

# A Versatile Workflow to Measure Plasma Renin Activity and Aldosterone for Clinical Research Using Automated On-line Extraction Coupled to LC-MS/MS

## INTRODUCTION

Researchers studying the renin-angiotensin-aldosterone system need efficient ways to measure renin activity and aldosterone concentration in plasma sample preparations. We report a workflow to accomplish this utilizing automated on-line extraction coupled to LC-MS/MS, which simplifies sample preparation.

## METHODS

### Plasma Renin Activity

In order to measure angiotensin I (Ang I), donor plasma specimens (100 µL), calibrators and Lyphochek hypertension markers controls (QCs, Bio-Rad Laboratories, Berkeley, CA) were prepared per Carter et al (1) and incubated at 37°C in the autosampler tray of a Thermo Scientific™ Prelude™ SPLC system.

25 µL injections were made at the beginning of the incubation and between 12 and 17 hours later (overnight) into a Thermo Scientific™ Cyclone™ TurboFlow™ column (0.5 x 50 mm) on the same channel of the Prelude SPLC system.

Extracted Ang I and its internal standard were automatically transferred to a 50 x 2.1 mm analytical column – either a XBridge BEH C18, 5 µm (Waters Corp., Milford, CA) or a Thermo Scientific™ Accucore™ aQ, 2.6 µm column – heated to 30°C for final isolation using a mobile phase gradient from water to methanol, both containing 0.2% formic acid (Figure 1).

**Figure 1. Angiotensin I LC parameters.**

Column 1										Column 2									
Cyclone 0.5x50mm										Accucore aQ 2.6µm, 50x2.1mm									
Loading Pump										Eluting Pump									
Tertiary										Binary									
A H2O + 0.2% Formic Acid										A H2O + 0.2% Formic Acid									
B MeOH + 0.2% Formic Acid										B MeOH + 0.2% Formic Acid									
C 2:2:1 ACN:IPA:Acetone										C									
Step	Start	Sec	Flow	Grad	%A	%B	%C	Tee	Loop	Flow	Grad	%A	%B						
1	0.00	30	1.50	Step	90.0	10.0	-	=====	out	0.50	Step	90.0	10.0						
2	0.50	60	0.20	Step	90.0	10.0	-	T	in	0.40	Step	90.0	10.0						
3	1.50	15	1.00	Step	-	-	100.0	=====	in	0.50	Ramp	70.0	30.0						
4	1.75	15	0.50	Step	100.0	-	-	=====	in	0.50	Ramp	50.0	50.0						
5	2.00	15	1.00	Step	-	-	100.0	=====	in	0.50	Ramp	20.0	80.0						
6	2.25	15	0.50	Step	100.0	-	-	=====	in	0.50	Ramp	5.0	95.0						
7	2.50	30	1.00	Step	50.0	50.0	-	=====	in	0.50	Step	5.0	95.0						
8	3.00	30	1.00	Step	50.0	50.0	-	=====	in	0.50	Step	90.0	10.0						
9	3.50	45	0.70	Step	-	-	100.0	T	out	0.00	Step	90.0	10.0						
10	4.25	15	0.50	Step	90.0	10.0	-	T	out	0.20	Step	90.0	10.0						
11	4.50	60	1.25	Step	90.0	10.0	-	=====	out	0.70	Step	90.0	10.0						

The analytes were eluted into a heated electrospray interface of a Thermo Scientific™ TSQ Endura™ mass spectrometer (MS) and subjected to positive-ion selected-reaction monitoring of their +3 charge states (Figure 2).

**Figure 2. Angiotensin MS/MS parameters**

SRM Table						
Compound	Start Time (min)	End Time (min)	Polarity	Precursor (m/z)	Product (m/z)	Collision Energy (V)
Ang I	0	1.4	Positive	433.2	619.4	20
Ang I	0	1.4	Positive	433.2	647.4	17
IS	0	1.4	Positive	437.3	631.4	22
IS	0	1.4	Positive	437.3	660.4	18
Electro-clean	1.4	1.5	Negative	437.3	660.4	18

Ang I amounts for each sample were measured using a <sup>13</sup>C, <sup>15</sup>N – labeled Ang I internal standard (AnaSpec Inc., Fremont, CA) and calibrators in each batch.

Plasma renin activity of each specimen and QC was expressed as the increase in Ang I formed from initial to final incubation (ng/mL/hr) using the date/time stamp of respective data files.

### Plasma Aldosterone

Donor plasma specimens (150 µL) as well as calibrators and the Lyphocheck QCs were mixed with 50 µL of internal standard solution (2 ng/mL water plus 10% methanol and 2% acetonitrile). 90 µL injections of each sample preparation were made into a Cyclone 0.5 x 50 mm TurboFlow column on the other channel of the Prelude SPLC with aqueous mobile phase (Figure 3.)

**Figure 3. Aldosterone LC parameters.**

Column 1										Column 2									
Two Cyclone 0.5x50mm										Accucore aQ 2.6um, 50x2.1mm									
Loading Pump										Eluting Pump									
Tertiary										Binary									
A H2O + 0.1% Formic Acid										A H2O+5% IPA+ 0.02% HAC & AmOH									
B MeOH pure										B MeOH pure									
C 2:2:1 ACN:IPA:Acetone										C									
Step	Start	Sec	Flow	Grad	%A	%B	%C	Tee	Loop	Flow	Grad	%A	%B						
1	0.00	30	1.50	Step	100.0	-	-	=====	out	0.50	Step	100.0	-						
2	0.50	60	0.20	Step	100.0	-	-	=====	in	0.50	Step	100.0	-						
3	1.50	15	1.00	Step	-	-	100.0	=====	in	0.40	Ramp	70.0	30.0						
4	1.75	15	0.50	Step	100.0	-	-	=====	in	0.40	Ramp	65.0	35.0						
5	2.00	15	1.00	Step	-	-	100.0	=====	in	0.40	Ramp	60.0	40.0						
6	2.25	15	0.50	Step	100.0	-	-	=====	in	0.40	Ramp	50.0	50.0						
7	2.50	30	1.00	Step	40.0	60.0	-	=====	in	0.40	Ramp	20.0	80.0						
8	3.00	30	1.00	Step	40.0	60.0	-	=====	in	0.50	Step	100.0	-						
9	3.50	75	0.90	Step	100.0	-	-	=====	out	0.50	Step	100.0	-						

The analytes were eluted into the heated electrospray interface of the TSQ Endura MS and subjected to negative-ion selected-reaction monitoring (Figure 4). Mobile phase conditions enhanced ionization.

**Figure 4. Aldosterone MS/MS parameters.**

SRM Table						
Compound	Start Time (min)	End Time (min)	Polarity	Precursor (m/z)	Product (m/z)	Collision Energy (V) RF Lens (V)
Aldosterone	0	1.5	Negative	359.2	189.1	19 120
Aldosterone	0	1.5	Negative	359.2	331.05	17 120
IS	0	1.5	Negative	366.25	276.25	23 130
IS	0	1.5	Negative	366.25	338.25	16 130
Electro-clean	1.5	1.6	Positive	366.25	338.25	16 130

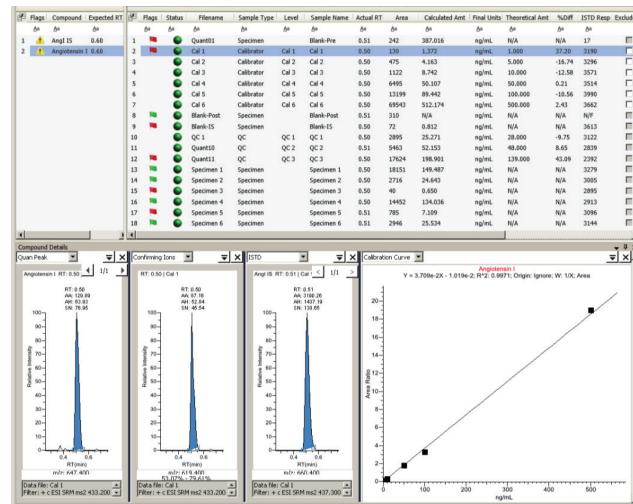
Aldosterone was quantitated using Aldosterone-D<sub>4</sub> internal standard (Cerilliant Corp., Round Rock, TX).

## RESULTS

### Plasma Renin Activity

The Accucore aQ and XBridge BHE columns produced virtually identical results. Figure 5 results involved the Accucore column.

**Figure 5. Typical Angiotensin I quantitation results**



The 5.5 minute method had a throughput of 10 samples per hour, and a linear (1/X weighting) analytical range of 1 to 500 ng/mL, with less than 0.5% carryover. Internal standard (IS) peak areas among calibrators varied no more than 7% in each batch. Matrix interferences among plasma samples presumably caused IS peak area reductions of 20 to 40%.

Overnight incubation provided reliable plasma renin activity (PRA) results (Table 1).

**Table 1. Plasma Renin Activity (PRA) of QCs**

	QC 1	QC 2	QC 3
Initial Ang I (ng/mL)	26	52	198
Final Ang I (ng/mL)	144	270	831
Difference (ng/mL)	118	218	633
PRA (ng/mL/hr)	7.3	13.4	39.0

PRA results for donor plasma samples have not yet been compared to those from a reference lab.

## CONCLUSIONS

Both Ang I and aldosterone research methods provided the desired analytical performance and throughput as they ran independently on a two-channel HPLC system equipped with on-line extraction. Sample preparations were easily accomplished in a matter of minutes. Setting the autosampler tray holder to 37°C conveniently allowed running initial and final-incubation batches to quantitate Ang I and calculate plasma renin activity (PRA). Aldosterone batches conveniently ran before or after Ang I batches. PRA and aldosterone results for each donor specimen were efficiently acquired and reported using this versatile workflow.

## REFERENCES

1. S Carter, LJ Owen, MN Kerstens, RPF Dullaart and BG Keevil. A liquid chromatography tandem mass spectrometry assay for plasma renin activity using online solid-phase extraction. *Ann Clin Biochem* 2012; 49: 570–579.

**For Research Use Only. Not for use in diagnostic procedures.**

Find out more at [www.thermofisher.com](http://www.thermofisher.com)

©2017 Thermo Fisher Scientific Inc. All rights reserved. Lyphocheck is a registered trademark of Bio-Rad Laboratories, Berkeley, CA. XBridge BEH C18 column is a registered trademark of Waters Corp., Milford, MA. All other trademarks are the property of Thermo Fisher Scientific and its subsidiaries. This information is presented as an example of the capabilities of Thermo Fisher Scientific products. It is not intended to encourage use of these products in any manners that might infringe the intellectual property rights of others. Specifications, terms and pricing are subject to change. Not all products are available in all countries. Please consult your local sales representatives for details. **PN65047-EN 0817S**

**ThermoFisher**  
SCIENTIFIC