# gibco

# CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> CD4, CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> CD8, CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> Release Buffer (Manual Workflow)

Catalog Numbers A56994, A56995, A5588303, A5588301, A5588302

Pub. No. MAN1000203 Rev. A

**WARNING!** Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves. Safety Data Sheets (SDSs) are available from thermofisher.com/support.

## Product description

Gibco<sup>™</sup> CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> CD4 and CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> CD8 positively isolate human T cell subsets. These CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> magnetic beads are intended to be used with the CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> Release Buffer that enables active release of magnetic beads at any time. This results in bead-free T cells for downstream processes such as gene-editing and activation.

This user guide provides a test protocol for a small-scale, manual process for isolation and active release of human T cells using the  $CTS^{\mathbb{T}}$ Detachable Dynabeads<sup>TM</sup> magnetic beads. For clinical T cell manufacturing, see  $CTS^{\mathbb{T}}$  Detachable Dynabeads<sup>TM</sup> CD4,  $CTS^{\mathbb{T}}$  Detachable Dynabeads<sup>TM</sup> CD8,  $CTS^{\mathbb{T}}$  Detachable Dynabeads<sup>TM</sup> Release Buffer (Automated Workflow) User Guide (Pub. No. MAN1000204) for a fully automated large-scale workflow. The manual protocol will be identical independent of using  $CTS^{\mathbb{T}}$  Detachable Dynabeads<sup>TM</sup> CD4 or  $CTS^{\mathbb{T}}$ Detachable Dynabeads<sup>TM</sup> CD8, individually, or a combination of the two  $CTS^{\mathbb{T}}$  Detachable Dynabeads<sup>TM</sup> magnetic beads together to isolate  $CD4^+$  and  $CD8^+$  T cells.

### Contents and storage

Product	Cat. No.	Volume	Capacity	Storage	
CTS <sup>™</sup> Detachable Dynabeads <sup>™</sup> CD4	A56994	15 mL	Can isolate and release up to $1.5\times10^9\text{CD4}^+\text{T}$ cells.	$5 \pm 3^{\circ}$ C; Store vial upright to keep	
CTS <sup>™</sup> Detachable Dynabeads <sup>™</sup> CD8	A56995	15 mL	Can isolate and release up to $1.5\times10^9\text{CD8}^+\text{T}$ cells.	beads in suspension.	
CTS <sup>™</sup> Detachable Dynabeads <sup>™</sup> Release Buffer	A5588303	212 mL	For use with up to 30 mL of CTS <sup>™</sup> Detachable Dynabeads <sup>™</sup> beads.	$5 \pm 3^{\circ}$ C; Protected from light.	
	IS <sup>™</sup> Detachable Dynabeads <sup>™</sup> A5588301		400 mL		
	A5588302	750 mL	For use with up to 120 mL of CTS <sup>™</sup> Detachable Dynabeads <sup>™</sup> beads.		

Table 1 Usage and storage for CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> (CD4 or CD8) and CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> Release Buffer

#### Table 2 Contents of CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> (CD4 or CD8) and CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> Release Buffer

Product	Contents
CTS <sup>™</sup> Detachable Dynabeads <sup>™</sup> (CD4 or CD8)	$4 \times 10^8$ beads/mL in phosphate buffered saline (PBS), pH 7.4, with 0.1% recombinant human albumin and 0.01% Tween <sup>™</sup> 80 detergent
CTS <sup>™</sup> Detachable Dynabeads <sup>™</sup> Release Buffer <sup>[1]</sup>	DPBS with 5 mM biotin and 0.5% recombinant human albumin, pH 7.2

<sup>[1]</sup> This product may develop minor protein aggregates after agitation, however, this does not affect product quality or performance.



# Required materials not supplied

Unless otherwise indicated, all materials are available through thermofisher.com. "MLS" indicates that the material is available from fisherscientific.com or another major laboratory supplier.

#### Table 3 Required materials not supplied

Item	Source				
Reagents					
CTS™ DPBS without calcium chloride, without magnesium chloride	A1285601				
Recombinant human albumin (rHA) or Human serum albumin (HSA)	MLS				
Equipment					
DynaMag <sup>™</sup> -5 Magnet, DynaMag <sup>™</sup> -15 Magnet, DynaMag <sup>™</sup> -50 Magnet	12303D, 12301D, 12302D				
Flow cytometer and fluorochrome labeled antibodies as required	MLS				
Vortex mixer	MLS				
Sample mixer allowing gentle tilting, rocking, or rotation	MLS				

# Generic protocol

Target cells = CD4<sup>+</sup> and/or CD8<sup>+</sup> cells

- 1. For optimal isolation, use a ratio of four beads per target cell, and a target cell concentration of 1 × 10<sup>7</sup> target cells/mL in DPBS with 1% rHA/HSA for isolation.
- 2. Incubate for 10 minutes with gentle tilting and rotation.

Bead-bound T cells are retained on a magnet and unbound cells can be washed away.

- 3. Incubate for 60 minutes with the CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> Release Buffer, which actively detaches the beads from the T cells.
- 4. Use a DynaMag<sup>™</sup> magnet to remove beads.

Released cells are bead-free and ready for downstream gene-modification and expansion.

# Protocol for manual small-scale isolation and activation of up to $5 \times 10^7$ T cells

1. Resuspend peripheral blood mononuclear cells (PBMCs) in DPBS with 1% rHA/HSA at a cell concentration of 1 × 10<sup>7</sup> target cells/mL. For examples of tubes and volumes see Table 4.

**Note:** If desired to isolate all T cells using a combination of the CTS<sup> $^{\text{M}}$ </sup> Detachable Dynabeads<sup> $^{\text{M}}$ </sup> CD4 and CTS<sup> $^{\text{M}}$ </sup> Detachable Dynabeads<sup> $^{\text{M}}$ </sup> CD8, considering the probable skewed ratio of target cells: adjust cell concentration to 1 × 10<sup>7</sup> CD3<sup>+</sup> cells.

- 2. Calculate the amount of CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> magnetic beads needed based on a recommended ratio of 4 beads per target cell.
- 3. Resuspend the magnetic beads by vortexing the vial for > 5 seconds. Then gently tilt and rotate for 15 minutes.
- 4. Wash the magnetic beads in a tube by following substep 4a to substep 4f.
  - a. Immediately after resuspension of beads, transfer the volume calculated in step 2 into a tube and place on a DynaMag<sup>™</sup> magnet. See Table 4.
  - b. Leave for 1 minute and remove supernatant.
  - c. Immediately, add 5–10x bead suspension volume of DPBS with 1% rHA/HSA.
  - d. Mix by vortexing 5 seconds and place on magnet.
  - e. Leave for 1 minute and remove washing buffer.
  - f. Resuspend beads in the same volume as originally added of bead suspension from step 2.
- 5. Add the washed magnetic beads to the cell suspension in step 1.
- 6. Incubate for 10 minutes with gentle tilting and rotation at room temperature.

- 7. Place tube in a DynaMag<sup>™</sup> magnet. Leave for 1 minute and transfer the supernatant with unbound cells to a new tube.
- Immediately wash the bead bound cells to improve purity of isolated cells. Add the same volume as cell suspension in step 1 of DPBS with 1% rHA/HSA to the tube, mix by 5 seconds vortexing.
- 9. Place tube in a DynaMag<sup>™</sup> magnet. Leave for 1 minute and transfer the supernatant to tube in step 7. The tube now contains your non-selected cells.
- 10. Repeat step 8 and step 9. The tube now contains your non isolated cells.
- 11. Immediately add CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> Release Buffer to the tube containing bead bound T cells.
- 12. Incubate for 60 minutes with gentle tilting and rotation at room temperature.
- 13. Resuspend magnetic beads and cells by pipetting up and down 10 times.
- 14. Place tube in a DynaMag<sup>™</sup> magnet. Leave for 1 minute and transfer the supernatant with the released cells to a new tube.
- 15. To increase the yield, wash the bead fraction by adding same volume as in step 11 with DPBS with 1% rHA/HSA and vortex for 5 seconds to mix.
- 16. Place tube(s) in a DynaMag<sup>™</sup> magnet. Leave for 1 minute and transfer the supernatant to the same tube as in step 14.
- 17. Repeat step 15 and step 16 (total of 3 washes). The tube now contains your selected target cells.
- **18.** Remove CTS<sup>™</sup> Detachable Dynabeads<sup>™</sup> Release Buffer from target cells by centrifugation at 350 × *g* for 5–10 minutes.
- 19. Resuspend T cells in culture media or any desired buffer and repeat centrifugation step.
- 20. Resuspend the target cells in culture media for downstream activation and expansion, or buffer for gene modification.

Table 4 Suggested volumes and equipment for isolation of target cells (manual workflow). Examples below are for single isolation of CD4<sup>+</sup> or CD8<sup>+</sup> cells.

Starting number of target cells and volume	CTS <sup>™</sup> Detachable Dynabeads <sup>™</sup> magnetic beads number and volume	Test tubes for isolation and release	DynaMag <sup>™</sup> magnet	CTS <sup>™</sup> Detachable Dynabeads <sup>™</sup> Release Buffer
1 × 10 <sup>7</sup> target cells in 1 mL	4 × 10 <sup>7</sup> beads (100 μL)	3.6 mL Nunc <sup>™</sup> tube (isolation) and 15 mL Nunc <sup>™</sup> tubes (release and wash)	DynaMag <sup>™</sup> -5 Magnet and DynaMag <sup>™</sup> -15 Magnet	2 mL
$5 \times 10^7$ target cells in 5 mL	$20 \times 10^7$ beads (500 µL)	15 mL Nunc <sup>™</sup> tube (isolation) and 50 mL Nunc <sup>™</sup> tubes (release and wash)	DynaMag <sup>™</sup> -5 Magnet and DynaMag <sup>™</sup> -50 Magnet	10 mL

### Limited product warranty

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For descriptions of symbols on product labels or product documents, go to thermofisher.com/symbols-definition.

#### Revision history: Pub. No. MAN1000203 A

Revision	Date	Description		
А	29 August 2024	New document for CTS <sup><math>^{\circ}</math></sup> Detachable Dynabeads <sup><math>^{\circ}</math></sup> CD4, CTS <sup><math>^{\circ}</math></sup> Detachable Dynabeads <sup><math>^{\circ}</math></sup> CD8, CTS <sup><math>^{\circ}</math></sup> Detachable Dynabeads <sup><math>^{\circ}</math></sup> Release Buffer (Manual Workflow)		

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