Hydrophilic Interaction Liquid Chromatography: Method Development Approaches

Monica Dolci, Luisa Pereira, and Tony Edge Thermo Fisher Scientific, Runcorn, Cheshire, UK





Abstract

This poster summarizes the results of an investigation into hydrophilic interaction liquid chromatography (HILIC), focusing on the retention mechanisms of eleven stationary phases. Additionally to a column characterization study, the effect on retention of changing acetonitrile content, column temperature, mobile phase pH and buffer concentration was investigated.

Introduction

The ability to retain and separate polar and hydrophilic molecules can be very challenging during method development. HILIC has proved to be a viable alternative for the analysis of polar compounds. Although it has been demonstrated that the organic modifier/aqueous ratio is the predominant factor in providing the necessary separation selectivity in HILIC [1], the choice of stationary phase is also very important. Unfortunately, HILIC materials and the roles of their functional groups are not always well understood and users are often under the impression that HILIC columns are interchangeable.

In this poster we are therefore proposing a HILIC column selection scheme, derived from a characterization regime developed by Tanaka et al. [2] which will allow to classify the HILIC retention and selectivity properties for Thermo Scientific[™] HILIC chemistries, according to the following :

- Selectivity for hydrophilic and hydrophobic groups, α (OH) and α (CH2);
- Configurational and positional isomer selectivity, α (V/A) and α (2dG/3dG);
- Anion and cation exchange properties, α (AX) and α (CX),
- Evaluation of the acidic-basic nature of the stationary phases, α $(T_{\rm b}/T_{\rm p})$
- Retention factor for uridine, k uridine.

Additionally, we investigated the chromatographic parameters that have a major role in the HILIC selectivity for acidic and basic compounds:

- Organic solvent content;
- Buffer concentration;
- Mobile phase pH;
- · Column temperature.

We will demonstrate how these various steps can help users during HILIC method development.

Methods

Instrumentation: Thermo Scientific[™] Accela[™] UHPLC *Columns* as given in Table1.

Column Characterization Study

Mobile phase: Acetonitrile:ammonium acetate (20 mM in the aqueous portion, pH 4.7) (90:10, v/v). Flow rate: 0.5 mL/min. UV: 254 nm. Inj Vol: 5 μ L. Column T: 30 C.

Chromatographic test mixtures:

- 1. Toluene, uridine, 5-methyluridine, α (CH2).
- 2. Toluene, uridine, 2'-deoxyuridine, α (OH).
- 3. Toluene, adenosine, vidarabine, α (V/A).
- 4. Toluene, 2'-deoxyguanosine, 3'-deoxyguanosine, $\alpha(2dG/3dG)$.
- 5. Toluene, uracil, sodium p-toluenesulfonate, α (AX).
- 6. Toluene, uracil, N,N,N-trimethylphenylammonium chloride, α (CX).
- 7. Toluene, theobromine, theophylline, $\alpha (T_p/T_p)$.

Acetone was used as to marker on Hypercarb.

Chromatographic Parameters Study

Mobile phases: Various mobile phases were prepared by mixing the desired volumes of acetonitrile and stock buffer solutions.

Flow rate: 1.0 mL/min. UV: 228 nm (for acid mixture) + 248 nm (for basic mixture). Inj Vol: 5 μ L. Column T: 30 C. *Basic test mixture:* 1. Uracil, 2. Adenosine, 3. Uridine, 4. Cytosine and 5. Cytidine

Acid test mixture: 1. Salicylamide, 2. Salicylic acid, 3. Aspirin and 4. 3,4 Dyhydroxyphenylacetic acid (dhpa)

TABLE 1. HILIC stationary phases characterized.

** Nanopolymer silica hybrid * Porous Graphitic Carbon

Column Name	Phase type	Column dimension (mm)	Surface area (m ² /g)	Pore size(Å)
Syncronis [™] HILIC (5 µm)	Zwitterion	100 x 4.6	320	100
Hypersil GOLD [™] HILIC (5 µm)	Polyethyleneimine	100 x 4.6	220	175
Hypersil GOLD [™] Silica (5 µm)	Unbonded Silica	100 x 4.6	220	175
Hypersil GOLD [™] Silica (1.9 µm)	Unbonded Silica	100 x 4.6	220	175
Syncronis [™] Silica (5 µm)	Unbonded Silica	100 x 4.6	320	100
Accucore [™] HILIC (2.6 µm)	Unbonded Silica	100 x 4.6	130	80
Acclaim [™] Mixed Mode HILIC-1 (5 µm)	Mixed Mode Diol	150 x 4.6	300	120
Acclaim [™] HILIC-10 (3 µm)	Proprietary	150 x 4.6	300	120
Acclaim [™] Trinity P1 (3 µm)	NSH**	150 x 3.0	100	300
Accucore [™] 150-Amide-HILIC (2.6 µm)	Polyacrylamide	100 x 2.1	80	150
Accucore [™] Urea-HILIC (2.6 µm)	Urea	100 x 2.1	130	80
Hypercarb [™] (5 µm)	PGC*	100 x 4.6	120	250

Results and Discussions

Characterization Study

The separation factors obtained from this study were displayed in radar plots, figure 1.



FIGURE 1. Radar plots for HILIC stationary phases

The degree of ion exchange interactions has the most significant influence on the radar plots shape, leading to the separation of the Thermo Scientific HILIC columns into two types:

TYPE 1

Columns with very little ion exchange interactions. Syncronis HILIC, Accucore-150-Amide-HILIC and Hypercarb show greater retention and better selectivity for all of the test compounds. Small hydrophilicity, but good selectivity are shown by Hypercarb, Accucore Urea-HILIC, Acclaim HILIC-10 and Acclaim Mixed-Mode HILIC-1.

TYPE 2

Columns with ion exchange interactions. The bare silica materials have strong cation exchange ability. Accucore HILIC and Syncronis Silica have greater α (CH2), α (OH) and k uridine selectivity than Hypersil GOLD Silica, due to their different pore size, surface area and particle technology. Acclaim Trinity P1 shows very strong anion exchange activity and so does (to a lesser extent) Hypersil GOLD HILIC.

Chromatographic Parameters Study

The effect of acetonitrile content on retention The percentage of acetonitrile in the mobile phase was varied whilst keeping ammonium acetate concentration constant at 10 mM. The logarithmic capacity factors (ln k) for a model compound (salicylic acid) were plotted against the acetonitrile content (Figure 2).

FIGURE 2. Effect of acetonitrile content on the retention of salicylic acid.



The effect of buffer concentration on retention Separation of the two test mixtures was carried out using mobile phase: acetonitrile:water (90:10, v/v), containing ammonium acetate, whose concentration was varied from 2.5 to 20 mM. The logarithmic capacity factors (ln k) for the two model compounds were plotted against the buffer molar concentrations. Figure 3 shows the data relative to the basic and acid test mixtures obtained on Hypersil GOLD HILIC.

FIGURE 3. Effect of buffer concentration on retention of basic and acidic mixtures, for Hypersil GOLD HILIC.



The effect of mobile phase pH on retention

The pH of a 100 mM ammonium formate stock solution (pH $^{-}$ 6.4) was adjusted with formic acid to pH 3.3, 4.0 and 4.8. Table 2 summarizes the retention data of the model compounds on four representative columns.

TABLE 2. Effect of buffer pH on retention of model compounds.

Column	рН	k salicylic	k aspirin	k cytosine	k cytidine	
Accucore HILIC		aciu				
	3.3	0.21	0.43	3.07	3.49	
	4.0	0.25	1.26	3.22	3.88	The retention is affected
	4.8	0.24	1.61	3.12	3.80	The retention is anected
	6.4	0.26	1.79	3.23	3.98	when the analyte ionisation
		0.70				state changes in the nH
Hypersil GOLD HILIC	3.3	3.73	0.50	3.34	6.50	state changes in the pri
	4.0	3.89	0.54	3.42	6.56	range considered. This is
	4.8	3.98	5.70	3.22	6.02	range considered. This is
	6.4	4.18	5.90	3.26	6.11	the case for aspirin, where
Syncronis HILIC	3.3	1.11	0.97	4.88	8.40	its retention decreases with
	4.0	1.17	2.15	4.90	8.82	
	4.8	1.06	2.60	4.62	8.13	the buffer pH on the silica
	6.4	1.06	2.60	4.68	8.23	materials and for salicylic
						materialo ana for banoyno
Hypersil GOLD Silica	3.3	0.40	0.61	2.41	2.55	acid on Hypersil GOLD HILIC
	4.0	0.43	1.20	2.49	2.76	
	4.8	0.42	1.43	2.41	2.66	
	6.4	0.40	1.50	2.37	2.64	

The effect of column temperature on retention The column temperature was varied from 20 to 70 °C for the separation of salicylic acid and cytosine. Mobile phase: acetonitrile:water (90:10, v/v), containing 10 mM ammonium acetate. Figure 4 shows van't Hoff plots for salicylic acid and for cytosine.

FIGURE 4. Effect of column temperature on retention of salicylic acid and cytosine on four representative columns.



Conclusion

Based on our observations made here, we propose the following procedure to help during HILIC method development:



References

Y. Guo, S. Gaiki, J. Chromatogr. A 1074 (2005) 71.
Y. Kawachi, T. Ikegami, H. Takubo, Y. Ikegami, M. Miyamoto, N. Tanaka, J.Chromatogr. A 1218 (2011) 5903.

www.thermoscientific.com/dionex

©2013 Thermo Fisher Scientific Inc. All rights reserved. ISO is a trademark of the International Standards Organization. All other trademarks are the property of Thermo Fisher Scientific Inc. and its subsidiaries. This information is presented as an example of the capabilities of Thermo Fisher Scientific Inc. products. It is not intended to encourage use of these products in any manners that might infringe the intellectual property rights of others. Specifications, terms and pricing are subject to change. Not all products are available in all countries. Please consult your local sales representative for details.

Australia +61 3 9757 4486 Austria +43 1 333 50 34 0 Belgium +32 53 73 42 41 Brazil +55 11 3731 5140 China +852 2428 3282 Japan +81 6 6885 1213 Korea +82 2 3420 8600 Netherlands +31 76 579 55 55 Singapore +65 6289 1190 Sweden +46 8 473 3380



Switzerland +41 62 205 9966 Taiwan +886 2 8751 6655 UK/Ireland +44 1442 233555 USA and Canada +847 295 7500

