Q Exactive HF-X QuickStart Guide

About this Guide

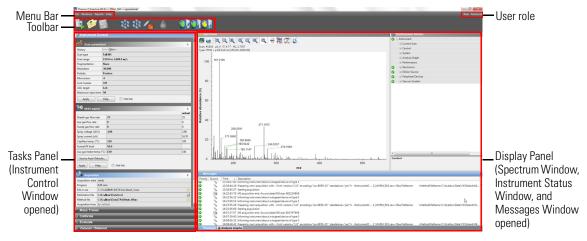
This *Q Exactive HF-X QuickStart Guide* gives an introduction on setting up and using the Thermo Scientific[™] Q Exactive[™] HF-X mass spectrometer. This guide is intended for all personnel who operate the Q Exactive HF-X mass spectrometer.

Please note that this guide does not contain any safety notices and precautionary statements. Thus, it is mandatory to read also the *Exactive Series Operating Manual* and the *Ion Max and Ion Max-S API Source Hardware Manual*.

Q Exactive HF-X Tune

Use Q Exactive HF-X Tune to operate the Q Exactive HF-X mass spectrometer with the data system computer. It has the following components:

- The *menu bar* provides drop-down menus with commands for operating the program. On its right side it shows the active user role.
- The *toolbar* provides symbol shortcuts for frequently used commands.
- The *tasks panel* provides five windows: Instrument Control, Mass Traces, Calibrate, Evaluate, and Vacuum/Bakeout. The *instrument control window* is divided into three additional windows for defining scan parameters, setting API source parameters, and acquiring and storing measurement data.
- The *display panel* provides various windows for showing real-time information about the current measurement and the instrument status.



To display the Q Exactive HF-X Tune program

Choose Start > Programs > Thermo Exactive Series > Tune.

Preparation

Before you go on, check that the following preconditions are fulfilled:

- The main power circuit breaker switch is in the On (|) position and the electronics service switch is in the Operating Mode position, the gas pressure is within the operational limit, and the vacuum levels are sufficient for operating the instrument.
- The Ion Max API source with H-ESI probe is installed on the mass spectrometer. You have checked the proper position of the H-ESI probe in the Ion Max API source housing.
- Power supply and communication between syringe pump and mass spectrometer are established. A clean 500 μ L Unimetrics syringe with 450 μ L of calibration solution for positive ion mode is placed in the syringe pump. An infusion line is installed between the syringe pump and the grounding union that is held by the grounding bar of the Ion Max API source.

Refer to the *Exactive Series Operating Manual* for instructions about preparing the positive-ion-mode calibration solution or buying the ready-to-use solution from Thermo Fisher Scientific (refer to www.fishersci.com, catalog number PI88323).





The data system computer with the Q Exactive HF-X Tune software is switched on and the mass spectrometer is placed in Standby mode.

For information about performing these tasks, refer to the *Exactive Series Operating Manual*, the *Q Exactive HF-X Software Manual* or the Q Exactive HF-X Tune Help, and the *Ion Max and Ion Max-S API Source Hardware Manual*.

This topic describes how to test the instrument before you can calibrate your Q Exactive HF-X mass spectrometer automatically.

Experiments Setting up Q Exactive HF-X

Getting lons

from Infusion



Tune

To set up the mass spectrometer in the Q Exactive HF-X Tune software for tuning and calibrating in the H-ESI/MS mode

- 1. If you have not already done so, choose **Start > Programs > Thermo Exactive Series> Tune** to start the Q Exactive HF-X Tune program.
- 2. In the Q Exactive HF-X Tune window, on the toolbar, click the **On/Off/Standby** button to take the mass spectrometer out of the Standby mode or Off mode and turn it On. When you turn the mass spectrometer to On, you initiate the following events:
 - The mass spectrometer begins scanning.
 - Nitrogen flows into the H-ESI probe.
 - The Q Exactive HF-X mass spectrometer applies a high voltage to the H-ESI probe.
 - The spectrum window of Q Exactive HF-X Tune shows a real-time display of the mass spectrum.
- 3. If necessary, open an existing tune file:
 - a. Choose File > Load Tune File to display the Open dialog box.
 - b. Browse for the folder C:\Xcalibur\methods\. Select a tune file (for example, the default tune file for H-ESI measurements).
 - c. Click **Open** to load the file. Q Exactive HF-X Tune downloads the Tune Method parameters to the mass spectrometer.

A tune file is specific to your particular analyte and solvent flow rate. You can load the tune file again and use it as a starting point for optimization of the mass spectrometer for a different analyte of interest or with a different flow rate.

- 4. Examine the pre-tune H-ESI source settings:
 - a. From the Instrument Control panel, open the H-ESI source window. See figure right.
 - b. Set the *sheath gas flow rate* to 12.
 - c. Leave the auxiliary gas flow rate at 0.
 - d. Leave the sweep gas flow rate at 0.
 - e. Set the spray voltage to 4 kV.
 - f. Set the *capillary temperature* to 320 °C.
 - g. Leave the Funnel RF level at 40.
 - h. Set the *auxiliary gas heater temperature* to 0 °C.
 - i. If you have made any changes in the window, click **Apply**.

NOTICE Select the Hot link check box to apply all changes in real time. A green frame around the parameter box indicates an active hot link. ▲

HESI source		\$				
		actual				
Sheath gas flow rate	12					
Aux gas flow rate	0					
Sweep gas flow rate	0					
Spray voltage (kV)	4.00					
Spray current (µA)						
Capillary temp. (°C)	320					
Funnel RF level	40.0					
Aux gas heater temp (°C)	0					
Source Auto-Defaults						
Apply Help 🔲 Hot link						

- 5. Set the scan parameters for tuning and calibration:
 - a. From the Instrument Control panel, open the Scan parameters window. See figure right.
 - b. Set Scan type to Full MS.
 - c. Enter a scan range between m/z 150 and 2000.
 - d. Set *Fragmentation* to *None* to specify that the ion source and HCD fragmentation options are turned off.
 - e. Select 15000 for Resolution.
 - f. Set the ion polarity mode to Positive.
 - g. Set the total number of *microscans* to 1.
 - h. Set Lock masses to Off.
 - i. Select 1e6 for AGC target.
 - j. Specify a *maximum injection time* of 30 ms.
 - k. If you have made any changes in the window, click Apply or select the Hot link check box.

The mass spectrometer is now properly set up for tuning and calibration in the H-ESI/MS mode.

In this section, you can test whether the mass spectrometer is operating properly. Infuse a low concentration calibration solution for positive ion mode—containing *n*-butylamine, caffeine, MRFA, and Ultramark 1621—into the H-ESI source with the syringe pump. Monitor the real-time display of the mass spectrum to ensure that a stable spray of solution enters the mass spectrometer.

Refer to the *Exactive Series Operating Manual* for instructions about preparing the positive-ion-mode calibration solution or buying the ready-to-use solution from Thermo Fisher Scientific (refer to www.fishersci.com, catalog number PI88323).

Syringe type

Volume (uL)

Apply

Syringe inner diameter (mm)

Help

Flow rate (µL / min)

Manual control

To test the operation of the mass spectrometer in H-ESI/MS mode

- 1. Click the arrow of the **Syringe Pump** button to display the Syringe Pump dialog box. See figure right.
- 2. Turn on the syringe pump and set an infusion flow rate of 5μ L/min, as follows:
 - a. Select *Unimetrics* to specify the proper *syringe type*.
 - b. Select *500* in the *Volume* list box to set the proper syringe volume. Note that the Q Exactive HF-X mass spectrometer automatically sets the syringe ID to its proper value of 3.257 mm.
 - c. Specify a *flow rate* of 5 µL/min.
 - d. Click **Apply** to apply the syringe parameters.
 - e. Click Start to start the syringe pump.

NOTICE Once you have set the syringe parameters, just click the **Syringe Pump** button to switch on or off the syringe pump. ▲

- 3. Test the efficiency and stability of the calibration solution spray into the mass spectrometer. Monitor the data for the calibration solution, as follows:
 - a. In the spectrum window of Q Exactive HF-X Tune, observe the mass spectra of the singly-charged ions of the positive ion mode calibration solution. See figure below. The ions are as follows:
 - *n*-Butylamine: *m*/*z* 74
 - Caffeine: *m/z* 138 (fragment), *m/z* 195
 - MRFA: *m/z* 524
 - Ultramark 1621: *m/z* 1022, 1122, 1222, 1322, 1422, 1522, 1622, 1722, 1822
 - b. At the top of the spectrum window, notice the values for the inject time (IT) in milliseconds and the normalized target level (NL). See figure below.

Scan parameters				
History				
Scan type	Full MS			
Scan range	150.0 to 2,000.0 m/z			
Fragmentation	None			
Resolution	15,000			
Polarity	Positive			
Microscans	1			
Lock masses	Off			
AGC target	1e6			
Maximum inject time	30			
Apply	Help 🔲 Hot link			

Unimetrics

500

.25

5.000

Stop

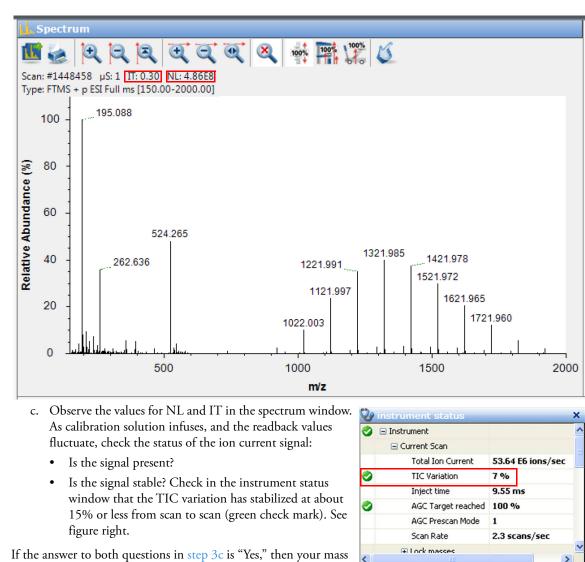
Start



Testing the Mass

Spectrometer





If the answer to both questions in step 3c is "Yes," then your mass spectrometer is operating properly.

If the answer to one of these questions is "No," then try the following troubleshooting measures:

- Make sure that the H-ESI probe is properly assembled and that the needle position is correct.
- Make sure that the entrance to the ion transfer capillary is clean.
- Make sure that the solution entering the probe is free of air bubbles and that the tubing and connectors are • free of leaks and that they are not clogged.

Current Scan

- Make sure that spray voltage is appropriate. •
- Make sure that discharging does not take place. •

Your mass spectrometer is operating properly in H-ESI mode. You are now ready to calibrate the mass spectrometer.

Mass Calibration	The Q Exactive HF-X mass spectrometer should be calibrated to ensure the mass accuracy of the results. Usually, it is sufficient to perform a mass calibration only. Before you start the calibration, check again that you have a stable spray: Focus on the injection time (IT) and on the normalized level (NL) in the header information on the spectrum window.
	NOTICE If the system was in Off mode before, put the instrument into On mode for at least 90 minutes before you perform a mass calibration. ▲
Positive	To calibrate the Q Exactive HF-X mass spectrometer for positive ion mode

ometer for positive ion mo

1. Place a clean 500 μ L Unimetrics syringe with 450 μ L of calibration solution for the positive ion mode into the syringe pump.

Ion Mode

Calibration

Thermo Scientific



- 2. Click the On/Off/Standby button to take the mass spectrometer out of the Standby mode or Off mode and turn it On.
- 3. Click the Syringe Pump button to start the syringe pump.
- 4. Click the title bar of the Calibrate window to display it.
 - 5. In the Calmix Calibration sub-window, select the Mass Calibration (pos) check box to specify a mass calibration for the positive ion mode. See figure right.
 - 6. In the Calibrate window, click Calibrate to start the automatic calibration procedure:
 - The Calibrate window indicates the completed percentage of the calibration process by a progress bar.
 - Q Exactive HF-X Tune displays a message box that informs about the active calibration procedure and the currently calibrated parameter.

Help

Mass Calibra

olation Mass and Re

OK (2017-03-16 16:44) ок

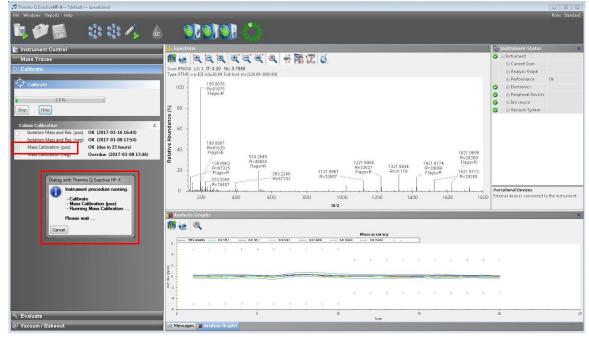
OK (due in 25 h rdue (2017-03-08 17:46)

(2017-03-08 17:54)

- The analysis graphs window displays a graphic representation of the calibration process for the currently tuned parameter.
- The messages window shows a record of the calibration.



7. An icon appears on the toolbar to indicate an active procedure.



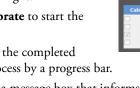
8. A message box appears when the calibration procedure is completed. The message box also informs you that Q Exactive HF-X Tune has automatically saved the calibration results. Click **OK** to close the message box.

Before you can start analyzing samples, you must ensure that the quadrupole isolation performance for the narrow isolation window is sufficient. Therefore, evaluate the isolation calibration for the positive ion mode as described on page 7.

Negative Ion Mode Calibration

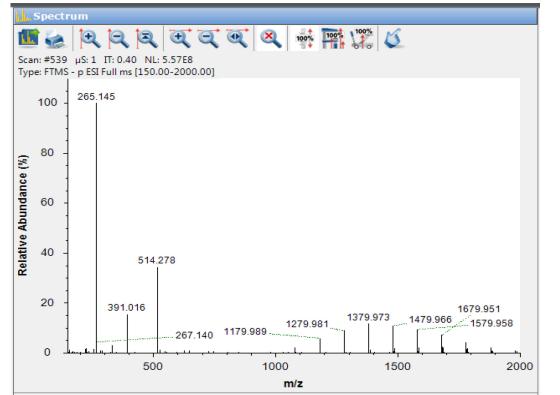
Before you perform measurements in the negative ion mode, the mass spectrometer must be calibrated with a calibration solution containing sodium dodecyl sulfate, sodium taurocholate, and Ultramark 1621. Refer to the Exactive Series Operating Manual for instructions about preparing the negative-ion-mode calibration solution or buying the ready-to-use solution from Thermo Fisher Scientific (refer to www.fishersci.com, catalog number PI88324).

- To calibrate the Q Exactive HF-X mass spectrometer for negative ion mode
- 1. Place a clean 500 μ L Unimetrics syringe with 450 μ L of calibration solution for the negative ion mode into the syringe pump.
- 2. Click the On/Off/Standby button to take the mass spectrometer out of the Standby mode or Off mode and turn it On.

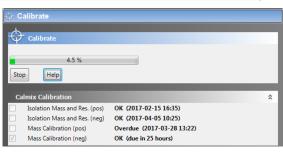




- 3. Click the **Syringe Pump** button to start the syringe pump.
- 4. In the spectrum window of Q Exactive HF-X Tune, observe the mass spectra of the singly-charged ions of the negative ion mode calibration solution. See figure below. The ions are as follows:
 - Sodium dodecyl sulfate: *m/z* 265
 - Sodium taurocholate: *m/z* 514
 - Ultramark 1621: *m/z* 1280, 1380, 1480, 1580, 1680, 1780



- 5. Click the title bar of the Calibrate window to display it.
- 6. In the Calmix Calibration sub-window, select the Mass Calibration (neg) check box to specify a mass calibration for the negative ion mode.
- 7. In the Calibrate window, click **Calibrate** to start the automatic calibration procedure.
- 8. The Calibrate window indicates the completed percentage of the calibration process by a progress bar. See figure right.



Isolation Mass and **Resolution Calibration** If the evaluation of the quadrupole isolation as described on page 7 fails, you must calibrate isolation mass and resolution of the quadrupole. This ensures that the quadrupole isolation performance for the narrow isolation window is sufficient for analyses. The following example describes this calibration procedure for the positive ion

mode.

To calibrate isolation mass and resolution of the quadrupole

1. Place a clean 500 μ L Unimetrics syringe with 450 μ L of calibration solution for the positive ion mode into the syringe pump.

9. A message box informs you when the calibration procedure is completed. Click **OK** to close the message box.

- 2. Click the **On/Off/Standby** button to take the mass spectrometer out of the Standby mode or Off mode and turn it On.
- 3. Click the Syringe Pump button to start the syringe pump.

- 4. Click the title bar of the Calibrate window to display it.
- 5. In the Calmix Calibration sub-window, select the Isolation Mass and Res. check box. See figure right.
- 6. In the Calibrate window, click **Calibrate** to start the automatic calibration procedure.
- 7. The Calibrate window indicates the completed percentage of the calibration process by a progress bar.
- Process by a progress bar.
 A message box informs you when the calibration procedure is completed. Click **OK** to close the message box.

The Q Exactive HF-X mass spectrometer is now ready to perform your measurements.

Use the Evaluate window of the Tasks panel to perform an automatic check of the instrument calibration.

The following example describes an evaluation of the positive mass calibration.

* To evaluate mass calibration for positive ion mode

- 1. Place a clean 500 μ L Unimetrics syringe with 450 μ L of calibration solution for positive ion mode into the syringe pump.
- 2. Click the **On/Off/Standby** button to take the mass spectrometer out of the Standby mode or Off mode and turn it On.
- 3. Click the Syringe Pump button to start the syringe pump.
- 4. Click the title bar of the Evaluate window to display it.
- 5. In the Calmix Evaluation sub-window, select the Mass Check (pos) check box to evaluate the mass calibration for the positive ion mode. See figure right.
- 6. In the Evaluate window, click **Evaluate** to start the automatic evaluation procedure.
- 7. The Evaluate window indicates the completed percentage of the evaluation process by a progress bar.
- 8. A message box informs you when the evaluation procedure is completed. Click **OK** to close the message box.

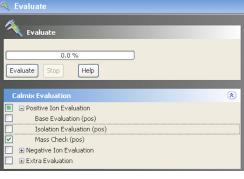
If this evaluation fails, calibrate the Q Exactive HF-X mass spectrometer as described on page 4.

This example describes the evaluation of the isolation calibration for the positive ion mode.

* To evaluate the isolation calibration for positive ion mode

- 1. Place a clean 500 μL Unimetrics syringe with 450 μL of calibration solution for positive ion mode into the syringe pump.
- 2. Click the **On/Off/Standby** button to take the mass spectrometer out of the Standby mode or Off mode and turn it On.
- 3. Click the **Syringe Pump** button to start the syringe pump.
- 4. Click the title bar of the Evaluate window to display it.
- 5. In the Calmix Evaluation sub-window, select the Isolation Evaluation (pos) check box. See figure right.
- 6. In the Evaluate window, click **Evaluate** to start the automatic evaluation procedure.
- 7. The Evaluate window indicates the completed percentage of the evaluation process by a progress bar.
- 8. A message box informs you when the evaluation procedure is completed. Click **OK** to close the message box.

2	Evaluate
2	🔍 Evaluate
E	0.0 % /aluate Stop Help
C	almix Evaluation
	Positive Ion Evaluation Base Evaluation (pos) Isolation Evaluation (pos)
	Mass Check (pos) Regative Ion Evaluation



🔶 Calibrate	
🕁 Calibrate	
0.0 %	
Calibrate Stop Help	
Calmix Calibration	*
Base Calibration	OK (2017-02-06 14:26)
📝 🗄 Isolation Mass and Res. (pos)	OK (2017-03-16 16:44)
Isolation Mass and Res. (neg)	OK (2017-03-08 17:54)
Mass Calibration (pos)	Overdue (2017-03-30 15:51)
Mass Calibration (neg)	Overdue (2017-03-08 17:46)



Evaluation

Evaluation of

Positive Mass Calibration



Evaluation of

Isolation Calibration If this evaluation fails, please perform an "Isolation Mass and Resolution" calibration as described on page 6.

Other Experiment Types

Performing SIM Scans In the previous examples, the Q Exactive HF-X mass spectrometer operates in the full scan mode. The following examples show the mass spectrometer operating in other scan modes.

In the following example, the quadrupole of the Q Exactive HF-X mass spectrometer works as mass filter and is used to perform a Selected Ion Monitoring (SIM) scan of a specified *m/z* range.

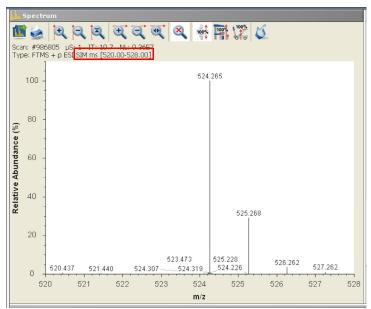
To perform a SIM scan

- 1. If necessary, calibrate your Q Exactive HF-X mass spectrometer as described above.
- 2. Set the scan parameters in the Scan parameters window:
 - a. Click into the Scan Range field to display the Scan Range dialog box.
 - b. Enter a value for the Center mass (for example, m/z 524). See figure right.
 - c. Enter a width not larger than 10 amu (for example, *m/z* 8).

Note that the scan type changes to SIM.

- d. If necessary, adjust the other scan parameters according to your requirements.
- e. Click **Apply** or select the Hot link check box.
- 3. Perform the data acquisition as described on page 11.

The figure below shows the typical spectrum of a SIM view (m/z 524 with 8 amu width).



Performing AIF Experiments

The Q Exactive HF-X mass spectrometer allows performing All Ion Fragmentation (AIF) measurements. The following example describes an AIF fragmentation experiment.

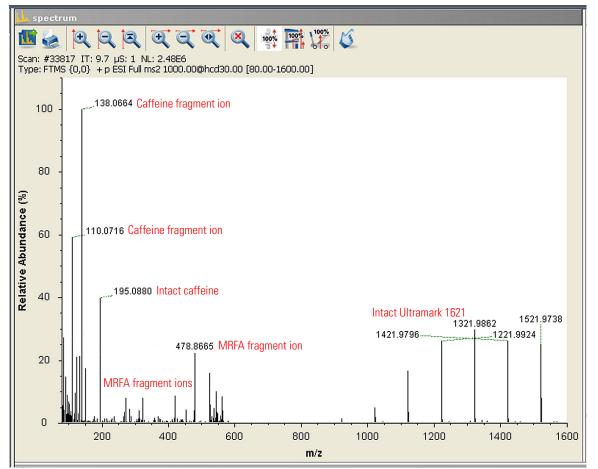
* To set the scan parameters for an AIF fragmentation experiment with NCE

1. If it is necessary, calibrate your Q Exactive HF-X mass spectrometer as described above.

M/VIV Scan param	eters		
History	\rightarrow		
Scan type	SIM		
Scan range	520.0 to 3	528.0 m/z	
Fragmentation	Minimum	520.0 🗢 m/z	
Resolution	Maximum	528.0 🍮 m/z	
Polarity	Maximum	02010 V III/2	
Microscans	Center	524.0 🛟 m/z	
Lock masses	Center		
AGC target	Width	8.0 😂 m/z	
Maximum inject time	30		
Apply Help	🗹 Hot link		

- 2. Set the scan parameters in the Scan parameters window:
 - a. In the Scan type field, select AIF-MS/MS.
 - b. Click into the Scan Range field to display the Scan Range dialog box.
 - c. Adjust the scan range according to your requirements (from *m/z* 150 to *m/z* 2000, for example). See figure right.
 - d. Click into the Fragmentation field to display the Fragmentation dialog box.
 - e. Select the NCE option button and enter the value for the Normalized Collision Energy (35, for example) into the spin box. See figure right.
 - f. Set the charge state appropriately, if necessary.
 - g. If necessary, adjust the other scan parameters according to your requirements.
 - h. Click **Apply** or select the Hot link check box.
- 3. Perform the data acquisition as described on page 11.

The figure below shows a typical spectrum of an AIF fragmentation experiment.



Performing MS/MS Scans

The following example shows an MS/MS experiment. Precursor ions that were selected with the quadrupole are sent to the HCD collision cell of the Q Exactive HF-X mass spectrometer. Here, they are fragmented with Normalized Collision Energy (NCE) or Collision Energy (CE)—selectable by the user.

To perform an MS/MS experiment

1. If it is necessary, calibrate your Q Exactive HF-X mass spectrometer as described above.

💦 Scan param	otors		
ya v Scan param	eters		
listory	\rightarrow		
ican type	AIF 150.0	– 2,000.0 m/z	
ican range	150.0 to 2	,000.0 m/z	
ragmentation	Minimum	\$50.0 🗢 m/	lz
tesolution			.
olarity	Maximum	2000.0 😂 m/	z
Aicroscans			. 1
ock masses	Center	1075.0 😂 mj	z
AGC target	Width	1850.0 😂 m/	lz I
	30		
Aaximum inject time	30		
Apply Help	Hot link		

Fragmentation	NCE 35.0 (z=1)	
Resolution	🔲 In-source CID	25.0 🔶 eV
Polarity	🔽 HCD	
Microscans	NCE	35 🌩
Lock masses	0.00	
AGC target	© CE	•
Maximum inject time	Charge	1
Annha		

- 2. Set the scan parameters in the Scan parameters window:
 - a. In the Scan type field, select AIF-MS/MS. See figure right.
 - b. In the Isolation area, enter a value for the precursor mass (m/z 524, for example).
 - c. Enter a width not larger than 50 amu (m/z 8, for example).

Note that the scan type changes to MS/MS.

- d. Click into the Fragmentation field to display the Fragmentation dialog box.
- Select the NCE (or CE) option button and enter the desired value for e. the collision energy (45, for example) into the spin box. See figure right.
- Set the Charge state appropriately, if necessary. f.
- Click into the Scan Range field to display the Scan Range dialog box. g.
- h. Adjust the scan range according to your requirements (from m/z 50 to m/z 600, for example). See figure right.
- i. If necessary, adjust the other scan parameters according to your requirements.
- j. Click **Apply** or select the Hot link check box.
- 3. Perform the data acquisition as described on page 11.

The figure below shows an example of a typical MS/MS spectrum.

🔊 Scan param	Scan parameters				
History	\rightarrow				
Scan type	M5/M5 of	M5/M5 of 524.0 ± 4.0 m/z			
Scan range	O Full MS -	🔘 Full MS – SIM			
Fragmentation	AIF – M:	AIF – MS/MS			
Resolution					
Polarity	Isolation:	Isolation:			
Microscans	Minimum	520.0 😂 m	n/z		
Lock masses	Maximum	528.0 😂 m	n/z		
AGC target					
Maximum inject time	Precursor	524.0 😂 m	n/z		
Apply Help	Width	8.0 🗘 m	n/z		

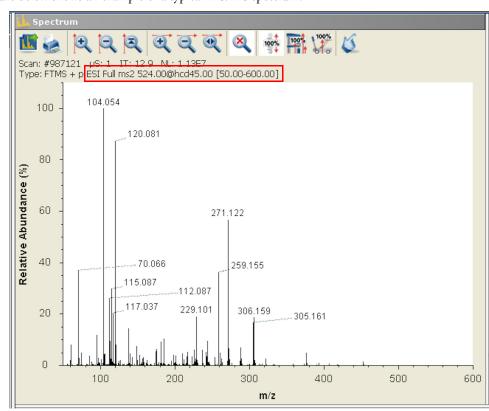
			-		
Scar	an type MS/I		VIS of 524.0 ± 4.0 m/z		
Scar	n range	50.0 to 600.0 m/z			
Frag	mentation	NCE	45.0 (z=1)		
Reso	olution	l 🗐	n-source CID	25.0 🔷 eV	
Pola	rity	V F	ICD		
Mic	roscans				
Lock	masses				
AGC target			⊙ CE		
Max	imum inject time		harge	1	
	Annhe	1			
	Scan type		M5/M5 of	524.0 ± 4.0 m/z	
	Scan range		50.0 to 60	0.0 m/z	
	Fragmentation		Minimum	50.0 🗢 m/z	
	Resolution		Maximum	600.0 🛟 m/z	
	Polarity		Maximum	00010 🗸 111/2	
	Microscans		Contor	325.0 🌨 m/z	
	t a di manana		Center	325.0 😂 m/z	

Width

550.0 😂 m/z

Lock masses

AGC target



Data Acquisition

After you have entered the scan parameters of your experiment, you can finally start to acquire data.

To perform data acquisition

- 1. Specify the acquisition parameters in the Acquisition window:
 - a. In the Destination file field, type the full path (Drive:\path\file name) of the raw file that is used to store the acquired data. Alternatively, click the button to the right of the text box to open a dialog box to browse your directories. See figure right.
 - b. Click into the Acquisition time field to open the dialog box. Select the continuously option button to specify that the acquisition will continue until you click Stop. The Acquisition time field shows continuously.
- 2. Click the **On/Off/Standby** button to take the mass spectrometer out of the Standby mode or Off mode and turn it On.
- 3. Click the Syringe Pump button to start the syringe pump. 4. In the Acquisition window, click **Start** to initiate the data
- acquisition. The spectrum window shows a real-time display of the mass spectrum. The data are saved in the selected raw file.
- 5. Click **Stop** to stop the acquisition.

The following example describes how to create an instrument method in the Instrument Setup view of Xcalibur. You can then use the instrument method to acquire data with the Q Exactive HF-X mass spectrometer. The tune file that is used in the instrument method is used for data acquisition. In the Q Exactive HF-X Tune software, make sure that your tune file has appropriate settings and save it accordingly.

To set up an instrument method

- 1. Display the Instrument Setup page in Xcalibur:
 - a. Choose Start > Programs > Thermo Xcalibur > Xcalibur to display the Xcalibur Home Page window.
 - b. Choose GoTo > Instrument Setup to display the Instrument Setup window.
 - c. If necessary, click **Q Exactive HF-X Orbitrap MS** in the View bar to display the Method Editor.
- 2. In the Method Editor, drag an experiment symbol (for example, Full MS SIM) from the Workflows pane (bottom left) to the gray bar in the Graph pane. A corresponding time bar is displayed in the Scan Groups pane.
- 3. Edit the fields of the Properties pane (right) to set up the experiment parameters. Refer to the Q Exactive HF-X Software Manual or the Q Exactive HF-X Tune Help for a description of the available parameters.
- 4. Assign a tune file:
 - a. On the External Hardware pane, click the (\checkmark) **Tune Files** icon to expand the Tune Files pane.
 - b. In the Properties of Tune Files pane, click the Base Tunefile field.
 - c. Click the induction to open a dialog box.
 - d. Browse for the tune file (for example, ab.mstune) and assign it to the experiment.
- 5. Click 📕 on the toolbar to save the new instrument method:
 - a. In the Save As dialog box, enter the name (Full MS.meth, for example) and the location for your instrument method. Then click Save.
 - b. In the File Summary Information Dialog Box, enter a comment as header information for your instrument method. Then click OK to close the dialog box.

The instrument method can now be used to acquire data with your Q Exactive HF-X mass spectrometer, either from the Xcalibur Sequence Setup or from the Q Exactive HF-X Tune window. The next figure shows the example of an instrument method that was set up as described above.

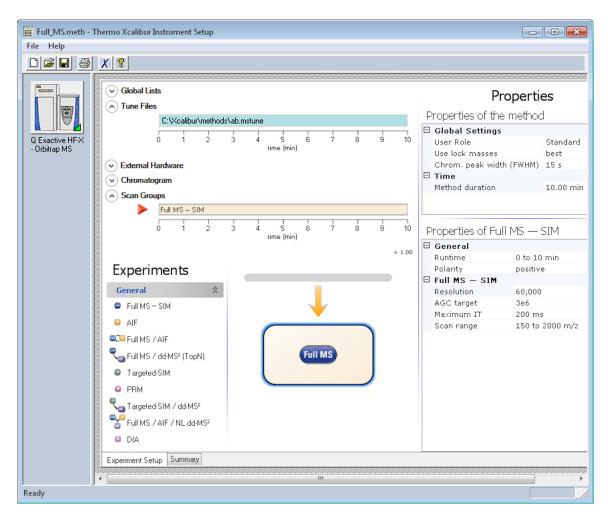
💵 Acquisitio	n	*
Acquisition state	off	
Progress	0.00 min	
File in use	unknown	12
Destination file	C:\Xcalibur\data\ESI_HCD	8
Method file	by time	12
Acquisition time	continuously	
Sample		
Comment		
On start	don't wait	
After acquisition	stay on	
Start	Pause Help	





Setting up and Running **Methods**

Creating the Instrument Method



Running the MS

This section shows an example of how to use an instrument method to run the Q Exactive HF-X mass spectrometer.

To run the Q Exactive HF-X mass spectrometer with an instrument method

- 1. Specify the acquisition parameters in the Acquisition window of Q Exactive HF-X Tune, as follows:
 - a. In the Destination file field, type the full path (Drive:\path\file name) of the raw file that is used to store the acquired data. See figure right.
 - b. In the Method file field, click the *button* to the right of the field to browse the computer for the instrument method file that was created in the previous topic. The Acquisition time field shows *by method*.
- 2. Click the **On/Off/Standby** button to take the mass spectrometer out of the Standby mode or Off mode and turn it On.
- 3. Click the **Syringe Pump** button to start the syringe pump or start a separate pump for infusion experiments.
- 4. In the Acquisition window, click **Start** to initiate the data acquisition:
 - The Start button is replaced by a Stop button.
 - The Acquisition state changes to *running*.
 - A progress bar indicates the completed percentage of the acquisition. See figure right.

🔟 Acquisitio	n	\$	
Acquisition state	off		
Progress	0.00 min		
File in use	unknown	1	
Destination file	C:\Xcalibur\data\ESI_full+CID	8	
Method file	C:\Xcalibur\methods\ESI_posfull+posCID		
Acquisition time	by method		
Sample			
Comment			
On start	don't wait		
After acquisition	stay on		
Start	Pause Help		

C Acquisiti	DN	۲
Acquisition state	running	
Progress	0.33 min 32.5 %	
File in use	C:\Xcalibur\data\ESI_full+CID.RAW	1
Destination file	C:\Xcalibur\data\ESI_full+CID	
Method file	C:\Xcalibur\methods\ESI_posfull+posCID	1
Acquisition time	by method	





- Icons appear on the toolbar to indicate an ongoing data acquisition.
- If necessary, Q Exactive HF-X Tune appends a time stamp to the name of the destination raw file to prevent overwriting existing files.

When the Scan parameters window and the Spectrum window are monitored, it is possible to view how Q Exactive HF-X Tune changes the values according to the settings defined in the instrument method.

The acquisition ends when the acquisition time defined in the instrument method has expired. To display the raw file with the acquired data in Qual Browser, click the \longrightarrow button in the File in use field.

After Operation

High Mass Range (HMR) Mode When the measurement is finished, Thermo Fisher Scientific recommends that you leave the Q Exactive HF-X mass spectrometer in Standby mode and not in Off mode to provide the best mass accuracy for the next analysis.

This mode of operation enables characterization of intact proteins, in particular mAbs and ADCs in native-like conditions.

NOTICE HMR Mode is only available with License 'High Mass Range.' ▲

To perform an experiment in the HMR mode

- 1. Prepare an HMR calibration solution containing up to 1 mg/mL ammonium hexafluorophosphate (AHFP or NH₄PF₆) in isopropanol/water 50/50%.
- 2. User role 'Advanced': Set the instrument to HMR mode:
 - a. If it is not already open, display the instrument status window.
 - b. In the tree view, expand the Instrument > Control > Settings item.
 - c. Set HMR mode to **On**.
- 3. In the Calmix Calibration window, select the HMR Mode Calibration (pos) check box.

Calmix Calibration	\$
🔲 🕀 Base Calibration	OK (2015-12-1415:28)
📃 🕀 Isolation Mass and Res. (pos)	OK (2016-02-08 16:27)
📃 🕀 Isolation Mass and Res. (neg)	Due (void)
Mass Calibration (pos)	Overdue (2016-02-08
Mass Calibration (neg)	Overdue (2015-12-14
📝 😑 HMR Mode Calibration (pos)	Due (default values)
HMR eFT Parameter (pos)	Due (default values)
HMR Mass Calibration (pos)	Due (default values)

The following parameter values are used for this calibration:

Scan parameters window		HESI source window	
Scan range	630.0 to 6670.0 m/z	Sheath gas flow rate	5
In-source CID	100 eV	Spray voltage	3.8 kV
Resolution	120000	Capillary temperature	320 °C
AGC target	3e6	Funnel RF level	150

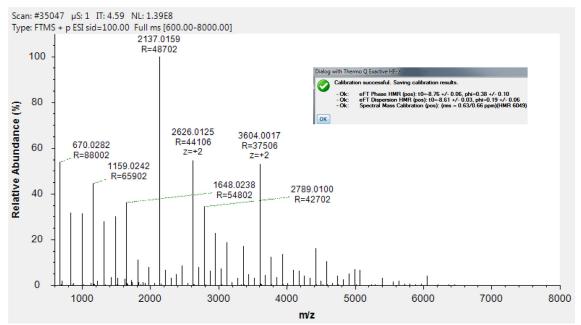
History		
Scan type	Full MS	
Scan range	630.0 to 6,670.0 m/z	
Fragmentation	In-source CID 100.0 eV	
Resolution	120,000	
Polarity	Positive	
Microscans	1	
Lock masses	Off	
AGC target	3e6	
Maximum inject time	20	

HESI source		*
		actua
Sheath gas flow rate	5	
Aux gas flow rate	0	
Sweep gas flow rate	0	
Spray voltage (kV)	3.80	
Spray current (µA)		
Capillary temp. (°C)	320	
Funnel RF level	150.0	
Aux gas heater temp (*C)	30	
Source Auto-Defaults		
Apply Help	🔽 Hot link	

- 4. Start injecting the AHFP solution.
- 5. In the Calibrate window, click **Calibrate** to start the calibration.

NOTICE The high concentration of the AHFP solution might cause contamination of the ion source. Try to spray this solution as short as possible. Rinse and spray the source with pure solvent or calmix right afterwards.

An example of a measurement of an AHFP solution is given in the figure below.



With active High Mass Range license, two additional experiments are available in Method Editor: HMR - Full MS and HMR - AIF. See the figure below.

Experiments

