Real-Time Library Search on Orbitrap IQ-X Tribrid MS enhances metabolite profiling

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Abstract

Purpose: To demonstrate a use case of the "Real-Time Library Search" intelligent data accuisition strategy on the Thermo Scientific" Orbitrap (1-X" Tribrid" Mass Spectrometer for enhanced small molecule structure identification.

Methods: Model compounds were incubated in human hepatocytes and metabolites identified using an Orbitrap IQ-X Tribrid mass spectrometer and the Met-IQ workflow via the new "Real-Time Library Search" (RTLS) filter.

Results: The Met-IQ workflow enhanced MS² sampling while simultaneously retaining generation of MS³ scans for ions of interest to assist in the structural characterization of metabolites.

Introduction

Drug metabolite profiling is an integral part of drug discovery. High resolution mass spectrometry with effective data acquisition methodology plays an essential role in the identification and structural characterization of metabolites from compounds of interest.

Here, we present a case study using the Orbitrap IQ-X Tribrid MS, leveraging the real-time decision-making Met-IQ workflow via the new RTLS filter for metabolite profiling. By using RTLs with the parent compound MS² spectral library, we selectively trigger MS³ for precursors which display fragmentation similarly with the parent drug, allowing for structural characterization of drug-related compounds, but simultaneously freeing the duty cycle to enhance Here, we present a case study using the Orbitrap IQ-X Tribrid MS. MS² sampling of additional precursors

Materials and methods

Sample preparation

The model compounds losartan and montelukast (obtained from Sigma Aldrich) were dissolved and diluted in catabolism buffer at a concentration of 2 µM and incubated at 37°C for 0 and 4 hours in the presence of human hepatocytes. Samples were crashed with an equal volume of cold acetonitrile containing 2% formic acid, centrifuged for 10 minutes at 10°C, and the supernatant transferred to sample vials for LC/MS analysis.

Liquid chromatography

Chromatographic separations were carried out with the Thermo Scientific* Vanquish* UHPLC system consisting of. Thermo Scientific* Vanquish* Binary pump Thermo Scientific* Vanquish* Column Compartment Thermo Scientific* Vanquish* Column Compartment Thermo Scientific* Vanquish* Diode Array Detector

LC separations were performed using a Thermo Scientific* Hypersil* GOLD C18 column (2.1x100 mm, 1.9 µm) with mobile phases composed of (A) 0.1% formic acid in water and (8) 0.1% formic acid in acetonitrile and gradient elution as described below Column temp: 40 °C.

Injection volume: 15 µl

Flow rate: 0.3 ml /mir

Table 1. LC gradients

Time (min)		0.5	9.0	11.0	12.0	13.0	15.0
%B (montelukast)	1	1	75	95	99	1	1
%B (losartan)	1	1	50	95	99	1	1

Mass spectrometry

Mass spectrometric data were acquired on a Orbitrap IQ-X Tribrid Mass Spectrometer via electrospray ionization in positive mode. The HRAM full scan followed by data-dependent product ion scan were collected at resolution settings of 120,000 and 15,000 at were collected at resolution s FWHM m/z 200 respectively.

Normalized stepped HCD collision energy (%): 20, 40, 60. Positive Ion Spray Voltage (V): 3400 Sheath Gas (Arb): 40 Aux Gas (Arb): 5 Sweep Gas (Arb): 1 Ion Transfer Tube Temp (°C): 300 Vaporizer Temp (°C): 400

Data processing

The data was processed using Thermo Scientific[™] Compound Discovers[™] 3.3 (CD 3.3) software, a small molecule structure analysis software which employs a flexible and customizable node-based processing workflow.

Compound Discoverer 3.3 node-based processing workflow enables the detection of targeted metabolites through biotransformation list, as well as detection of unexpected metabolites using "Compound Class Scoring" node.

In this study, the processing workflow was built using the pre-defined workflow template "MetID w Expected and Unknown vn w Background Removal", see Figure 1

Figure 1. Compound Discoverer Data Processing workflow



Learn more at thermofisher.com/IQX

Results

Mass spectrometer and novel data acquisition workflows The Orbitrap IQ-X^m Tribrid^{av} mass spectrometer is dedicated to small molecule structure analysis. The tribrid architecture with ultra-high field orbitrap analyzer and state of the art hardware guarantee the highest performance. As illustrated by the instrument schematic in Figure 2, this system offers exceptional versatility for enhanced small molecule structural analysis through an array of novel data acquisition workflows, including Met-IQ workflow using real-time library search (RTLS) and AcquireX workflows for automatic heartment of the structural analysis through an ethol and acquirement before the structural analysis through an ethol and acquires the structural analysis through the structure of the structure of the structural analysis through the structure of the structure workflows for automatic background exclusion and in-depth MSⁿ triggering

Met-IQ workflow using the RTLS filter for metabolite profiling Met-IQ workflow using the RTLS filter for metabolite profiling For small molecule analysis, the Real-Time Library Search (RTLS) filter compares experimental MS² to a spectral library and generates a score based on similarity. Compounds that meet the scoring criteria set in the method can trigger additional scan behaviors, including MS³ or alternate fragmentation. By selectively triggering MS³ only on precursors with spectral similarity to the parent or compound(s) of interest, additional structural information sandie or compound(s) of interest, additional structural informati is gained on the most important compounds while maintaining a high MS² scan rate.

In this study, the model compounds' HCD/CID spectra at multiple fragmentation energies were collected using the Library Builder template. The spectral libraries were constructed through mzVault" spectral library curation software. Existing MassBa record spectral libraries can be converted to the .db mzVault . sBank format if applicable. The Met-IQ workflow methods were set up using the RTLS template in method editor, as shown in Figures 3 and 4.

Figure 3. Method setup for mzVault library gene tion





Figure 4. Met-IQ workflow MS method

Efficient ion transfer Broad tuning curves



Figure 2. Schematic of the Orbitrap IQ-X Tribrid Mass Spectrometer Segmented design improves Real-Time Library Search Spectral library directed MSⁿ 1M Option solution 1M improves transmission at Offers resolution 1M FWHM @ m/z 200 Advanced Active Ion Beam Guide Prevents neutrals and high velocity clusters from entering mass resolving quadrupole higher resolution symmetric transmission across the window . . EASY- IC Based on Townsend discharge; reliable and easy to use Ion Routing Multipole Auto-Ready Extended Mass Ran Enables parallel analysis; performs HCD at any MSⁿ stage Automated Increased MSⁿ calibration ragmentatio Electrodynamic Ion Funnel

Round Bore Ion Transfer Tube Optimized for labile compounds

n coverage with MSⁿ low

mass range *m/z* 40

Data processing and performance evaluation

Data processing and performance evaluation The CD processing results showed that with Met-IQ workflow, more metabolites were detected and triggered MS³ than using conventional DDA acquisition without RTLS. For instance, RTLS data acquisition accurately identified fragmentation ievel similarity within a low-abundant dealkylated metabolite of losartan. Despite being masked by a background lon's isotope peak, the RTLS filter observed sufficient MS² similarity to trigger MS³ acquisition on the fragment *m*² 171.06042. This metabolite was confidently identified based on MS³, HRAM and fine isotope pattern, as shown in Figures 5 and 6.

Figure 5. Dealkylated metabolite of Losartan identified by $\rm MS^2$ spectral similarity in t=4h RTLS Run



Figure 6. The auto-annotated MS² spectrum in the result view ows the same fragment at *m*/z 171.06802 as the parent npound. The MS² spectral similarity triggered acquisition of MS³ spectra for the Top3 fragment ions.



By comparison, fewer metabolite MS3 were triggered in DDA runs for compounds related to losartan and montelukast due to the un targeted triggering, especially in the presence of complex biological matrices, as shown in Tables 2 and 3 below. By leveraging RTLS, selective MS³ data acquisition resulted in freed duty cycle time and increased MS² triggering.

Table 2. Overview of 10 identified "Expected Compounds" metabolites of Losartan by DDA

-									
Peak	RT [min]	formula	Transformations	Composition C	eltaMass (ppn	Calc. MW	m/1	RSh Coverage	MS Dep
1	671	C22H23CIN602	Deidation	10)	025	493.15713	499.16440	30.8	3
2	6.97	(22H21CIN603	Secara ration, Oxidation, Oxidation	0421+)023	-011	452.13636	453,14965	49.5	3
ž	6.98	C22H28CIN602	2x dation	100	0.8	498.15723	499.16652	29.3	3
ĥ	7.44	(22H01CIN603	Secaru ration, Oxidation, Oxidation	0421+)023	017	492.1864	631098	36.4	3
5	2.29	C22H23C1N602	Oxidation	-(O)	0.18	623 15718	679.16656	22.9	3
6	2.96	C22H23C1N6O2	Audation	-(0)	0.18	628 15718	679.16456	27.2	3
7	8.04	C28 H31 C1 N 6 O 8	Oxidation, Gucuronide Conjugation	+(C6 H8:07)	0.13	614.18927	615.19653	62.5	3
8	8.51	C28 HB1 CI N6 07	Glucuronide Conjugation	+(C6 H8:06)	0.36	598.19443	599.20172	19.8	3
	8.64	C22 H23 CI NE 0	Locaritan Parent Compound		0.54	402.16245	423.16970	18.2	3
	9.28	C22 H21 CI N6 02	Decaturation, Oxidation	-(H2)+(0)	0.16	436.11132	437.16880	34.8	- 2
		and the second second second							

Table 3. Overview of 24 identified "Expected Compounds"

Peak	RT (min)	formula	Transformations	Composition Change	Anna. [ppm]	Calc. NW	m/s	HSh Coverage	MS Depth
1	1.16	CEHLECIN2D		(CILHODM)	-1.29	188.07542	289.07870	29.5	1
1	6.22	C22 H28 C MEOR	Dedation, Dedation	+1021	0.19	454 15211	65.1380	63	1
1	6.72	C22 H23 CI MILO2	Desiducion	10	0.26	408.15713	499.36440	36.4	1
4	6.72	C22 H28 C MEOR	Dedation, Dedation	+1021	034	454 15204	65.1388	401	1
1	6.58	C22 H23 CI MILO2	Destation	10	-0.06	438.15729	499.36437	28.1	1
6	6.98	C22 H21 CI NEQ3	Decatoration, Oxidation, Oxidation	(101+)02(-0.14	492.1859	68.3689	43	1
7	2.09	C28 HE1 CI MICR	Deidation, Glucuranide Conjugation	+)C6 H803	-0.36	615.18909	613.296.ID	1003	1
8	7.26	C221H21 CI NEO2	Decaturation, Oxidation	-(10)+(0)	-0.18	46.1637	687.36865	617	1
•	2.44	221421121508008	Decisivation, Oxidation, Oxidation	(10) +(02)	-0.26	021821	03.1079	39.2	1
10	7.81	C22 H28 CI NE 02	Destation	+101	0.25	48 19 11	629,36640	263	1
11	2.80	C28 HE1 CI MACR	Chidation, discurpride Conjugation	+)C6 H80/5	0.19	655.18931	613.29639	46.2	1
12	2.88	C22 H21 CI NEQ3	Decatoration, Oxidation, Oxidation	(101+)02(0.16	492.1864	68,34836	68.0	1
13	2.85	221423111102	Deidation	+)이	0.16	68.15717	679.3500	23.1	1
14	2.86	C22 H2FCI N6	Dehydration, Devaluation	(HED)	028	402.1800	408.16838	57.5	1
15	8.06	C28 HE1 CI MILCOR	Chidation, discurphide Conjugation	+)C\$ H8:0(7)	022	605.1802	\$23.29647	64.3	1
16	8.88	C28HEECI NEOE	Buccode Conjugation	+105 H 30 010	0.21	58122513	585.22241	100.0	1
17	3.17	222 H21 CI NEOR	Deciducation, Oxidation, Oxidation	(101+102)	-0.08	492.13635	63.30%	37.0	1
18	8.51	C28 HE1 CI MILO7	Steparanide Cargagation	+(CEH806)	0.26	598,19631	599.2006	20.8	1
19	8.54	C221421 CI MILO2	Decatorization, Christiation	(10)+(0)	0.34	68.1047	687.36870	63.4	1
	1.64	C22 H23 C MEO	Localtan Parent Compound		0.65	422 16218	013807	201	1
20	8.75	(22 H21 CI MAG)	Decaturation, Oxidation, Oxidation	(H2) +(02)	-0.05	02.1834	63.3692	36.9	1
21	9.28	222 H21 CI MILO2	Decanuation, Oxidation	1921+121	0.22	436.1655	417.34888	26.3	1
22	3.05	C2814291CI No.O.8	Decatoration, Oxidation, Studynomide Con	+105 H6(07)	0.27	612.17971	613.18298		1
23	3.61	C221K21 CI NEO2	Decaturation, Oxidation	-(10)+(0)	0.10	48.1010	687.36877	27.1	1

	RT (min)	formula	frandomations	Composition Charge	Anna. [ppm]	Calc. NW	m/s	HSh Coverage	MS Depth
	3.36	CEHOLCIN2D		-CM ROOM	-1.29	188.07542	299.01830	19.5	1
	6.22	C22 H28 C MEOR	Dedation, Dedation	+1021	0.19	451,15217	65.1380	63	1
	6.72	C22 H23 CI MILO2	Desiducion	10	0.26	48.15713	499.36440	36.4	1
	6.72	C22 H28 C MEOR	Dedation, Dedation	+1021	034	45115201	65.1388	401	1
	6.58	C22 H23 CI MILO2	Destation	10	-0.06	48.15709	499.36437	28.1	1
	6.98	C22 H21 CT MIG R	Decaturation, Oxidation, Oxidation	(92)+(22)	-0.14	452,18632	6113079	42.1	1
1	2.09	C28 HE1 CI NILCOR	Chidation, discurphide Conjugation	+)C8 H8/2/5	-0.36	605.18909	923.29635	1003	1
	2.26	C22 H21 CI MILO2	Decaturation, Oxidation	0121+121	-0.18	48.1037	617.308/5	41.7	1
	2.44	22142112150803	Decinutation, Oxidation, Oxidation	(19) +(02)	-0.26	012.18629	03.1079	39.2	1
	7.81	C22 H23 CT MEO2	Deduction	+101	025	48 15211	699,36680	26.1	1
	2.80	C28 HE1 CI MACR	Chidation, discurpride Conjugation	+)Cá H80/5	0.19	615.18931	613.29639	46.2	1
	2.88	(22 H21 CI MILO 8	Decatoration, Oxidation, Oxidation	(101+)22)	0.16	452.18644	63.1878	68.0	1
	2.85	22142311502	Deidation	+)이	0.16	68.15717	679.35505	23.1	1
	2.86	C22 H29 CI M6	Dehydration, Decaturation	(HEO)	028	452.1802	401.1033	57.1	1
	8.06	C28 HE1 CI MA CR	Chidation, Glucuranide Conjugation	+)CS H80/7	011	655.18922	613.29647	64.3	1
	1.11	C28 HER CI MILOS	Shecoride Conjugation	+)Cá H 32 C/I)	0.21	38122513	585 22242	120.2	1
	8.17	C221H21 CI NEO 8	Decatoration, Oxidation, Oxidation	-(101+)02)	-0.08	492.18635	63.1035	37.0	1
	8.51	C28 HE1 CI MILOT	Steparanide Cargagation	+)C6 H8:C9)	029	598,19031	599.2006	20.8	1
	8.54	C221421 CI MILO2	Decaturation, Oxidation	(H2)+(D)	0.34	68.1047	687.36876	63.6	1
	1.64	C22 H23 C MEO	Localtan Parent Compound		0.65	422.16238	013807	201	1
	8.75	(22 H21 CI MAG)	Decaturation, Oxidation, Oxidation	(102) +(02)	-0.05	452.18534	63.3692	36.9	1
	9.28	222 H21 CI MILO2	Decaturation, Oxidation	1921+121	0.22	48.1655	417.34888	26.3	1
	3.05	C2814291CI NSICR	Decatoration, Oxidation, Studynomide Con	+105 H6(07)	0.27	612.17971	613.18298		1
				10 million					_



..., or multiple fragmentation types: CID, HCD

Blue: New on IQ-X

Table 4. Overview of 2 identified "Expected Compounds" metabolites of Montelukast by DDA



Table 5. Table 2. Overview of 2 identified "Expected Compounds" metabolites of Montelukast with RTLS

Peak	RT [min]	Formula	Transformations	Composition Change	DeltaMa	Calc. MW	m/2	FiSh Covera	MS Depth
1	7.26	C41H66CIN0115	Oxidation, Oxidation, Gucanonide	H(CEHBOB)	-0.09	793.22229	794.22656	100.0	2
2	7.63	C41 H64 CI N 011 S	Oxidation, Oxidation, Gucanonide	H(CEHBOB)	-0.18	793.22221	796.22650	75.0	2
3	7.83	C35 H36 CI N 0 5 5	Oxidation, Oxidation	-103	0.17	617.20038	618.20764	56.4	2
4	804	C41 H64 CI N O SO S	Oxidation, Glucuronide Conjugation	H(CEHEO7)	-0.09	777.23737	778.24653	£7.1	ž
5	8.44	C41 HH4 CI N 0:00 S	Oxidation, Glucuronide Conjugation	H(C6 H807)	0.05	777.23748	778.24675	2.0	2
6	\$79	C25 HBSCI N 04 S	Oxidation	-(0)	0.19	621.20547	602.21277	61.9	2
7	885	C25 H34CI N 03 S	Desisturation	-(H2)	0.25	\$83,19699	586.20227	54.8	2
1	9.03	C36 H28 CI N 05 S	Oxidation, Oxidation, Methylation	H[C H2 02]	-0.18	6121581	632.22908	30.8	2
	9.15	C25 HB4CI N 05 S	Desaturation, Oxidation, Oxidation	- (42) +(02)	0.09	615.18468	616.19196	75.9	2
20	9.34	CISHIECI NO45	Oxidation	-101	0.41	601,20560	602.21289	49.6	2

Figure 7. Overview of Montelukast Expected Metabolite structures identified by DDA with RTLS







Conclusions

The study results show that the Orbitrap IQ-X Tribrid MS Met-IQ workflow uses an intelligent data acquisition strategy 'Real-Time Library Search' to guide data acquisition based on structural similarity and improves efficiency for metabolite identification. RTLS can also be used for other small molecule structure ID applications, such as impurity and degradation profiling

The Real-Time Library Search-based intelligent MS3 triggering can be combined with AcquireX workflows for automated

background exclusion to further improve efficiency of analysis. Compound Discoverer 3.3 software facilitates effective data mining and generates confident metabolite ID and structure aracterization

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