Protocol

Phenol/Chloroform Extraction and Ethanol Precipitation

This protocol is for the Phenol/Chloroform Extraction and Ethanol Precipitation

1. Mix your sample with 1 volume of Tris-saturated phenol and 1 volume of chloroform. Centrifuge at 10,000 rpm for 5 min at room temperature.
2. Transfer the upper aqueous phase to a fresh tube. Add an equal volume of chloroform and mix. Centrifuge at 10,000 rpm for 5 min at room temperature. Repeat.
3. Transfer the upper aqueous phase to a fresh tube. Add 1/10 the volume of 3 M Sodium Acetate Solution or 2 M sodium chloride.
4. Add 2.5 volumes of ethanol or an equal volume of isopropanol to precipitate DNA.
5. Incubate the mixture for 30 min at -20°C.
6. Centrifuge for 10 min at 10,000 rpm. Then discard the supernatant and rinse the pellet with 70% cold ethanol.
7. Air-dry the pellet. Dissolve in Water, nuclease-free or TE buffer.

Note
Use Glycogen to maximize the yield of DNA during precipitation.

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