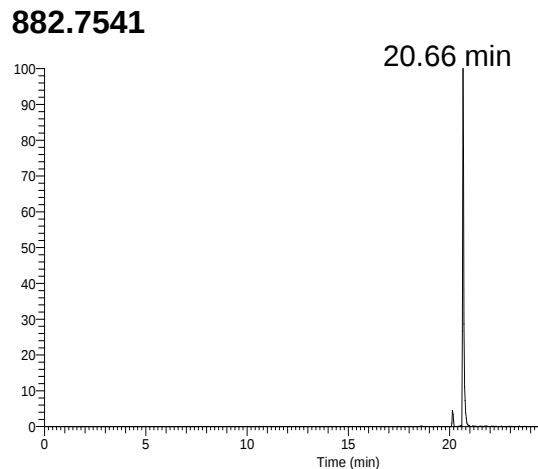


Luteolin 7-rutinoside



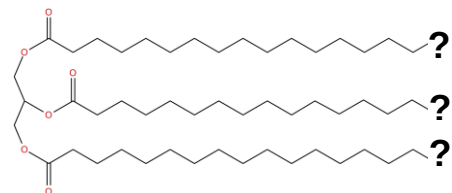
Complete Characterization of Co-eluting Isomeric Lipids Using MS³

MS¹

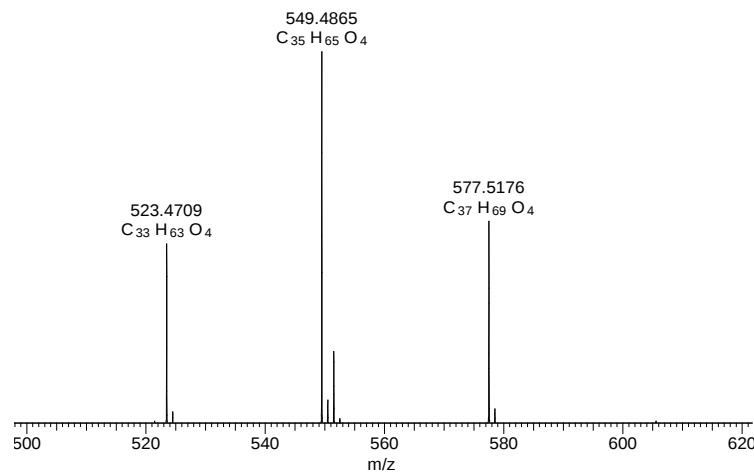


TG 48:1 [M+NH₄]⁺
C₅₁H₉₆O₆

Triglyceride
Total 48 acyl carbons and 1 double bond



MS²

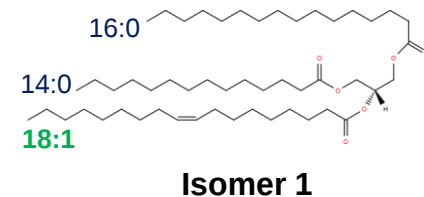
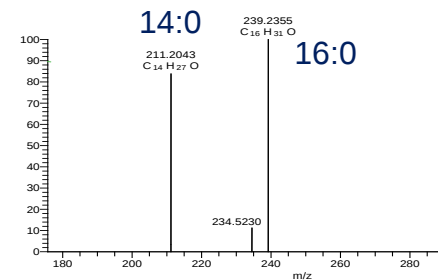


Mixture of neutral losses

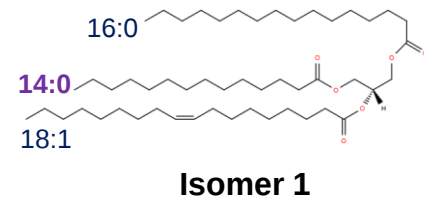
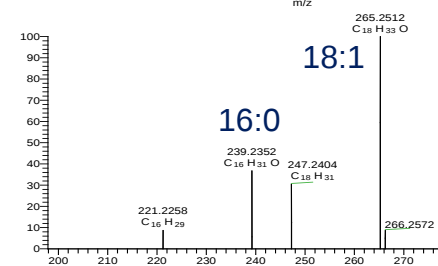
18:1 FA + NH₃
16:0 FA + NH₃
14:0 FA + NH₃
+ more...

MS³

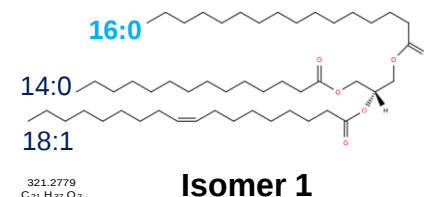
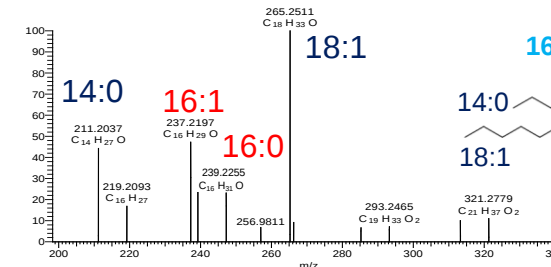
523.4709
NL 18:1



577.5176
NL 14:0

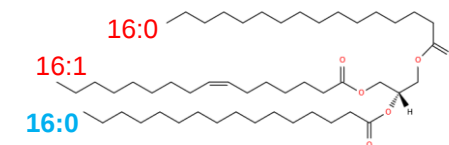


549.4685
NL 16:0



TG 14:0_18:1_16:0
TG 16:0_16:1_16:0

Two isomers



Small Molecule Dedicated Mass Spectrometer on Thermo Scientific Tribrid Platform



**Thermo Scientific™ Orbitrap ID-X™
Tribid™ Mass Spectrometer System**

Improved
Instrumentation

Orbitrap
ID-X
MS

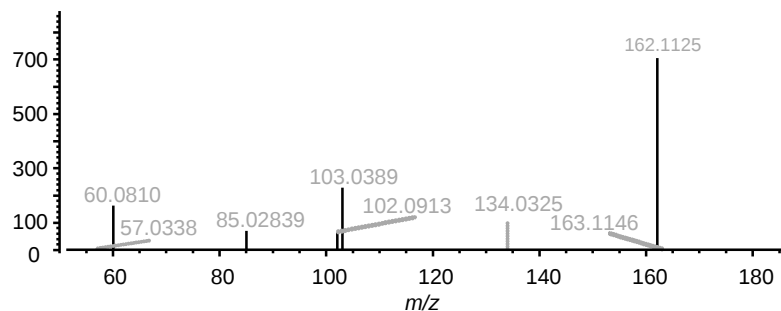
Advanced
Data Processing

Novel Data
Acquisition

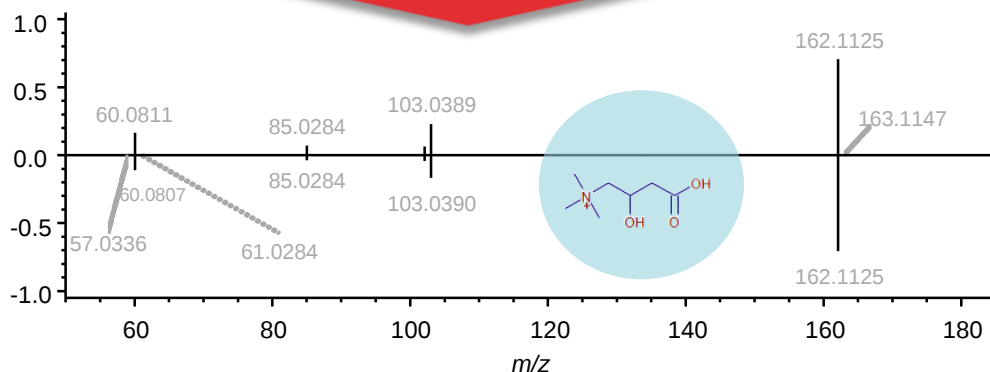
Small Molecule Data Processing Challenges

Ideally

MS² of unknown compound



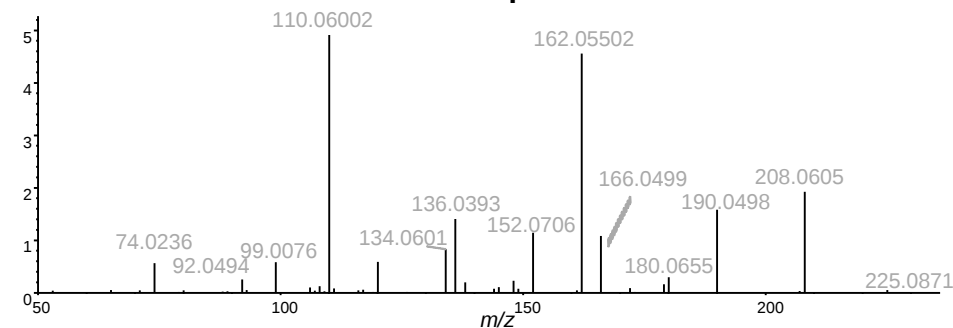
Spectral Library Search



Reference data found in spectral library
→ **Compound identified**

But what if ...

MS² of unknown compound



Spectral Library Search



NO reference data found in spectral library

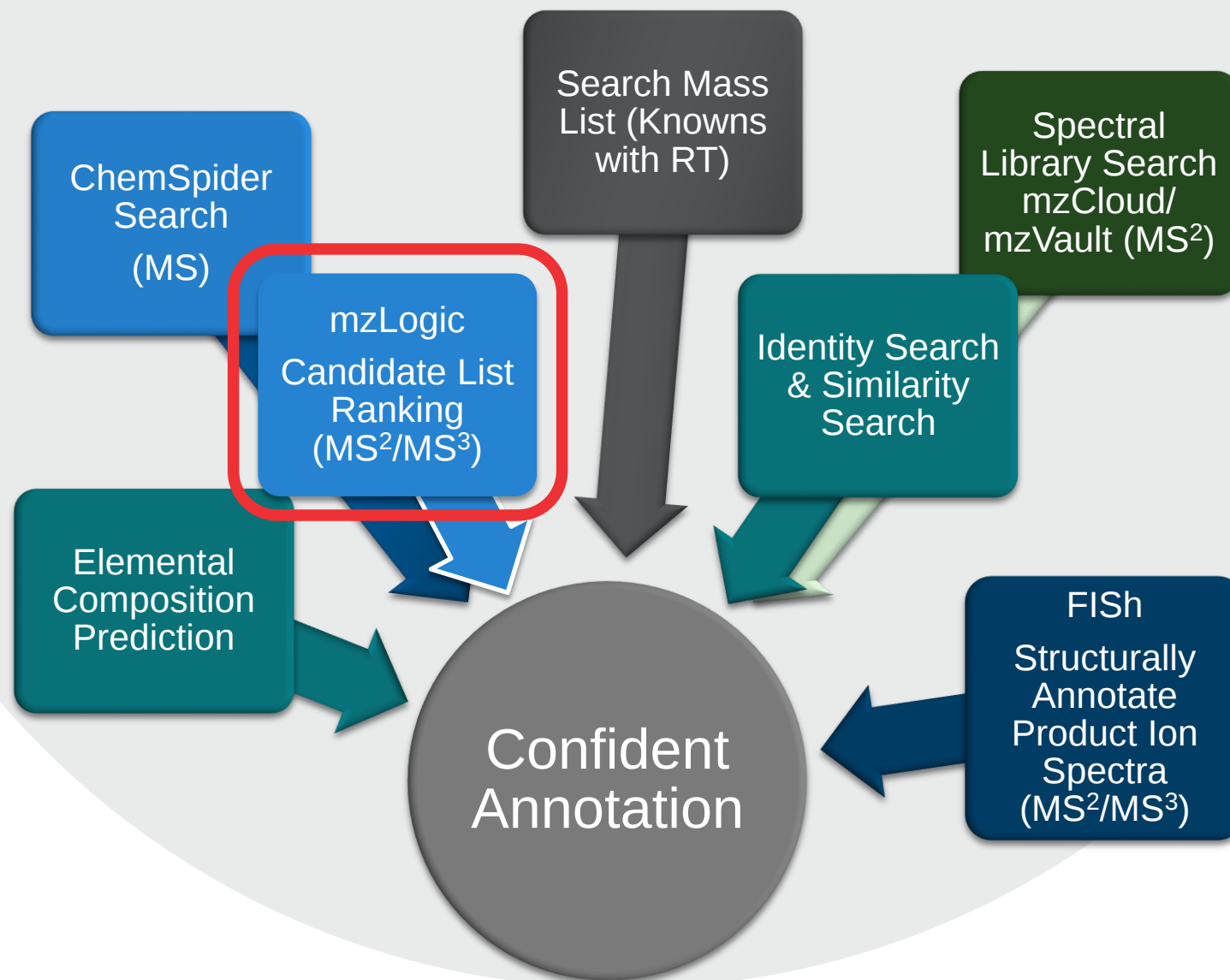
Database Search
(HMDB,KEGG,BioCyc,PubChem)

441 hits
(based on formula)

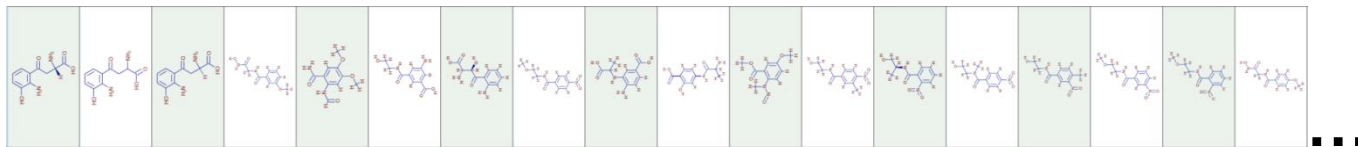
Diverse
Chemical Space

new mzLogic
algorithm

Rank-ordering
of putative
structures



mzLogic: Allows Ranking Putative Candidate Structures



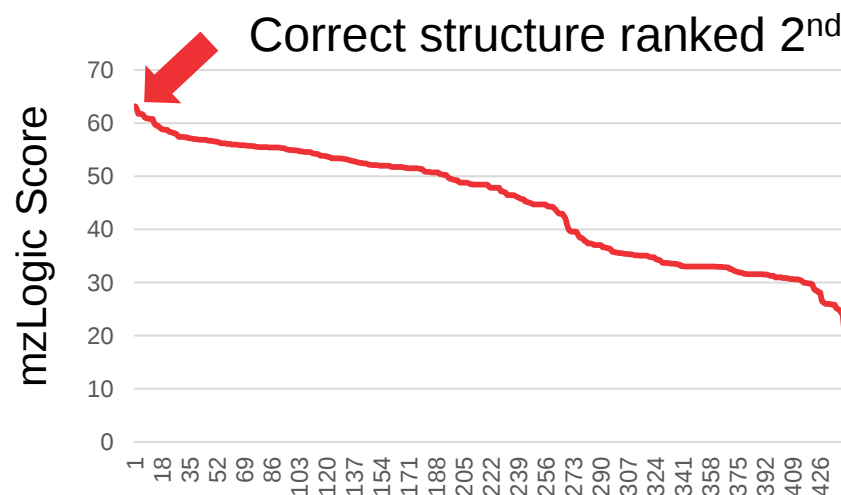
441 putative candidates for identification



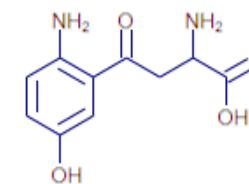
Curated spectral library
HCD/CID/MSⁿ 2.8M spectra

mzLogic in Compound Discoverer 3.0 software

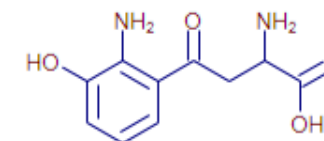
With the new mzLogic algorithm, you can use the extensive fragmentation spectral information in mzCloud to rank-order putative database results



#1 5-hydroxy-kynurenine



#2 3-hydroxy-kynurenine



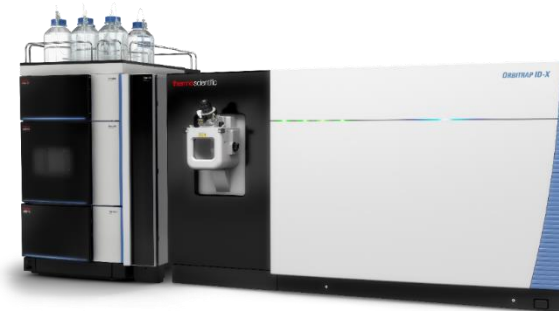
Acknowledgements

Thermo Scientific

Alexander Makarov	Iain Mylchreest	Mark Sanders	Romain Huguet
Amanda Souza	Iman Mohtashemi	Maroun El Khoury	Ryo Komatsuzaki
Andreas Huhmer	Ioanna Ntai	Martin Zeller	Sally Webb
August Specht	Ipsita Basu	Mary Blackburn	Scott Peterman
Balaram Barange	Jack Cunniff	Mike Senko	Sean Malynn
Beena Raman	Jacob Schwartz	Milla Neffling	Seema Sharma
Caroline Ding	Jean Jacques Dunyach	Neal Borelli	Shannon Eliuk
Chris Mullen	Jenny Berryhill	Nick Duczak	Stephanie Samra
Dat Lam	Jesse Canterbury	Nigama Ekkad	Susan Yung
David Peak	Jonathan Josephs	Paul Hendricks	Terry Olney
Derek Bailey	Jonathan McNally	Phil Remes	Thomas Moehring
Edward Gonzalez	Karen Johnson	Priya Rajamani	Tim Stratton
Edward Denisov	Kate Comstock	Rajesh Raina	Tony Ziberna
Eugen Damoc	Ken Miller	Ralf Tautenhahn	Tran Howard
Evett Kruka	Kien Le	Randy Porch	Uma Kota
George DeShong	Linda Lin	Ray Chen	Vlad Zabrouskov
Graeme McAlister	Lisa Thomas	Reiko Kiyonami	Wei Wei
Helene Cardasis	Manish Doshi	Rich Klein	Wilfried Voorhorst
Howard Tran	Marcia Mendonca	Rick Carberry	Willy Bjorklund
	Mark Hardman	Rodney McCoy	

Collaborators

Daniel Gachotte (Corteva Agriscience)
Fenghe Qiu (Boehringer Ingelheim)
Fuad Nasir (Washington University)
Gary Patti (Washington University)
Jeff Gilbert (Dow AgroSciences)
Jesse Balcer (Dow AgroSciences)
Kevin Cho (Washington University)
Mark Viant (Univ of Birmingham)
Martin Jones (Univ of Birmingham)
Shuguang Ma (Genentech)
Warwick Dunn (Univ of Birmingham)
William Nash (Univ of Birmingham)
Yelena Adelfinskaya (Dow AgroSciences)



A 3D visualization of an Orbitrap ion trap. It features a central cylindrical structure with two endcap rings, all rendered in a translucent red. Numerous white dots, representing ions, are distributed throughout the trap, with many having thin white lines extending from them, suggesting their trajectories or detection signals. The background is a blurred, dark red, and a semi-transparent dark grey band runs horizontally across the middle of the image.

thermofisher.com/orbitrapID-X