Plants, Food, Environment and Microbes

Profiling soybeans using UPLC-UV-CAD-MSn with UVPD to build and curate comprehensive spectral libraries for saponins and flavonoids Eric D. Tague, Thermo Fisher Scientific, 355 River Oaks Parkway, San Jose, CA, 95134, USA

Abstract

Soybeans and their derivatives are widely used for livestock feed and protein sources in consumer foods. It's well known that two classes of compounds (flavonoids and saponins) are abundant in soy-based products. Flavonoids and saponins have been touted for their health benefits but some compounds in these classes could have negative side effects. With many people adopting a more plant-based diet, consumer products are beginning to utilize a wider variety of protein alternatives.

By leveraging a new inline LC universal detector, charged aerosol, and an advanced fragmentation technique, ultraviolet photo dissociation, a more comprehensive library focused on these two compound classes can be created with only a few injections and limited sample quantity. This library can then be applied to other plants to accelerate annotation of compounds in these classes during untargeted experiments. A UPLC was used with a Hypersil GOLD[™] aQ column to separate a wide variety of analytes. Following separation, the flow was split to 2 different detectors, a CAD and a VWD. Eluent was sent to the Thermo Scientific[™] Orbitrap[™] IQ-X[™] Tribrid[™] mass spectrometer where different data fragmentation techniques were performed including HCD, CID, and UVPD, within the same injection. Using this plethora of data collected from the VWD, CAD, and different fragmentation types, a curated spectral library of 12 unique saponins, and unique 6 flavonoids was curated using Thermo Scientific[™] Mass Frontier[™] 8.1 software. This library was then used to identify the other related compounds within the soy sample. The library was used to search a sample of hops for related flavonoids or saponins.

Instrumental design

Figure 2. A.- Thermo Scientific Vanquish[™] Horizon UHPLC System. B.- Column used for separation.



Results

Figure 5. Different flavonoids of interest¹

Mass Frontier 8.1 is capable to

detection for a sample with 3

built in detection algorithms

each with user customizable

Results using Joint Component

Detection and searches against

mzCloud are shown in Table 1.

targeted method with a focus on

a list of known flavonoids and

This method was as semi

known saponins (Table 2).

performing component

parameters.



daidzein (R1 = H, R2 = H) glycitein (R1 = H, R2 =OMe) genistein (R1 = OH, R2 = H) In plant tissues:

Daidzin (7-O-glucosyl-daidzein) Glycitin (7-O-glucosyl-glycitein) Genistin (7-O-glucosyl-genisteir

6"-O-malonyl-7-O-glucosyl-daidzein 6"-O-malonyl-7-O-glucosyl-glycitein 6"-O-malonyl-7-O-glucosyl-genisteir

ound in processed food produ

6"-O-acetyl-7-O-glucosyl-daidzein 6"-O-acetyl-7-O-glucosyl-glycitein

Results

Figure 8. Additional views of Mass Frontier 8.1



Introduction

High resolution mass spectrometry is as great way to collect a wealth of information rapidly but it's only useful if it can be easily accessed and mined in later experiments. Library building and spectral curation are essential but can be very time consuming. By leveraging sophisticated tools aimed at optimizing time spent curating data, users can spend more time interrogating data.

Many times, just a full MS1 scan and single fragmentation mode isn't enough to confidently identify unknown compounds especially when working with plant-based compounds that have multiple isomers and isobars. This works adds 3 different fragmentation techniques (HCD, CID, and UVPD) to capture subtle differences in structure change in how compounds fragment.

Column: Thermo Scientific[™] Hypersil[™] Gold AQ C18 **selectivity**, 1.9 µm, 150 x 2.1 mm

Column Temperature: 45 °C

Autosampler Tray Temperature: 20 °C

Injection Volume: 2 µL

- Mobile Phase: A = 0.1% Formic acid in Water
 - B = 0.1% Formic acid in Methanol
- 0.3 ml / min Flow rate: UV: 280 nm
- 35 °C CAD:

Figure 3. LC gradient used in separation



Figure 4. A.- Thermo Scientific Orbitrap[™] IQ-X [™] **Tribrid mass spectrometer. B.- Mass spectrometry** method



6"-O-acetyl-7-O-glucosyl-genistein

Figure 6. Different saponins of interest²



Table 1: Detected compound summary

Soy meal Compounds			
Total detected compounds	1,697		
Identity search matches (MS2)	488		
Tree Search matches (MSn)	512		

From the 1,697 compounds that had at least 1 MS2, 22 flavonoid and saponin based compounds were chosen for library curation using the Curator module in Mass Frontier 8.1. Since data was collected for both ionization modes, the library will include data for these along with all three fragmentation types.

 Table 2: Specific flavonoids and saponins chosen
for library curation.

Soymeal Comounds of Interest						
Compound Class	Name	RT (max)	М-Н	RT (max)	M+H	
Isofalvones	Daidzin	8.19	415.1035	8.17	417.118	
Isofalvones	Genistein	9.37	269.0457	9.36	271.0601	
Isofalvones	6"-O-malonyl-genistin	10.98	517.193	11.05	519.1133	
Isofalvones	6"-O-acetyl-genistin	12.05	473.1092	12.09	475.1235	
Saponins Group A	A2	13.9	1105.503	13.91	1107.515	
Saponins Group A	A1 w/ 4 Ac (22-gluc)	15.41	1435.638	15.41	1437.654	
Saponins Group A	A3 w/ 4 Ac (22-gluc)	16.1	1243.577	16.08	1245.59	
Saponins Group E	Bd	17.39	955.492	17.39	957.5063	
Saponins Group B	III (Bb')	18.34	795.4553	18.39	797.4691	
Saponins DMPP soy	betta a	18.99	1037.534	19	1039.546	
Isofalvones	Daidzein	8.19	253.0507	8.19	255.0652	
Isofalvones	Genistin	9.37	431.0985	9.36	433.1129	
Misc	Isoflavone realated 2	12.26	467.2135	12.26	469.2807	
Saponins Group A	A2 w/ 4 Ac (22-gluc)	15.92	1273.591	15.95	1275.602	
Saponins Group B	V (Ba)	17.96	957.5076	17.99	959.5222	
Saponins Group B	l (Bb)	18.39	941.5162	18.41	943.5273	
Saponins Group E	Ве	18.8	939.4986	18.81	941.5108	
Saponins Group B	II (Be)	18.56	911.5029	18.56	913.516	
Saponins Group B	IV	18.47	765.4437	18.41	767.4948	
Saponins DMPP soy	betta g	18.84	1067.545	18.89	1069.558	
Misc	Isoflavone realated 1	8.19	461.1091		ND	
Misc	Isoflavone realated 3	9.37	477.1047		ND	

Figure 9. Comparison of chromatogram for TIC, CAD, and UV traces



NB: Delay time of 0.02 and 0.12 min to align CAD and UV trace

By comparing the TIC, CAD, and UV signals

- Region 1: No MS response but significant UV and CAD signals
- Region 2: response on all channels, especially UV
- Region 3: CAD, MS response, no UV

Conclusion

Leveraging the power of the Orbitrap IQ-X and Mass Frontier 8.1, it was possible to quickly and thoroughly create a comprehensive library of 22 flavonoids and saponins. This library contains multiple fragmentation types, polarities, and putative fragment structures. Future work enables this library to be used when searching for these 2 classes of compounds in other samples.

Orthogonal to the MS data, the standard LC retention time was collected in addition to an inline UV detector and split flow to a Charged Aerosol Detector (CAD). This will help collect as much information as possible with a single injection.

Materials and methods

Sample Preparation

Samples used in this project were received as defatted dry soymeal. In brief, preparation involved grinding the whole germ to a fine powder. Soxhlet extraction with hexane was allowed to proceed overnight. The resulting powder was allowed to thoroughly dry before metabolite extraction was completed⁴.

Metabolite Extraction

Approximately 150 mg of sample was aliquoted into 5 replicates samples. Each extracted with 4 mL of methanol. Samples were sonicated for 30 minutes and allow to extract overnight. After 10 minutes of centrifugation the supernatant was evaporated to dryness under a stream of nitrogen.

Samples were reconstituted in 300 µL of methanol.

Figure 1. A. whole soy meal B. processed and weighed sample





A single injection was focused on the m/z range of 300-1250, HCD from 300- 500 *m/z*, CID from 550-1250 *m/z*, and UVPD fragmentation types for all MS2 data. MS3 and MS4 with CID fragmentation were acquired using NL triggers for 6 different sugars. This method was used for library searching and building using Mass Frontier 8.1 software. Data was collected in both positive and negative mode using separate injections.

Data Processing





Several of the compounds had a match to mzCloud but matched multiple saponins with similar structures. By leveraging additional UVPD fragmentation, targeted NL, and advanced curation a highly annotated library can be created.

Mass Frontier 8.1 provides several ways to expedite library generation.

- New SledgeHammer fragmentation algorithms
- Importing SMILES or InChI strings for structure drawing
- Improved curation speeds
- Quick filtering and searching of user generated libraires
- Improved negative mode fragmentation

Using semi automated curation workflow, a library of 22 unique compounds was created in a single afternoon. An example of the curation workflow is shown in Figures 7 and 8.

Figure 7. Example of curation process in Mass Frontier 8.1



References

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Thermo Scientific Mass Frontier 8.1 software was used to for all other processing. This included component detection, library building, curation, lı.l. fragment annotation, recalibration, spectral averaging, and searching.



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