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Fast analysis of multi-class Pesticides panel in wine and olive oil extracts using Single Run LC-Triple Quadrupole Mass Spectrometry

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ABSTRACT

Purpose: To present LC-MS/MS multi-class pesticide residue analysis methods in wine and olive oil that are robust, rapid, easy to use, and have the sensitivity, accuracy, and precision that is required in order to meet EU regulatory guidelines. All aspects of these methods represent a 'workflow' from sample to final report for food safety laboratories.

Methods: Various wines and olive oils were processed and analyzed to test the core methodology- from sample preparation using a modified QuEChERS protocol (Quick, Easy, Cheap, Effect, Rugged, and Safe) to analysis, data processing, and reporting with triple quadrupole LC-MS/MS and comprehensive data handling software. A compound database of over 700 compounds with optimized SRMs and liquid chromatography conditions was created, along with a screening method for over 550 residues and quantitative methods with over 250 residues.

Data Analysis

Individual standards for 720 pesticides were purchased from Ultra Scientific, Inc., and solutions of each were prepared at 1 ppm in acetonitrile or methanol for optimization of MS/MS conditions. Up to five SRMs with collision energies were obtained for each pesticide using an automated routine, in which large numbers of the standards are simply placed into the autosampler and infused unattended into the LC-MS/MS. This information combined with compound retention times and other meta-data was then used to create a comprehensive compound data base in Thermo Scientific[™] TraceFinder[™] software. A quantitative method containing 250 pesticides and a screening method of over 550 were easily created from the database and used for method validation.

Figure 5 shows some typical results of calibration curves from 0.5 to 100 ppb. Over 95% of the pesticides studied had calibration curves with $r^2 > 0.990$. Confirmation ions are displayed in the middle of each panel at 1 ppb for each pesticide, with indicator colors (green) easily visible to show passing ion ratio criteria. The ISVEA laboratory then wanted to expand the list of analytes beyond the original scope, and decided to create a comprehensive screening method using the SRM compound database. A method of over 550 compounds was developed and optimized to ensure that at least two SRM transitions were detected per compound (one for quantitation and the other for confirmation), and still maintain polarity switching throughout the run. LODs and LOQs were determined as outlined by the EU guidelines, with results shown in Figure 6. The method allowed ISVEA to quickly screen samples with confidence at or below the EU MRL for a wide variety of pesticides, giving their customers added confidence in the safety of their products.

Results: Results demonstrate that the methods are fit-for-purpose for both quantitative and broad spectrum pesticide residue screening that can be easily implemented in food safety testing labs. Calibrations with matrix matched standards (MMS) were performed, and matrix extracted spikes (MES) were used to evaluate extraction efficiency. Acceptable results were obtained for the key figures of merit: Limit of Detection (LODs), Limit of Quantitation (LOQs), calibration range/linearity, and recovery.

INTRODUCTION

Food pesticide residue laboratories face significant analytical challenges. Growing target compound lists, large numbers of samples, wide varieties of matrices, and decreasing limits of detection are pressuring labs to become more efficient than ever before. In addition, customers often require more information on contaminants that are not on any target lists that may be a threat. Increasing food safety concerns and growing agricultural trade has resulted in more stringent pesticide regulations globally. To comply with such regulatory standards, quantitative and screening methods for large numbers of pesticide residues are becoming more common in the routine food safety laboratory. Tandem quadrupole mass spectrometry offers a highly sensitive and selective detection in complex matrices. This poster describes a method for analysis of multi-class pesticides in food samples (wine and olive oil) using liquid chromatography coupled with a triple quadrupole mass spectrometer.

MATERIALS AND METHODS

Sample Preparation

Sample preparation involves a protocol that was optimized to be easy to implement and also reduce matrix co-extractives, resulting in enhanced sensitivity and robustness in electrospray ionization LC-MS/MS. The basic elements of the preparation procedures are described in Figure 1.

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d Name and All Collapse All	F	Compound Name	ņ	Peak Label 4	Peak Workflow		ssociated arget Peak	₽ ↔	MS Order	Precursor m/z	Product m/z	⊨ m/z +⊐	Addu	ct 🕂 Pola	rity	👳 Charge Sta
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2,4-dimethylaniline		azinphos-ethyl		T1: 346.044->76.917	-	•				346.044	76.917	346.044	None			• 1
2,6-dichlorobenzamide 2-methyl-4-6-dinitrophen		azinphos-ethyl		T1C1: 346.044->131.	-	• T	1: 346.044->76.			346.044	131.917	346.044	None			• 1
3,4,5-trimethacarb 3-hydroxycarbofuran		azinphos-ethyl		T1C2: 346.044->260.	Confirming	• T	1: 346.044->76.			346.044	260.887	346.044	None	• ▼ Posit	tive	• 1
6-chloro-4-hydroxy-3-phe		aziprotryne		T1: 226.086->124.92	-	•				226.086	124.929	226.086	None			• 1
abamectin acephate		aziprotryne		T1C1: 226.086->155.	-		1: 226.086->124			226.086	155.917	226.086	None			• 1
acetamiprid				T1C2: 226.086->195.	-		1: 226.086->124			226.086	197.857	226.086	None			• 1
acetochlor acibenzolar-s-methyl		aziprotryne			-	•	1: 220.000-2124									
aclonifen		Azocyclotin		T1: 430.195->178.04/	-					430.195	178.042	430.195	None			• 1
acrinathrin-NH4 alachlor		Azocyclotin		T1C1: 430.195->248.	_		1: 430.195->178			430.195	248.917	430.195	None			• 1
albendazole		Azocyclotin		T1C2: 430.195->274.	-		1: 430.195->178			430.195	274.887	430.195	None			• 1
aldicarb aldicarb-sulfone-NH4+		azoxystrobin		T1: 404.124->328.85		•				404.124	328.857		None			• 1
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Amisulbrom	134	benalaxyl		T1: 326.175->148.00(TargetPeak	•		-	ms2 •	326.175	148.000	326.175	None	• • Posit	tive	- 1
amitrole ancymidol	135	benalaxyl		T1C1: 326.175->207.9	Confirming	• T	1: 326.175->148	•	ms2 •	326.175	207.929	326.175	None	• • Posit	tive	• 1
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Add 10 g of wine to a 50 mL polypropylene centrifuge tube	Add 5 g of olive oil to a 50 mL polypropylene centrifuge tube
Add 10 mL of acetonitrile	Add 10 mL of acetonitrile
Add 100 µL of internal standard and 50 µL of control	Add 6 mL of water
standard	
•	Add 100 µL of internal standard and 50 µL of control
Cap and shake vigorously for 1 minute	standard
Add salt pouch: 4000mg MgSO4, 1000mg NaCl,	Cap and shake vigorously for 5 minute
500mg Na2Citrate, 1000mg Na3Citrate	
•	Add salt pouch: 4000mg MgSO4, 1000mg NaCl,
Shake for approximately 1 minute	500mg Na2Citrate, 1000mg Na3Citrate
•	
Centrifuge at 4000 rpm for 5 minutes	Shake for approximately 1 minute
Filter at least 1.5 mL of supernatant through a 0.45 µm filter	Centrifuge at >3300 rpm for 15 minutes
Inject 1 µL for analysis	Filter at least 2.0 mL of supernatant through a 0.45
	µm filter
	Pass 1.5 mL thru lipid removal cartridge
	Inject 1 µL for analysis

Figure 1: Extraction procedures for wine and olive oil samples. No dispersive solid phase extraction clean-up was required. Only 1 µL of the extracts were injected (pure acetonitrile), which provided excellent sensitivity and robustness for the method.

695	fomenafen-MH4	5.46	0.5	Positive	456	299.815	25.43		4100	
696	fomesafen-NH4	5.46	0.5	Positive	456	343.845	13.37			Show Visualization
697	domazone	5.47	0.5	Positive	240.078	88,899	46.28		5 100	Copy Esperiment Time
6/38	clomazone	5.47	0.5	Positive	240.078	98.845	46.32			
699	clomazone	5.47	0.5	Positive	240.078	124.845	21.11		2 50	
700	fenobucarb	5.5	0.5	Positive	208.133	76.97	35.98		- 1	
701	fenobucarb	5.5	0.5	Positive	208 133	94,917	15.08		2 4 6 8	10
702	fenobucarb	5.5	0.5	Positive	208 133	151.97	10.23	Ψ.	Time (min)	10
4			111							

Figure 3: Large pesticide residue panels are easily created in TraceFinder directly from A (database) to B (acquisition method). Visualization tools within the tune program of the TSQ Quantis triple quadrupole mass spectrometer show SRM density throughout the run, ensuring that enough scans are acquired across each peak with appropriate dwell times for accurate quantitation.

RESULTS

Wine (white and red) as well as olive oil samples from various sources were obtained for method validation. Typically, matrix matched standards (MMS) are required for calibration, and matrix extracted spikes (MES) are used to assess recovery. For excellent quantitation, there must be a sufficient number of scans across the target quantitation peak. At least and one or two SRMs can be used for confirmation. Figure 4 shows a 10 ppb MMS for a method containing over 250 pesticides with positive and negative polarity switching occurring throughout the run. Plenty of scans across each peak are present for accurate quantitation.



Figures 6: LODs and LOQs in white wine obtained following the SANTE guidelines for the screening method of over 550 pesticides at the ISVEA laboratory.

Method robustness is key to any laboratory. The screening method showed excellent reproducibility in terms of a) consistent peaks shapes and long column lifetime, with over 1000 injections (and still going strong) and b) consistent peak response over time. Figure 7 shows some select pesticides across the retention time range of the method (1-10 minutes), for approximately 300 injections.



Figures 7: LC-MS/MS extracted ion chromatograms of spiked white wine of select pesticides (overlay of injection #1 and injection # 300) demonstrates good robustness of the analytical system and API source.

CONCLUSIONS

Large pesticide panels for quantitative analysis and screening at levels below EU MRLs have been shown to provide excellent sensitivity and robustness in a routine laboratory

Analytical Methods

Analytical condition use for the analysis are described here below in Figure 2

	\sim
<u> Thermo Scientific™ Vanquish™ Flex binary</u>	<u>Thermo Scientific™ TSQ</u>
UHPLC pump:	<u>Quantis™ triple quadrupole MS:</u>
Mobile phase:	Negative Voltage 2500V
A: Water + 5mM Ammonium formiate & 0.1% FA	Positve Voltage 3500V
B: MeOH + 5mM Ammonium formiate & 0.1% FA	Sheath Gas 30 units
<u>Injection volume:</u> 1 µl	Auxiliary Gas 6 units
<u>Column:</u> Accucore aQ 100 x 2.1 mm x 2.6 µm	Sweep Gas 1 units
<u>Column temperature: 25°C</u>	lon transfer tube T 290°C
<u>Flow rate: 300 µl/min</u>	<u>Vaporizer T</u> 350°C
<u>Run time: 15 min</u>	

Figure 2: LC gradient, mobile phase, column, and API source conditions for the Vanquish Flex Binary UHPLC pump with TSQ Quantis triple quadrupole mass spectrometer.

Figures 4: White wine MMS standard at 10 ppb. The peak at 5.37 minutes is Methabenzthiazuron, showing over 15 scans across the quantitation ion used for the Large pesticide panels of extracted SRMs are easily displayed in analysis. **TraceFinder Software.**



Figures 5: Quantitation ions and confirming ion at 1 ppb in a MES, along with calibration range from 0.5 to 100 ppb in TraceFinder software for Flusilazole in white wine. The technique allows for confident screening with confirmation well below the **MRL concentration.**

setting for both red and white wines.

The QuEChERS extraction procedure demonstrated good recovery and precision, with only 1 µL required for injection to meet EU SANTE validation guidance.

On-going work is required to evaluate the performance of the residues in olive oil samples

REFERENCES

1.EUROPEAN COMMISSION- SANTE/11813/2017, November 2017

ACKNOWLEDGEMENTS

We would like to thank Thermo Fisher Scientific local team for the support in developing the method

TRADEMARKS/LICENSING

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