

# SiteClick™ Antibody Azido Modification Kit \*1 mg labeling\*

Catalog No. S10900

Pub. No. MAN0018872

Rev. A.0

## Product information

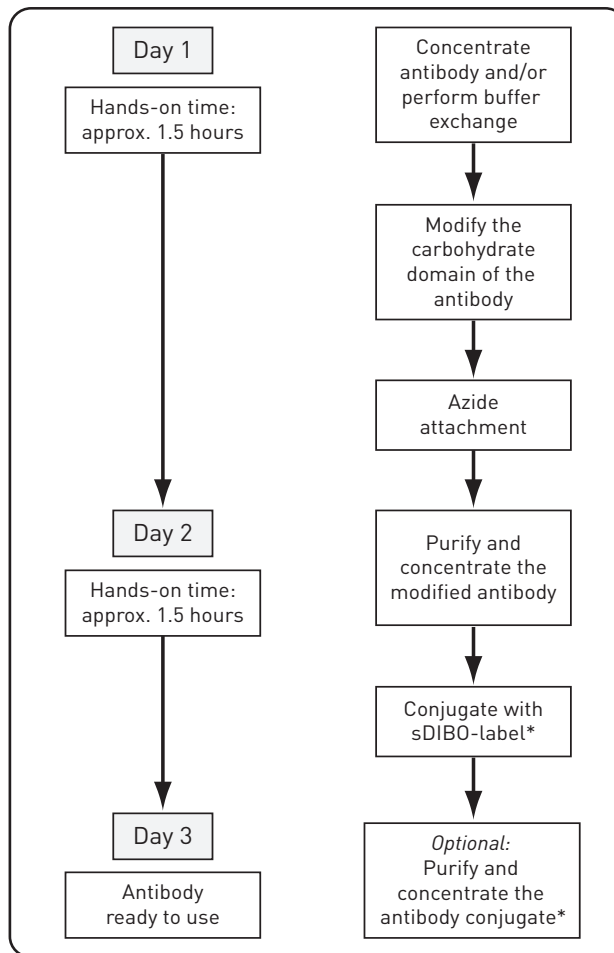
The SiteClick™ Antibody Azido Modification Kit allows you to specifically attach an azide moiety to the heavy chains of an unlabeled IgG antibody, ensuring that the antigen binding domains of the antibody remain unaltered for binding to your antigen target. The azide-modified antibody can then be covalently linked to SiteClick™ sDIBO Alkyne labels (available separately; see Table 2, page 3) in a copper-free click reaction without reducing the protein. This gives you the option to choose different fluorescent labels for your antibody, attach another molecule via streptavidin, or attach your own molecule via amine-reactive or amine-containing moieties depending on your assay.

Each SiteClick™ Antibody Azido Modification Kit contains sufficient reagents to perform one azido modification reaction starting with 1 mg of whole IgG produced in eukaryotic cells from any host species. The antibody concentrators provided in the kit are used to purify and concentrate the antibody at each step of the SiteClick™ antibody labeling workflow (Figure 1, page 2)

Table 1 Contents and storage

Material	Amount	Storage*
Antibody preparation buffer (Component A)	1.8 mL	<ul style="list-style-type: none"> <li>• 2–8°C</li> <li>• DO NOT FREEZE</li> </ul>
Antibody concentrator (small) (Component B)	each	
Collection tube (Component C)	each	
β-Galactosidase (Component D)	24 μL	
UDP-GalNAz (Component E)	440 μg	
20X Tris pH 7.0 (1M) (Component F)	1.8 mL	
Buffer additive (Component G)	60 μL	
β-1,4-galactosyltransferase (GalT) (Component H)	176 μL	
Antibody concentrator (large) (Component I)	each	

\* When stored as directed, this kit is stable for at least 6 months.



\* Requires the use of the SiteClick™ sDIBO Alkyne for SiteClick™ Antibody Labeling kits (available separately).

**Figure 1** SiteClick™ antibody azido modification and antibody labeling workflow. The SiteClick™ Antibody Azido Modification Kit is designed to be used with the SiteClick™ sDIBO Alkyne for SiteClick™ Antibody Labeling kits (available separately; see Table 2) for a complete antibody labeling workflow.

## Before you begin

- |  |  |
|--|--|
| <b>Equipment required</b>              | <ul style="list-style-type: none"> <li>• Centrifuge with fixed angle rotor that can accommodate 1.5-mL centrifuge tubes</li> <li>• Centrifuge with swinging bucket rotor that can accommodate 17 mm × 100 mm centrifuge tubes</li> </ul>   |
| <b>Required Materials not supplied</b> | <ul style="list-style-type: none"> <li>• 1 mg of whole IgG antibody produced in eukaryotic cells, preferably at a concentration of 2–20 mg/mL in a Tris-based buffer, free of carrier proteins and/or azide</li> <li>• Centrifuge tubes: 1.5-mL and 15-mL</li> <li>• Distilled water (dH<sub>2</sub>O)</li> <li>• PBS or TBS</li> <li>• SiteClick™ sDIBO Alkyne label (sDIBO-dye, sDIBO-biotin, or sDIBO-chelator) (Table 2, page 3).</li> </ul> |

**Table 2** SiteClick™ sDIBO Alkynes for SiteClick™ Antibody Labeling. The SiteClick™ sDIBO Alkyne labels (available separately) are used in conjunction with the SiteClick™ Antibody Azido Modification Kits (sufficient for 1 mg azide-modified antibody) or with engineered antibodies containing azido moieties to create high-quality antibody conjugates.

Product	Catalog No. <sup>[1]</sup>		
	100 µg kit	1 mg kit	5 mg kit
SiteClick™ Biotin sDIBO Alkyne	C20030	S10902	S10907
SiteClick™ pHrodo™ iFL Red sDIBO Alkyne	C20034	S10903	S10908
SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne	C20027	S10904	S10909
SiteClick™ Alexa Fluor™ 555 sDIBO Alkyne	C20028	—	—
SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne	C20029	S10906	S10911

<sup>[1]</sup> See Table 3 (page 9) for the amount of SiteClick™ sDIBO Alkyne label required to label 100 µg, 1 mg, and 5 mg azide-modified antibody with the SiteClick™ sDIBO Alkyne Kits available from Thermo Fisher Scientific.

- Caution**
- **IMPORTANT!** Avoid sodium azide throughout the protocol.
  - β-Galactosidase (Component D) may cause an allergic skin reaction, and it may cause allergy or asthma symptoms or breathing difficulties, if inhaled. Read the Safety Data Sheet (SDS), available at [thermofisher.com](http://thermofisher.com), before handling this reagent.
  - Dispose of the reagents in compliance with all pertaining local regulations. In case of contact with eyes, rinse immediately with plenty of water and seek medical advice. Always wear suitable laboratory protective clothing and gloves when handling these reagents.

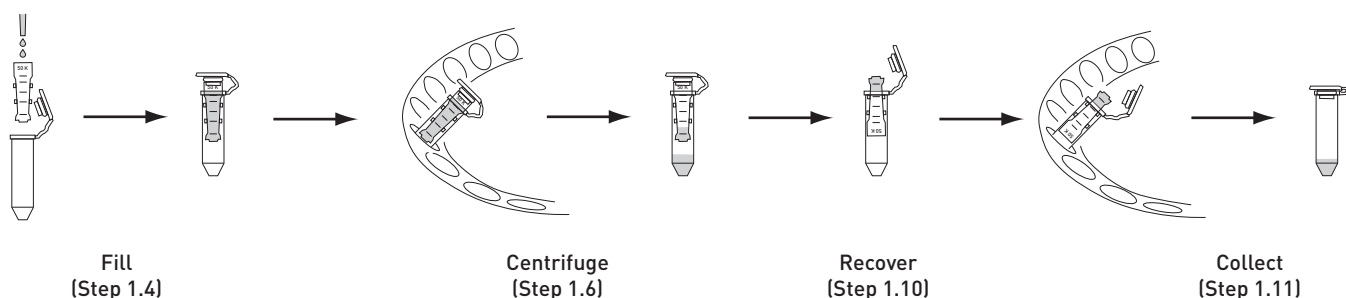
## Step 1. Concentrate antibody and/or perform buffer exchange

**Time required:** 1 hour

This antibody concentration and buffer exchange step is required if:

- Your antibody concentration is less than 10 mg/mL, and/or
- Your antibody is in a phosphate-based buffer (e.g. PBS), and/or
- Your antibody is in a buffer containing azide.

Before you begin, briefly centrifuge the tubes containing enzymes, substrates, or dyes to ensure all material is at the bottom of the tubes.



**Figure 2** Antibody concentration and/or buffer exchange

### Wash the antibody concentrator

- 1.1 Add 500  $\mu\text{L}$  of  $\text{dH}_2\text{O}$  to the small antibody concentrator (Component B) and cap the device as shown in Figure 2 (page 3).
- 1.2 Ensure that the cap strap and one membrane panel of the concentrator faces the center of the rotor, then centrifuge at  $5000 \times g$  for 6 minutes.
- 1.3 Discard the flow-through.

### Concentrate antibody and/or perform buffer exchange

- 1.4 Add a sufficient volume of antibody solution to contain 1 mg of antibody to the small antibody concentrator (Component B).
- 1.5 Dilute the added antibody to 500  $\mu\text{L}$  using the antibody preparation buffer (Component A).
- 1.6 Ensure that the cap strap and one membrane panel of the concentrator faces the center of the rotor, then centrifuge at  $5000 \times g$  for 6 minutes.
- 1.7 Discard the flow-through.
- 1.8 Add antibody preparation buffer (Component A) to the small antibody concentrator (Component B) so that the total volume in the concentrator is 500  $\mu\text{L}$ .
- 1.9 Ensure that the cap strap and one membrane panel of the concentrator faces the center of the rotor, then centrifuge at  $5000 \times g$  for 6 minutes.

**Note:** If the antibody volume in the concentrator is greater than 100  $\mu\text{L}$  following Step 1.9, centrifuge at  $5000 \times g$  for an additional 3 minutes or until the appropriate volume is achieved.

- 1.10 Invert the small antibody concentrator (Component B) into the collection tube (Component C) as shown in Figure 2.
- 1.11 Centrifuge for 3 minutes at  $1000 \times g$  to collect the concentrated antibody. Following collection, you should have approximately 100  $\mu\text{L}$  of concentrated antibody in the collection tube.

## Step 2. Modify the carbohydrate domain of the antibody

**Time required:** 5 minutes hands-on, then 6 hours incubation

### Add $\beta$ -galactosidase

- 2.1 Add 20  $\mu$ L of  $\beta$ -galactosidase (Component D) to the antibody collected in Step 1.11, as shown in Figure 3.
- 2.2 Wrap the tube cap with Parafilm™ laboratory film or similar, then incubate for 6 hours to overnight at 37°C.

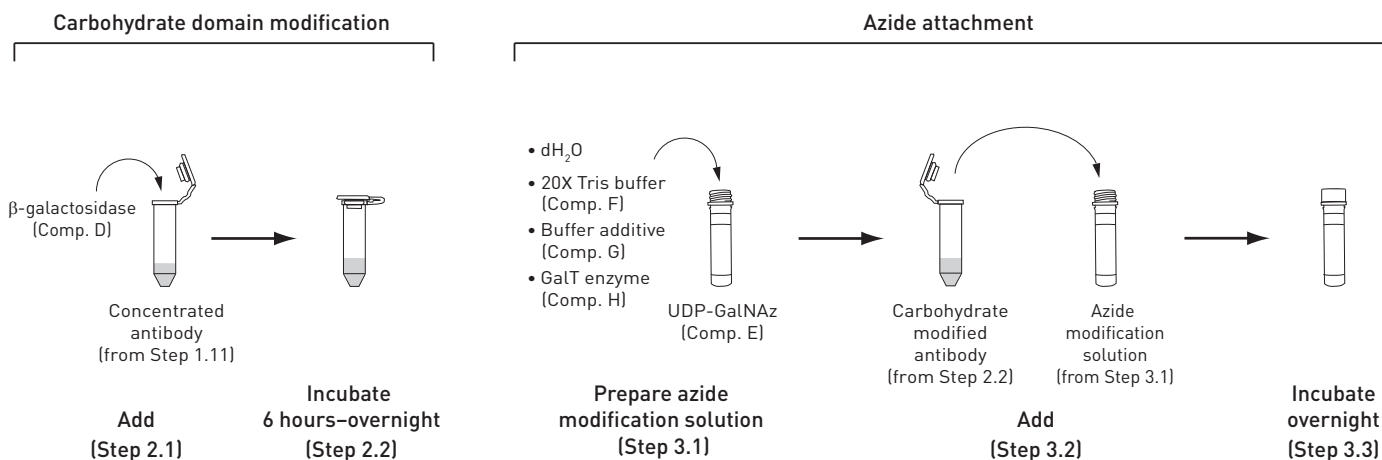


Figure 3 Modification of antibody carbohydrate domain and azide attachment

## Step 3. Azide attachment

**Time required:** 10 minutes hands-on, then overnight incubation

### Add GalT enzyme

- 3.1 Prepare the azide modification solution by adding the following components to the tube containing UDP-GalNAz (Component E), as shown in Figure 3 (page 5):
  - 12  $\mu$ L of  $dH_2O$
  - 18  $\mu$ L of 20X Tris buffer, pH 7.0 (Component F)
  - 40  $\mu$ L of buffer additive (Component G)
  - 160  $\mu$ L of GalT enzyme (Component H)
- 3.2 Vortex the reaction components, then add the modified antibody from Step 2.2 (120  $\mu$ L) to the tube.
- 3.3 Briefly centrifuge the tube, wrap the tube cap with Parafilm™ laboratory film or similar, then incubate overnight at 30°C.

## Step 4. Purify and concentrate the azide-modified antibody

**Time required:** 2 hours

- This step removes any excess substrate UDP-GalNAz.
- You can also use TBS or other phosphate-free buffers for purification and collection of the modified antibody (Steps 4.2–4.12). 20X Tris, pH 7.0 is provided for your convenience.

4.1 Prepare 10 mL of 1X Tris, pH 7.0 by adding 500  $\mu$ L of 20X Tris, pH 7.0 (Component F) to 9.5 mL of dH<sub>2</sub>O in a 15-mL conical tube. Vortex briefly to mix.

### Wash the antibody concentrator

- 4.2 Remove the conical collection tube from the large antibody concentrator (Component I).
- 4.3 Add 2 mL of 1X Tris, pH 7.0 (or TBS) to the large antibody concentrator (Component I) as shown in Figure 4 (page 6).
- 4.4 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at  $1200 \times g$  for 10 minutes. Discard the flow-through.

**IMPORTANT!** To avoid damage to the antibody concentrator during centrifugation, ensure that it is properly assembled and seated at the bottom of the rotor. The rim of the concentrate collection tube should be inside the rotor well. Check clearance before centrifugation.

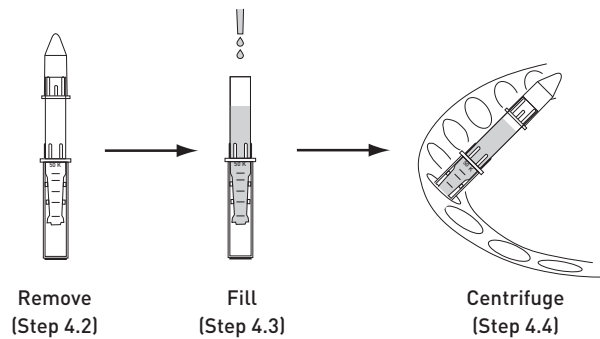


Figure 4 Wash the antibody concentrator

### Purify the antibody

4.5 Add 1.5 mL of 1X Tris pH 7.0 (or TBS) and 350  $\mu$ L of the azide-modified antibody from Step 3.3 to the large antibody concentrator (Component I) as shown in Figure 5.

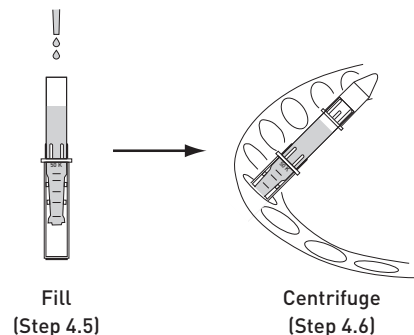


Figure 5 Purification and concentration of azide-modified antibody

- 4.6 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at  $1200 \times g$  for 8 minutes. Discard the flow-through.
- 4.7 Add 1X Tris pH 7.0 (or TBS) to a total volume of 2 mL to the large antibody concentrator (Component I). Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at  $1200 \times g$  for 11 minutes.
- 4.8 Discard the flow-through and repeat Step 4.7 two more times.

**Note:** If the antibody volume in the concentrator is greater than  $\sim 450 \mu\text{L}$  or if an antibody concentration of more than  $\sim 2.0 \text{ mg/mL}$  is desired, you can reduce the volume in the concentrator by additional centrifugation (e.g., at  $1200 \times g$  for an additional 5 minutes or until the appropriate volume is achieved).

**Note:** If you intend to conjugate the azide-modified antibody to macromolecule-sDIBO, measure the  $\text{OD}_{260}$  in the flow-through (with  $\epsilon_{260} = 9900 \text{ M}^{-1} \text{ cm}^{-1}$ ) following the removal of UDP-GalNAz. In this case, an additional Step 4.7 might be necessary to remove free UDP-GalNAz sufficiently.

#### Collect the azide-modified antibody

- 4.9 Invert the antibody concentrator into the conical collection tube as shown in Figure 6.
- 4.10 Centrifuge at  $1000 \times g$  for 3 minutes to collect the concentrated antibody.
- 4.11 Transfer the antibody from the conical collection tube to a 1.5-mL centrifuge tube.
- 4.12 Measure the  $\text{OD}_{280}$  (with  $\text{OD}_{280}$  at 1.4 =  $1 \text{ mg/mL}$ ) to determine the antibody concentration. Expected concentration is  $\sim 2\text{--}5 \text{ mg/mL}$ .

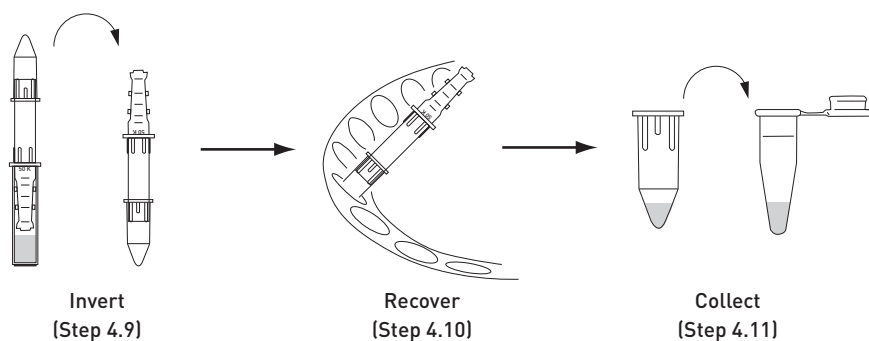


Figure 6 Collection of purified and concentrated azide-modified antibody

#### Store the azide-modified antibody

At this point, you can store the azide-modified antibody at  $2\text{--}8^\circ\text{C}$  until needed. Do not freeze the azide-modified antibody.

**IMPORTANT!** If you wish to perform a click reaction to conjugate your azide-modified antibody to a SiteClick™ sDIBO Alkyne label, do not add sodium azide to your modified antibody. Sodium azide must be avoided throughout the protocol.

## Step 5. Attach sDIBO-modified label to azide-modified antibody

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**Time required:** 5 minutes hands-on, then overnight incubation

This section provides instructions to covalently link the azide-modified antibody to a SiteClick™ sDIBO Alkyne label in a copper-free click reaction.

See Table 3 (page 9) for the amount of SiteClick™ sDIBO Alkyne label required to label 100 µg, 1 mg, and 5 mg azide-modified antibody with the SiteClick™ sDIBO Alkyne Kits available from Thermo Fisher Scientific.

### Materials required but not provided

- Azide-modified antibody (from Step 4.11) in a Tris-based buffer, free of carrier proteins and/or azide
- SiteClick™ sDIBO Alkyne label (available separately; see Table 3, page 9)
- Anhydrous DMSO (only required for dissolving SiteClick™ pHrodo™ iFL Red sDIBO Alkyne; included in Cat. No. S10903)
- Distilled water (dH<sub>2</sub>O)
- PBS or TBS
- 1.5-mL centrifuge tubes

### Caution

- **IMPORTANT!** Sodium azide must be avoided throughout the protocol.
- DMSO is known to facilitate the entry of organic molecules into tissues. Handle reagents containing DMSO using equipment and practices appropriate for the hazards posed by such materials.
- Read the Safety Data Sheet (SDS), available at [thermofisher.com](http://thermofisher.com), before handling the reagents.
- Dispose of the reagents in compliance with all pertaining local regulations. In case of contact with eyes, rinse immediately with plenty of water and seek medical advice. Always wear suitable laboratory protective clothing and gloves when handling these reagents.

### Add SiteClick™ sDIBO Alkyne to azide-modified antibody

- 5.1 Bring 1 mg azide-modified antibody (from Step 4.11) to a volume of 450 µL with 1X Tris pH 7.0 (or TBS) in the 1.5-mL centrifuge tube, then add 50 µL of SiteClick™ sDIBO Alkyne label.

**Note:** The SiteClick™ pHrodo™ iFL Red sDIBO Alkyne for SiteClick™ Antibody Labeling (Cat. No. S10903) is supplied lyophilized as a solid powder. Before use, dissolve the SiteClick™ pHrodo™ iFL Red sDIBO Alkyne in 50 µL of anhydrous DMSO, which is included in the kit.

Other SiteClick™ sDIBO Alkynes for SiteClick™ Antibody Labeling are supplied as 50-µL solutions in DMSO and do not need to be dissolved.

- 5.2 Vortex the reaction mixture, briefly centrifuge, and incubate overnight at 25°C.

**Note:** Following incubation, you can store the antibody conjugate at 2–8°C until needed (see “Store the antibody conjugate”, page 11) or purify it of the excess SiteClick™ sDIBO Alkyne label (Step 6, optional).



**Table 3** SiteClick™ sDIBO Alkynes for SiteClick™ Antibody Labeling. The SiteClick™ sDIBO Alkyne labels (available separately) are used in conjunction with the SiteClick™ Antibody Azido Modification Kits or with engineered antibodies containing azido moieties to create high-quality antibody conjugates.

Product	Catalog No. <sup>[4]</sup>			100 µg antibody		1 mg antibody		5 mg antibody	
	100 µg kit	1 mg kit	5 mg kit	in TBS <sup>[5]</sup>	sDIBO <sup>[6]</sup>	in TBS <sup>[5]</sup>	sDIBO <sup>[6]</sup>	in TBS <sup>[5]</sup>	sDIBO <sup>[6]</sup>
SiteClick™ Biotin sDIBO Alkyne	C20030	—	—	90 µL	10 µL	—	—	—	—
SiteClick™ pHrodo™ iFL Red sDIBO Alkyne <sup>[1]</sup>	C20034	—	—	90 µL	10 µL	—	—	—	—
SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne	C20027	—	—	90 µL	10 µL	—	—	—	—
SiteClick™ Alexa Fluor™ 555 sDIBO Alkyne	C20028	—	—	90 µL	10 µL	—	—	—	—
SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne	C20029	—	—	90 µL	10 µL	—	—	—	—
SiteClick™ Biotin sDIBO Alkyne	—	S10902	—	45 µL	5 µL	450 µL	50 µL	—	—
SiteClick™ pHrodo™ iFL Red sDIBO Alkyne <sup>[2]</sup>	—	S10903	—	45 µL	5 µL	450 µL	50 µL	—	—
SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne	—	S10904	—	45 µL	5 µL	450 µL	50 µL	—	—
SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne	—	S10906	—	45 µL	5 µL	450 µL	50 µL	—	—
SiteClick™ Biotin sDIBO Alkyne	—	—	S10907	45 µL	5 µL	450 µL	50 µL	2.25 mL	250 µL
SiteClick™ pHrodo™ iFL Red sDIBO Alkyne <sup>[3]</sup>	—	—	S10908	45 µL	5 µL	450 µL	50 µL	2.25 mL	250 µL
SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne	—	—	S10909	45 µL	5 µL	450 µL	50 µL	2.25 mL	250 µL
SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne	—	—	S10911	45 µL	5 µL	450 µL	50 µL	2.25 mL	250 µL

<sup>[1-3]</sup> Dissolve SiteClick™ pHrodo™ iFL Red sDIBO Alkyne in 25 µL<sup>[1]</sup>, 50 µL<sup>[2]</sup>, and 250 µL<sup>[3]</sup> DMSO, respectively.  
<sup>[4]</sup> 100 µg kits, 1 mg kits, and 5 mg kits contain sDIBO in 25 µL, 50 µL, and 250 µL DMSO, respectively, enough to label 2.5 × 100 µg, 10 × 100 µg, and 50 × 100 µg azide tagged antibody.  
<sup>[5]</sup> Antibody can be in 50 mM Tris (pH 7.0), TBS, PBS, or other thiol-free and sodium azide-free buffer.  
<sup>[6]</sup> sDIBO-derivatives are dissolved in DMSO.

## Step 6. Purify and concentrate the antibody conjugate (optional)

**Time required:** 1 hour

- The purification step removes any excess antibody that has not been conjugated with the sDIBO-modified label.
- You may use TBS or PBS for the purification and collection of the modified antibody (Steps 6.2–6.7)

**Materials required but not provided**

- Antibody conjugate (from Step 5.2)
- Antibody concentrator, large (included in the SiteClick™ sDIBO Alkyne for 1 mg SiteClick™ Antibody Labeling kits)

**Note:** The antibody concentrator included in the SiteClick™ sDIBO Alkyne kits (Component B; Component C in Cat. No. S10903) is identical to the large antibody concentrator (Component I) supplied with the 1 mg SiteClick™ Antibody Azido Modification Kit.

- Distilled water (dH<sub>2</sub>O)
- PBS or TBS
- 1.5-mL centrifuge tubes

## Wash the antibody concentrator

- 6.1 Remove the conical collection tube from the large antibody concentrator.
- 6.2 Add 2 mL of 1X Tris, TBS, or PBS to the large antibody concentrator (Component B in SiteClick™ sDIBO Alkyne kits) as shown in Figure 4 (page 6).
- 6.3 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at  $1200 \times g$  for 10 minutes. Discard the flow-through.

## Purify the antibody conjugate

- 6.4 Add 1.3 mL of 1X Tris, TBS, or PBS and 500  $\mu$ L of the sDIBO-modified antibody (from Step 5.2) to the large antibody concentrator (Component B).
- 6.5 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at  $1200 \times g$  for 12 minutes. Discard the flow-through.
- 6.6 Add 1X Tris, TBS, or PBS to a total volume of 2 mL to the large antibody concentrator (Component B).
- 6.7 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at  $1400 \times g$  for 12 minutes. Discard the flow-through.
- 6.8 Repeat Steps 6.6 and 6.7 at least three more times.

**Note:** If an antibody concentration of more than  $\sim 2\text{--}4$  mg/mL is desired, you can reduce the volume in the concentrator by additional centrifugation (e.g., at  $1400 \times g$  for an additional 3 minutes or until the appropriate volume is achieved).

## Collect the purified antibody conjugate

- 6.9 Invert the antibody concentrator into the conical collection tube as shown in Figure 7.
- 6.10 Centrifuge at  $1000 \times g$  for 3 minutes to collect the concentrated antibody.
- 6.11 Transfer the purified antibody conjugate from the conical collection tube to a new 1.5-mL centrifuge tube.

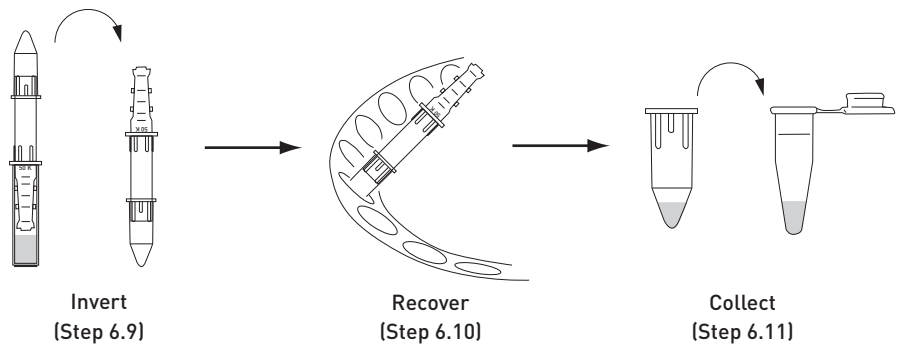


Figure 7 Optional purification and concentration of the labeled antibody conjugate

Store the antibody conjugate      Store the antibody conjugate at 2–8°C until needed. DO NOT FREEZE.

You can add sodium azide or thimerosal at this stage to a final concentration of 0.02% (w/v) for long term storage, if preferred.

## Step 7. Determine the Degree of Labeling (DOL) of sDIBO alkyne-labeled antibody (optional)

7.1 Determine the DOL from the  $A_{\text{dye}}/A_{280}$  ratio. Use Correction Factor ( $CF_{280}$ ) of the label at  $A_{280}$  to calculate (see Table 4).

$$(\text{Moles/L})_{\text{dye}} = A_{\text{dye}} / \epsilon_{\text{dye}}$$

$$(\text{Moles/L})_{\text{IgG}} = [A_{280} - (CF_{280} \times A_{\text{dye}})] / 203,000$$

$$\text{DOL} = (\text{Moles})_{\text{dye}} / (\text{Moles})_{\text{IgG}}$$

**Table 4** Molecular weight [MW], emission maxima ( $\lambda_{\text{max}}$ ), molar extinction coefficient ( $\epsilon_{\text{dye}}$ ), and Correction Factor ( $CF_{280}$ ) for the SiteClick™ sDIBO Alkynes for SiteClick™ Antibody Labeling.

Product	~MW	$\lambda_{\text{max}}$	$\epsilon_{\text{dye}}$ <sup>[1]</sup>	$CF_{280}$ <sup>[2]</sup>
SiteClick™ pHrodo™ iFL Red sDIBO Alkyne	~1800	560 nm	65,000	0.221
SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne	~1450	495 nm	73,000	0.134
SiteClick™ Alexa Fluor™ 555 sDIBO Alkyne	~1850	555 nm	145,000	0.091
SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne	~1900	655 nm	234,000	0.037

<sup>[1]</sup> Extinction coefficient at  $\lambda_{\text{max}}$  in  $\text{M}^{-1}\text{cm}^{-1}$ .  
<sup>[2]</sup> Correction factor for absorption readings ( $A_{280}$ ) at 280 nm; e.g.  $A_{280,\text{actual}} = A_{280,\text{observed}} - (CF_{280} \times A_{\text{dye}})$ .

### Example calculation with SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne

Determine the DOS from the  $A_{495}/A_{280}$  ratio, using  $CF_{280} = 0.134$  for Alexa Fluor™ 488 at  $A_{280}$  (see Table 4) for the calculation:

$$(\text{Moles/L})_{\text{dye}} = A_{495} / 73,000$$

$$(\text{Moles/L})_{\text{IgG}} = [A_{280} - (0.134 \times A_{495})] / 203,000$$

$$\text{DOL} = (\text{Moles})_{\text{dye}} / (\text{Moles})_{\text{IgG}}$$

## Appendix: Assay principle

In the first step of SiteClick™ conjugation, terminal galactose residues on the N-linked sugars in the Fc region of the antibody are removed by  $\beta$ -Galactosidase. The azide-containing sugar, GalNAz, is then added to the modified carbohydrate domain of the antibody via the  $\beta$ -1,4-galactosyltransferase (Gal-T)-catalyzed reaction targeting the terminal GlcNAc residues. This specific targeting maintains the integrity of the antigen binding site on the antibody. Finally, the antibody (now containing an azide moiety) is conjugated to the sDIBO-modified label in a copper-free click reaction with simple overnight incubation (Figure 8).

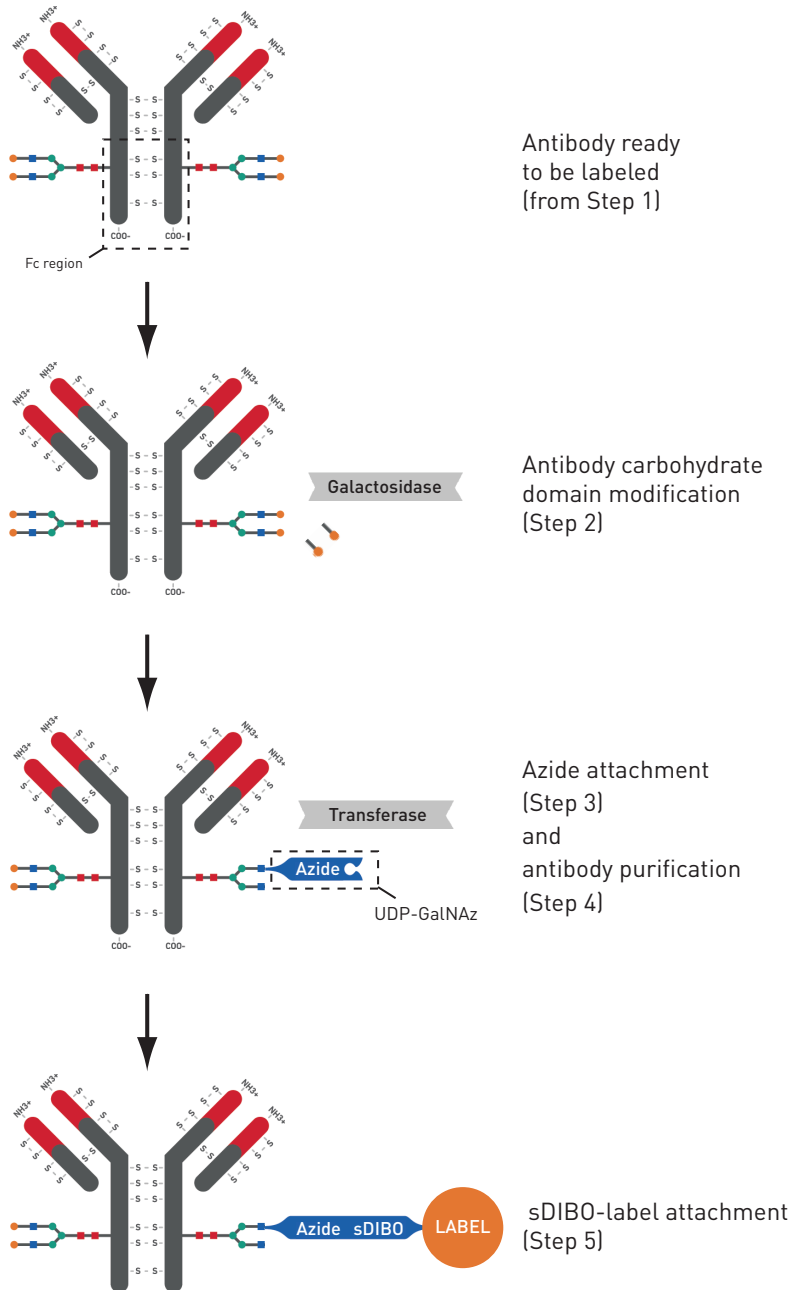


Figure 8 SiteClick™ conjugation reaction

## Ordering information

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Cat. No.	Product	Unit size
S10900	SiteClick™ Antibody Azido Modification Kit *1 mg labeling* . . . . .	1 kit

### Related products

#### 1 mg SiteClick™ sDIBO labels for azido-modified antibodies:

S10902	SiteClick™ Biotin sDIBO Alkyne for SiteClick™ Antibody Labeling. . . . .	1 kit
S10903	SiteClick™ pHrodo™ iFL Red sDIBO Alkyne for SiteClick™ Antibody Labeling. . . . .	1 kit
S10904	SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne for SiteClick™ Antibody Labeling. . . . .	1 kit
S10906	SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne for SiteClick™ Antibody Labeling. . . . .	1 kit

#### 100 µg SiteClick™ sDIBO labels for azido-modified antibodies:

C20027	SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne for SiteClick™ Antibody Labeling. . . . .	1 kit
C20028	SiteClick™ Alexa Fluor™ 555 sDIBO Alkyne for SiteClick™ Antibody Labeling. . . . .	1 kit
C20029	SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne for SiteClick™ Antibody Labeling. . . . .	1 kit
C20030	SiteClick™ Biotin sDIBO Alkyne for SiteClick™ Antibody Labeling. . . . .	1 kit
C20031	SiteClick™ Amine sDIBO Alkyne for SiteClick™ Antibody Labeling . . . . .	1 kit
C20032	SiteClick™ SDP Ester sDIBO Alkyne for SiteClick™ Antibody Labeling . . . . .	1 kit
C20034	SiteClick™ pHrodo™ iFL Red sDIBO Alkyne for SiteClick™ Antibody Labeling. . . . .	1 kit
S20033	SiteClick™ Biotin Antibody Labeling Kit . . . . .	1 kit
S20026	SiteClick™ Antibody Azido Modification Kit. . . . .	1 kit

## Customer and technical support

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- Order and web support
- Product information
  - User guides, manuals, and protocols
  - Certificates of Analysis
  - Safety Data Sheets (SDSs; also known as MSDSs)

**Note:** For SDSs for reagents and chemicals from other manufacturers, contact the manufacturer.

### Limited Product Warranty

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