

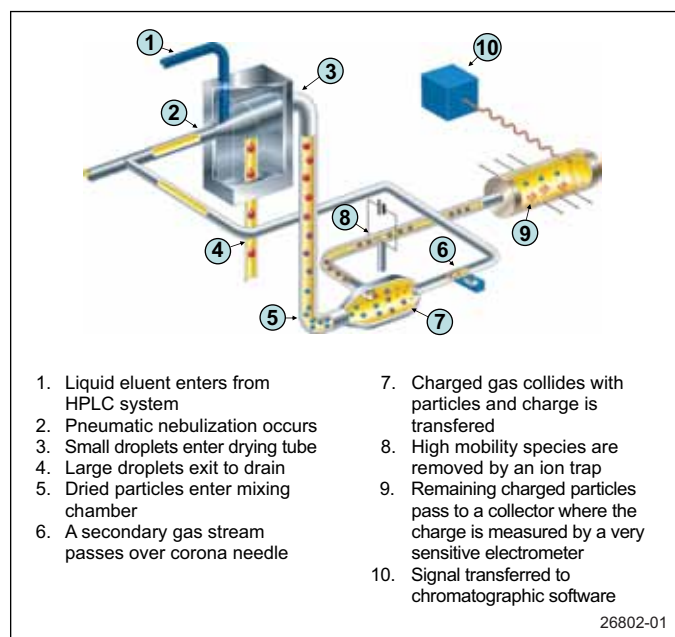
# Determination of the Composition of Natural Products by HPLC with Charged Aerosol Detection

Ian Acworth, Bruce Bailey, Paul Gamache, and John Waraska  
Thermo Fisher Scientific, Chelmsford, MA, USA

## Introduction

Natural products contain a great diversity of compounds that can show extreme variation in their physicochemical properties. Analysis of active components can be challenging, as not all contain a chromophore or can be ionized, thereby limiting the use of UV absorbance and mass spectrometry, respectively. Charged aerosol detection is a sensitive, universal (nonselective) approach that can measure any nonvolatile and many semivolatile compounds. A number of isocratic and gradient HPLC-Thermo Scientific Dionex Corona™ CAD™ Charged Aerosol Detector methods have been developed and evaluated for the measurement of analytes from a variety of natural products, including black cohosh, ginkgo biloba, ginseng, soy, and stevia. Analytes showed consistent response independent of chemical structure (typically < 10% variability between compounds). All methods had a wide dynamic range (four orders of magnitude), good sensitivity (typically low ng levels of detection), and excellent reproducibility (RSDs typically < 2%) even at low detection levels.

FIGURE 1. Charged aerosol detection principle.



## Black Cohosh

### Corona Detector Parameters

Gas: 35 psi using nitrogen generator  
Filter: None  
Range: 100 pA

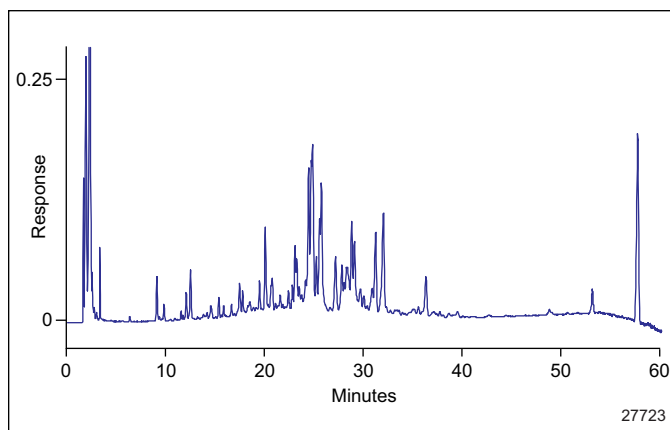
### HPLC Parameters

Mobile Phase A: 0.1% Formic acid  
Mobile Phase B: 100% Acetonitrile  
Gradient: Time (min) 0 60 65 66 80  
                  % A 80 0 0 80 80  
                  % B 20 100 100 20 20  
Flow Rate: 1.0 mL/min  
Column: Shiseido MG 100 C18,  
          4.6 × 250 mm, 5 μm  
Column Temp.: Ambient  
Injection Volume: 10 μL

### Sample Preparation

The sample (1 g powder from capsule) was sonicated in 10 mL methanol for 30 min. The mixture was then passed through a Centrex™ 0.2 μm nylon filter by centrifugation.

FIGURE 2. Separation of metabolites in black cohosh.



## Ginseng

### Corona Detector Parameters

Gas: 35 psi using nitrogen generator  
Filter: None  
Range: 100 pA

### HPLC Parameters

Mobile Phase A: Water  
Mobile Phase B: Acetonitrile  
Gradient: 

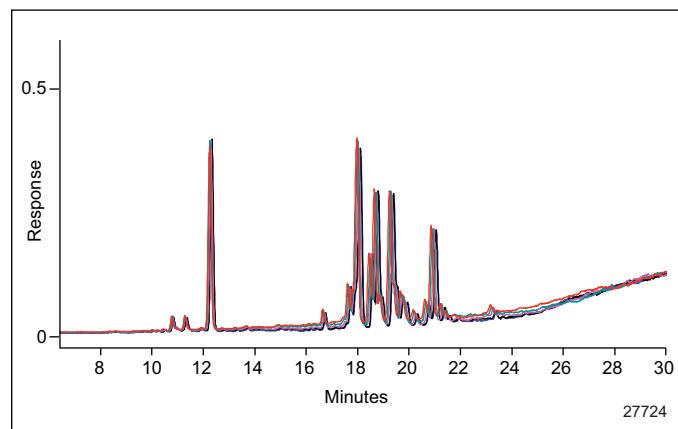
Time (min)	0	30.00	30.01	35.00
% A	80	40	80	80
% B	20	60	20	20

  
Flow Rate: 0.8 mL/min  
Column: Shiseido MG 100 C18,  
4.6 × 250 mm, 5 μm  
Column Temp.: Ambient  
Injection Volume: 10 μL

### Sample Preparation

The sample (1 g powder from capsule) was sonicated in 10 mL methanol for 30 min. The mixture was then passed through a Centrex 0.2 μm nylon filter by centrifugation.

**FIGURE 3. Reproducibility of ginseng analysis (five replicates).**



## Ginkgo Biloba

### Corona Detector Parameters

Gas: 35 psi using nitrogen generator  
Filter: None  
Range: 100 pA

### HPLC Parameters

Mobile Phase A: 5% Acetonitrile in 0.1% trifluoroacetic acid  
Mobile Phase B: 70% Acetonitrile in 0.1% trifluoroacetic acid  
Gradient: 

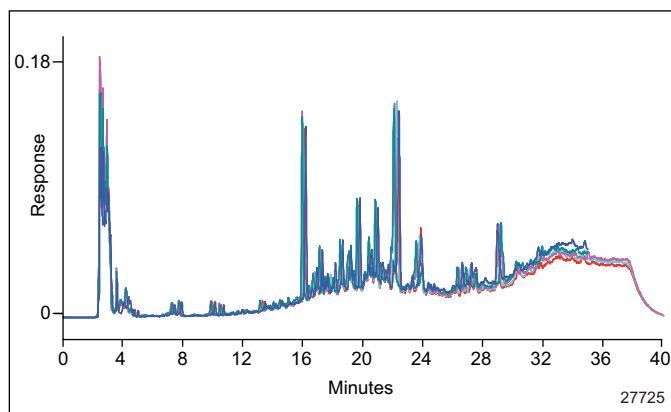
Time (min)	0	30	35	40
% A	100	25	25	100
% B	0	75	75	0

  
Flow Rate: 1.0 mL/min  
Column: Shiseido MG 100 C18,  
4.6 × 250 mm, 5 μm  
Column Temp.: Ambient  
Injection Volume: 10 μL

### Sample Preparation

The sample (1 g powder from capsule) was sonicated in 10 mL methanol for 30 min. The mixture was then passed through a Centrex 0.2 μm nylon filter by centrifugation.

**FIGURE 4. Reproducibility of ginkgo biloba analysis (five replicates).**



## Soy Saponins

### Corona Detector Parameters

Gas: 35 psi using nitrogen generator  
Filter: None  
Range: 100 pA

### HPLC Parameters

Mobile Phase A: 0.1% Trifluoroacetic acid  
Mobile Phase B: 100 % Acetonitrile  
Gradient: 

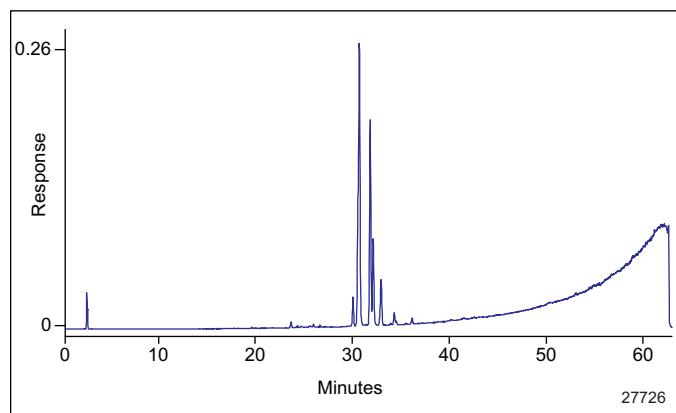
Time (min)	0	60	65
% A	90	10	90
% B	10	90	10

  
Flow Rate: 1.0 mL/min  
Column: Shiseido MG 100 C18,  
4.6 × 250 mm, 5 μm  
Column Temp.: Ambient  
Injection Volume: 10 μL

### Sample Preparation

The sample (50 mg powder) was dissolved in 25 mL water in a 50 mL volumetric flask. The mixture was sonicated for 10–15 min. Ethanol (24 mL) was then added and the now warm solution was cooled. The solution was brought to 50 mL by the addition of ethanol. The solution was then passed through a Centrex 0.2 μm nylon filter by centrifugation.

FIGURE 5. Separation and detection of soy saponins.



## Phytoestrogens—Isoflavones

### Corona Detector Parameters

Gas: 35 psi using nitrogen generator  
Filter: None  
Range: 100 pA

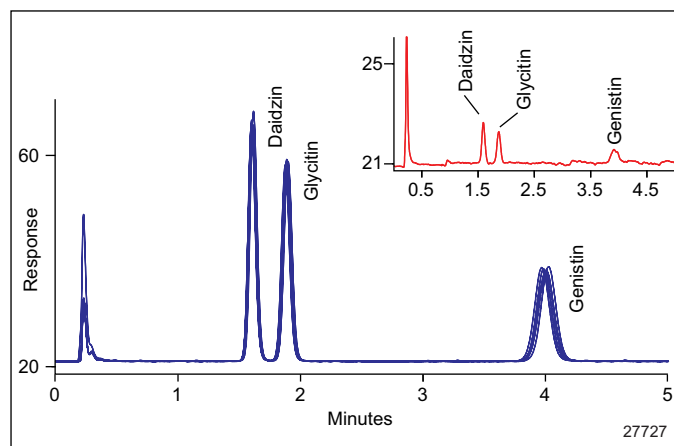
### HPLC Parameters

Mobile Phase: 15% (v/v) Acetonitrile in  
0.1% (v/v) acetic acid  
Flow Rate: 1.5 mL/min  
Column: Shiseido MG 100 C18,  
4.6 × 35 mm, 3 μm  
Column Temp.: Ambient  
Injection Volume: 10 μL

### Sample Preparation

Standards (1 mg/mL) were prepared in DMSO. Dilutions were made in mobile phase

FIGURE 6. Phytoestrogen standards (200 ng each on column, five replicate injections). Inset: 10 ng injections of each standard.



## Phytoestrogens—Coumestans and Mammalian Lignans

### Corona Detector Parameters

Gas: 35 psi using nitrogen generator  
 Filter: None  
 Range: 100 pA

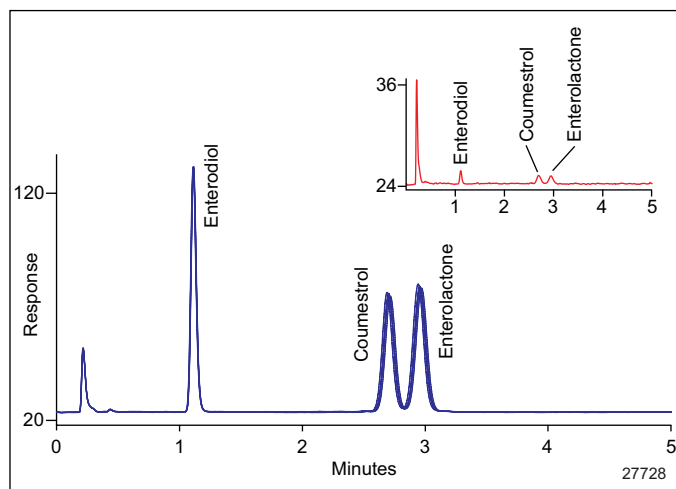
### HPLC Parameters

Mobile Phase: 30% (v/v) Acetonitrile in 0.1% (v/v) acetic acid  
 Flow Rate: 1.5 mL/min  
 Column: Shiseido MG 100 C18, 4.6 × 35 mm, 3 μm  
 Column Temp.: Ambient  
 Injection Volume: 10 μL

### Sample Preparation

Standards (1 mg/mL) were prepared in DMSO. Dilutions were made in 20% acetonitrile.

**FIGURE 7.** Phytoestrogen standards (200 ng each on column, five replicate injections). Inset: 5 ng injections of each standard.



## Stevia

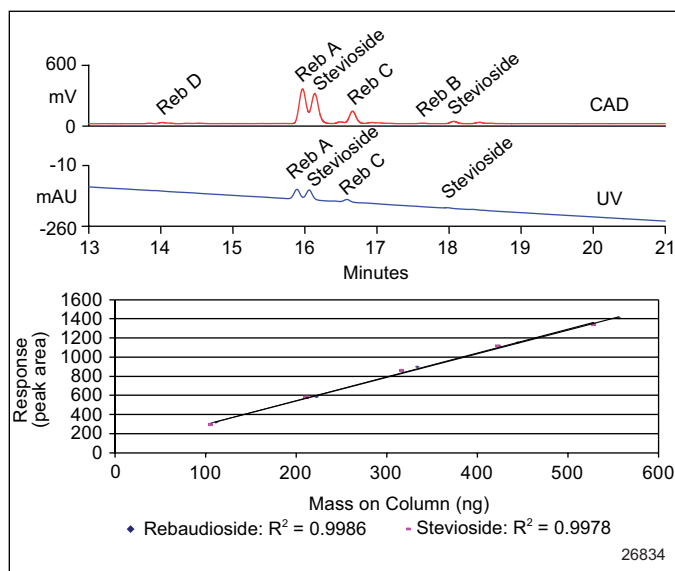
### Corona Detector Parameters

Gas: 35 psi using nitrogen generator  
 Filter: None  
 Range: 100 pA

### HPLC Parameters

Mobile Phase A: DI water, acetonitrile, TFA (95:5:0.1)  
 Mobile Phase B: Acetonitrile, DI water (95:5)  
 Gradient: Time (min) 0 3 30 35 40 45  
           % A 95 95 10 10 95 95  
           % B 5 5 90 90 5 5  
 Flow Rate: 1.0 mL/min  
 Column: Shiseido MG 100 C18, 4.6 × 250 mm, 5 μm  
 Column Temp.: Ambient  
 Injection Volume: 10 μL  
 UV Detection: 210 nm

**FIGURE 8.** Selected portion of chromatogram of SweetLeaf® Stevia Extract at ~860 ng on column run with UV at 210 nm and the Charged Aerosol Detector in series (top). Overlay of response curves for rebaudioside A (Reb A) and stevioside from ~500 to 100 ng on column each (bottom). Average of 3 injections, each fit to a linear regression.



Detector	Linear Correlation Coefficients	
	Rebaudioside A	Stevioside
CAD	0.9986	0.9978
UV at 210	0.9992	0.9994

Detector	Limit of Detection, s/n = 3 (mass on column)		
	Rebaudioside A	Stevioside	Isosteviol
CAD	4 ng	4 ng	60 ng
UV at 210	65 ng	65 ng	> 900 ng

## Conclusions

The need for universal HPLC detection in analytical laboratories is widespread. While several detection technologies (e.g., low wavelength UV, refractive index, evaporative light scattering, chemiluminescent nitrogen detectors) are currently being used, there is significant room for improvement in performance characteristics (i.e., sensitivity, dynamic range, consistency of response factors, and gradient or solvent compatibility). The Corona CAD detector was developed to help address the many challenges of universal detection. This novel technology offers many benefits to analytical scientists, including universal detection of nonvolatile analytes with response independent of chemical properties, a wide dynamic response range, high sensitivity, and good precision. These characteristics, along with reliability and simple operation, make this a superior detector for the measurement of numerous natural products.

©2011 Thermo Fisher Scientific, Inc.  
 SweetLeaf is a registered trademark of United American Industries, Inc.  
 Centrex is a trademark of Whatman International, Ltd.  
 All other trademarks are the property of Thermo Fisher Scientific Inc. and its subsidiaries.  
 This information is not intended to encourage use of these products in any manners that might infringe the intellectual property rights of others.

**Dionex Products:** 1228 Titan Way, PO Box 3603, Sunnyvale, CA 94088-3603, (408) 737-0700  
**North America:** U.S./Canada (847) 295-7500  
**South America:** Brazil (55) 11 3731 5140  
**Europe:** Austria (43) 616 51 25, Benelux (31) 20 683 9768 (32) 3 353 4294  
 Denmark (45) 36 36 90 90, France (33) 1 39 30 01 10, Germany (49) 61125 991 0  
 Ireland (353) 644 0064, Italy (39) 02 51 62 1267, Sweden (46) 8 473 3380,  
 Switzerland (41) 62 205 9966, United Kingdom (44) 1276 691722  
**Asia Pacific:** Australia (61) 2 9420 5233, China (852) 2428 3282, India (91) 22 2764 2735,  
 Japan (81) 6885 1213, Korea (82) 2 2653 2580, Singapore (65) 6289 1190,  
 Taiwan (886) 2 875 6655