

Validation of the RapidINTEL™ Plus Cartridge for Investigative Lead Samples and to Meet the SWGDAM Rapid Guidelines

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Abstract

Although DNA profiles from *reference samples* run on rapid DNA instruments are allowed into CODIS, the FBI does not currently allow upload of DNA profiles from *crime scene samples* run on rapid DNA instruments. In 2020, SWGDAM, ENFSI, and the FBI Rapid DNA Crime Scene Technology Advancement Task Group outlined rapid DNA requirements that, when met, would allow DNA profiles from crime scene samples run on rapid DNA instruments to be uploaded into CODIS. In early 2024, Thermo Fisher released a new sample cartridge, RapidINTEL™ Plus, designed to meet these requirements while improving on the performance of crime scene samples run on the RapidHIT™ ID system.

The RapidINTEL™ Plus sample cartridge was developed in conjunction with the RapidHIT™ ID System Software v2.0 and RapidLINK™ Software v2.0. In addition to new features required for CODIS, improvements were made to the RapidINTEL™ Plus cartridge to produce high quality and reliable data from crime scene samples through the introduction of Internal Quality Control markers, Quantification markers, and PCR enhancements to increase sensitivity and improve peak height and inter-locus balance.

We performed a developmental validation to assess the newly optimized RapidHIT™ ID System v2.0 for the analysis of investigative lead samples with our RapidINTEL™ Plus cartridge. The system demonstrated substantial improvements in first pass success rate, sensitivity, and suitability for law enforcement personnel to generate investigative leads and identify suspects faster.

Introduction

The Applied Biosystems™ RapidINTEL™ Plus sample cartridge is an updated version of the RapidINTEL™ sample cartridge specifically developed for RapidHIT™ ID System v2.0 and RapidLINK™ Software v2.0 for use with single source crime scene samples. It was developed to increase first pass success and to meet the requirements outlined in the position statement by ENFSI, SWGDAM and the Rapid DNA Crime Scene Technology Advancement Task Group.

The RapidINTEL™ Plus sample cartridge includes the following improvements:

- Amplification chemistry** – Based on GlobalFiler™ Express chemistry, but with the following additions:
 - Internal Quality Control (IQC) markers to identify samples that contain PCR inhibitors
 - Quantification markers to estimate the DNA quantity in the PCR and to determine if the sample is degraded (QTS, QTL)
 - Master mix and primer mix optimizations for improved DNA profile balance
- Dual instrument protocols** – Promotes flexibility by providing a dual protocol system which includes a Specialized protocol and General protocol. The Specialized protocol, in conjunction with the use of a micro/mini swab, enables higher sensitivity and is intended for samples that are expected to contain limited amounts of DNA.
- System performance** – Increased DNA capture in PCR and reduced stochastic amplification by decreasing the number of PCR cycles.

Materials and Methods

The study included a total of 135 samples that were prepared or collected to simulate biological evidence found at crime scenes, including blood, saliva, touch, and other types of samples such as tooth, bone, nail, etc. A few samples contained measured DNA input amounts, and many samples were unmeasured input amounts from stains, cast-off blood, and used or worn items. Most of the samples were collected using a micro HydraFlock swab (Puritan, 25-3318) and were left to air-dry at ambient temperature overnight before running on the RapidHIT™ ID instrument using the Specialized protocol. Other samples such as the nail, tooth, gum, and fabric cuttings were run neat using either or both Specialized and General protocols. A total of 12 RapidHIT™ ID instruments were used for this study.

All results were taken from the RapidLINK™ v2.0 Software or GeneMapper™ ID-X v1.7.

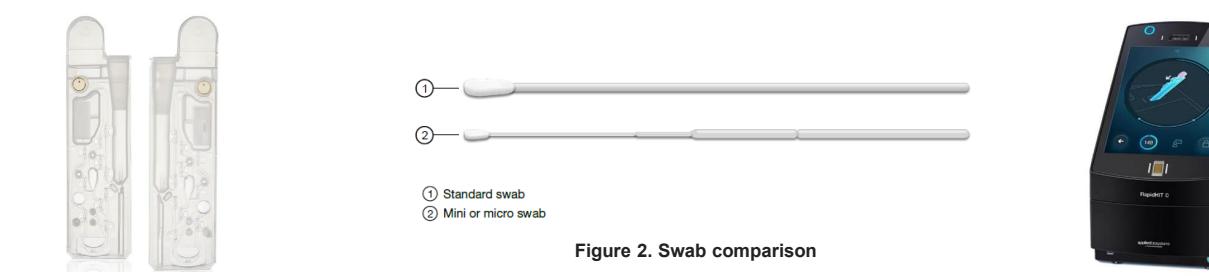


Figure 1. RapidINTEL™ Plus sample cartridge



Figure 2. Swab comparison



Figure 3. RapidHIT™ ID

Results

RapidINTEL™ Plus Chemistry

Figure 4 is an example of an electropherogram of the RapidINTEL™ Plus chemistry containing the Quantification/Quality Small (QQS) and Quantification/Quality Large (QQL) markers located at the front and back of the FAM (blue) dye channel.

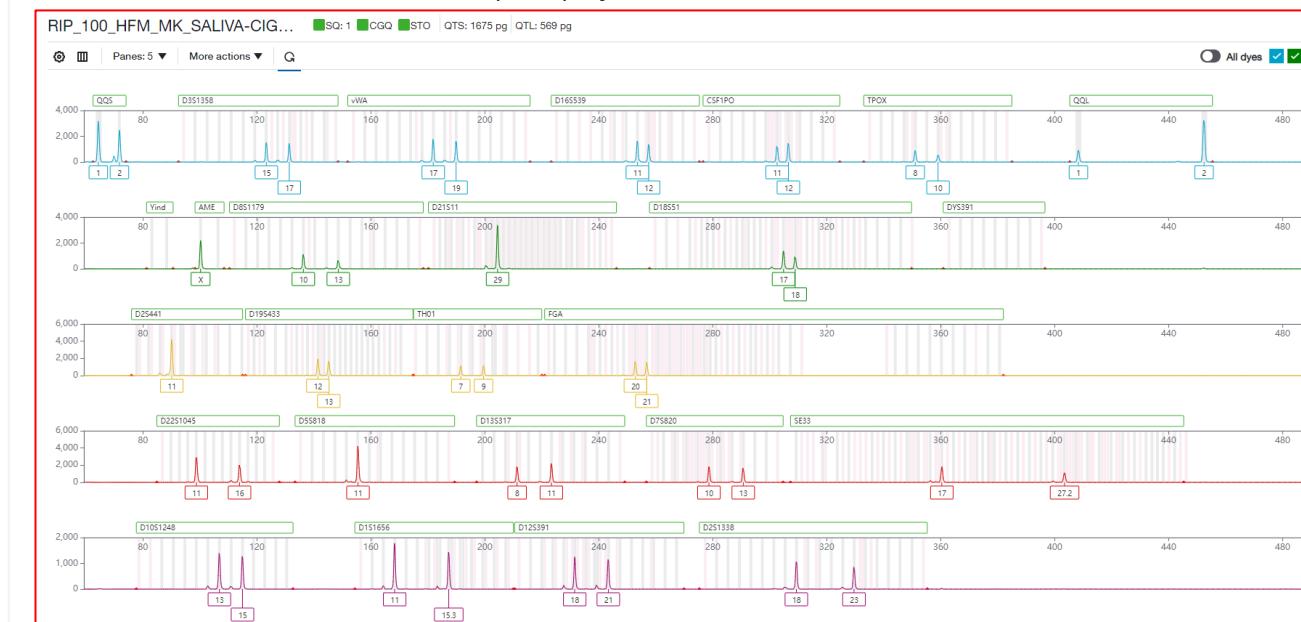


Figure 4. RapidINTEL™ Plus electropherogram of a cigarette butt sampled using HydraFlock micro swab and run under the Specialized protocol

Quantification System

Figure 4 displays the accuracy and precision of the quantification system incorporated into the RapidINTEL™ Plus chemistry. The blue dots are the estimated amount of DNA loaded on the swab in the blood sample after back calculating from the QTS value. The red boxes are the theoretical DNA quantities for the specified volumes of blood on the swab (x-axis). Theoretical values are based on a typical WBC of 4,500-11,000/ μ L (27-66 ng/ μ L). The dotted green boxes are the adjusted expected DNA quantities based on 50% recovery on cotton swab. The data indicates that a flocked swab is more efficient at releasing sample than the standard cotton swab.

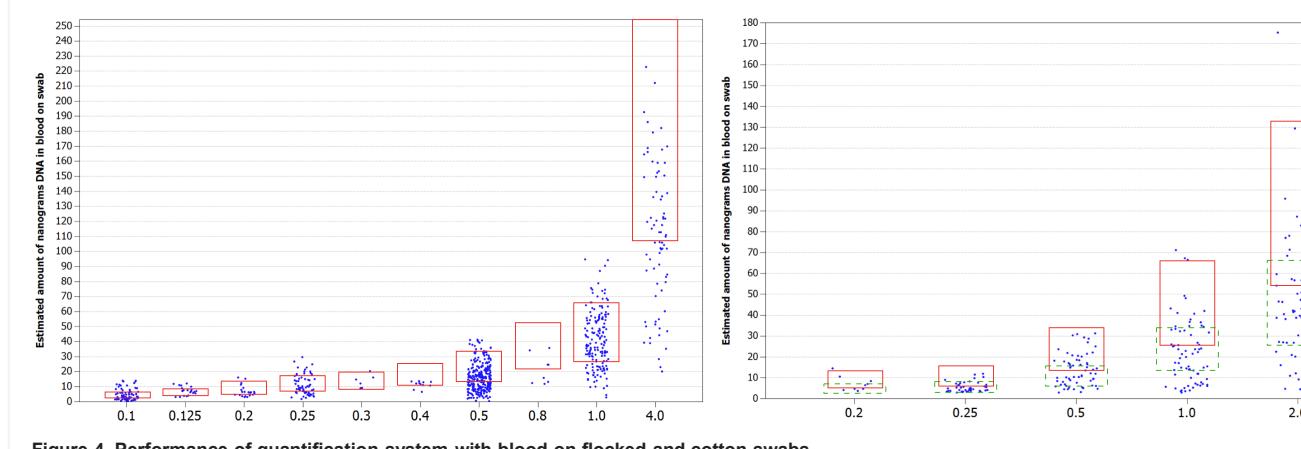


Figure 4. Performance of quantification system with blood on flocked and cotton swabs

Stochastic Flagging

Table 1 shows the percentage of samples that exhibited a red STO flag which contain allelic dropout (all samples run during development of RapidINTEL™ Plus).

Table 1. STO flag indicator correlation to allelic dropout

STO Indicator flag	Meaning	Number of samples	Percent of samples with dropout
■	Extremely rare dropout	1,379	0.15
▲	Rare dropout	432	1.4
●	Frequent dropout	531	75.3

Table 2. Percent alleles recovered (%) for each mock case sample															
Sample Type	Substrate Category	Detailed Substrate/Sample Information	Protocol	Loading Method	Input Information	Percentage of Alleles Recovered	Unflagged Loci (Autosomal)	Allele Dropouts	QTS (ng)	QTL (ng)	Mix	Deg	Inh	STO	RL Review Status
OTHER	Hair board	Spec	DL	Plucked	100%	20	0	8,442	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass w. review
Hair	Hair eyebrow	Spec	DL	Plucked	100%	21	0	6,552	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass w. review
Hair	Hair head	Spec	DL	Plucked	100%	19	0	4,964	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass w. review
Semen	Colored felt	Spec	HL	Plucked	100%	21	0	2,809	862	862	Pass	Pass	Pass	Pass	Pass
Hair	Hair head	Spec	HL	Plucked	100%	21	0	10,054	2,203	2,203	Pass	Pass	Pass	Pass	Pass
Hair	Hair head	Spec	HL	Plucked	100%	21	0	10,054	2,203	2,203	Pass	Pass	Pass	Pass	Pass
Hair	Hair head	Spec	HL	Plucked	100%	21	0	10,054	2,203	2,203	Pass	Pass	Pass	Pass	Pass
Hair	Hair head	Spec	HL	Plucked	100%	21	0	10,054	2,203	2,203	Pass	Pass	Pass	Pass	Pass
Bone	Tibia, powdered	Gen	DL	Extracted	100%	21	0	10,054	2,203	2,203	Pass	Pass	Pass	Pass	Pass
Tooth	Whole tooth ground	Spec	DL	Extracted	100%	21	0	10,054	2,203	2,203	Pass	Pass	Pass	Pass	Pass
Semen	Denim	Spec	HL	Stain	100%	21	0	1,043	902	902	Pass	Pass	Pass	Pass	Pass
Fingernail	Fingernail, clipping	Spec	DL	Clipped	100%	21	0	2,462	1,089	1,089	Pass	Pass	Pass	Pass	Pass
Fingernail	Fingernail, clipping cleaned with EtOH	Spec	DL	Clipped	100%	21	0	1,057	422	422	Pass	Pass	Pass	Pass	Pass
Fingernail	Fingernail, clipping cleaned with EtOH	Spec	DL	Clipped	100%	21	0	667	126	126	Pass	Pass	Pass	Pass	Pass
Hair	Hair head	Spec	DL	Plucked	100%	21	0	292	644	644	Pass	Pass	Pass	Pass	Pass
Fabric	Colored cotton, 4-mm cutting	Spec	HL	Stain	100%	21	0	4,734	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	White cotton, 4-mm cutting	Spec	DL	5 μ l	100%	21	0	5,518	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Glass slide	Spec	HL	1 μ l	100%	21	0	2,824	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Cement	Spec	HL	1 μ l	100%	21	0	2,047	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Drywall	Spec	DL	5 μ l	100%	21	0	1,873	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Fabric, 4-mm cutting	Gen	DL	7.5 μ l	100%	21	0	1,969	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Denim, 4-mm cutting	Spec	HL	1 μ l	100%	21	0	2,840	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Colored cotton, 4-mm cutting	Spec	DL	2.5 μ l	100%	21	0	3,660	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	White cotton, 4-mm cutting	Spec	DL	2.5 μ l	100%	21	0	1,032	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Cement	Spec	HL	5 μ l	100%	21	0	2,047	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Drywall	Spec	HL	5 μ l	100%	21	0	2,045	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Fabric, 4-mm cutting	Gen	DL	7.5 μ l	100%	19	0	1,871	60	60	Pass	Pass	Pass	Pass	Pass
Fabric	Denim, 4-mm cutting	Spec	HL	10 μ l	100%	21	0	2,056	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Colored cotton, 4-mm cutting	Spec	HL	10 μ l	100%	21	0	1,052	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	White cotton, 4-mm cutting	Spec	HL	10 μ l	100%	21	0	2,050	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Cement	Spec	HL	10 μ l	100%	21	0	1,050	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Drywall	Spec	HL	10 μ l	100%	21	0	2,052	>1,600	>1,600	Pass	Pass	Pass	Pass	Pass
Fabric	Fabric, 4-mm cutting	Spec	HL	10 μ l	100%	21	0	1,050	>1,						