

Quant-iT™ microRNA Reagent

Catalog no. Q32883

Table 1 Contents and storage

Material	Amount	Concentration	Storage	Stability
Quant-iT™ microRNA reagent	1.0 mL	200X in DMSO	<ul style="list-style-type: none"> • Room temperature • Protect from light • Dessicate 	When stored as directed, product is stable for 1 year.
Number of assays: Sufficient material is provided for 1000 assays using 200 µL assay volume in a 96-well microplate format. The Quant-iT™ microRNA reagent can be adapted for use in cuvettes or 384-well microplates.				
Approximate fluorescence excitation/emission maxima: 500/525 nm.				

Introduction

The Quant-iT™ microRNA reagent allows easy and accurate quantification of small RNA (~20 nucleotides or base pairs), even in the presence of common contaminants such as salts, free nucleotides, solvents, detergents, and protein (*Appendix, Table 2, page 4*). The assay is highly selective for small RNA over rRNA or large mRNA (>1000 nt). We have been able to reproducibly quantify small RNA in pure samples at levels as low as 0.5 ng in the assay tube following the supplied protocol below.

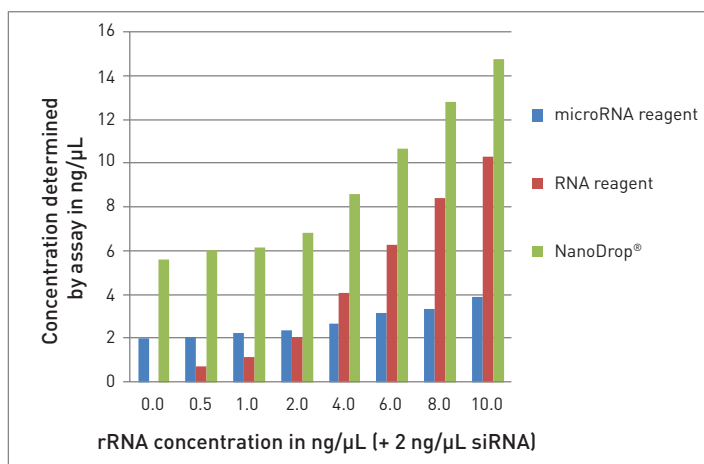


Figure 1 Comparison of detection techniques for accurate quantification of small RNA in the presence of ribosomal RNA. rRNA at the concentrations listed was spiked into solutions containing 2 ng/µL siRNA, then read using the Quant-iT™ microRNA reagent or the Quant-iT™ RNA reagent on the Qubit® fluorometer, or by 260 nm absorbance [A260] on the NanoDrop® spectrophotometer.

For Research Use Only. Not for use in diagnostic procedures.

The assay accurately detects as little as 0.5 ng small RNA and has a dynamic range of 5 ng/mL to 500 ng/mL (1–100 ng) in the core assay range. The assay is accurate for initial sample concentrations from 0.05 ng/ μ L to 100 ng/ μ L. To perform the assay, simply dilute the reagent, add your sample, and read the concentration using a fluorescent microplate reader.

Before you begin

Handling the Quant-iT™ microRNA reagent

There is no data available addressing the mutagenicity or toxicity of the Quant-iT™ microRNA reagent. This reagent is known to bind nucleic acid and is provided as a solution in DMSO; treat the reagent with the same safety precautions as all other potential mutagens. Dispose of the dye in accordance with local regulations.

Remove the Quant-iT™ microRNA reagent from storage, allow it to equilibrate to room temperature, and mix well. During all steps, protect the Quant-iT™ microRNA reagent concentrate and the working solution from light as much as possible.

Materials required

- 20X TE Buffer*, pH 7.5 (Cat. no. T11493)
- 1X TE Buffer, pH 7.5, with 0.01% CHAPS
- small RNA standard (e.g., *Silencer*® SelectGAPDH Positive Control siRNA, Cat. no. 4390849)
- 96-well microplates

*200 mM Tris, 1 mM EDTA (pH 7.5)

Experimental protocols

Prepare buffers and reagents

- 1.1 Allow all reagents to come to room temperature.
- 1.2 Prepare sufficient 1X TE Buffer (10 mM Tris, 1 mM EDTA, pH 7.5) for standard dilutions. This may be prepared by diluting 20X TE Buffer in E-pure™ water. This buffer solution also acts as the “0 small RNA standard” in the microRNA assay.
- 1.3 Prepare sufficient 1X TE Buffer + 0.01% CHAPS (w/v) assay buffer. All dye working solutions should be prepared in this buffer. For convenience, you may prepare an aqueous stock solution of 1% CHAPS (w/v) and dilute it to 0.01% (w/v) accordingly.

A solution of 200 mL buffer (20 mg of CHAPS in 200 mL of 1X TE buffer) is sufficient for 1000 assays following the protocol below.

In the presence of CHAPS, dye working solution is stable for at least 3 hours at room temperature.

- 1.4 Prepare small RNA standards. In the development of the microRNA assay reagent, a range of 0–10 ng/μL was used. A volume of 10 μL will be used for each reaction.

If you are using *Silencer*[®] Select siRNA, prepare a 100 ng/μL solution by adding 650 μL of 1X TE Buffer only (no CHAPS) to 5 nmoles of *Silencer*[®] Select siRNA solution. Prepare a 10 ng/μL solution by diluting the 100 ng/μL siRNA solution 10-fold in 1X TE buffer (no CHAPS) (i.e., add 100 μL of 100 ng/μL siRNA solution to 900 μL of 1X TE buffer).

Perform the microRNA assay

The following protocol describes a Quant-iT[™] microRNA assay with a total volume of 200 μL per microplate well.

- 2.1 Prepare the Quant-iT[™] microRNA working solution by diluting the Quant-iT[™] microRNA reagent 1:200 in 1X TE Buffer + 0.01% CHAPS (w/v). For a 96-well microplate, approximately 20 mL of the Quant-iT[™] microRNA working solution will be required for the samples and standard curve.
- 2.2 Load 200 μL of the Quant-iT[™] microRNA working solution (prepared in step 2.1) into the wells containing the standard and samples.
- 2.3 Pipet 10 μL of the standards prepared in step 1.4 into separate wells of a microplate and mix well.
- 2.4 Pipet 1–20 μL of each unknown small RNA sample to separate wells of a microplate and mix well. Duplicates or triplicates of the unknown samples are recommended. Some contaminating substances may interfere with the assay; see Table 2 in the *Appendix*, page 4.
- 2.5 Incubate the reactions for 2–5 minutes at room temperature.
- 2.6 Measure the fluorescence using a microplate reader (excitation/emission maxima are 500/525 nm). The fluorescence signal is stable for 3 hours.
- 2.7 Use a standard curve to determine the small RNA amounts. For the small RNA standards, plot amount vs. fluorescence, and fit a straight line to the data points.

Appendix

Contaminating substances A number of common contaminants have been tested in the Quant-iT™ microRNA assay, and most are well tolerated (Table 2, below). For untested contaminating substances, and, in general, for highest accuracy, the standards should be assayed under the same conditions as the unknowns. For example, if the experimental samples are in an unusual buffer and if 10 µL volumes of these samples are used, then add 10 µL volumes of the unusual buffer (lacking RNA) to the wells containing the standards.

Table 2 Effects of contaminants in the Quant-iT™ microRNA assay.

Contaminant	Final Concentration in the Assay	Concentration in 20 µL sample	Concentration in 10 µL sample	Result*
Sodium chloride	5 mM	50 mM	100 mM	OK
Magnesium chloride	1 mM	10 mM	20 mM	OK
Sodium acetate	5 mM	50 mM	100 mM	OK
Ammonium acetate	1 mM	10 mM	20 mM	OK
Ethanol	1%	10%	20%	OK
Phenol	0.1%	1%	2%	OK
Chloroform	0.2%	2%	4%	OK†
SDS	0.01%	0.1%	0.2%	NR
Triton® X-100	0.001%	0.01%	0.02%	OK
NTPs**	1:1 NTP:miRNA	1:1 NTP:miRNA	1:1 NTP:miRNA	OK
dsDNA	10:1 miRNA:dsDNA	10:1 miRNA:dsDNA	10:1 miRNA:dsDNA	OK†
ssDNA	10:1 miRNA:ssDNA	10:1 miRNA:ssDNA	10:1 miRNA:ssDNA	OK†
Oligo DNA	10:1 miRNA:Oligo	10:1 miRNA:Oligo	10:1 miRNA:Oligo	NR

* Results are given either as OK, usually less than 10% perturbation, or as NR (not recommended).

** A mixture of ATP, CTP, GTP, and UTP.

† Some distortion at the high end of the assay; for best results dilute the sample so concentration is ≤ 300 ng/mL.

‡ Immiscible.

Product List

Current prices may be obtained from our website or from our Customer Service Department.

Cat no.	Product Name	Unit Size
Q32883	Quant-iT™ microRNA reagent	1 mL
Related products		
4390849	Silencer® Select GAPDH Positive Control siRNA (Hs, Mm, Rn)	5 nmoles
T11493	20X TE Buffer, RNase-free	100 mL
ZC10003	Zoom® CHAPS	5 g

Purchaser Notification

These high-quality reagents and materials must be used by, or directly under the supervision of, a technically qualified individual experienced in handling potentially hazardous chemicals. Read the Safety Data Sheet provided for each product; other regulatory considerations may apply.

Obtaining Support

For the latest services and support information for all locations, go to www.lifetechnologies.com.

At the website, you can:

- Access worldwide telephone and fax numbers to contact Technical Support and Sales facilities
- Search through frequently asked questions (FAQs)
- Submit a question directly to Technical Support (techsupport@lifetech.com)
- Search for user documents, SDSs, vector maps and sequences, application notes, formulations, handbooks, certificates of analysis, citations, and other product support documents
- Obtain information about customer training
- Download software updates and patches

SDS

Safety Data Sheets (SDSs) are available at www.lifetechnologies.com/sds.

Certificate of Analysis

The Certificate of Analysis provides detailed quality control and product qualification information for each product. Certificates of Analysis are available on our website. Go to www.lifetechnologies.com/support and search for the Certificate of Analysis by product lot number, which is printed on the product packaging (tube, pouch, or box).

Limited Product Warranty

Life Technologies Corporation and/or its affiliate(s) warrant their products as set forth in the Life Technologies' General Terms and Conditions of Sale found on Life Technologies' website at www.lifetechnologies.com/termsandconditions. If you have any questions, please contact Life Technologies at www.lifetechnologies.com/support.

Disclaimer

LIFE TECHNOLOGIES CORPORATION AND/OR ITS AFFILIATE(S) DISCLAIM ALL WARRANTIES WITH RESPECT TO THIS DOCUMENT, EXPRESSED OR IMPLIED, INCLUDING BUT NOT LIMITED TO THOSE OF MERCHANTABILITY, FITNESS FOR A PARTICULAR PURPOSE, OR NON-INFRINGEMENT. TO THE EXTENT ALLOWED BY LAW, IN NO EVENT SHALL LIFE TECHNOLOGIES AND/OR ITS AFFILIATE(S) BE LIABLE, WHETHER IN CONTRACT, TORT, WARRANTY, OR UNDER ANY STATUTE OR ON ANY OTHER BASIS FOR SPECIAL, INCIDENTAL, INDIRECT, PUNITIVE, MULTIPLE OR CONSEQUENTIAL DAMAGES IN CONNECTION WITH OR ARISING FROM THIS DOCUMENT, INCLUDING BUT NOT LIMITED TO THE USE THEREOF.

Important Licensing Information

This product may be covered by one or more Limited Use Label Licenses. By use of this product, you accept the terms and conditions of all applicable Limited Use Label Licenses.

All trademarks are the property of Thermo Fisher Scientific and its subsidiaries, unless otherwise specified.

NanoDrop is a registered trademark of NanoDrop Technologies, LLC.

Triton is a registered trademark of Union Carbide Corporation.

Axygen is a registered trademark of Axygen, Inc.

©2015 Thermo Fisher Scientific Inc. All rights reserved.

