

#### **ThermoFisher** SCIENTIFIC

# **GlycanAssure**<sup>™</sup>: A High Throughput & High Resolution Glycan Analysis Platform

# Agenda

#### Introduction

- Glycosylation and Glycan Diversity
- Importance of Protein Glycosylation
- Glycan Labeling Chemistry
- Current Glycan Analysis Challenges

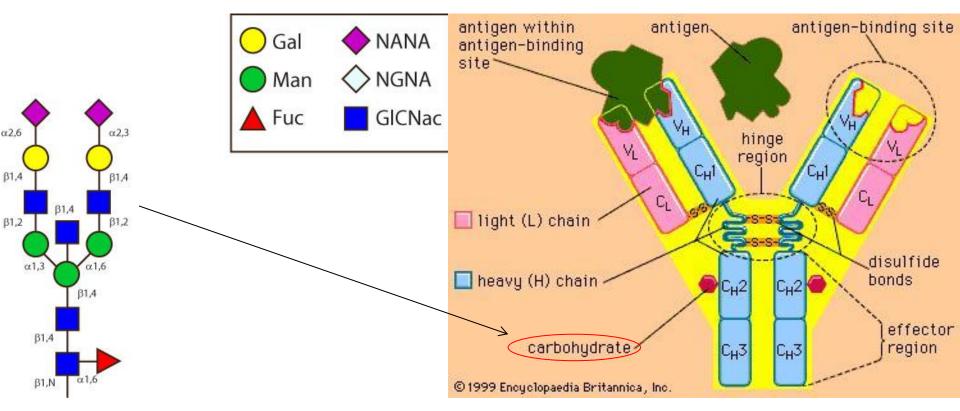
#### Thermo Fisher GlycanAssure Platform

- Sample Prep
- Instrumentation
- Software
- Multicap CE for Glycan Analysis Literature
- GlycanAssure Workflow

#### Sample Data Sets

- Magnetic Bead Based Workflow vs. Carbon Column
- Consistency Across Varying Glycoprotein Inputs
- Reproducibility of CE Separation
- Variability Across Capillaries
- Variability Across Instruments & Capillary Arrays
- Glycan Spike Studies
- Improved Glycan Separation Using Thermo Fisher Dyes
- Thermo Fisher 3500 Vs. Competition
- Summary

## **Glycans - Introduction**



- Glycosylation Attachment of Glycans (Carbohydrates or Oligosaccharides) to proteins
- Glycans are made up of Monosaccharides
- More than 70% of biotherapeutics are glycosylated
- Glycans are important for protein stability, folding, assembly, signaling, etc.
- Glycosylation is a critical quality attribute (CQA) of biotherapeutics



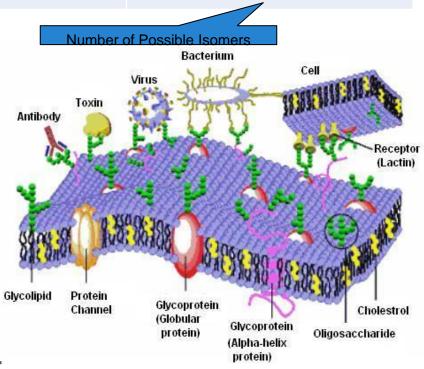
Oligomer	Composition	Peptide	Oligosaccharide
Dimer	AA/AB	1/2	11 / 20
Trimer	AAA / ABC	1 / 6	120 / 720
Tetramer	AAAA / ABCD	1 / 24	1424 / 34560
Pentamer	AAAAA / ABCDE	1 / 120	17872 / 2144640

**Glycoconjugate Biosynthesis** 

- Glycans are not templated
- Glycan structure is determined by sequential glycosyltransferase action

More than 50% of Human Proteins are Glycosylated

- Glycosylation is Heterogeneous
  - Structures of Sugars Attached
  - Sites To Which They are attached
  - Anomericity



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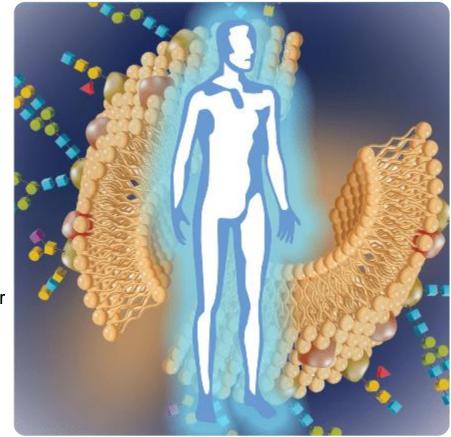
# Many Conditions Affecting Humans Involve Glycans

Pregnancy N-glycosylation of human serum transcortin and thyroxine-binding globulin

Aging Outer arm galactosylation of human serum IgG

#### Alcoholism

Increase in relative number of serum asialo-tranferrin glycoforms



Cancer Unique oligosaccharides Such as CA 125 (Ovarian) CA 19-9 (Colon)

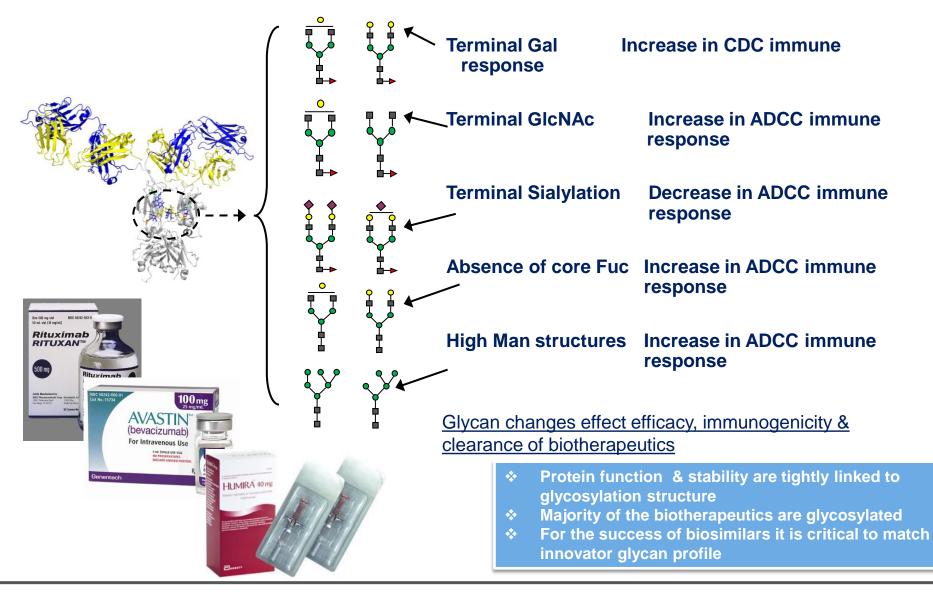
Liver Disease Increase in number of sialic acid residues on fibrinogens

Tuberculosis agalactosyl IgG

Ref: Dwek et al., Ann. Rev. Biochem., 57 (1988) 785-838



#### Biotherapeutics: Glycosylation is a CQA





#### **Current Regulatory Guidelines**

Ensures glycans/glycosylation is well characterized in biotherapeutics

- FDA (USA) and EMA (Europe) define the regulatory requirements
- Biotherapeutic manufactures are legally obligated to comply with these

#### US FDA, November 3, 2010

Many complex biologics have sugar chains that impact their efficacy and safety, and industry experts have noted that the FDA has made several statements over the years suggesting a primary concern that biosimilars have the same sugar structure for interchangeability and those sugar structures do not change with time.

#### EMEA Guidelines 2009

On the development, production, characterization and specifications for monoclonal antibodies and related products 'glycan structures should be characterized, and particular attention should be paid to their degree of of mannosylation, galatosylation, fucosylation and sialylation'.

European Medicines Agency EMEA/CHMP/BWP/157653/2007 (2009)



#### Draft Guidance for Analysis of Biosimilars

As monoclonal antibody molecules come off patent, biosimilars will start entering the market.

FDA (USA) and EMA (Europe) consider the degree of glycosylation to be a critical factor in determining the degree of "similarity" to the original approved drug

**Similar**: Additional analytical data or other studies are necessary to determine if observed differences are within an acceptable range to consider the proposed biosimilar. For example, the agency says that "glycosylation plays an important role in the PK of certain protein products. Manufacturing process conditions may impact glycosylation. Comparative PK and PD studies of the proposed biosimilar product and the reference product help resolve that some difference in glycosylation identified in analytical studies would be within an acceptable range to consider the proposed biosimilar product to be highly similar to the reference product."

**Reference:** 

Draft Guidance for Industry - Clinical Pharmacology Data to Support a Demonstration of Biosimilarity to a Reference Product

U.S. Department of Health and Human Services, Food and Drug Administration

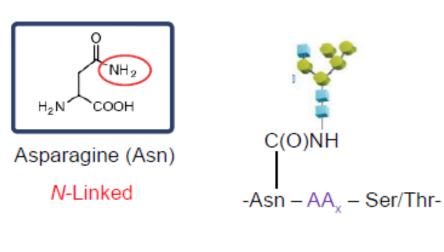
Center for Drug Evaluation and Research (CDER) & Center for Biologics Evaluation and Research (CBER) May 2014 Biosimilars

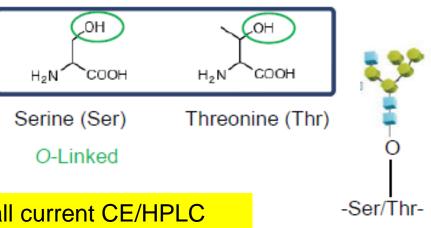
European Medicines Agency EMEA/CHMP/BWP/157653/2007 (2009)



# Which Glycans are Analyzed?

- N-linked glycans
  - Attached to the amide side chain of Asparagine in peptide sequence: Asn- AA<sub>x</sub>-Ser/Thr
- O-linked glycans
  - Attached to the hydroxyl group on Serine or Threonine residues
- Other forms (less studied)
  - Glycophosphatidiylinositol anchors
    - Attached to protein C-terminus
  - C-glycosylation on tryptophan residues
  - S-linked glycosylation
    - Linked through S atom in Cys and Met residues

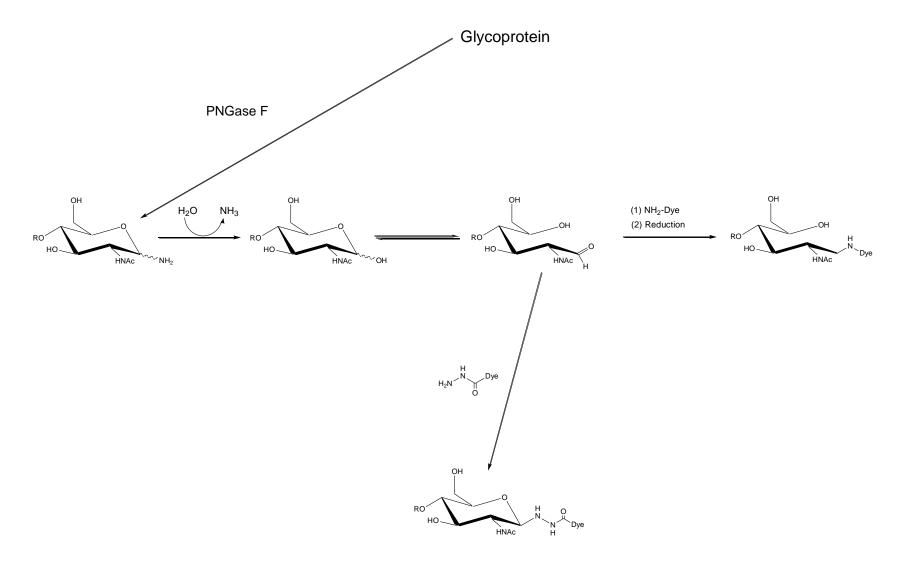




Our method (and almost all current CE/HPLC methods) analyze ONLY N-glycans



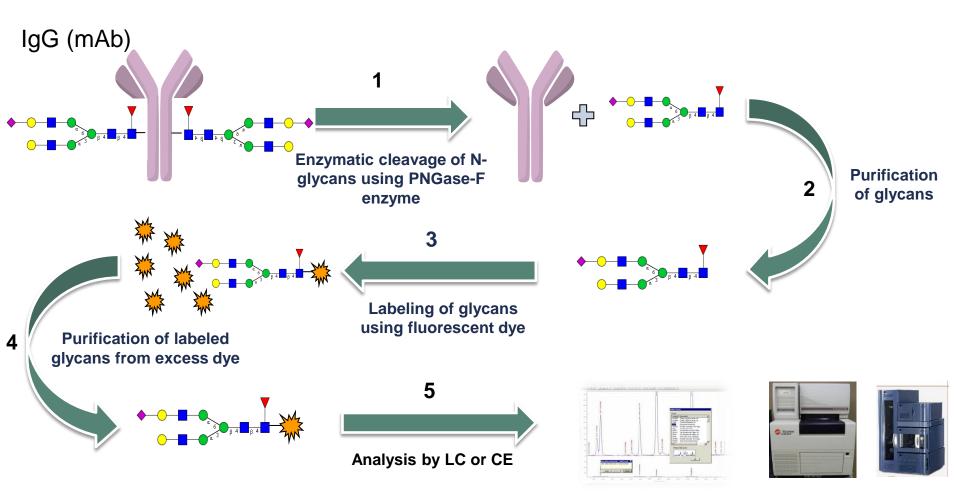
#### **Glycan Labeling Chemistries**



Predominantly Ring Closed B-Glycoside



## How N-Glycans are Analyzed?





## Current Glycan Analysis Challenges



- Single channel separation with low throughput (HPLC/UHPLC & single cap CE)
- · Poor data quality of high throughput methods (Chip based instruments)
- Labor Intensive workflow (multiple pipetting steps, use of spin columns, etc.)
- Use of toxic Sodium cyanoborohydride chemistry
- Use of lengthy vacuum centrifugation steps
- Generic software taking long analysis time
- Non-integrated solution from multiple vendors
- · Commercial sample prep kits with high cost per sample
- No validation support for result/solution



#### Thermo Fisher GlycanAssure



- Very high throughput (96 samples/7-9hrs)
- No compromise in CE separation time
- Easy magnetic bead based workflow
- Fewer pipetting steps & hands-on time
- Multiple fluorescent dyes for glycan labeling
- No use of Sodium cyanoborohydride
- No vacuum centrifugation steps
- Parallel analysis of 24 samples on AB DNA Analyzer
- Fit for use "app" style software
- Complete integrated solution with sample
  prep kits, CE instrument & software
- Reduced cost per sample

#### First Fully Integrated System Combining Throughput & Data Quality



#### **Product Overview**

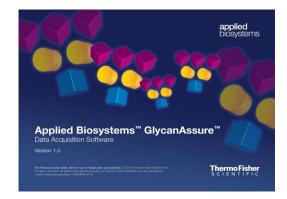


Turquoise Kit PN: A28678

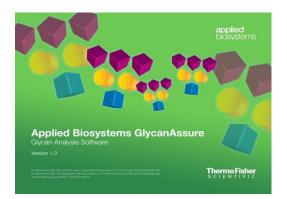




3500 & 3500XL PN: A30467 & A30556



#### Data Acquisition Software PN: A30750



Data Analysis Software PN: A30751



# General

First in class glycan analysis platform that combines both high throughput and data quality

# Specific\*

- Faster
- Simpler
- High Quality
- Cost-effective
- Integrated solution

\*Compared to other Commercial & Homebrew Products



### GlycanAssure Sample Prep Kits



- Three fluorescent dyes with distinct properties
- Traditional APTS and two Thermo proprietary dyes
- Faster glycan labeling with Teal & Turquoise dyes (30min)
- Magnetic beads for glycan purification and excess dye removal
- No need for excess dye removal with Teal (96 samples in 7hrs)
- Each kit with PNGase-F enzyme and beads for 96 samples

9-Aminopyrene-1,4,6-trisulfonic acid (APTS)



#### **GlycanAssure CE Instrumentation**



#### Key Features:

• 8 & 24-capillary systems for medium & high throughput analysis

• Single-line, 505 nm, solid-state, long-life laser that utilizes a standard power supply and requires no heat-removal

- Powerful, integrated data acquisition and analysis software that provides real-time assessment of data
- Radio frequency identification (RFID) technology that tracks key consumables data and records administrative information
- Advanced multiplexing capabilities for glycan analysis with up to six unique dyes
- Unrivaled application flexibility—one array and one polymer are used for most applications
- Simple setup, operation, and maintenance



#### **GlycanAssure CE Instrumentation**

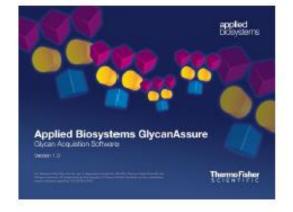


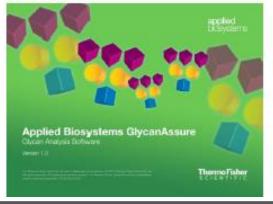


- Industry "Gold Standard" CE instrument
- Parallel analysis of 8 or 24 samples (< 2min/sample)
- Robust capillary array
- Calibration across capillaries using internal standard

## **GlycanAssure Software**

- Acquisition Dashboard
  - · Quick access to instrument status and data
- Run Setup
  - · Simplified and Intuitive chevron based, step by step workflow
- Reports
  - · Easily create reports with choice of templates
- Library
  - Stores historical information for plates, experiments, methods and reference
- Analysis Dashboard
  - Quick view of analyzed and un-analyzed samples. Allows to create data analysis projects
  - Quick access to Favorite Projects
- Data Processing
  - Quick comparison of analysis methods, alignment, normalization, and smoothing
  - · Library of analysis methods and recommendations for glycan analysis
- Analysis
  - Simplified graphics interface for manual integrations







#### **GlycanAssure: Data Acquisition Software**

# **Applied Biosystems GlycanAssure**

Glycan Acquistion Software

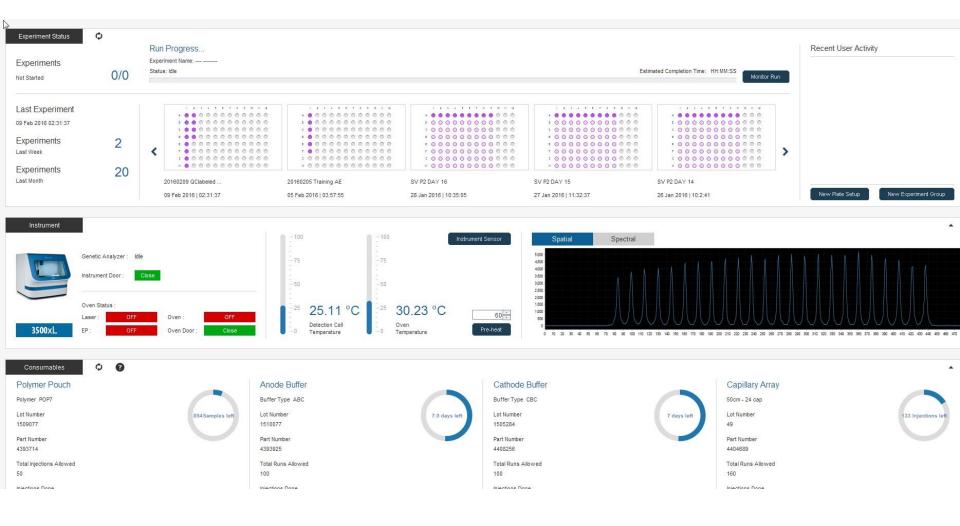
Version 1.0

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applied biosystems

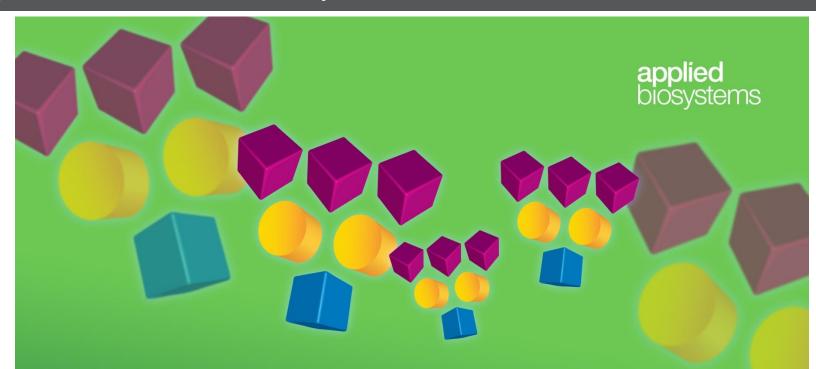


#### **GlycanAssure: Data Acquisition Software**





#### GlycanAssure: Data Analysis Software



#### Applied Biosystems<sup>™</sup> GlycanAssure<sup>™</sup> Data Analysis Software

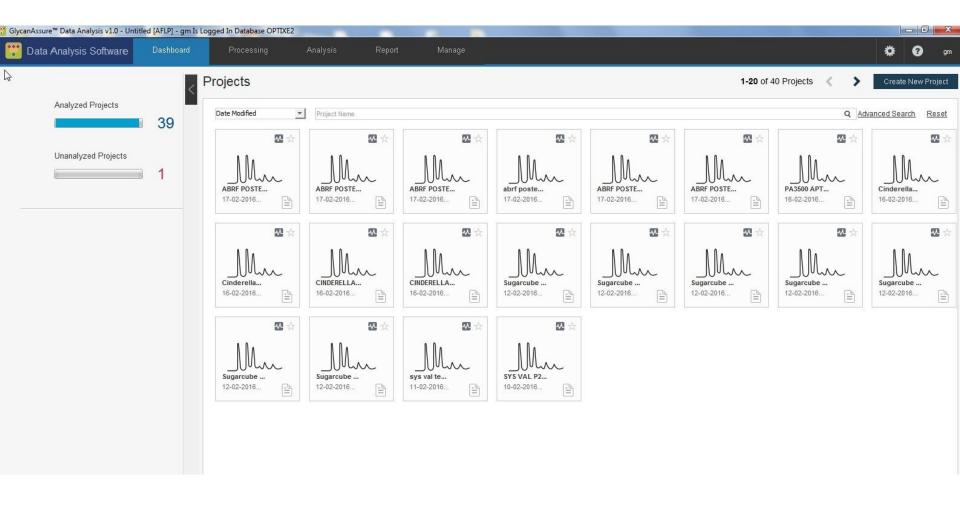
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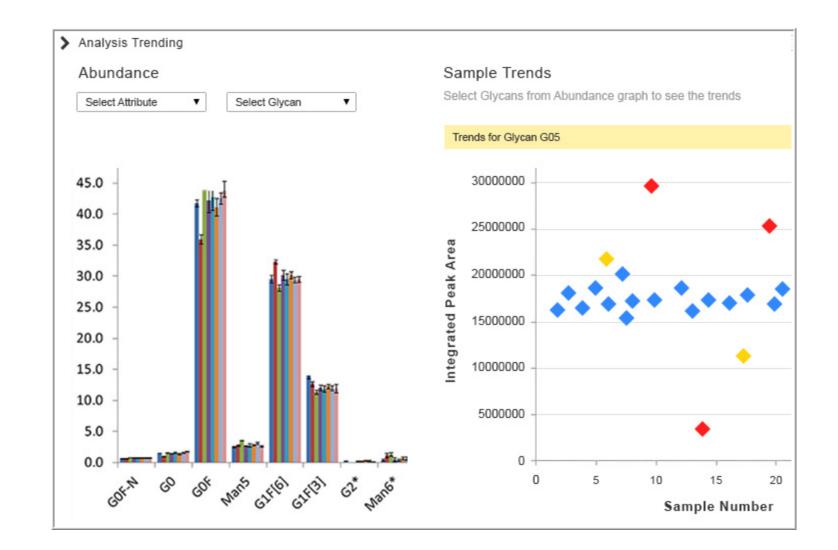


#### GlycanAssure: Data Analysis Software





#### GlycanAssure: Data Analysis Software

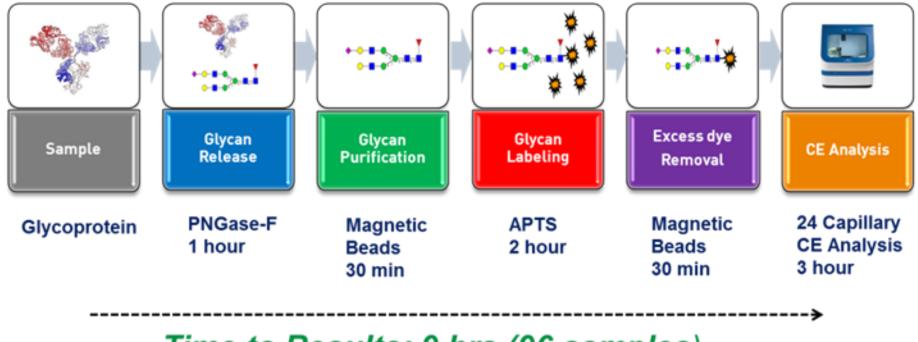




## Reports of AB Multi Capillary CE for Glycan Analysis

Glycobiology vol. 11 no. 4 pp. 275-28	1, 2001						
Ultrasensitive profilin DNA-sequencing equ		d oligosaccharides using standard					
	-	Display Settings: V Abstract	<u>Send to:</u> ⊘				
		<u>J Proteome Res.</u> 2010 Dec 3;9(12):6655-64. doi: 10.1021/pr100802f. Epub 2010 Nov 2.					
		Optimized workflow for preparation of APTS-labeled N-glycans allowing high-through plasma glycomes using 48-channel multiplexed CGE-LIF.	out analysis of human				
Nico Callewaert, Steven Geys Roland Contreras <sup>1</sup>							
Unit of Fundamental and Applied Mol Molecular Biology, Ghent University a for Biotechnology, K.LLedeganckstra	ind Flanders Interuniversity Institute For	Author information					
Received on July 21, 2000; revised on November 16, 2000	·	High-throughput methods for oligosaccharide analysis are required when searching for glycan-based biomarkers. Next to mass spectrometry-base methods, which allow fast and reproducible analysis of such compounds, further separation-based techniques are needed, which allow for quantit analysis. Here, an optimized sample preparation method for N-glycan-profiling by multiplexed capillary gel electrophoresis with laser-induced fluorescence detection (CGE-LIF) was developed, enabling high-throughput glycosylation analysis. First, glycans are released enzymatically from denatured plasma glycoproteins. Second, glycans are labeled with APTS using 2-picoline borane as a nontoxic and efficient reducing agent. Rea conditions are optimized for a high labeling efficiency, short handling times, and only limited loss of sialic acids. Third, samples are subjected to					
Electrophoresis 2006, 27, 1363–1367 Kay Vogel Joachim Kuhn	Short Communicatio						
Knut Kleesiek Christian Götting	A novel ultra-sen	sitive method for the					
Institut für Laboratoriums-	quantification of glycosaminoglycan disaccharides using an a <u>utomated DNA</u>						
und Transfusionsmedizin, Herz- und Diabeteszentrum							
Nordrhein-Westfalen, Universitätsklinik der Ruhr- Universität Bochum,	sequencer	mAbs 6:1, 185–196; January/February 2014; © 2014 Landes Bioscience	REPORT				
Bad Oeynhausen, Germany	Analysis of glycosaminoglycan alterations in extracellular matrix membrane. In this report we des	High-throughput glycosylation analysis					
Received August 8, 2005 Revised October 8, 2005	as an example of GAG ∆disa						
Accepted October 10, 2005	equipment (DNA sequencer-ass presented methodology allows						
	sulfonic acid (APTS)-derived G	AG disacchari Dietmar Reusch, <sup>1,*</sup> Markus Haberger, <sup>1</sup> Tobias Kailich, <sup>1</sup> Anna-Katharina Heidenreich, <sup>1</sup> Michael Kampe, <sup>1</sup> Patrick Bulau and Manfred Wuhrer <sup>2,3</sup>	J <sup>1</sup>				
		Pharma Biotech Development Penzberg; Roche Diagnostics GmbH; Penzberg, Germany; "Center for Proteomics and Metabolomics; Leiden University Medical Centr The Netherlands; "Division of BioAnalytical Chemistry; Department of Chemistry and Pharmaceutical Sciences; VU University Amsterdam; Amsterdam; The Nether Comparison of BioAnalytical Chemistry; Department of Chemistry and Pharmaceutical Sciences; VU University Amsterdam; Amsterdam; The Nether Comparison of BioAnalytical Chemistry; Department of Chemistry and Pharmaceutical Sciences; VU University Amsterdam; Amsterdam; The Nether Comparison of BioAnalytical Chemistry; Department of Chemistry and Pharmaceutical Sciences; VU University Amsterdam; Amsterdam; The Nether Comparison of BioAnalytical Chemistry; Department of Chemistry and Pharmaceutical Sciences; VU University Amsterdam; Amsterdam; The Nether Comparison of BioAnalytical Chemistry; Department of Chemistry and Pharmaceutical Sciences; VU University Amsterdam; Amsterdam; The Nether Chemistry Chemistry; Chemist					
		Keywords: monoclonal antibody (mAb), IgG glycosylation, automation, multiplexed capil- lary electrophoresis, DNA analyzer, HILIC-UPLC, APTS labeling, LC-MS					

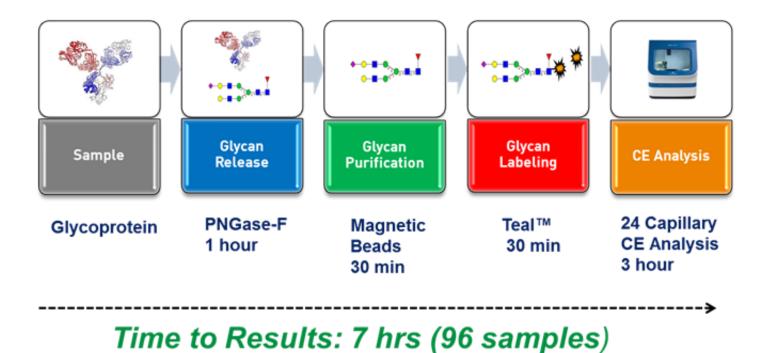
## GlycanAssure APTS Workflow



Time to Results: 9 hrs (96 samples)

\*Total 9hrs taking into account initial glycoprotein prep time and time between steps. Total time of the workflow is only 7hrs. CE takes 45min to analyze 24 samples

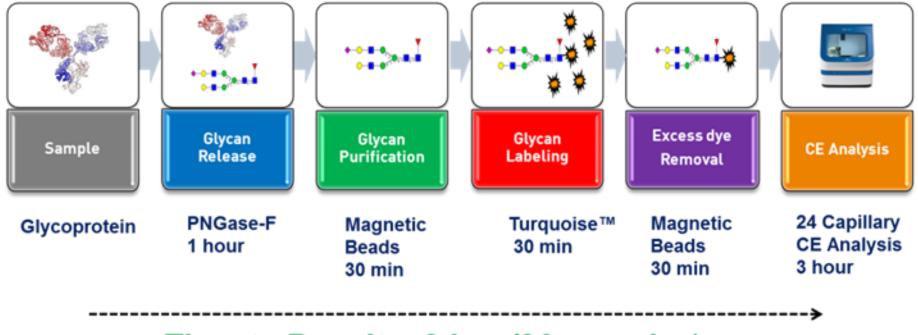




\*Total 7hrs taking into account initial glycoprotein prep time and time between steps. Total time of the workflow is only 5hrs. CE takes 45min to analyze 24 samples



## GlycanAssure: Turquoise Workflow



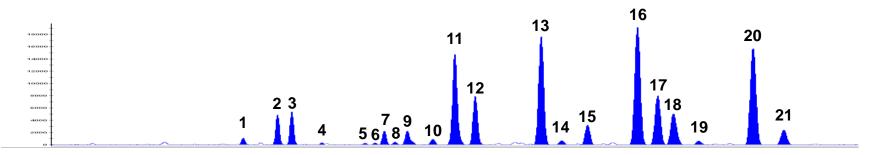
Time to Results: 8 hrs (96 samples)

\*Total 8hrs taking into account initial glycoprotein prep time and time between steps. Total time of the workflow is only 5.5hrs. CE takes 45min to analyze 24 samples



### Magnetic Bead Workflow Vs Carbon Spin Columns

- Glycans purified by <u>carbon column</u> or <u>magnetic beads</u> in triplicate
- Glycans were labeled with APTS for separation on 3500xL

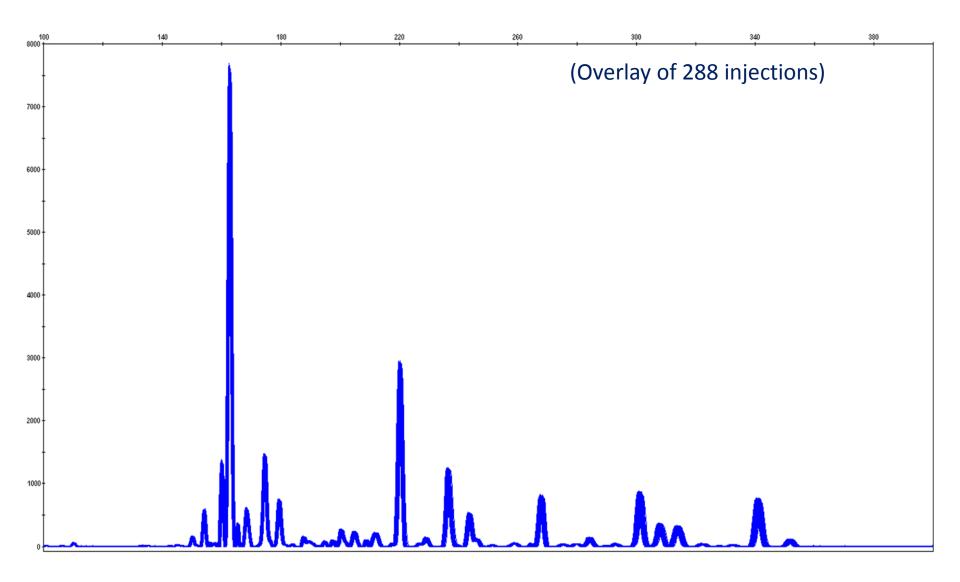


	Peak	1	2	3	4	5	6		7	8	9	10
Carbon	Rel % Area	0.69%	3.11%	3.54%	0.20%	<b>6 0.1</b>	<b>6% 0.</b> 1	<b>16%</b> 1	.55%	0.29%	1.96%	0.68%
column	CV%	5.12%	4.88%	5.75%	4.31%	6 5.3	8% 3.2	29% 3	8.80%	0.39%	2.52%	1.76%
Magnetic	Rel % Area	0.68%	2.82%	3.09%	0.23%	<b>6</b> 0.1	<b>6% 0.</b> 1	1 <b>8%</b> 1	.66%	0.26%	1.94%	0.68%
Beads	CV%	1.91%	3.63%	4.13%	1.69%	<b>%</b> 1.1	7% 1.5	52% (	).50%	2.28%	1.37%	1.12%
	Peak	11	12	13	14	15	16	17	18	3 19	20	21
Carbon	Rel % Area	12.28%	6.49%	15.64%	0.51%	2.80%	18.15%	7.58%	6 5.2	.5% 0.5	6% 15.9	5% 2.43%
column	CV%	2.64%	1.76%	0.95% <sup>~</sup>	18.63%	1.26%	1.19%	2.15%	6 1.4	1% 1.5	7% 1.6	8% 2.62%
Magnetic Beads	Rel % Area	12.77%	6.44%	15.76%	0.53%	2.66%	18.38%	7.67%	6 5.1	6% 0.5	4% 16.0	0% 2.39%
	CV%	0.88%	1.46%	1.07%	1.53%	0.85%	0.59%	1.00%	<b>6 0.4</b>	6% 2.6	9% 0.6	0% 0.64%

# **Consistent Glycan Quantitation**

	10μց		50µ	g	100µg		
Peak #	%Area	%CV	%Area	%CV	%Area	%CV	
1	0.90%	4.05%	0.72%	3.06%	0.64%	5.07%	
2	0.61%	6.31%	0.45%	3.31%	0.39%	4.77%	
3	1.64%	2.22%	1.69%	2.46%	1.66%	2.68%	
4	0.42%	7.09%	0.42%	4.67%	0.38%	4.26%	
5	8.54%	2.47%	8.31%	2.19%	8.18%	1.48%	
6	0.76%	5.89%	0.55%	8.97%	0.42%	6.47%	
7	22.51%	2.04%	23.07%	4.25%	23.71%	1.60%	
8	0.63%	4.50%	0.71%	9.54%	0.71%	3.36%	
9	3.73%	0.83%	3.82%	2.60%	3.85%	1.28%	
10	23.52%	0.43%	23.33%	2.52%	23.84%	0.65%	
11	9.98%	0.52%	10.16%	3.37%	9.91%	0.88%	
12	5.37%	1.00%	5.46%	3.50%	5.42%	1.00%	
13	0.43%	4.27%	0.44%	4.53%	0.43%	2.12%	
14	19.58%	1.16%	19.53%	2.91%	19.20%	0.85%	
15	1.38%	2.25%	1.33%	4.83%	1.27%	1.84%	

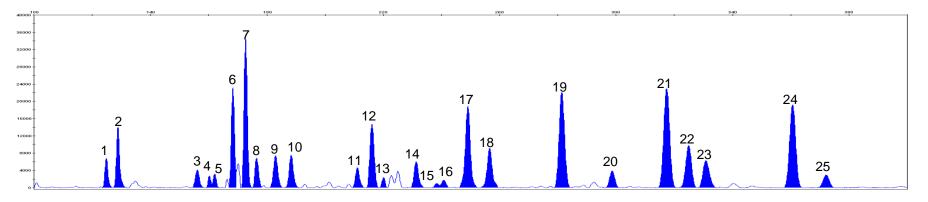
# Reproducibility of Glycan Separation





# Variability Across Capillaries of an Array

- Mixture of glycans from purified human serum IgG and bovine Fetuin
- Labeled with APTS & run on 3500xL CE (24 capillaries)



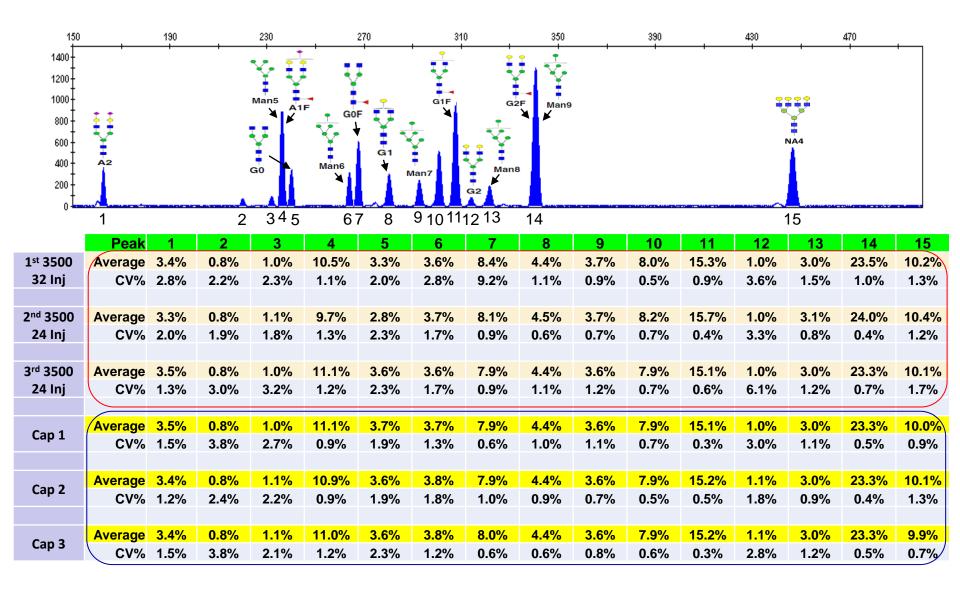
	Average	STDEV	CV%		Average	STDEV	CV%
1	1.8%	0.03%	1.5%	11	1.7%	0.03%	2.0%
2	3.9%	0.08%	2.1%	12	5.4%	0.08%	1.4%
3	1.4%	0.02%	1.5%	13	0.7%	0.08%	11.3%
4	0.6%	0.02%	3.1%	14	2.4%	0.04%	1.5%
5	0.8%	0.02%	2.3%	15	0.3%	0.02%	4.8%
6	5.8%	0.43%	7.4%	16	0.7%	0.01%	1.4%
7	9.1%	0.19%	2.1%	17	8.2%	0.07%	0.9%
8	2.3%	0.22%	9.8%	18	4.2%	0.04%	1.0%
9	2.7%	0.02%	0.8%	19	11.0%	0.09%	0.8%
10	2.7%	0.02%	0.7%	20	1.8%	0.03%	1.7%

	Average	STDEV	CV%
20	1.8%	0.03%	1.7%
21	11.7%	0.08%	0.7%
22	5.0%	0.05%	1.0%
23	3.6%	0.03%	0.8%
24	10.6%	0.09%	0.9%
25	1.7%	0.02%	1.1%

\*Data analysis was done using GeneMapper software that won't allow manual integration



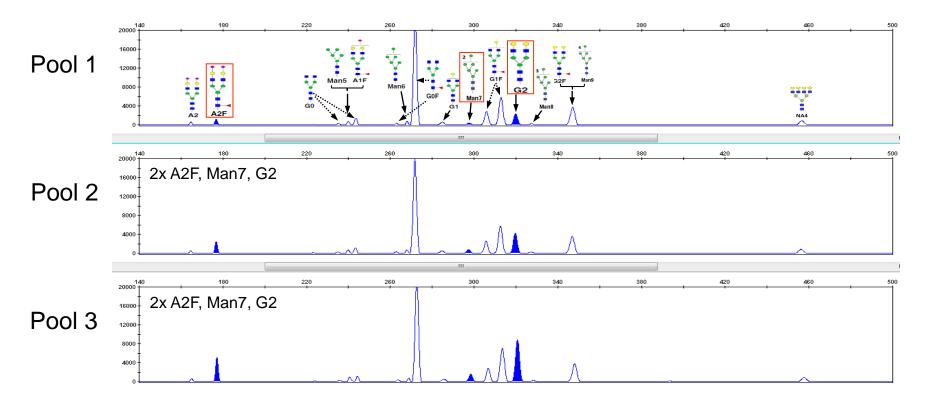
# Variability Across Multiple CE Instruments & Arrays





# Glycan Spike Recovery

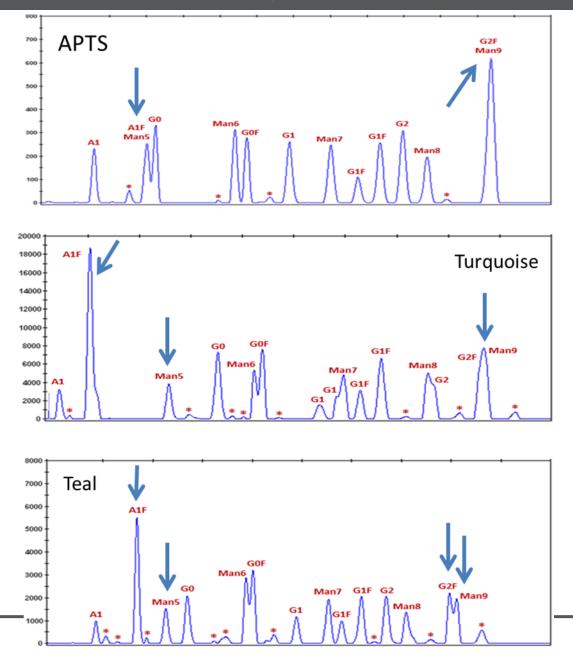
- 15 diverse types of purified glycans labeled with APTS
- Mixed to create large difference in concentrations (50% to <1%)</li>
- A2F (sialylated), Man7 (high mannose), and G2 (complex) glycans were increased serially by 2x



Increased signals of three specific glycans while the rest remain constant

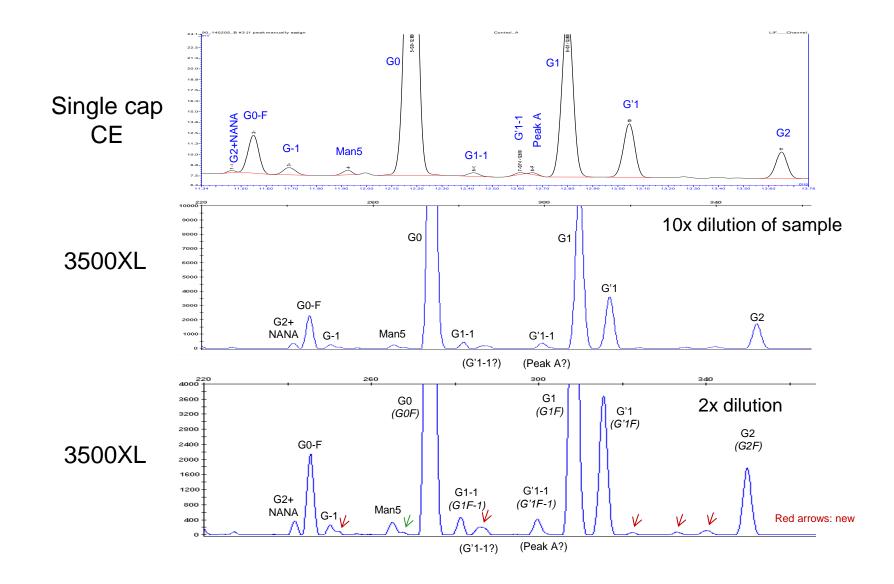


#### Better Glycan Separation Using Thermo Fisher Dyes



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#### Thermo Fisher 3500 Vs Leading Single Capillary CE





# GlycanAssure: Summary

#### Easy sample prep

- Magnetic bead based sample prep
- Hands-on-time <3 hrs for 96 samples</li>
- Less no. of pipetting steps
- No use of Sodium cyano borohydride
- No vacuum centrifugation steps
- Throughput
  - Sample prep & data of 96 samples in 7-9hrs
- Resolution
  - Sialyated glycans
  - Structural isomers
  - Fucose species
  - High Mannose species
- Dye Labeling
  - Multiple dyes with superior sensitivity
- Sensitivity
  - Low glycoprotein input
- Low Cost of Analysis
  - Robust instrument and capillaries with low running cost
- Software
  - Data Acquisition & Analysis software with novel features
- Integration
  - Full sample prep, hardware & software solution









