



## Acclaim Mixed-Mode WAX-1 Columns

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**Thermo**  
SCIENTIFIC

*For Research Use Only. Not for use in diagnostic procedures.*

## **Product Manual**

**for**

### **Acclaim<sup>®</sup> Mixed-Mode WAX-1 Columns**

5 $\mu$ m, 10 x 250 mm, P/N 088785

5 $\mu$ m, 4.6 x 150 mm, P/N 064984

5 $\mu$ m, 4.6 x 250 mm, P/N 064985

5 $\mu$ m, 2.1 x 150 mm, P/N 067084

3 $\mu$ m, 3.0 x 150mm, P/N 070088

3 $\mu$ m, 3.0 x 50mm, P/N 071908

3 $\mu$ m, 2.1 x 150mm, P/N 070089

### **Acclaim<sup>®</sup> Mixed-Mode WAX-1 Guards**

4.3 x 10 mm, P/N 064986

2.1 x 10 mm, P/N 069686

3.0 x 10mm, P/N 071909

4.6 x 10mm, P/N 069704

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## SECTION 1 – INTRODUCTION

The Acclaim® Mixed-Mode WAX-1 column is based on a new mixed-mode silica-based packing material that incorporates both hydrophobic and weak anion-exchange properties. Unlike traditional reversed-phase stationary phases, the new packing features an alkyl long chain with an ionizable terminus, and demonstrates great potentials for separating a wide range of anionic compounds containing mixtures, including pharmaceuticals, food & beverage, chemical, and more.

### 1.1. Comparison of Mixed-Mode Chromatography with Reversed-Phase, Ion-exchange and Ion-pairing Chromatography

Reversed-phase (RP) silica columns (e.g. C18) are the most widely used stationary phases for a wide range of liquid chromatography (LC) separations. However, hydrophilic ionic compounds such as small organic acids or inorganic ions are poorly retained and separated on these columns.

Ion exchange columns are used to separate ionic or ionizable compounds such as proteins, nucleic acids, inorganic ions, small organic acids, etc. Because most conventional ion-exchange stationary phases provide inadequate hydrophobic retention for neutral molecules, they have limited applications in small molecules separations.

Ion pairing chromatography is a method for separating ionic or ionizable compounds on a conventional RP medium, which requires hydrophobic ionic compounds, typically comprised of an alkyl chain with an ionizable terminus, are added to the mobile phase. Generally, retention of neutral analytes is nearly unaffected, while analytes with charges complementary to the ion pairing reagent are retained for a longer period of time and analytes with the same charge as the ion pairing reagent are retained for a shorter period of time. Limitations of ion pairing chromatography include long column equilibration times and the quantity of solvent and time needed to elute the ion pairing reagent from the column.

Mixed-mode chromatography combines aspects of ion exchange chromatography and conventional reversed-phase chromatography. A mixed-mode stationary phase has both hydrophobic and ion-exchange properties. These two strong interactions of the phase with analytes allow for controlling retention of ionizable and neutral molecules independently. As a result, many application challenges involving hydrophilic ionizable compounds that are difficult for C18 columns, can be easily tackled on a mixed-mode column.

### 1.2. Features

1. Adjustable selectivity.
2. Selectivity complementary to reversed-phase columns.
3. Simultaneous separation of acidic, basic, and neutral molecules.
4. High capacity and unique selectivity for anionic molecules.
5. Multi-mode retention mechanisms: reversed-phase, anion-exchange, and HILIC modes.

### 1.3. Specifications and Recommended Operating Conditions

Shipping Solution 90 / 10 Acetonitrile / 100mM Ammonium Acetate

Storage Solution: 90 / 10 Acetonitrile / 100mM Ammonium Acetate

Buffer pH Range: pH 2.5 - 7.5

Temperature Range < 50°C

Stationary Phase	Particle Size	Column Dimensions	P/N	Max Recommended Pressure	Typical Flow Rate
Mixed-Mode WAX-1	3 $\mu$ m	2.1x150 mm	070089	6000 psi	0.2 - 0.4 mL/min
		3.0x50 mm	071908	4500 psi	0.4 – 0.8 mL/min
		3.0x150 mm	070088	6000 psi	0.4 – 0.8 mL/min
	5 $\mu$ m	2.1x150 mm	067084	6000 psi	0.2 - 0.4 mL/min
		4.6x150 mm	064984	6000 psi	0.8 – 1.6 mL/min
		4.6x250 mm	064985	6000 psi	0.8- 1.6 mL/min
		10 x 250 mm	088785	6000 psi	3.7 – 7.5 mL/min

### 1.4. Physical Characteristics

Bonding Chemistry: Proprietary alkyl

Silica Substrate: Spherical, high-purity

Particle sizes 5  $\mu$ m and 3  $\mu$ m

Surface area 300 m<sup>2</sup>/g

Pore size 120 Å

### 1.5. Acclaim Mixed-Mode WAX-1 Products

Part number Description Dimensions

Acclaim Mixed-Mode WAX-1	Particle size	Column Dimensions	P/N	
Analytical	3 $\mu$ m	2.1x150 mm	070089	
		3.0x50 mm	071908	
		3.0x150 mm	070088	
	5 $\mu$ m	2.1x150 mm	067084	
		4.6x150 mm	064984	
		4.6x250 mm	064985	
Semi-preparative		10x250 mm	088785	
Guard	5 $\mu$ m	4.3 x 10mm	064986	Requires Holder p/n 059456
		2.1 x 10mm	069686	Requires Holder V-2 p/n 069580
		3.0 x 10mm	071909	Requires Holder V-2 p/n 069580
		4.6 x 10mm	069704	Requires Holder V-2 p/n 069580

## SECTION 2 – INSTALLATION: STEP-BY-STEP USER GUIDE

### Step 1 – Validating Column performance

Dionex recommends that you perform an efficiency test on your Acclaim Mixed-Mode HILIC column before use. The purpose of column performance validation is to ensure no damage has occurred during shipping. Test the column using the conditions described on the Quality Assurance (QA) Report enclosed in the column box. Repeat the test periodically to track the column performance over time. Note that slight variations may be obtained on two different HPLC systems due to system electronic, plumbing, operating environment, reagent quality, column conditioning, and operator technique. The steps below outline the steps in preparing your system and using your column.

### Step 2 – Mobile phase preparation

Obtaining reliable, consistent and accurate results require mobile phases that are free of ionic and spectrophotometric impurities. Chemicals, solvents and de-ionized water used to prepare mobile phase should be of the highest purity available. Maintaining low trace impurities and low particle levels in mobile phases helps to protect your columns and system components. DIONEX cannot guarantee proper column performance when the quality of the chemicals, solvents and water used to prepare the mobile phase has been compromised.

#### De-ionized Water

The de-ionized water used to prepare the mobile phase should be Type 1 Reagent Grade Water, or HPLC Grade Water. The de-ionized water should be free of ionized impurities, organics, microorganisms and particulate matter larger than 0.2  $\mu\text{m}$ . Many commercial water purifiers are designed for HPLC applications and are suitable for these applications.



#### NOTE

*Degas the aqueous component of the mobile phase and then add the solvent component. Avoid excessive purging or degassing of mobile phases containing solvents, if possible, since the volatile solvent can be 'boiled' off from the solution.*

#### Solvents

The solvents used must be free from ionic and UV-absorbing impurities. Use of ultrahigh purity solvents, HPLC grade, will usually ensure that your chromatography is not affected by impurities in the solvent.

#### Mobile Phase for Column Performance Test:

Depending on specific application, the mobile phase system consists of an organic modifier (e.g. acetonitrile or methanol) and a buffer (e.g. phosphate buffer).

#### **Example A.** Preparation of 50 mM, pH phosphate buffer.

1. Weigh  $7.90 \pm 0.05$  g of sodium phosphate monobasic monohydrate and  $0.25 \pm 0.01$  g of tetrasodium pyrophosphate decahydrate\*.
2. Completely dissolve above two salts in  $1000 \pm 10$  g of D.I. water.
3. Add  $0.96 \pm 0.01$  g of 50% sodium hydroxide solution and mix thoroughly.
4. Check that the pH is 5.95 - 6.15
5. Filter through a 0.5  $\mu\text{m}$  (or smaller) membrane.
6. Discard the buffer after two weeks.

\* Pyrophosphate is used to eliminate metal interference from stainless steel instrument (pump head, autosampler, tubing, etc) and column hardware.

- Example B.** Preparation of a mobile phase containing 50 mM, pH 6 phosphate buffer and acetonitrile (50:50 v/v).
- Pre-mixed: mix  $500 \pm 1$  g of 50 mM, pH 6 phosphate buffer and  $390 \pm 1$  g of acetonitrile; mix thoroughly and degas using an ultrasonic bath.
  - HPLC proportioning: 50% acetonitrile, 50% buffer. Avoid precipitation of the phosphate buffer; do not exceed 70% acetonitrile.



*These two mobile phases could give slightly different results due to the ways they are prepared.*

**NOTE**

**Step 3 – Set up the LC system**

Use a standard LC system equipped with a LC pump, a column oven, a UV detector, and an injector (or an auto-sampler). The system should be thoroughly primed before use.

**Step 4 – Condition the column**

The column is shipped in an acetonitrile-water mixture, therefore when a new column is used for the first time, it should be washed thoroughly with the mobile phase (e.g., for at least 45 min at the normal flow rate) before any injection is made. When switching to a new mobile phase, make sure that the new mobile phase is compatible with the previous mobile phase in the column to avoid column clogging due to precipitation. The column should be fully conditioned before any injection is made (e.g. at least 45 min at the normal flow rate).

**Step 5 – Reproduce the chromatogram in the Quality Assurance Report**

Perform the column performance test using the conditions described in the Quality Assurance Report and compare the result with the one in the report. After the column is fully equilibrated, multiple injections should be made until the reproducible retention is obtained. Keep a record of the column performance for future reference.



*Due to various reasons, such as differences in LC systems, mobile phases oven temperature control, etc, you may observe slightly different retention time for the iodide peak from that in the report.*

**NOTE**

**Step 6 – Real sample analysis**

Contamination of the column by particulate matter in the sample is a leading cause of column failure. Symptoms may include high pressure, poor symmetry, extraneous peaks, or low efficiency. Pass the sample through a  $0.5\mu\text{m}$  or smaller porosity filter before injection. Biological samples are especially troublesome due to dissolved proteins that may precipitate some time after the initial preparation.

## SECTION 3 – METHOD DEVELOPMENT

To optimize chromatographic methods, mobile phase ionic strength, pH, and organic modifier are three key variables that can be adjusted either independently or concurrently.

(Figure 1 see pg. 8, Figure 2 see pg.9, Figure 3 see pg.10)

### 3.1. Ionic Strength

Ionic strength is crucial for changing retention of charged molecules.

An increase in ionic strength can create the following results:

- a) Retention decrease for acidic molecules
- b) Retention increase for basic molecules
- e) Minimal effect for neutral molecules

### 3.2. Organic Modifier

Hydrophobic retention is markedly affected by organic modifier composition in the mobile phase. In general, all types of molecules (acids, bases, and neutrals) are less retained on this column with increased organic content in the mobile phase, when keeping other conditions constant (e.g. ionic strength, pH, temperature, etc).

### 3.3. Mobile Phase pH

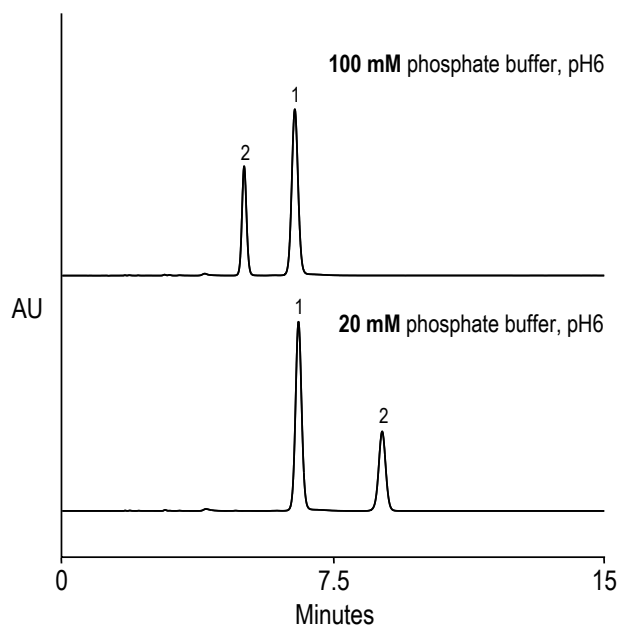
Although pH has little effect on retaining neutral molecules, it affects anionic molecules significantly. For example, with pH decrease, molecules containing carboxylic groups are less negatively charged, giving rise to decreased ion-exchange retention.

### 3.4. Isocratic vs. Gradient

For many applications that involve small number of analytes, it is usually easier to develop an isocratic method on the Acclaim Mixed-Mode WAX-1 column than a reversed-phase column. For more complicated separations, such as one that concerns a mixture of molecules with different types and numbers of charge, as well as different hydrophobicity, a gradient method could be advantageous. In practical, ionic strength gradient, organic modifier gradient, or a combination of both has proven to be satisfactory with respect to reproducibility and simplicity.



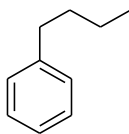
**Figure 1**  
**Adjustable Selectivity - Ionic Strength Effect**



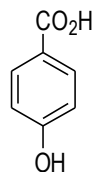
Column: Acclaim Mixed-Mode WAX, 5  $\mu$ m  
Dimension: 4.6x150 mm  
Mobile Phase: 50/50 v/v acetonitrile/phosphate buffer  
Temperature: 30  $^{\circ}$ C  
Flow Rate: 1 mL/min  
Inj. Volume: 2  $\mu$ L  
Detection: UV @ 210 nm

Peaks:

1. Butylbenzene (0.1 mg/mL)
2. 4-Hydroxybenzoic acid (0.5 mg/mL)

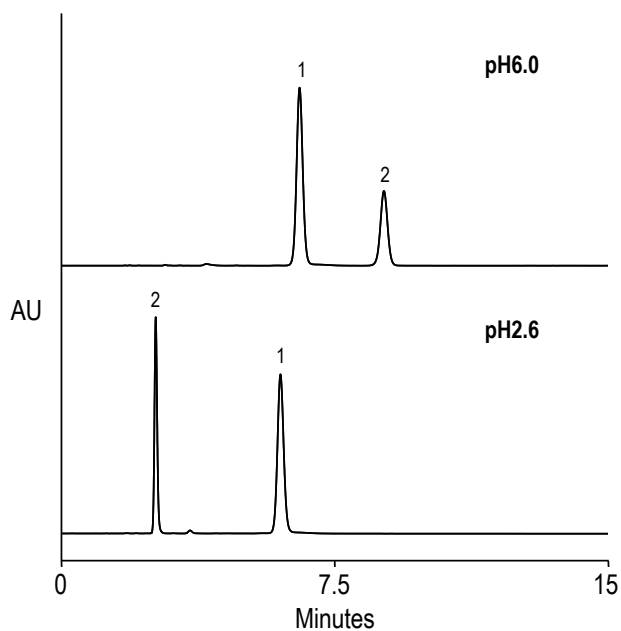


Butylbenzene



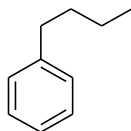
4-Hydroxybenzoic acid

**Figure 2**  
**Adjustable Selectivity - pH Effect**

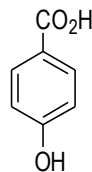


Column: Acclaim Mixed-Mode WAX, 5  $\mu$ m  
Dimension: 4.6x150 mm  
Mobile Phase: 50/50 v/v acetonitrile/20 mM phosphate buffer  
Temperature: 30  $^{\circ}$ C  
Flow Rate: 1 mL/min  
Inj. Volume: 2  $\mu$ L  
Detection: UV @ 210 nm  
Peaks:

1. Butylbenzene (0.1 mg/mL)
2. 4-Hydroxybenzoic acid (0.5 mg/mL)

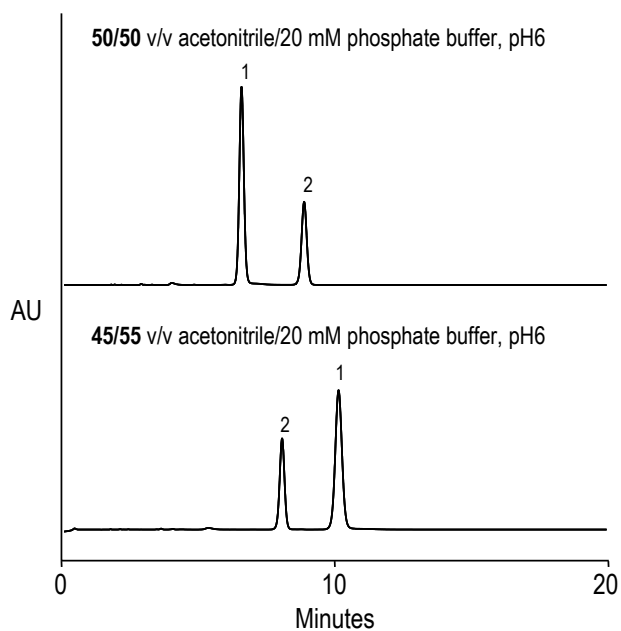


**Butylbenzene**



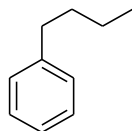
**4-Hydroxybenzoic acid**

**Figure 3**  
**Adjustable Selectivity- Organic Modifier Effect**

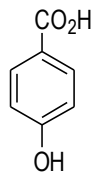


Column: Acclaim Mixed-Mode WAX, 5  $\mu$ m  
Dimension: 4.6x150 mm  
Mobile Phase: see chromatograms  
Temperature: 30  $^{\circ}$ C  
Flow Rate: 1 mL/min  
Inj. Volume: 2  $\mu$ L  
Detection: UV @ 210 nm  
Peaks:

1. Butylbenzene (0.1 mg/mL)
2. 4-Hydroxybenzoic acid (0.5 mg/mL)



**Butylbenzene**

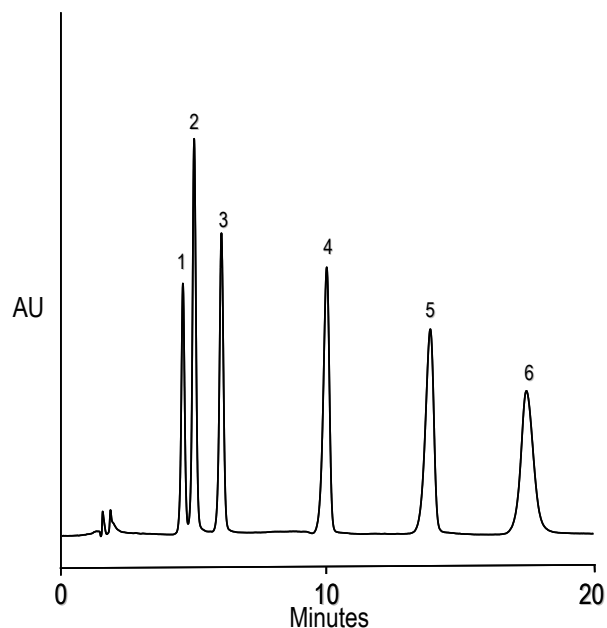


**4-Hydroxybenzoic acid**

### 3.5. HILIC Mode

The Acclaim Mixed-Mode WAX-1 column can operate in HILIC mode. In this mode, acetonitrile (not methanol) should be used in a range of 70 to 100% acetonitrile. The elution power can be modified by the employment of a polar solvent, such as an aqueous buffer. Using this column in HILIC mode provides increased retention for highly polar molecules. In general the higher the organic content in mobile phase, the longer the retention for a highly polar compound.

#### Separation of Amino Acids in HILIC Mode



Column: Acclaim Mixed-Mode WAX, 5  $\mu$ m  
Dimensions: 4.6 x 150 mm  
Mobile phase: 25/75 v/v 25 mM phosphate buffer, pH6.0/acetonitrile  
Temperature: 30°C  
Flow rate: 1 mL/min  
Injection vol.: 10  $\mu$ L  
Detection: UV, 210 nm

Peaks:

1. Leucine
2. Isoleucine
3. Valine
4. Alanine
5. Serine
6. Glycine

**Figure 4**

### 3.6. Buffer Types

The Acclaim Mixed-Mode WAX-1 column should not be used with non-buffered aqueous mobile phase, whether alone or with an organic modifier. Phosphate buffers have proven to be useful for most HPLC applications. An acetate or formate buffer may also be applicable, depending on applications.

## SECTION 4 – COLUMN CARE

### 4.1. Mobile phases

Mobile phases should be freshly prepared. All chemicals and solvents should be at the highest available quality. All mobile phases should be filtered before use. In-liner filters are recommended.

### 4.2. Guard cartridges

It is highly recommended that a guard cartridge be used with the analytical column, and replaced periodically depending on the nature of the sample. Failing to do so may result in rapid deterioration of column performance, and short column lifetime.

### 4.3. Column storage

The column can be stored in the mobile phase for a short period of time. For long term storage, use a buffered solution with higher organic content, such as 90 / 10 Acetonitrile / 100mM Ammonium Acetate pH between 3.8 to 5.8.

### 4.4. Column pH range – pH 2.5 to 7.0

To obtain better column lifetime, it is highly recommended to use “silica friendly” mobile phases. While the pH limit of the column is pH 2.5 to 7 the recommended operating pH range is between 2.8 - 6.5.

### 4.5. Recommended operating temperature limit <50 °C

Although our experimental results indicated that the column could be used at 50 °C, the separation is usually optimized by modifying mobile phase ionic strength, pH, and/or organic modifier content. Elevated temperature is not recommended and should be avoided.

### 4.6. Flow rate and pressure limit

Usually, good column efficiency for a 4.6mm internal diameter column can be obtained at 1 mL/min while a higher flow rate (2 mL/min) can be used for fast analysis provided that the pressure limit is not exceeded. The 2.1mm internal diameter columns are normally used at 0.1 to 0.8 mL/min provided that the pressure limit is not exceeded. It is extremely important not to impose sudden column pressure surge. Thus increase flow rate gradually from 0.5 mL/min up to the desired flow rate. The pressure limit for the column is 4000 psi.

### 4.7. Column washing procedure

When the column washing practice is needed, such as deteriorated column performance and/or excessively high backpressure, the following procedure can be used as a guideline:

1. Wash the column with 20 mM phosphate buffer, pH 3/acetone nitrile v/v 50/50 for 3 column volumes at a flow rate between 0.5 to 1 mL/min.
2. Wash the column with 150 mM phosphate buffer, pH 3 /acetone nitrile v/v 50/50 for 30 column volumes at a flow rate between 0.5 to 1 mL/min (to remove strongly retained anionic compounds).
3. Wash the column with 20 mM phosphate buffer, pH 3 /acetone nitrile v/v 50/50 for 3 column volumes at a flow rate between 0.5 to 1 mL/min.
4. Wash the column with 20 mM phosphate buffer, pH 3 /acetone nitrile v/v 75/25 for 30 column volumes at a flow rate between 0.5 to 1 mL/min (to remove strongly retained hydrophobic compounds).
5. Equilibrate the column with the mobile phase.



*Before any injection is made, the column should be equilibrated with a mobile phase for at least 30 column volumes.*

*If the above treatment fails to improve the column performance, replace it with a new one.*

#### NOTE

*For a 2.1 mm i.d. column, the flow rate should be reduced to 20% of that for 4.6 mm i.d. column.*

*If the above treatment fails to revive the column, the column should be replaced.*

## SECTION 5 – FREQUENTLY ASKED QUESTIONS

### 1. What is the Acclaim Mixed-Mode WAX-1 column?

The Acclaim Mixed-Mode WAX-1 column is a new mixed-mode silica column that incorporates both hydrophobic and weak anion-exchange properties. Its surface chemistry features an alkyl long chain with a weak anion-exchange terminus. This column has demonstrated great potentials for separating a wide range of anionic compounds containing mixtures, including pharmaceuticals, food & beverage, chemical, and more.

### 2. Why do I need the Acclaim Mixed-Mode WAX-1 column?

The mixed mode separation mechanism of the Acclaim Mixed-Mode WAX-1 column allows for controlling retention of ionizable and neutral molecules by change mobile phase ionic strength, pH, and organic composition, either independently or collaboratively. As a result, many application challenges involving hydrophilic ionizable compounds that are difficult for C18 columns, can be easily accomplished on this column.

### 3. When do I need the Acclaim Mixed-Mode WAX-1 column?

Here are some situations among others you may consider using the Acclaim Mixed-Mode WAX-1 column:

- 1) Separation of hydrophilic organic acids.  
(Figures 5 see pg.15, Figure 6 see pg.16, Figure 7 see pg.17)
- 2) Simultaneous separation of acidic, neutral, and hydrophobic pharmaceuticals.  
(Figures 8 see pg.18, Figure 9 see pg.19)
- 3) Fast analysis of soft drinks. (Figure 10 see pg. 20)
- 4) When you need selectivity orthogonal to a reversed-phase column.  
(Figure 11 see pg.2, Figure 12 see pg.22)

### 4. What factors should I consider for method development using this column?

There are three main factors that affect column selectivity: mobile phase ionic strength, mobile phase pH, and mobile phase organic composition. You can optimize your separation by changing one, two, or all three factors. See Section 3 - Method Development for details.

### 5. What mobile phases should I use with this column?

Phosphate buffer works satisfactorily in many applications. Depending on the application, the buffer concentration range can be between 10 mM to 250 mM, and pH range should be in the range 2.5 to 7. When an organic modifier is used, make sure to keep it miscible with the buffer solution. A formate or acetate buffer can also be used in some applications.

### 6. What should I do before starting using Acclaim Mixed-Mode WAX-1 column?

Read this User Guide carefully, and contact Dionex Technical Support if you have any questions regarding using this column.

### 7. Can I use this column for separating hydrophilic organic acids?

Yes. You can use this column to separate a wide range of organic acids that are difficult to separate on reversed-phase columns.

### 8. How to store the column?

Refer to “Section 4.3. Column storage” for details.

### 9. Can I use this column to analyze basic molecules?

This column is suitable to analyze basic molecules with intermediate to high hydrophobicity under proper chromatographic conditions. Good peak shape is usually expected on this column.

### 10. Can I use this column to analyze neutral molecules?

Yes. This column provides intermediate hydrophobic retention so that neutral molecules with intermediate to high hydrophobic retention can be retained sufficiently. For highly hydrophilic/polar molecules, a HILIC mode separation using this column should be considered.

**11. Can I use this column to separate a mixture of basic, acidic, and neutral molecules?**

Yes. As shown in Figures 8 and 9 the Acclaim Mixed-Mode WAX-1 separates a mixture of basic, neutral, and acidic molecules in a single, with excellent peak shape and resolution. It provides higher degree of flexibility for application method development compared to both conventional reversed-phase and ion-exchange columns.

**12. Do I need a guard cartridge with an Acclaim Mixed-Mode WAX-1 analytical column?**

Yes. It is highly recommended to use guard cartridges with an Acclaim Mixed-Mode WAX-1 analytical column. The guard cartridge protects the more expensive analytical column by trapping highly retained components and particulates from the mobile phase or the sample.

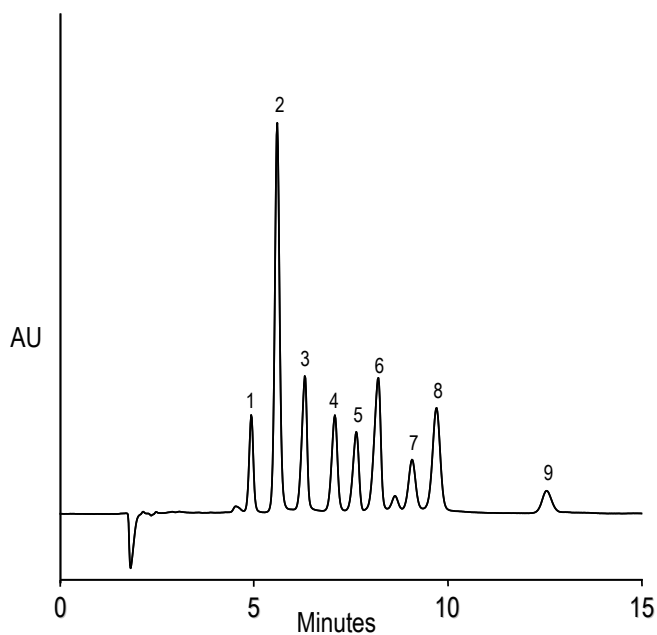
**13. What should I do if the column shows deteriorated performance?**

Refer to “Section 4.7. Column washing procedure” for details.

**14. What should I do if the column exhibits excessively high backpressure?**

First, make sure that the mobile phase is freshly prepared and filtered before use, and that the sample are free of particulates. Then, back flush the column for certain amount of time while monitoring the change in column pressure. If problem persists, try to replace the inlet bed support. If all above fail, purchase a new column.

**Figure 5**  
**Separation of Mono-Valent Carboxylic Acids**

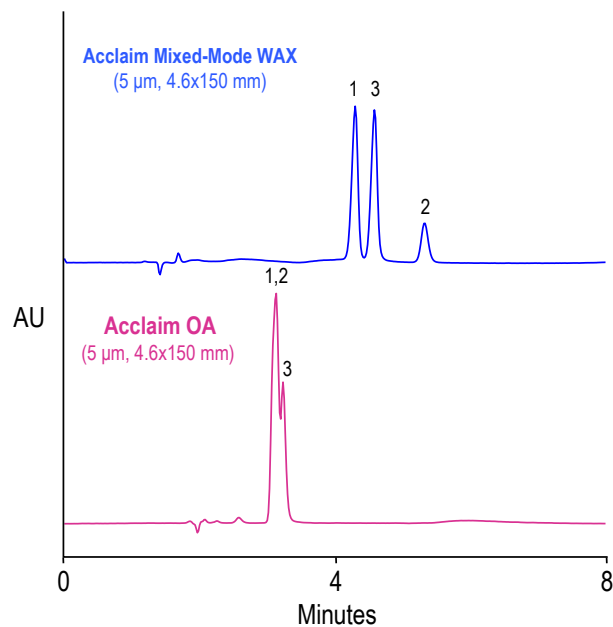


Column: Acclaim Mixed-Mode WAX, 5  $\mu$ m  
Dimensions: 4.6 x 150mm  
Mobile phase: 25 mM phosphate buffer, pH6  
Temperature: 30°C  
Flow rate: 0.8 mL/min  
Injection vol.: 10  $\mu$ L  
Detection: UV @ 210 nm  
Peaks:

1. Quinic acid
2. Shikimic acid
3. Glycolic acid
4. Lactic acid
5. Acetic acid
6. Formic
7. Ascorbic acid (Vitamin C)
8. Iso-ascorbic acid
9. Propionic acid

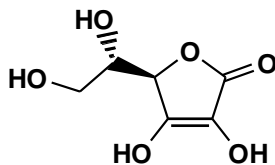


**Figure 6**  
**Separation of Lactate, Acetate and Ascorbate**

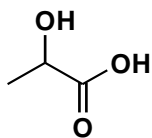


Mobile Phase: 20 mM phosphate buffer, pH6.0 for Acclaim Mixed-Mode WAX  
 50 mM phosphate buffer, pH2.6 for Acclaim OA  
 Temperature: 30 °C  
 Flow Rate: 1 mL/min  
 Inj. Volume: 5 μL  
 Detection: UV @ 210 nm  
 Peaks:

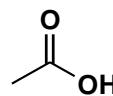
1. Lactic acid (0.8 mg/mL)
2. Ascorbic acid (Vitamin C) (0.25 mg/mL)
3. Acetic acid (0.8 mg/mL)



**Ascorbic acid (Vitamin C)**

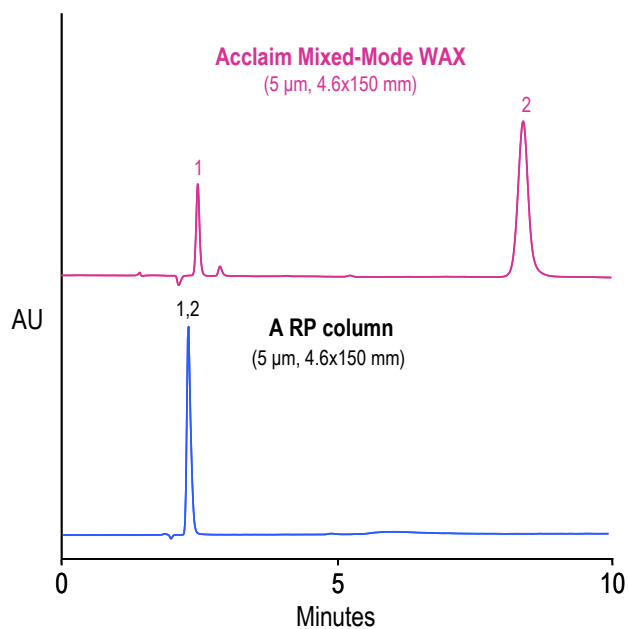


**Lactic acid**



**Acetic acid**

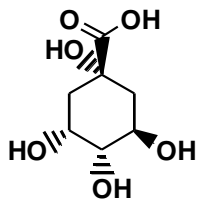
**Figure 7**  
**Separation of Quinic Acid and Tartaric Acid**



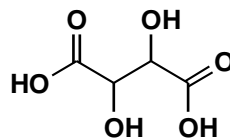
Mobile Phase: 40/60 v/v acetonitrile/50 mM phosphate buffer, pH6.0 for  
Acclaim Mixed-Mode WAX  
50 mM phosphate buffer, pH2.6 for a RP column  
Temperature: 30 °C  
Flow Rate: 1 mL/min  
Inj. Volume: 5  $\mu$ L  
Detection: UV @ 210 nm

Peaks:

1. Quinic acid (1 mg/mL)
2. Tartaric acid (1 mg/mL)

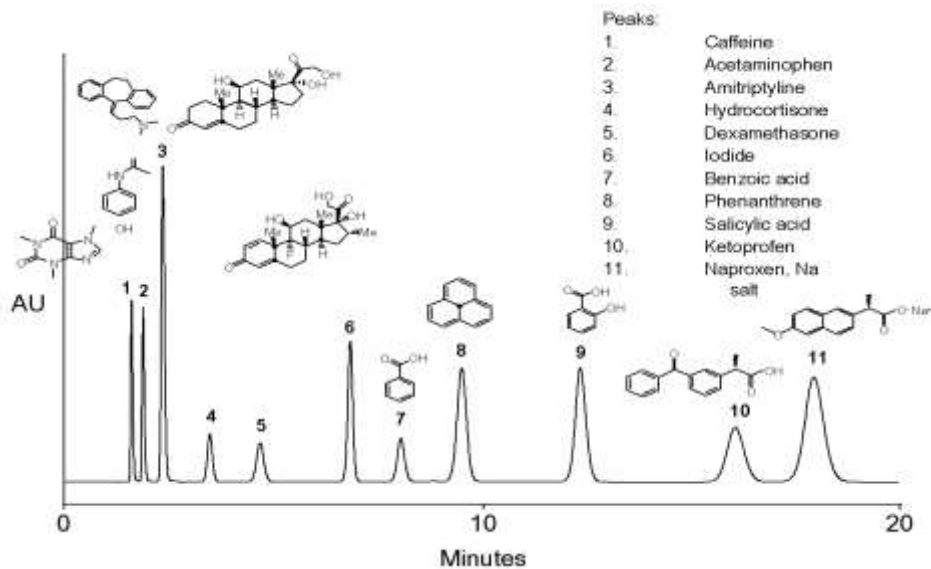


**Quinic acid**



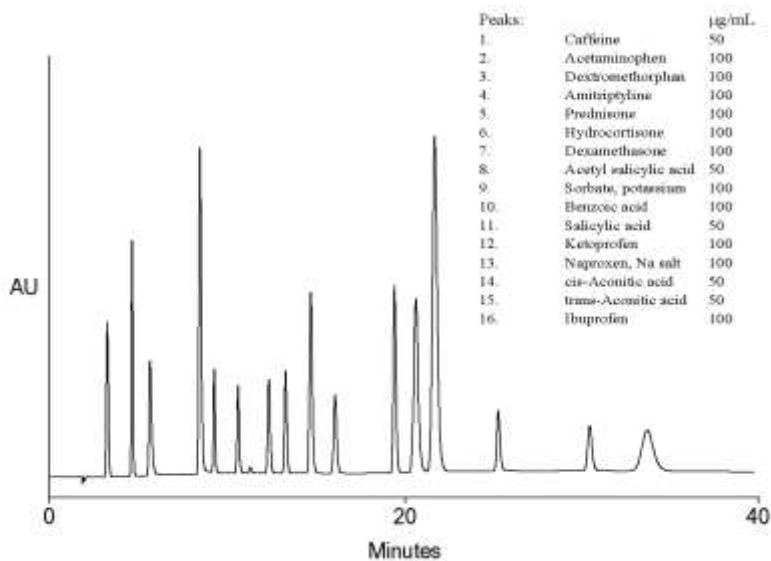
**Tartaric acid**

**Figure 8**  
**Isocratic Separation of Basic, Neutral and Acidic Molecules**



Column: Acclaim Mixed-Mode WAX, 5  $\mu$ m  
 Dimension: 4.6 x 150 mm  
 Mobile Phase: 50/50 v/v Acetonitrile/buffer (6.8 g potassium monophosphate and 0.5 g pyrophosphate in 1000 g D.I. H<sub>2</sub>O, pH is adjusted to 6.0 with NaOH)  
 Temperature: 30 °C  
 Flow Rate: 1 mL/min  
 Inj. Volume: 5  $\mu$ L  
 Detection: UV @ 220 nm

**Figure 9**  
**Gradient Separation of Basic, Neutral and Acidic Pharmaceuticals**

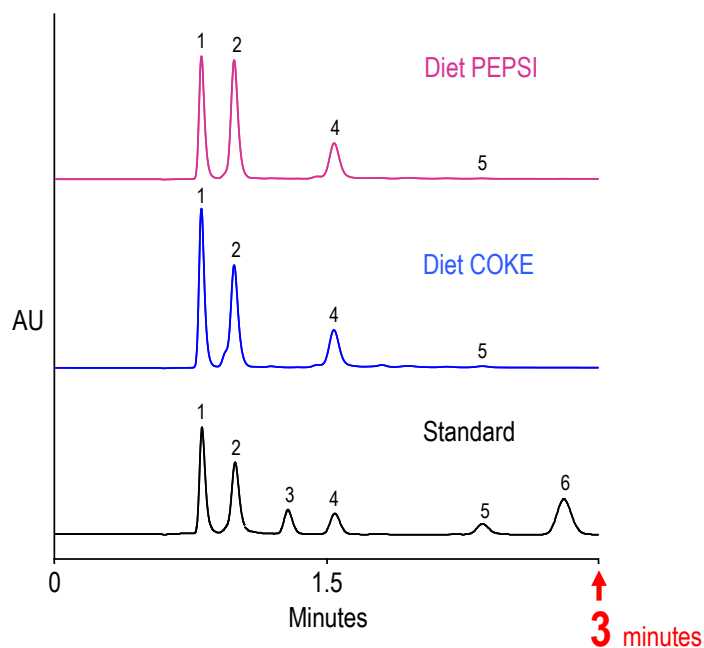


Column: Acclaim Mixed-Mode WAX, 5 µm  
 Dimension: 4.6 x 150 mm  
 Mobile Phase: A – Acetonitrile;  
 B – D.I. water;  
 C – 150 mM phosphate buffer, pH6.0

Gradient:	Time	%A	%B	%C
	-15	10	80	10
	0	10	80	10
	10	50	40	10
	12	50	40	10
	25	50	0	50
	40	50	0	50

Temperature: 30 °C  
 Flow Rate: 1 mL/min  
 Inj. Volume: 15 µL  
 Detection: UV @ 220 nm

**Figure 10**  
**Rapid Analysis of Soft Drinks**

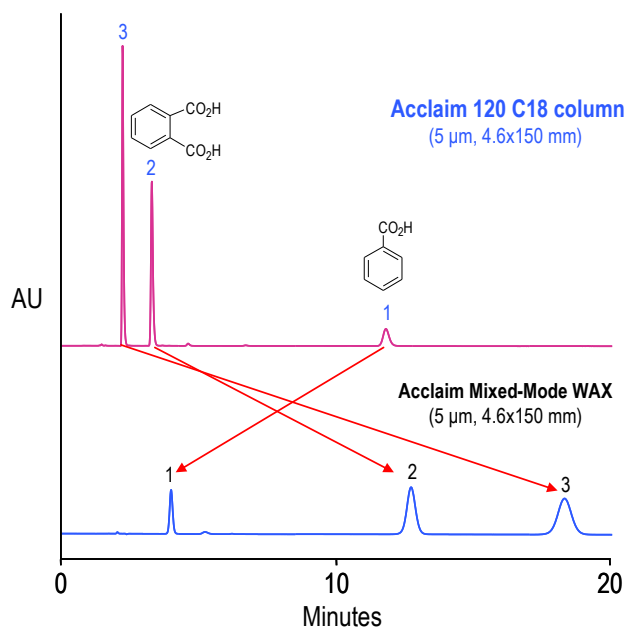


Column: Acclaim Mixed-Mode WAX, 5  $\mu$ m  
 Dimension: 4.6 x 150 mm  
 Mobile Phase: 57/43 v/v Acetonitrile/ 120 mM phosphate buffer, pH2.9  
 Temperature: 30  $^{\circ}$ C  
 Flow Rate: 2 mL/min  
 Inj. Volume: 2.5  $\mu$ L  
 Detection: UV @ 210 nm  
 Sample: **Direct injection of degassed sample**  
 Peaks:

1. Caffeine
2. Aspartame
3. Sorbate
4. Benzoate
5. Citrate
6. Acesulfame

Diet Pepsi is a registered trademark of Pepsi-Cola Company.  
 Diet Coca-Cola is a registered trademark of the Coca-Cola Company.

**Figure 11**  
**Orthogonal Selectivity to Reversed-Phase Columns – I**

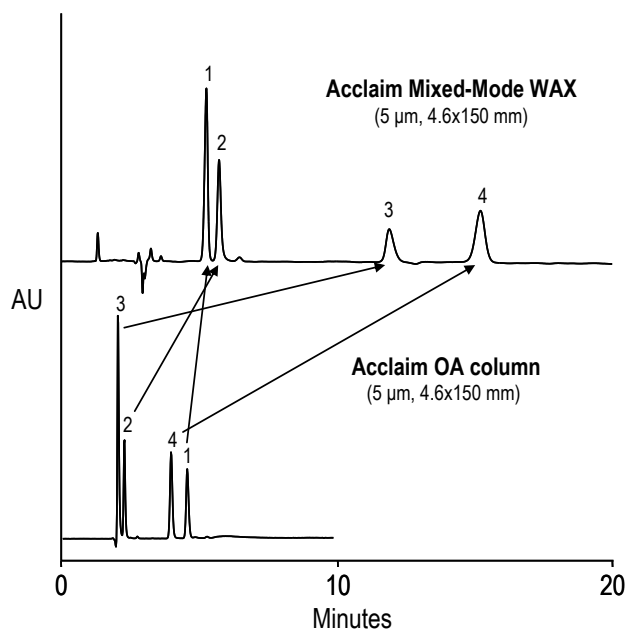


Mobile Phase: 50/50 v/v acetonitrile/50 mM phosphate buffer, pH2.8 for  
Acclaim Mixed-Mode WAX  
20/80 v/v acetonitrile/50 mM phosphate buffer, pH2.8 for  
Acclaim 120 C18 column

Temperature: 30 °C  
Flow Rate: 1 mL/min  
Inj. Volume: 2  $\mu$ L  
Detection: UV @ 220 nm  
Peaks:

1. Benzoic acid (0.2 mg/mL)
2. Phthalic acid (0.2 mg/mL)
3. 1,2,3-Tricarboxylic benzene (0.2 mg/mL)

**Figure 12**  
**Orthogonal Selectivity to Reversed-Phase Column – II**



Mobile Phase: 50/50 v/v acetonitrile/100 mM phosphate buffer, pH6.0 for  
Acclaim Mixed-Mode WAX

50 mM phosphate buffer, pH2.6 for Acclaim OA column

Temperature: 30 °C

Flow Rate: 1 mL/min

Inj. Volume: 5  $\mu$ L

Detection: UV @ 210 nm

Peaks:

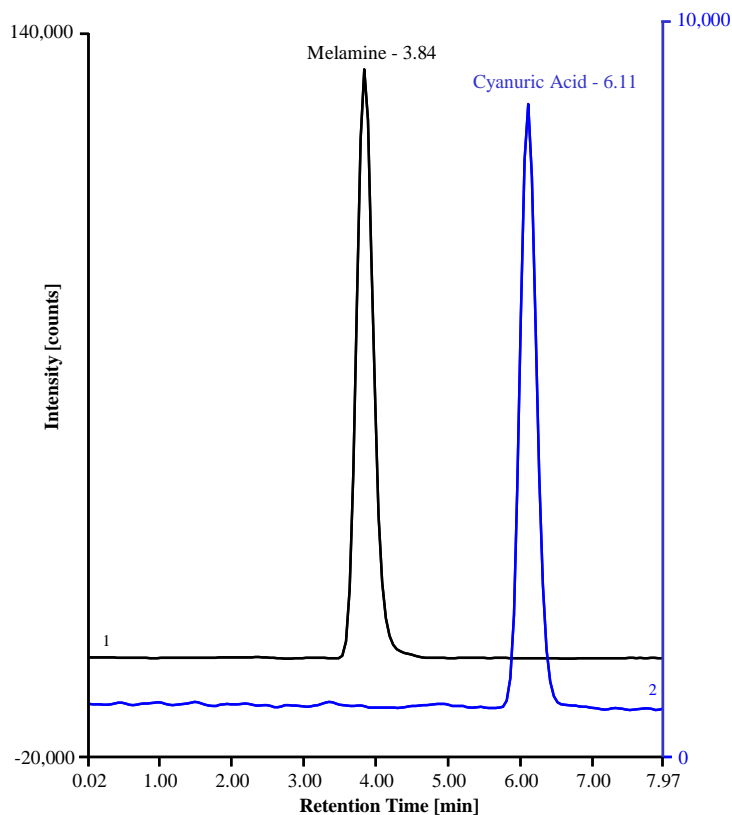
1. Succinic acid (0.5 mg/mL)

2. Tartaric acid (0.5 mg/mL)

3. Oxalic acid (0.1 mg/mL)

4. Citric acid (0.5 mg/mL)

**Figure 13**  
**Melamine and Cyanuric Acid on Mixed-Mode WAX-1**



Column:	Mixed-Mode WAX-1,	
Dimensions:	2.1 x 150 mm, 5 $\mu$ m	
Mobile Phase:	90/4/6 v/v acetonitrile/50 mM ammonium formate buffer (pH 4)/water	
Flow Rate:	0.25 mL/min	
Column Temp.:	30 °C	
Inj. Volume:	5 $\mu$ L	
Detector:	MSQ Plus mass spectrometer with Monitoring (SIM)	Selected Ion
Melamine SIM:	Positive ESI +127 $m/z$ @ 40V	
Cyanuric Acid SIM:	Negative ESI -128 $m/z$ @ 40V	
Dwell time:	0.5 second	