

Real-Time Library Search on Orbitrap IQ-X Tribrid MS enhances metabolite profiling

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Abstract

Purpose: To demonstrate a use case of the “Real-Time Library Search” intelligent data acquisition strategy on the Thermo Scientific™ Orbitrap IQ-X™ Tribrid™ Mass Spectrometer for enhanced small molecule structure identification.

Methods: Model compounds were incubated in human hepatocytes and metabolites identified using an Orbitrap IQ-X Tribrid mass spectrometer and the Met-IQ workflow via the new “Real-Time Library Search” (RTLS) filter.

Results: The Met-IQ workflow enhanced MS² sampling while simultaneously retaining generation of MS³ scans for ions of interest to assist in the structural characterization of metabolites.

Introduction

Drug metabolite profiling is an integral part of drug discovery. High resolution mass spectrometry with effective data acquisition methodology plays an essential role in the identification and structural characterization of metabolites from compounds of interest.

Here, we present a case study using the Orbitrap IQ-X Tribrid MS, leveraging the real-time decision-making Met-IQ workflow via the new RTLS filter for metabolite profiling. By using RTLS with the parent compound MS² spectral library, we selectively trigger MS² for precursors which display fragmentation similarity with the parent drug, allowing for structural characterization of drug-related compounds, but simultaneously freeing the duty cycle to enhance MS² sampling of additional precursors.

Materials and methods

Sample preparation

The model compounds losartan and montelukast (obtained from Sigma Aldrich) were dissolved and diluted in catabolism buffer at a concentration of 2 μM and incubated at 37°C for 0 and 4 hours in the presence of human hepatocytes. Samples were crashed with an equal volume of cold acetonitrile containing 2% formic acid, centrifuged for 10 minutes at 10°C, and the supernatant transferred to sample vials for LC/MS analysis.

Liquid chromatography

Chromatographic separations were carried out with the Thermo Scientific™ Vanquish™ UHPLC system consisting of:

- Thermo Scientific™ Vanquish™ Binary pump
- Thermo Scientific™ Vanquish™ Autosampler
- Thermo Scientific™ Vanquish™ Column Compartment
- Thermo Scientific™ Vanquish™ Diode Array Detector

LC separations were performed using a Thermo Scientific™ Hypersil™ GOLD C18 column (2.1x100 mm, 1.9 μm) with mobile phases composed of (A) 0.1% formic acid in water and (B) 0.1% formic acid in acetonitrile and gradient elution as described below. Column temp: 40 °C. Injection volume: 15 μl. Flow rate: 0.3 mL/min.

Table 1. LC gradients

Time (min)	0	0.5	9.0	11.0	12.0	13.0	15.0
%B (montelukast)	1	1	75	95	99	1	1
%B (losartan)	1	1	50	95	99	1	1

Mass spectrometry

Mass spectrometric data were acquired on a Orbitrap IQ-X Tribrid Mass Spectrometer via electrospray ionization in positive mode. The HRAM full scan followed by data-dependent product ion scan were collected at resolution settings of 120,000 and 15,000 at FWHM m/z 200 respectively.

Normalized stepped HCD collision energy (%): 20, 40, 60.

Source parameters:
Positive Ion Spray Voltage (V): 3400
Sheath Gas (Arb): 40
Aux. Gas (Arb): 5
Sweep Gas (Arb): 1
Ion Transfer Tube Temp (°C): 300
Vaporizer Temp (°C): 400

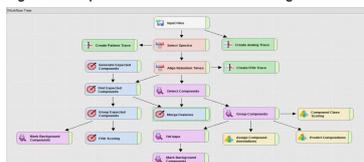
Data processing

The data was processed using Thermo Scientific™ Compound Discoverer™ 3.3 (CD 3.3) software, a small molecule structure analysis software which employs a flexible and customizable node-based processing workflow.

Compound Discoverer 3.3 node-based processing workflow enables the detection of targeted metabolites through biotransformation list, as well as detection of unexpected metabolites using “Compound Class Scoring” node.

In this study, the processing workflow was built using the pre-defined workflow template “MetID v Expected and Unknown w Background Removal”, see Figure 1.

Figure 1. Compound Discoverer Data Processing workflow



Results

Mass spectrometer and novel data acquisition workflows

The Orbitrap IQ-X™ Tribrid™ mass spectrometer is dedicated to small molecule structure analysis. The tribrid architecture with ultra-high field orbitrap analyzer and state of the art hardware guarantee the highest performance. As illustrated by the instrument schematic in Figure 2, this system offers exceptional versatility for enhanced small molecule structural analysis through an array of novel data acquisition workflows, including Met-IQ workflow using real-time library search (RTLS) and AcquireX workflows for automatic background exclusion and in-depth MSⁿ triggering.

Met-IQ workflow using the RTLS filter for metabolite profiling

For small molecule analysis, the Real-Time Library Search (RTLS) filter compares experimental MS² to a spectral library and generates a score based on similarity. Compounds that meet the scoring criteria set in the method can trigger additional scan behaviors, including MS³ or alternate fragmentation. By selectively triggering MS³ only on precursors with spectral similarity to the parent or compound(s) of interest, additional structural information is gained on the most important compounds while maintaining a high MS² scan rate.

In this study, the model compounds' HCD/CID spectra at multiple fragmentation energies were collected using the Library Builder template. The spectral libraries were constructed through mzVault™ spectral library curation software. Existing MassBank record spectral libraries can be converted to the .db mzVault format if applicable. The Met-IQ workflow methods were set up using the RTLS template in method editor, as shown in Figures 3 and 4.

Figure 3. Method setup for mzVault library generation

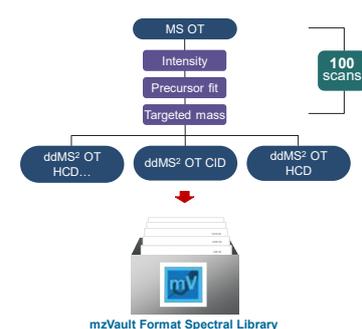
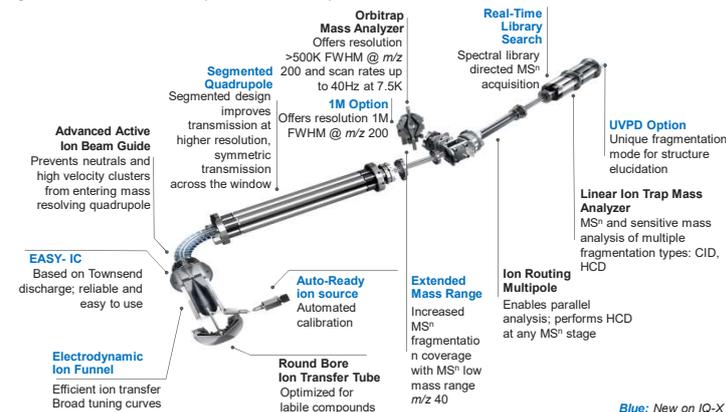


Figure 4. Met-IQ workflow MS method



Figure 2. Schematic of the Orbitrap IQ-X Tribrid Mass Spectrometer



Data processing and performance evaluation

The CD processing results showed that with Met-IQ workflow, more metabolites were detected and triggered MS³ than using conventional DDA acquisition without RTLS. For instance, RTLS data acquisition accurately identified fragmentation level similarity within a low-abundant dealkylated metabolite of losartan. Despite being masked by a background ion's isotope peak, the RTLS filter observed sufficient MS² similarity to trigger MS³ acquisition on the fragment m/z 171.06802. This metabolite was confidently identified based on MS², HRAM and fine isotope pattern, as shown in Figures 5 and 6.

Figure 5. Dealkylated metabolite of Losartan identified by MS² spectral similarity in t=4H RTLS Run

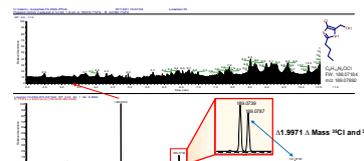
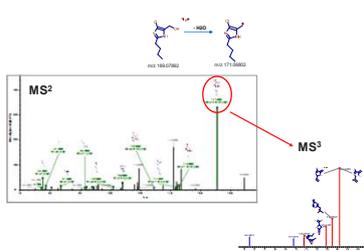


Figure 6. The auto-annotated MS² spectrum in the result view shows the same fragment at m/z 171.06802 as the parent compound. The MS² spectral similarity triggered acquisition of MS³ spectra for the Top3 fragment ions.



By comparison, fewer metabolite MS³ were triggered in DDA runs for compounds related to losartan and montelukast due to the untargeted triggering, especially in the presence of complex biological matrices, as shown in Tables 2 and 3 below. By leveraging RTLS, selective MS³ data acquisition resulted in freed duty cycle time and increased MS² triggering.

Table 2. Overview of 10 identified “Expected Compounds” metabolites of Losartan by DDA

Peak	RT (min)	Formula	Transformation	Compound Charge	Subst. Type	Cell. Mass	MS ²	MS ³	MS ² Coverage	MS ³ Coverage
1	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
2	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
3	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
4	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
5	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
6	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
7	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
8	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
9	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
10	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%

Table 3. Overview of 24 identified “Expected Compounds” metabolites of Losartan with RTLS

Peak	RT (min)	Formula	Transformation	Compound Charge	Subst. Type	Cell. Mass	MS ²	MS ³	MS ² Coverage	MS ³ Coverage
1	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
2	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
3	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
4	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
5	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
6	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
7	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
8	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
9	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
10	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
11	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
12	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
13	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
14	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
15	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
16	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
17	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
18	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
19	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
20	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
21	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
22	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
23	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%
24	1.22	C ₁₆ H ₁₅ N	Losartan	1	1	213.1266	171.0680	171.0680	100%	100%

Table 4. Overview of 2 identified “Expected Compounds” metabolites of Montelukast by DDA

Peak	RT (min)	Formula	Transformation	Compound Charge	Subst. Type	Cell. Mass	MS ²	MS ³	MS ² Coverage	MS ³ Coverage
1	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
2	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%

Table 5. Table 2. Overview of 2 identified “Expected Compounds” metabolites of Montelukast with RTLS

Peak	RT (min)	Formula	Transformation	Compound Charge	Subst. Type	Cell. Mass	MS ²	MS ³	MS ² Coverage	MS ³ Coverage
1	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
2	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
3	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
4	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
5	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
6	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
7	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
8	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
9	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
10	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
11	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
12	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
13	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
14	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
15	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
16	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
17	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
18	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947	171.0680	171.0680	100%	100%
19	1.22	C ₂₃ H ₂₇ N ₂ O ₂ S	Montelukast	1	1	375.1947				