**PRODUCT INFORMATION**

**Taq DNA Polymerase (recombinant)**

<table>
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<tbody>
<tr>
<td>Taq DNA Polymerase, 5 µL</td>
<td>100 U</td>
<td>500 U</td>
<td>5 x 500 U</td>
<td>10 x 500 U</td>
</tr>
<tr>
<td>10X Taq Buffer with KCl</td>
<td>0.6 mM</td>
<td>2x1.25 mL</td>
<td>10x1.25 mL</td>
<td>20x1.25 mL</td>
</tr>
<tr>
<td>10X Taq Buffer with (NH₄)₂SO₄</td>
<td>0.6 mM</td>
<td>2x1.25 mL</td>
<td>10x1.25 mL</td>
<td>20x1.25 mL</td>
</tr>
<tr>
<td>25 mM MgCl₂</td>
<td>0.6 mL</td>
<td>2x1.25 mL</td>
<td>10x1.25 mL</td>
<td>20x1.25 mL</td>
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**Description**

Taq DNA Polymerase is a highly thermostable DNA polymerase of the thermophilic bacterium *Thermus aquaticus*. The enzyme catalyzes $5'\rightarrow3'$ synthesis of DNA, has no detectable $3'\rightarrow5'$ exonuclease (proofreading) activity and possesses $5'\rightarrow3'$ exonuclease activity. In addition, Taq DNA Polymerase exhibits deoxyribonuclease activity, which frequently results in the addition of extra adenosine at the 3'-end of PCR products. Recombinant Taq DNA Polymerase is ideal for standard PCR of amplicons 5 kb or shorter.

**Applications**

- Routine PCR amplification of DNA fragments up to 5 kb (1).
- Generation of PCR product for TA cloning.
- DNA labeling (2-4).
- DNA sequencing (5).

**Source**

*Escherichia coli* cells with a cloned *pol1* gene from *Thermus aquaticus YT1*.

**Definition of Activity Unit**

One unit of the enzyme catalyzes the incorporation of 10 nmol of deoxyribonucleotides into a polynucleotide fraction in 30 min at 74°C.

**Storage Buffer**

The enzyme is supplied in: 20 mM Tris-HCl (pH 8.0), 1 mM DTT, 0.1 mM EDTA, 100 mM KCl, 0.5% (v/v) Nonidet® P40, 0.5% (v/v) Tween® 20 and 50% (v/v) glycerol.

**Inhibition and Inactivation**

- Inhibitors: ionic detergents (deoxycholate, sarkosyl and SDS) at concentrations higher than 0.06, 0.02 and 0.01%, respectively (6).
- Inactivated by phenol/chloroform extraction.

**Protocol**

To prepare several parallel reactions and to minimize the possibility of pipetting errors, prepare a PCR master mix by mixing water, buffer, dNTPs, primers and Taq DNA Polymerase. Prepare sufficient master mix for the number of reactions plus one extra. Aliquot the master mix into individual PCR tubes and then add template DNA.

1. Gently vortex and briefly centrifuge all solutions after thawing.

2. Place a thin-walled PCR tube on ice and add the following components for each 50 µL reaction:

   - **10X Taq Buffer**: 5 µL
   - **dNTP Mix, 2 mM each (#R0241)**: 5 µL (0.2 mM of each)
   - **Forward primer (#R0581)**: 0.1-1.0 µM
   - **Reverse primer (#R0581)**: 0.1-1.0 µM
   - **25 mM MgCl₂**: 1-4 mM
   - **Template DNA**: 10 pg - 1 µg
   - **Tag DNA Polymerase**: 1.25 U
   - **Water, nuclease-free (#R0581)**: to 50 µL

   **Total volume**: 50 µL

   *Volumes of 25 mM MgCl₂ required for specific final MgCl₂ concentration:

   - Final concentration of MgCl₂, mM: 1.5, 2, 2.5, 3, 4
   - Volume of 25 mM MgCl₂ to be added for 50 µL reaction, µL: 2, 3, 4, 5, 6, 8

3. Gently vortex the samples and spin down.

4. If using a thermal cycler that does not use a heated lid, overlay the reaction mixture with 25 µL of mineral oil.

5. Perform PCR using recommended thermal cycling conditions:

<table>
<thead>
<tr>
<th>Step</th>
<th>Temperature, °C</th>
<th>Time</th>
<th>Number of cycles</th>
</tr>
</thead>
</table>
   | Initial denaturation | 95              | 1-3 min | 1
   | Denaturation     | 95              | 30 s  |                 |
   | Annealing       | Tm-5            | 30 s  | 25-40            |
   | Extension       | 72              | 1 min/kb |              |
   | Final Extension | 72              | 5-15 min | 1              |

**Guidelines for Preventing Contamination of PCR Reaction**

During PCR more than 10 million copies of template DNA are generated. Therefore, care must be taken to avoid contamination with other templates and amplicons that may be present in the laboratory environment. General recommendations to lower the risk of contamination are as follows:

- Prepare your DNA sample, set up the PCR mixture, perform thermal cycling and analyze PCR products in separate areas.
- Set up PCR mixtures in a laminar flow cabinet equipped with an UV lamp.
- Wear fresh gloves for DNA purification and reaction setup.
- Use reagent containers dedicated for PCR. Use positive displacement pipettes, or use pipette tips with aerosol filters to prepare DNA samples and perform PCR setup.
- Use PCR-certified reagents, including high quality water (e.g., Water, nuclease-free, (#R0581)).
- Always perform "no template control" (NTC) reactions to check for contamination.

**Guidelines for Primer Design**

Use the ThermoScientific REviewer primer design software at www.thermoscientific.com/reviewer or follow general recommendations for PCR primer design as outlined below:

- PCR primers are generally 15-30 nucleotides long.
- Optimal GC content of the primer is 40-60%. Ideally, C and G nucleotides should be distributed uniformly along the primer.
- Avoid placing more than three G or C nucleotides at the 3'-end to lower the risk of non-specific priming.
- If possible, the primer should terminate with a G or C at the 3'-end.
- Avoid self-complementary primer regions, complementarities between the primers and direct primer repeats to prevent hairpin formation and primer dimerization.
- Check for possible sites of undesired complementary between primers and template DNA.
- When designing degenerate primers, place at least 3 conserved nucleotides at the 3'-end.
- When introducing restriction enzyme sites into primers, refer to the table “Cleavage efficiency close to the termini of PCR fragments” located on www.thermoscientific.com/onebio to determine the number of extra bases required for efficient cleavage.
- Differences in melting temperatures (Tm) between the two primers should not exceed 5°C.

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COMPONENTS OF THE REACTION MIXTURE

Template DNA
Optimal amounts of template DNA in the 50 µL reaction volume are 0.01-1 ng for both plasmid and phage DNA, and 0.1-1 µg for genomic DNA. Higher amount of template increases the risk of generation of non-specific PCR products. Lower amount of template reduces the accuracy of the amplification. All routine DNA purification methods are suitable for template preparation e.g., Thermo Scientific GeneJET Genomic DNA Purification Kit (#K0721) or GeneJET™ Plasmid Miniprep Kit (#K0502). Trace amounts of certain agents used for DNA purification, such as phenol, EDTA and proteinase K, can inhibit DNA polymerases. Ethanol concentration and repeated ethanol precipitation of DNA pellet with 70% ethanol normally removes trace contaminants from DNA samples. 

MgCl₂ concentration
Due to the binding of Mg²⁺ to dNTPs, primers and DNA templates, MgCl₂ concentration needs to be optimized for maximal PCR yield. The recommended concentration range is 1-4 mM. If the MgCl₂ concentration is too low, the yield of PCR product could be reduced. On the contrary, non-specific PCR products may appear and the PCR fidelity may be reduced if the MgCl₂ concentration is too high.

Taq DNA Polymerase is supplied with two buffers: Taq buffer with KCl and Taq buffer with (NH₄)₂SO₄. K+ stabilizes primer annealing whereas NH₄⁺ has a destabilizing effect especially on weak hydrogen bonds between mismatched primer-template base pairs. Therefore for standard PCR with Taq DNA Polymerase and 0.2 mM dNTPs the recommended MgCl₂ concentrations are in general lower 1.5x to 2.5x than when using Taq buffer with KCl. Due to antagonistic effects of NH₄⁺ and Mg²⁺, Taq buffer with (NH₄)₂SO₄ offers higher primer specificity in a broad range of magnesium concentrations at variety of annealing temperatures. If the DNA samples contain EDTA or other metal chelators, the Mg²⁺ ion concentration in the PCR mixture should be increased accordingly (1 molecule of EDTA binds one Mg²⁺).

dNTPs
The recommended final concentration of each dNTP is 0.2 mM. In certain PCR applications, higher dNTP concentrations may be necessary. Due to the binding of Mg²⁺ to dNTPs, the MgCl₂ concentration needs to be adjusted accordingly. It is essential to have equal concentrations of all four nucleotides (dATP, dCTP, dGTP and dTTP) present in the reaction mixture.

To achieve 0.2 mM concentration of each dNTP in the PCR mixture, use the following volumes of dNTP mixes:

<table>
<thead>
<tr>
<th>Volume of PCR mixture (µL)</th>
<th>dNTP Mix 2 mM each (µL)</th>
<th>dNTP Mix 10 mM each (µL)</th>
<th>dNTP Mix 25 mM each (µL)</th>
</tr>
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<tbody>
<tr>
<td>50 µL</td>
<td>0.25 µL</td>
<td>5 µL</td>
<td>1 µL</td>
</tr>
<tr>
<td>25 µL</td>
<td>1 µL</td>
<td>2.5 µL</td>
<td>0.5 µL</td>
</tr>
<tr>
<td>20 µL</td>
<td>2 µL</td>
<td>2 µL</td>
<td>0.4 µL</td>
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Primers
The recommended concentration range of the PCR primers is 0.1-1 µM. Excessive primer concentrations increase the probability of mispriming and generation of non-specific PCR products. For degenerate primers higher primer concentrations in the range of 0.3-1 µM are often favorable.

CYCLING PARAMETERS

Initial DNA denaturation
It is essential to completely denature the template DNA at the beginning of PCR to ensure efficient utilization of the template during the first amplification cycle. If the GC content of the template is 50% or less, an initial 1-3 min denaturation at 95°C is sufficient. For GC-rich template this step should be prolonged up to 10 min. If longer initial denaturation step is required, or the DNA is denatured at a higher temperature, Taq DNA Polymerase should be added after the initial denaturation step to avoid a decrease in its activity.

Denaturation
A DNA denaturation time of 30 seconds per cycle at 95°C is normally sufficient. For GC-rich DNA templates, this step can be prolonged to 3-4 min. DNA denaturation can also be enhanced by the addition of either 10-15% glycerol or 10% DMSO, 5% formamide or 1-1.5 M betaine. The melting temperature of the primer-template complex decreases significantly in the presence of these reagents. Therefore, the annealing temperature has to be adjusted accordingly. In addition, 10% DMSO and 5% formamide inhibit DNA polymerases by 50%. Thus, all the enzyme of the amplification should be increased if these additives are used.

Primer annealing
The annealing temperature should be 5°C lower than the melting temperature (Tm) of the primers. Annealing for 30 seconds is normally sufficient. If non-specific PCR products appear, the annealing temperature should be optimized stepwise in 1°C increments. When additives which change the melting temperature of the primer-template complex are used (glycerol, DMSO, formamide and betaine), the annealing temperature must also be adjusted.

Extension
The optimal extension temperature for Taq DNA Polymerase is 70-75°C. The recommended extension step is 1 min at 72°C for PCR products up to 2 kb. For larger products, the extension time should be prolonged by 1 min/kb.

Number of cycles
The number of cycles may vary depending on the amount of template DNA in the PCR mixture and the expected PCR product yield. If less than 10 copies of the template are present in the reaction, about 40 cycles are required. For higher template amounts, 25-35 cycles are sufficient.

Final extension
After the last cycle, it is recommended to incubate the PCR mixture at 72°C for additional 5-15 min to fill-in any possible complete reaction products. If the PCR product will be cloned into TA vectors (for instance, using Thermo Scientific InstAClone PCR Cloning Kit (#K1123)), the final extension step may be prolonged to 30 min to ensure the highest efficiency of 3'-dA tailing of PCR product. If the PCR product will be used for cloning using Thermo Scientific CloneJET PCR Cloning Kit (#K1231), the final extension step can be omitted.

Troubleshooting
For troubleshooting please visit www.thermoscientific.com/onebio

References

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