## SuperScript™ IV One-Step RT-PCR System

**USER GUIDE** Pub. No. MAN0016518 Rev. A.0

		_	
1	7	긔	
1	>	3	
W		Ш	
		4	7

#### Package contents

Catalog No. Size

12594025 25 reaction 12594100 100 reaction

12595025 25 reaction (with ezDNase<sup>™</sup> Enzyme)

100 reaction (with ezDNase<sup>™</sup> Enzyme) 12595100



# conditions

Store all contents at -20°C (non-frost-free)



Required materials Template: RNA

Gene specific primers (forward and reverse)



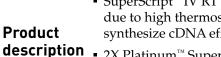
#### Timing

Product

Preparation time: 5 minutes

Total cycling time: varies with target size (1–3.5 hours)

- The Invitrogen<sup>™</sup> SuperScript<sup>™</sup> IV One-Step RT-PCR System is designed for sensitive end-point detection and analysis of RNA by RT-PCR. The convenient formulation enables both cDNA synthesis and PCR amplification to be performed in a single reaction tube using gene-specific primers.
- The system can amplify RNA targets up to 14 kb in length, and is compatible with multiplex RT-PCR.



- SuperScript<sup>™</sup> IV RT Mix allows high efficiency cDNA synthesis due to high thermostability, processivity, and ability to synthesize cDNA efficiently from a variety of RNA samples.
- 2X Platinum<sup>™</sup> SuperFi<sup>™</sup> RT-PCR Master Mix contains Platinum<sup>™</sup> SuperFi<sup>™</sup> DNA Polymerase which has high specificity, offers high yield, and is ideally suited for PCR applications that require sequence accuracy. The master mix uses a buffer system optimized for both RT and PCR reactions in the same tube.
- ezDNase<sup>™</sup> enzyme (available separately as Cat. No. 11766051) is included with the SuperScript<sup>™</sup> IV One-Step RT PCR System with ezDNase<sup>™</sup> Kits for protocols that require removal of residual gDNA contamination.



#### Online resources

Visit our product page for additional information and protocols. For support, visit thermofisher.com/support.

#### **Guidelines for RNA samples**

- This kit is optimized for use with 0.01 pg to 1 μg of total RNA, and is compatible with total RNA, mRNA, viral RNA, or *in vitro* transcribed RNA.
- High-quality, intact RNA is essential for RT-PCR, particularly for long targets. RNA must be devoid of RNase contamination and handled using aseptic conditions.
- Isolate total RNA with TRIzol<sup>™</sup> Reagent, the PureLink<sup>™</sup> RNA Mini Kit, or the MagMAX<sup>™</sup>-96 Total RNA Isolation Kit.
- Determine RNA quality using a bioanalyzer or by agarose gel electrophoresis.

#### **Guidelines for primers**

Kit

contents

- Use gene-specific primers (GSPs) with the SuperScript<sup>™</sup> IV One-Step RT-PCR System. Oligo(dT) or random primers are not recommended, because nonspecific products can be generated, thereby reducing the amount of target RT-PCR product.
- A final concentration of 0.5 µM for each primer is recommended, but further optimization may be necessary.
- Design primers that anneal to the mRNA sequence in exons on both sides of an intron or exon/exon boundary, to allow differentiation between the amplified cDNA and potential contaminating genomic DNA. If this approach is not feasible, use the protocol that involves residual gDNA removal with ezDNAse™ enzyme.
- Ensure that primers are not self-complementary or complementary to each other at the 3' ends.
- To calculate primer T<sub>m</sub> and estimate appropriate annealing temperatures for PCR, use the T<sub>m</sub> calculator at thermofisher.com/tmcalculator

## Limited product warranty

### Important licensing information

These products may be covered by one or more Limited Use Label Licenses. By use of these products, you accept the terms and conditions of all applicable Limited Use Label Licenses.

Disclaimer: TO THE EXTENT ALLOWED BY LAW, LIFE TECHNOLOGIES AND/OR ITS AFFILIATE(S) WILL NOT BE LIABLE FOR SPECIAL, INCIDENTAL, INDIRECT, PUNITIVE, MULTIPLE OR CONSEQUENTIAL DAMAGES IN CONNECTION WITH OR ARISING FROM THIS DOCUMENT, INCLUDING YOUR USE OF IT.

Corporate entity: Life Technologies | Carlsbad, CA 92008 USA | Toll Free in USA 1.800.955.6288

©2017 Thermo Fisher Scientific Inc. All rights reserved. All trademarks are the property of Thermo Fisher Scientific and its subsidiaries unless otherwise specified.



#### **Guidelines for cDNA synthesis and PCR**

- Keep all components, reaction mixes, and samples on ice.
- After preparing reaction mixes, transfer them to the preheated thermal cycler and start the RT-PCR program.
- The optimal conditions for RT-PCR depend on primer and target sequences.
- Efficient cDNA synthesis can be accomplished by a 10-minute incubation at 45–60°C. A 50°C incubation is recommended as a general starting point.
- For GC-rich or structurally complex RNA templates, increasing the cDNA synthesis incubation temperatures up to 55–60 °C may improve RT-PCR results.
- Use 40 cycles of amplification for RT-PCR products ≤3 kb, and 35 cycles for RT-PCR products >3 kb.
- For very low RNA input or rare targets, increasing the number of PCR cycles to 40 may improve results.
- The PCR extension time varies with the size of the amplicon (recommended extension time is approximately 30 seconds per 1 kb of amplicon).

## SuperScript™ IV One-Step RT-PCR System protocol

Step	Action	Procedure details				
1		Step	Temp	Time	No. cycles	
		Reverse transcription	45-60°C	10 min	1	
		RT inactivation/initial denaturation	98°C	2 min	I	
	1		98°C	10 s		
	Program thermal cycler	Amplification	55-72°C [1]	10 s	35–40 [2]	
			72°C	30 s/kb		
		Final extension	72°C	5 min	1	
		[1] <b>IMPORTANT!</b> Use the $T_m$ calculator at <b>thermofisher.com/tmcalculator</b> to determine actual annealing temperature. [2] Use 40 cycles for short ( $\leq 3$ kb) templates only.				
Prepare RT-PCR reaction mix		a. Combine the following components in a 0.2-mL, nuclease-free, thin-walled PCR tube on ice.				
		Component			Volume	
		2X Platinum™ SuperFi™ RT-PCR Master	25 μL			
	Forward primer (10 µM)	2.5 μL				
	Prepare RT-PCR	Reverse primer (10 µM)	everse primer (10 μM)			
	-	SuperScript™ IV RT Mix [1]			0.5 μL	
		Template RNA (0.01 pg to 1 μg total RNA)			varies	
		Nuclease-free water			to 50 μL	
		[1] Use all components except the SuperScript™ IV RT Mix for no RT control reactions.				
		b. Mix gently and ensure all the components are at the bottom of the amplification tube. Centrifuge briefly if needed.				
3	Run thermal cycler	Place the reaction in the pre-heated thermal cycler and run program set up in step 1.				

# SuperScript™ IV One-Step RT-PCR System with ezDNase™ Enzyme protocol

Step	Action		Procedure deta	ails				
		Prepare a 10 µL gDNA digestion reaction mix for each RT-PCR reaction. Mix the following components in a 0.2-mL, nuclease-free, thin-walled PCR tube on ice.						
Prepare gDNA digestion reaction mix	Component	Volu	me					
	10X ezDNase™ Buffer	1 <sub>L</sub>	ıL					
	ezDNase™ Enzyme	1 <sub>L</sub>	ıL					
	Template RNA (0.01 pg to 1 µg total RI	VA) var	ies					
	Nuclease-free Water	to 10	) μL					
2 5 min	Digest gDNA	<ul> <li>a. Gently mix and incubate at 37°C for 5 minutes.</li> <li>b. (Optional) For long RT-PCR products (&gt;3 kb) add 2 μL of 13.2 mM EDTA, then incubate at 65°C for 10 minutes.</li> <li>c. Briefly centrifuge the reaction mix and place the tube on ice.</li> </ul>						
3 Program thermal cycler		Step	Temp	Time	No. cycles			
		Reverse transcription	45-60°C	10 min				
		RT inactivation/initial denaturation	98°C	2 min	1			
			98°C	10 s				
	Program thermal cycler	Amplification	55-72°C [1]	10 s	35–40 [2]			
			72°C	30 s/kb				
		Final extension	72°C	5 min	1			
		[1] <b>IMPORTANT!</b> Use the T <sub>m</sub> calculator at [2] Use 40 cycles for short (≤3 kb) templa		<b>alculator</b> to determin	e actual annealing temperatu			
		a. Combine the following components in t	he tube on ice containi	ng the template RNA	٩.			
4 Prepare RT-PCR reaction mix		Component			Volume			
	Template RNA (from step 2)	10 μL						
	2X Platinum™ SuperFi™ RT-PCR Maste	25 µL						
	Forward primer (10 µM)	2.5 µL						
	reaction mix	Reverse primer (10 µM)	2.5 μL					
		SuperScript™ IV RT Mix [1]	0.5 μL					
		Nuclease-free water to 50 μL [1] Use all components except the SuperScript™ IV RT Mix for no RT control reactions.						
		b. Mix gently and ensure all the compone	•		ube. Centrifuge briefly if nee			
5	Run thermal cycler	Place the reaction in the pre-heated thermal cycler and run program set up in step 3.						