# An in-depth and high throughput plasma proteomics workflow powered by Orbitrap Exploris 480 mass spectrometer using multi nanoparticle-based workflow

# **ThermoFisher** SCIENTIFIC

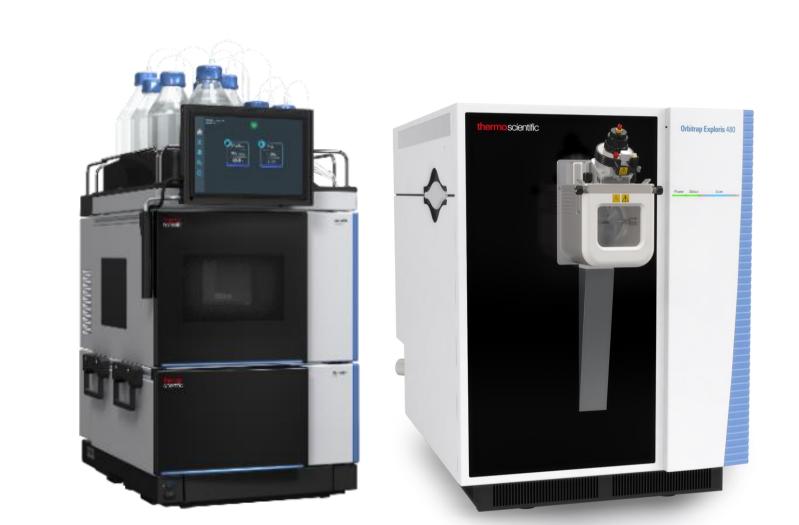
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# Introduction

Plasma proteomics using mass spectrometry (MS) remains a promising method to discover disease biomarkers. However, for large-scale plasma proteomics studies, a robust liquid chromatography-mass spectrometry (LC-MS) setup that does not compromise on protein identification, sequence coverage, dynamic range and analysis precision is required. Here we present a high-throughput and a maximum identification (Max-ID) workflow on an Orbitrap Exploris 480 mass spectrometer for in-depth analysis of plasma. The plasma samples were processed with Seer's Proteograph<sup>™</sup> XT Assay utilizing multi-nanoparticles (NPs) for an unbiased and deep proteomics enrichment and analysis at scale.

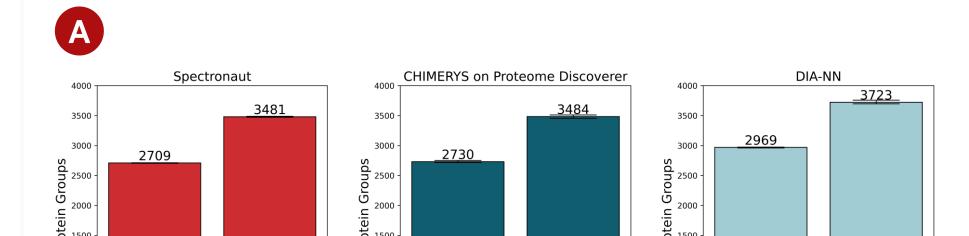




# Results

The optimized DIA method enabled the identification of > 3,000 protein groups, > 20,000 peptides from the high-throughput method (Figure 1 A and B) and > 3,800 protein groups, > 27,000 peptides in the Max-ID method (Figure 1C and D) using the Proteograph Analysis Suite with match between run (MBR).





# Methods

Human plasma sample (a low complex pooled healthy control plasma) was processed prepared using with an automated, scalable, and robust Seer Proteograph<sup>™</sup> Product Suite, SP100 automation system with Proteograph<sup>™</sup> XT Assay Kit.

### **Proteograph XT Assay**

Samples are processed using 100  $\mu$ L aliquots of each plasma sample, mixed with each of two NP wells included in the Proteograph XT Assay Kit. A one-hour incubation allows high-affinity proteins to displace highabundance proteins, resulting in a reproducible protein corona on each NP surface that probes the depth of the plasma proteome. A series of gentle washes remove non-specific and weakly bound proteins. The paramagnetic property of the NPs allows for accumulation of NPs with protein corona after each wash step. This results in a highly specific and reproducible protein corona that contains the high-affinity protein binding partners selected by the NPs. Protein coronas are reduced, alkylated,

and digested with Trypsin/Lys-C to generate tryptic peptides for LC-MS analysis. All steps are performed in a one-pot reaction directly on the NPs. The in-solution digestion mixture is then desalted, and all detergents are removed using a mixed-media filter plate and positive pressure (MPE) system. Clean peptides are eluted in a high-organic buffer into a deep-well collection plate. A 0.5 µg of tryptic peptides were analyzed on the Orbitrap Exploris 480 MS with both nanoflow and capillarity flow LC methods.





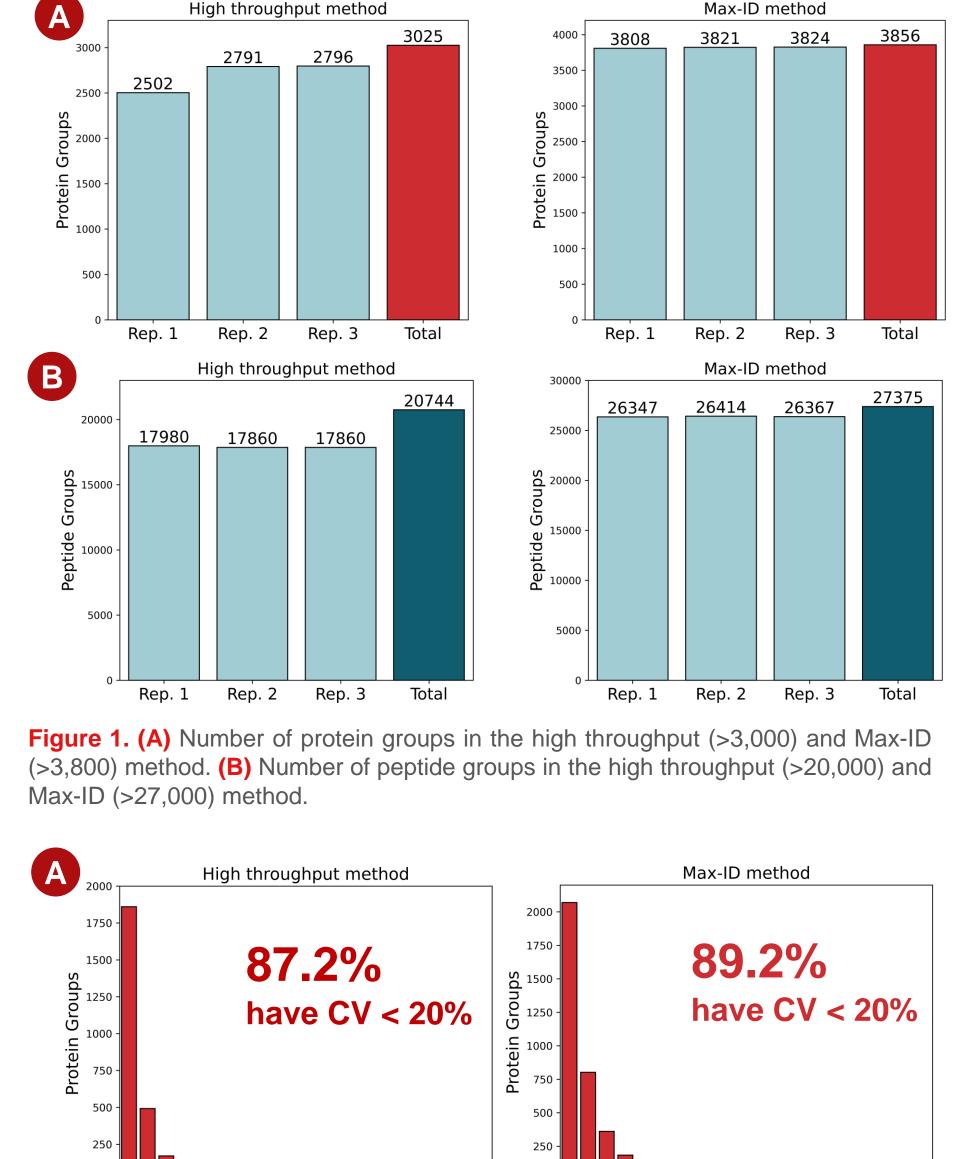
**LC Gradient** 



High throughput								Max-ID						
No	Time	Duration [min]	Flow [µl/min]	%В	Volume [µl]	No. of Column Volumes	No	Time	Duration [min]	Flow [µl/min]	%В	Volume [µl]	No. of Column Volumes	
1	0.000	Run						0.000	Run					
2	0.000	0.000	2.000	4.0	0.00	0.00	2	0.000	0.000	0.250	1.0	0.00	0.00	
3	0.200	0.200	2.000	8.0	0.40	0.23	3	0.100	0.100	0.250	6.0	0.03	0.01	
4	14.600	14.400	1.500	20.0	25.20	14.19	4	60.100	60.000	0.250	20.0	15.00	6.76	
5	21.500	6.900	1.500	35.0	10.35	5.83	5	90.100	30.000	0.250	35.0	7.50	3.38	
6	21.500	Column Wash						90.100	Column Wash					
7	21.900	0.400	2.000	99.0	0.70	0.39	7	91.100	1.000	0.250	99.0	0.25	0.11	
8	22.600	0.700	2.000	99.0	1.40	0.79	8	102.000	10.900	0.250	99.0	2.73	1.23	
9	22.600	Stop Run						102.000	Stop Run					
10	22.600	Column Equilibration						102.000	Column Equilibration					

# **MS Method**

The Proteograph XT Assay Kit generates two sets of enriched peptides at the end of analysis. For the high throughput method, 2 individual injections of 24 minutes each were used for a total instrument time of 48 minutes per sample. For the Max ID method, the two peptide wells were pooled and LC-MS analysis was done in single injection using a 102 minutes total gradient.



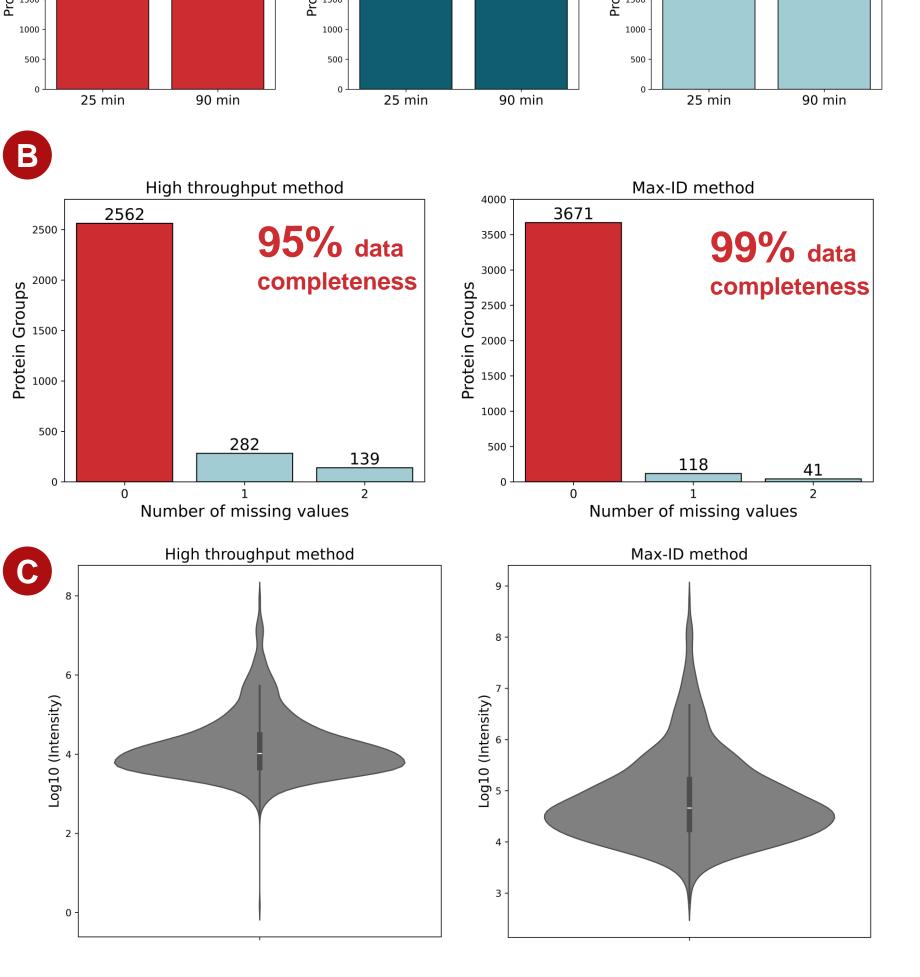


Figure 3. (A) Number of protein groups obtained from library free searched on Spectronaut, CHIMERYS on Proteome Discoverer and DIA-NN. (B) Data completeness of high throughput (95%) and Max-ID (99%) method across three injections. (C) Wide dynamic range covered by Orbitrap Exploris 480 mass spectrometer in both the high throughput and Max-ID method. Data was obtained from PAS.

# LC Method

High throughput method:

- Thermo Scientific<sup>™</sup> Vanguish<sup>™</sup> Neo UHPLC system utilizing "Fast Loading" and "Fast Equilibration" features.
- Thermo Scientific<sup>™</sup> EASY-Spray<sup>™</sup> PepMap<sup>™</sup> Column, 2 μm C18, 150 μm x 15 cm
- Thermo Scientific<sup>™</sup> PepMap Neo Trap Cartridge, 5 µm 300 µm x 5 mm

• Flow rate: 1.5 µL/min

• LC workflow: Trap & Elute

Max-ID method:

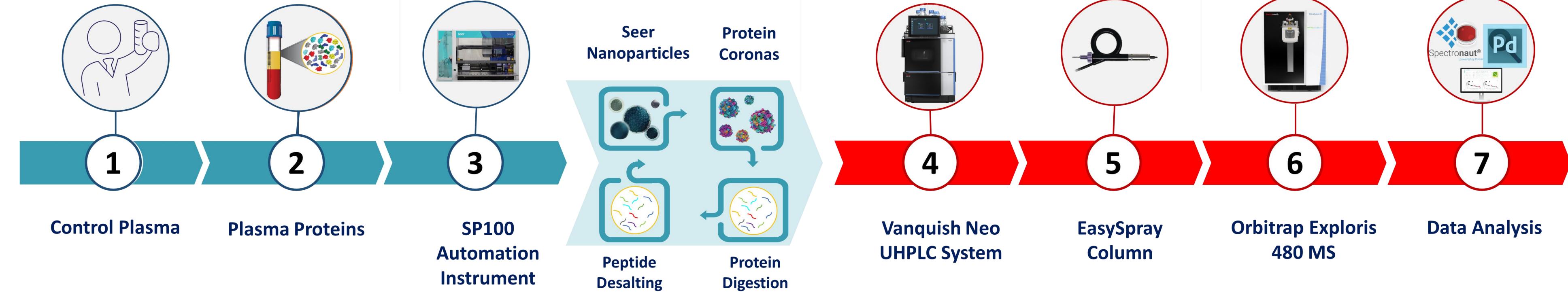
```
Thermo Scientific<sup>™</sup> EASY-Spray<sup>™</sup> PepMap<sup>™</sup> Neo Column, 2 µm C18, 75 µm
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x 75 cm

• Flow rate: 0.25 µL/min

• LC workflow: Direct Injection

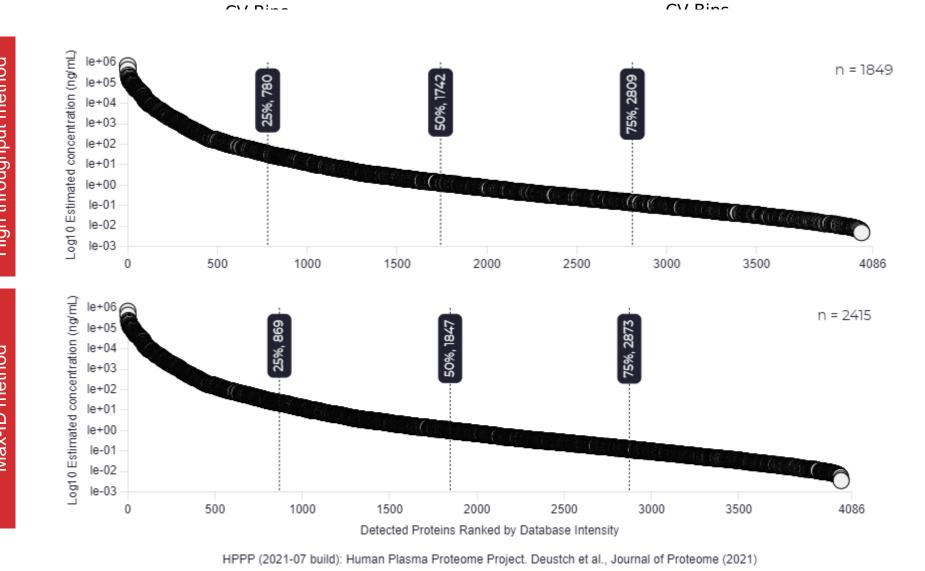
## • Mobile phases: [A] 0.1% FA in $H_2O$ , [B] 0.1% FA in 80% ACN



N	IS Parameter	Max ID Method	High Throughput Method		
	Resolution	60K	60K 300%		
MS1	AGC	300%			
	Max-IT	Auto	Auto		
	Resolution	15K	15K		
MS2	AGC	800%	1000%		
	Max-IT	Auto	28ms		
	Isolation Window	4Da	4Da		

# **Data Analysis**

Proteograph<sup>™</sup> Analysis Suite (PAS) with match between run (MBR) was used for data analysis of the resulting LC-MS files unless stated otherwise. Data was exported and processed by Pandas and Seaborn packages in Python for data visualization. Spectral library free search was done on Spectronaut 18 software, DIA-NN (v1.8.1) or Proteome Discoverer 3.1 software using CHIMERYS intelligent search algorithm by MSAID. In all software packages, 1% FDR cut off was applied at both protein and peptide levels.



5 10 15 20 25 30 35 40 45 50 55 60 65 70 75 80 85 90 95100

5 10 15 20 25 30 35 40 45 50 55 60 65 70 75 80 85 90 9510

Figure 2. (A) Protein group coefficient of variance (CV) distribution of the high throughput and Max-ID method demonstrate the excellent quantitation and precision from Orbitrap Exploris 480 mass spectrometer. (B) Dynamic range of proteins identified in one run compared to a deep plasma proteome coverage reported in human plasma proteome project (HPPP) index.

# **CONCLUSIONS**

- Enrichment of plasma proteins using Seer's Proteograph XT technology helps dig deeper into the plasma proteome dynamic range.
- LC-MS workflow with Vanquish Neo UHPLC system with a PepMap Easy-Spray column coupled to an Orbitrap Exploris 480 MS provides a robust and reproducible setup for identification and quantification of plasma proteins.
- High throughput capillary flow method enables identification of over 3,000 protein groups, of which 87.2% have a CV below 20%.
- Max-ID method allows for deeper proteome coverage along with excellent quantitation performance on Orbitrap Exploris 480 MS.

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