

Determination of Pesticide Residues in Drinking Water Using Automated SPE and GC-NPD

Aaron Kettle¹, Sergio Guazzotti¹, and Lina Liang², Thermo Fisher Scientific, Sunnyvale, CA, USA¹, Thermo Fisher Scientific, Beijing, China²

ABSTRACT

The Thermo Scientific[™] Dionex[™] AutoTrace[™] 280 Solid-Phase Extraction instrument automates all steps in the solid-phase extraction (SPE) process, including conditioning, loading, rinsing, and eluting. This system extracts organic pollutants from largevolume liquid samples ranging from 20 mL to 4 L and requires only 10 minutes of operator intervention to complete a batch of six samples. This poster provides a complete workflow for organophosphorous pesticides using automated SPE and gas chromatography (GC) with nitrogen phosphorus detection (NPD). Methods for both are provided and can be used immediately by any environmental or food laboratory that would like to test for these compounds in water samples.

INTRODUCTION

Organophosphorous pesticides have been widely used to prevent and treat plant diseases and pests in agricultural products. As a result, excessive residual pesticide content is frequently reported in drinking and surface water sources. A number of these compounds are acutely toxic to human and animal life, making the detection of residual organophosphorous pesticides a growing concern. This work addresses the detection of trace amounts of organophosphorous pesticides in water and optimizes the conditions for solid-phase extraction (SPE) and chromatographic analysis using the Dionex AutoTrace 280 Solid-Phase Extraction instrument and the Thermo Scientific[™] TRACE[™] 1300 Series gas chromatograph with a nitrogen-phosphorus detector (NPD). Eight different types of organophosphorous pesticide residues were detected in drinking water. The results show that this method achieves rapid, simple extraction with small quantities of solvent. In addition, the method has high sensitivity to the eight types of organophosphorous pesticides, a high recovery rate and good linear range.

Figure 2. Blank tap water sample chromatogram.



Figure 3. Spiked tap water sample chromatogram.

MATERIALS AND METHODS

Sample Preparation

Samples consisted of 500 mL of tap water from the Thermo Fisher Scientific Beijing Laboratory. Methanol and NaCl were added to reduce the solubility of the organophosphorous pesticides in water. This enables greater retention during the extraction process and higher recovery of the target analytes. These samples were then spiked with 50 µL of 10 µg/mL mixed standard solution containing eight organophosphorous pesticides (Dichlorvos, Demeton, Phorate, Disulfoton, Dimethoate, Methyl Parathion, Malathion, and Parathion). The sample was purified on a 6 mL SPE cartridge and eluted with ethyl acetate and dichloromethane. The collected liquid was concentrated to 1 mL by nitrogen stream and analyzed using gas chromatography.

SPE Conditions

Six spiked tap water samples were processed automatically on the Dionex AutoTrace 280 Solid-Phase Extraction instrument . All conditioning, loading, rinsing, and eluting were managed by the instrument without operator intervention. The condition solvents were methanol, ethyl acetate, and dichloromethane. The elution solvents were ethyl acetate and dichloromethane.

GC-NPD Conditions

The analysis was performed using a TRACE 1310 gas chromatograph with an SSL injector and a Thermo Scientific[™] AI/AS 1310 Series autosampler. The system configuration and method parameters are listed Table 1, below.

Table 1. System configuration and method parameters.

Parameter	Settings
Chromatography Column	TG-1701 MS, 30 m, 0.25 mm, 0.25 μm
Oven Temperature Program	80 °C (1 min), 20 °C/min to 250 °C (0 min), 5 °C/min
Detector Type and Conditions	Instant Connect NPD: 300 °C Air: 60 mL/min Hydrogen: 2.3 mL/min Make-up gas: Nitrogen 15 mL/min
Carrier Gas Type and Flow Rate	Nitrogen, constant current mode Flow Rate: 1 mL/min
Injection Method and Volume	Splitless, for 1 min, injector temperature 250 °C Sample volume: 1 mL



METHOD PERFORMANCE

A mixed standard solution was added to the water samples (spiked concentrations: 0.2, 0.4, and 1 µg/L) to investigate the spike recovery rates of the eight pesticides. The test results showed that spike recovery rates for each component are between 83% and 100% and are in compliance with routine analysis detection requirements. At the same time, water samples were spiked with 0.2 µg/L, 0.4 µg/L, and 2.0 µg/L and analyzed six times in parallel. The average RSD value was between 2.4% and 8.7%, in compliance with stability requirements. The detection limit for each component was calculated using three times the signal-to-noise ratio. The detection limits for each component were 0.02–0.1 μ g/L as demonstrated in Table 2, below.

Table 2. Detection limits and RSDs for spiked samples.

Compound	Spiked sample 0.2 µg/L Recovery rate (%)	Spiked sample 0.4 µg/L Recovery rate (%)	Spiked sample 1.0 µg/L Recovery rate (%)	Detection limit (µg/L)	RSD (%) (n=6)
Dichlorvos	93.2	88.4	98.6	0.01	4.1
Demeton	88.3	88.5	90.7	0.05	8.7
Phorate	98.7	91.5	95.4	0.05	6.3
Disulfoton	95.3	89.8	95.7	0.01	7.3
Dimethoate	88.5	89.3	99.8	0.01	2.4
Methyl parathion	94.0	90.9	89.3	0.05	3.8
Malathion	90.5	88.8	90.1	0.01	4.4
Parathion	93.2	95.7	96.4	0.01	3.5

RESULTS

Optimization of SPE Conditions

Chinese Method GB5749-2006 "Hygiene Standards for Domestic Drinking Water" requires that residual amounts of eight types of organophosphorous pesticide in drinking water be monitored. Therefore, the retention capacities and elution efficiencies of various SPE columns when extracting these organophosphorous pesticides was investigated. SPE columns tested included silicon, Waters Oasis[®] HLB, Agela Cleanert PEP, Agela Cleanert S C18 and Agela Cleanert C8. The test results showed that the Cleanert PEP column offered the best retention capacity.

In a separate experiment, the elution efficiencies of solvents on the organophosphorous pesticides retained on the Agela PEP SPE column were tested by using different solvents (methanol, ethyl acetate, DCM, and n-hexane). The test results showed that carrying out elution with ethyl acetate/ethyl acetate/dichloromethane (4 mL/4 mL/2 mL) produced the highest elution efficiency with the smallest quantity of solvent. The boiling points of Demeton and Phorate were low, the solubility in water was quite high, and the recovery rate produced by the direct use of SPE was not ideal. However, adding NaCI to the water increased the recovery rate and allows SPE to be used for this method.

Extraction efficiencies were tested by adding 2, 5, and 10 g of NaCl to the water. The recovery rate was highest when adding 5 g NaCl to the water. Figure 1 shows the effect of adding different amounts of NaCl on Phorate recovery rates. Another reason for low Demeton recovery rates is the low boiling point, which makes it degrade easily during testing. Large quantities of Demeton are lost easily during the concentration process. Therefore, lowering the nitrogen blowing temperature to 30 °C during the concentration process and reducing the flow rate to prevent rapid degradation are recommended.

Figure 1. The effect of the quantity of NaCl added on recovery rates.



CONCLUSIONS

- The Dionex AutoTrace 280 Solid-Phase Extraction instrument provides a means to accomplish an efficient, reproducible, and automated extraction for organophosphorous pesticides in environmental waters.
- Use of the TRACE 1310 gas chromatograph with NPD provides low detection limits for eight key organophosphorous compounds $(0.02-1.0 \mu g/L)$ and the excellent recovery ranges (83–100%) at three different spiked concentrations.
- The extraction and analytical methods presented this work meet the requirements of the Chinese EPA Method and can be used by other regulatory agencies for compliance monitoring of organophosphorous compounds in water samples.



Chromatograms and Method Performance

This method compared an unspiked tap water sample to a sample spiked with eight organophosphorous compounds. Both types of samples were run through the entire extraction and analytical procedure as outlined. Figure 2 is the chromatogram for the unspiked tap water blank. As observed, no residual organophosphorous pesticides are present. Figure 3 is the chromatogram for the spiked tap water samples. Each compound is separated according to the expected retention time and the chromatogram provides identifiable, well-resolved peaks for each of the pesticides.





Dionex AutoTrace 280 Solid-Phase Extraction Instrument

TRACE 1310 GC with NPD

TRADEMARKS/LICENSING

© 2016 Thermo Fisher Scientific Inc. All rights reserved. Oasis is a registered trademark of Waters Corporation. All other trademarks are the property of Thermo Fisher Scientific and its subsidiaries. This information is not intended to encourage use of these products in any manner that might infringe the intellectual property rights of others.

> **ThermoFisher** SCIENTIFIC