

# CellROX® Flow Cytometry Assay Kits

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Revision 1.0

Detailed protocol is available online at [www.lifetechnologies.com/manuals](http://www.lifetechnologies.com/manuals).

**WARNING!** For every chemical, read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves. Safety Data Sheets (SDSs) are available from [www.lifetechnologies.com/support](http://www.lifetechnologies.com/support).

| CellROX® Detection Reagent | Cat. No. | Ex/Em maxima (in nm) | Suggested Dead Cell Stain   |
|----------------------------|----------|----------------------|-----------------------------|
| CellROX® Deep Red          | C10491   | 640/665              | SYTOX® Blue Dead Cell Stain |
| CellROX® Green             | C10492   | 508/527              | SYTOX® Red Dead Cell Stain  |
| CellROX® Orange            | C10493   | 545/565              | SYTOX® Red Dead Cell Stain  |

For research use only. Not for use in diagnostic procedures.

## Quick Protocol

1. Grow cells to sub-confluency or prepare cell suspensions at  $10^4$ – $10^6$  cells/mL in complete medium.
2. Prepare negative and positive control (see detailed protocol).
3. Induce ROS in cells using the desired method.
4. Add the CellROX<sup>®</sup> Detection Reagent at a final concentration of 500–1000 nM (see detailed protocol); incubate for 30–60 minutes at 37°C, 5% CO<sub>2</sub>, protected from light.
5. During the final 15 minutes of staining, add 1 µL of the appropriate SYTOX<sup>®</sup> Dead Cell Stain solution per mL cell suspension.
6. Analyze samples on a flow cytometer equipped with appropriate lasers and filters for excitation and collection, applying standard fluorescence compensation (see detailed protocol).

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