

## PRODUCT INFORMATION

# AloI

**#ER1491** 100 U

**Lot:** \_\_\_\_ **Expiry Date:** \_\_

5'... ↓ 7(N)GAAC (N)<sub>6</sub>TCC (N)<sub>(12-13)</sub> ↓ ...3'  
3'... ↑<sub>(12-13)</sub> (N)CTTG (N)<sub>6</sub>AGG (N)<sub>7</sub> ↑ ...5'

Concentration: 2 U/μL  
Supplied with: 1 mL of 10X Buffer R  
1 mL of 10X Buffer Tango

**Store at -20°C**



In total 3 vials.

BSA included

[www.thermoscientific.com/onebio](http://www.thermoscientific.com/onebio)

## RECOMMENDATIONS

**1X Buffer R** (for 100% AloI digestion)

10 mM Tris-HCl (pH 8.5), 10 mM MgCl<sub>2</sub>, 100 mM KCl,  
0.1 mg/mL BSA.

**Incubation temperature**

30°C\*.

**Unit Definition**

One unit is defined as the amount of AloI required to digest 1 μg of lambda DNA in 1 hour at 30°C in 50 μL of recommended reaction buffer.

**Dilution**

Dilute with Dilution Buffer (#B19): 10 mM Tris-HCl (pH 7.4 at 25°C), 100 mM KCl, 1 mM EDTA, 1 mM DTT, 0.2 mg/mL BSA and 50% glycerol.

**Double Digests**

Thermo Scientific Tango Buffer is provided to simplify buffer selection for double digests. 98% of Thermo Scientific restriction enzymes are active in a 1X or 2X concentration of Tango™ Buffer. Please refer to [www.thermoscientific.com/doubledigest](http://www.thermoscientific.com/doubledigest) to choose the best buffer for your experiments.

1X Tango Buffer: 33 mM Tris-acetate (pH 7.9 at 37°C), 10 mM magnesium acetate, 66 mM potassium acetate, 0.1 mg/mL BSA.

\* Incubation at 37°C results in 20% activity.

## Storage Buffer

Alol is supplied in: 10 mM Tris-HCl (pH 7.4 at 25°C), 100 mM KCl, 1 mM DTT, 1 mM EDTA, 0.2 mg/mL BSA and 50% glycerol.

## Recommended Protocol for Digestion

- Add:  
nuclease-free water      16 µL  
10X Buffer R                2 µL  
DNA (0.5-1 µg/µL)        1 µL  
Alol                            0.5-2 µL\*\*
- Mix gently and spin down for a few seconds.
- Incubate at 30°C for 1-16 hours\*\*.

The digestion reaction may be scaled either up or down.

## Recommended Protocol for Digestion of PCR Products Directly after Amplification

- Add:  
PCR reaction mixture      10 µL (~0.1-0.5 µg of DNA)  
nuclease-free water        18 µL  
10X Buffer R                2 µL  
Alol                            1-2 µL\*\*
- Mix gently and spin down for a few seconds.
- Incubate at 30°C for 1-16 hours\*\*.

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\*\* See Star Activity.

## Thermal Inactivation

Alol is inactivated by incubation at 65°C for 20 min.

## ENZYME PROPERTIES

### Enzyme Activity in Thermo Scientific REase Buffers, %

B	G	O	R	Tango	2X Tango
0-20	0-20	0-20	100	20-50	100

## Star Activity

An excess of Alol (6 U/µg lambda DNA x 2 hours) may result in star activity.

## Methylation Effects on Digestion

Dam: may overlap— effect not determined.

Dcm: may overlap – no effect.

CpG: may overlap – cleavage impaired.

EcoKI: never overlaps – no effect.

EcoBI: never overlaps – no effect.

## Stability during Prolonged Incubation

A minimum of 0.1 units of the enzyme is required for complete digestion of 1 µg of lambda DNA in 16 hours at 30°C.

## Number of Recognition Sites in DNA

λ	ΦX174	pBR322	pUC57	pUC18/19	pTZ19R/U	M13mp18/19
7	0	0	0	0	1	1

For **CERTIFICATE OF ANALYSIS** see back page

## Note

- Alol produces double-strand cuts on both sides from interrupted recognition site. Its unique feature is a degenerate cleavage point on the 3' side of the recognition sequence (12 or 13 nt away).
- The presence of S-adenosylmethionine in a reaction mixture results in incomplete cleavage with Alol.
- At least two copies of Alol recognition site are required for efficient cleavage.
- Alol may remain associated with the cleaved DNA. This may cause DNA band shifting during electrophoresis. To avoid atypical DNA band patterns, use the 6X DNA Loading Dye&SDS Solution (#R1151) for sample preparation or heat the digested DNA in the presence of SDS prior to electrophoresis.

## CERTIFICATE OF ANALYSIS

### Overdigestion Assay

No detectable change in the specific fragmentation pattern is observed after a 10-fold overdigestion with Alol (5 U/μg lambda DNA x 2 hours) (*see* Star Activity).

### Ligation and Recleavage (L/R) Assay

The ligation and recleavage assay was replaced with LO test after validating experiments showed LO test ability to trace nuclease and phosphatase activities with sensitivity that is higher than L/R by a factor of 100.

### Labeled Oligonucleotide (LO) Assay

No detectable degradation of single-stranded or double-stranded labeled oligonucleotides occurred during incubation with 5 units of Alol for 4 hours.

### Blue/White (B/W) Cloning Assay

The B/W assay was replaced with LO test after validating experiments showed LO test ability to detect nuclease and phosphatase activities with sensitivity that equals to that of B/W test

Quality authorized by:



Jurgita Zilinskiene

**PRODUCT USE LIMITATION**

This product is developed, designed and sold exclusively *for research purposes and in vitro use only*. The product was not tested for use in diagnostics or for drug development, nor is it suitable for administration to humans or animals.

Please refer to [www.thermoscientific.com/onebio](http://www.thermoscientific.com/onebio) for Material Safety Data Sheet of the product.

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