applied biosystems

CytoScan[™] Assay SITE PREPARATION GUIDE

Manual Protocol

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Introduction

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Safety

Follow universal precautions in the laboratory. For waste disposal, follow federal, state, local, and within-country regulations.

Text alerts

Text alerts draw your attention to a particular piece of information. There are five types of text alerts: Note, Important, Caution, Warning, and Danger.

Note: Information that may be of interest or of help to a user but is not critical to the primary purpose of the text.

IMPORTANT! Information that is essential to the successful use of a product or the completion of a procedure and is not safety related.



CAUTION! Alerts the user of hazards that, if not avoided, can cause minor or moderate personal bodily injury and/or damage to an instrument or loss of data.



WARNING! Alerts the user to hazards that, if not avoided, can cause serious bodily injury or death, or produce potentially incorrect data that could cause harm to a patient.



DANGER! Alerts the user to an imminent hazard that, if not avoided, will cause serious bodily injury or death, or will produce a result that could cause serious harm to a patient.

Contamination prevention

Proper laboratory practice is necessary because previously amplified PCR product is the most likely potential source of contamination.

- Set up the laboratory areas for a single-direction workflow from Pre-PCR to Post- PCR.
- Use dedicated equipment for each area (e.g., thermal cyclers, microfuges, pipettes and tips, ice buckets, etc.).
- Place all reagents and master stocks in use area. *Do not move equipment between Pre- and Post-PCR Rooms, e.g., ice buckets, pipettes, etc.*
- Place separate copy of assay procedure in Pre- and Post-PCR areas.
- Follow the laboratory standard procedure for re-entry to Pre-PCR Clean area from post PCR Area.
- Use filter tips for all pipetting steps.

Equipment in Pre-PCR Clean Area

- Freezer, -25 to -15°C
- Ice bucket with ice
- Laminar flow cabinet or PCR cabinet
- Microfuge
- Pipettes on stand
- Plate centrifuge
- Refrigerator
- Thermal cycler
- Vortexer

Equipment in Post-PCR Area

- Electrophoresis gel box
- Electrophoresis power supply
- Freezer, -25 to -15°C
- Gel imager
- GeneChip[™] System 3000
 - Computer, monitor and keyboard
 - Fluidics station
 - Scanner
- GeneChip™ Hybridization Oven 645
- Ice bucket
- Magnetic stand
- · Pipettes on stand
- Spectrophotometer, UV/VIS, single or multichannel
- Vortexer
- Microcentrifuge
- Microfuge
- Refrigerated plate centrifuge
- Refrigerator
- Thermal cycler(s)
- Vortexer with foam tube adaptor

Equipment and calibration

Set up, maintain, and calibrate all equipment according to the manufacturer's instructions.

Reagent handling and storage

- Store all reagents at the indicated temperatures and conditions (see Chapter 3, Materials required)
- Do not store enzymes in a frost-free freezer
- Store reagents used for digestion, ligation, and PCR only in the Pre-PCR Clean Area

Thermal cyclers, 96-well plate, and adhesive seals

Use the thermal cyclers, 96-well plate, and adhesive films listed in Chapter 3, "Materials required".



Laboratory Setup

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Before you begin

Read Chapter 1, Introduction.

Configurations

Two setups are given following, one for two separate rooms and one for a single room with Pre- and Post-PCR separation. Always set up for a single-direction workflow.

Two separate rooms

This configuration greatly reduces the risk of sample contamination due to previously-amplified PCR products. These are the:

- Pre-PCR Clean Room
- Post-PCR Room

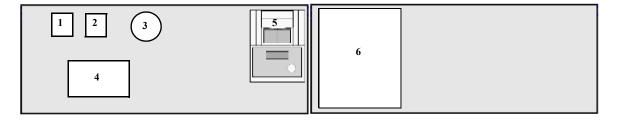
These rooms are set up to accommodate the steps in Table 1.

Table 1 Assay workflow when two separate rooms are used

Room	Template (Genomic DNA)	PCR Product
Pre-PCR Clean Room Assay steps: - Genomic DNA preparation - Digestion - Ligation - PCR setup only		0
Post-PCR Room Assay steps: - PCR thermal cycling and purification - Fragmentation - Labeling - Hybridization - Washing and staining - Scanning		

Pre-PCR Clean Room

The Pre-PCR Clean Room is a low-copy DNA template lab, and must be free of PCR product (amplicons). Setup and major pieces of dedicated equipment required are shown in Figure 1. After entering the Post-PCR Room, do not re-enter the Pre-PCR Clean Room without first showering and changing into freshly laundered clothes.



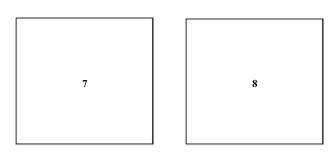


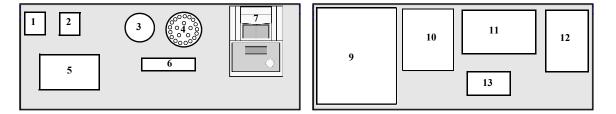
Figure 1 Pre-PCR Clean Room

Equipment Shown

- 1. Vortexer
- 2. Microfuge
- 3. Pipettes on stand
- 4. Ice bucket
- 5. Thermal cycler
- 6. Plate centrifuge
- 7. Freezer, -25 to -15°C
- 8. Refrigerator

Post-PCR Room

Setup and major pieces of dedicated equipment required are shown in Figure 2.



Equipment Shown

- 1. Vortexer
- 2. Microfuge
- 3. Pipettes on stand
- 4. Vortexer (with foam tube adaptor)
- 5. Ice bucket
- 6. Magnetic stand
- 7. Thermal cycler
- 8. GeneChip[™] Hybridization Oven 450
- 9. Refrigerated plate centrifuge
- 10.Microcentrifuge
- 11. Spectrophotometer, UV/VIS, single or multichannel
- 12.Gel Imager
- 13. Electrophoresis gel box
- 14. Computer, monitor, keyboard
- 15.Fluidics Station
- 16.Scanner
- 17.Refrigerator
- 18.Freezer

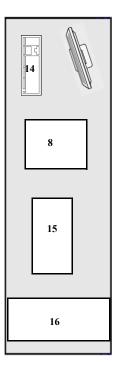


Figure 2 Post-PCR Room

18

17

One room with single-direction workflow

One room with two distinctly separated areas: *Pre-PCR Clean Area* and *Post-PCR Area* (see Figure 3).

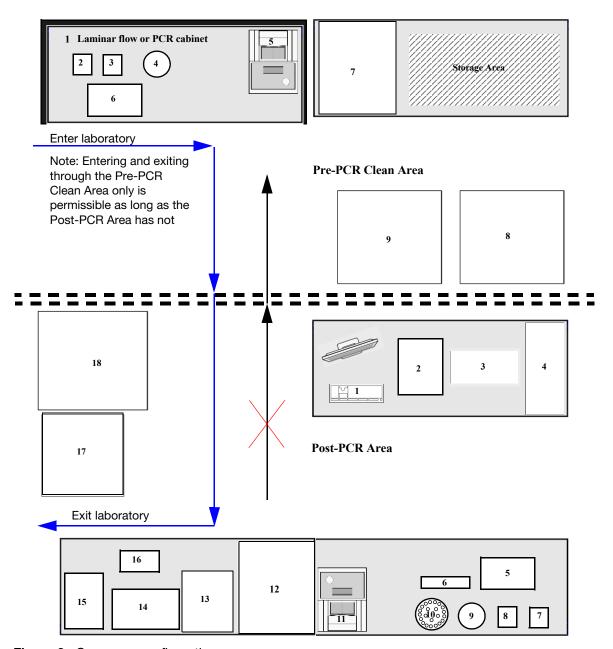


Figure 3 One room configuration

Equipment in Pre-PCR Clean Area

Setup and major pieces of dedicated equipment required are shown in Figure 3.

- 1. Laminar flow cabinet or PCR cabinet
- 2. Vortexer
- 3. Microfuge
- 4. Pipettes on stand
- 5. Ice bucket with ice
- 6. Thermal cycler
- 7. Plate centrifuge
- 8. Freezer, -25 to -15°C
- 9. Refrigerator

Equipment in Post-PCR Area

Setup and major pieces of dedicated equipment required are shown in Figure 3.

- 1. Vortexer
- 2. Microfuge
- 3. Pipettes on stand
- 4. Vortexer (with foam tube adaptor)
- 5. Ice bucket
- 6. Magnetic stand
- 7. Thermal cycler
- 8. GeneChip[™] Hybridization Oven 450
- 9. Refrigerated plate centrifuge
- 10. Microcentrifuge
- 11. Spectrophotometer, UV/VIS, single or multichannel
- 12. Gel imager
- 13. Electrophoresis gel box
- 14. Computer, monitor, keyboard
- 15. Fluidics station
- 16. Scanner
- 17. Refrigerator
- 18. Freezer, -25 to -15°C

GeneChip™ System 3000 installation

The system consists of the GeneChip Fluidics Station, scanner, barcode reader, computer, monitor, and keyboard. Install in the Post-PCR Room/Area on a bench that is free of possible vibration.



Materials required

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From Thermo Fisher Scientific

Equipment and software required

Table 2 Thermo Fisher Scientific equipment and software required

✓	Item	Qty	Cat. No.	
Equ	ipment			
	GeneChip [™] System 3000 - GeneChip [™] Scanner 3000 with AutoLoader - GeneChip [™] Fluidics Station 450 - Workstation	1	00-0210	
	GeneChip [™] Hybridization Oven 645		00-0331	
	Tubing, Silicone peristaltic for Fluidics Station 450		400110	
Sof	Software			
	Applied Biosystems [™] GeneChip Command Console Software		Version 3.2.2 or higher	
	Chromosome Analysis Suite (ChAS 2.1 or higher)		901394	

Optional equipment

 Table 3
 Optional Thermo Fisher Scientific equipment

✓	Item	Part Number
	GeneChip [™] System 3000Dx v.2 (running AMDS v1.1 or higher)	00-0334

CytoScan[™]Reagent Kit

Table 4 CytoScan[™] Assay Kit–24 Reaction Kit components (Cat. No. 901808)

Cap color	Component	Part No.	Storage
CytoSc	an™ Module 1: Pre-Lab Restriction and Ligation	904004	
	Nsp I	901718	–25°C to
	Nsp I Buffer	901719	–15°C
	100X BSA	901720	
	Adaptor,Nsp I	902702	
	DNA Ligase buffer	901722	
	DNA Ligase	901723	
\circ	REF DNA 103 (50 ng/µL)	900421	
0	PCR Primer	902674	
CytoSc	an™ Module 2: Pre-Lab TE Buffer and Water	904001	
•	Low EDTA TE Buffer	902979	2°C to 8°C
	Water, Nuclease-Free	902976	
CytoSc	an™ Module 3: Post-Lab Fragmentation, Labeling and Hybridization	904002	
	Fragmentation Reagent	902428	-25°C to
	Fragmentation Buffer	903001	–15°C
	TdT Enzyme	902675	
	TdT Buffer	902676	
	DNA Labeling Reagent	902677	
	Oligo Control Reagent	902678	
	Hyb. Buffer Part 1	901725	
	Hyb. Buffer Part 2	901726	
	Hyb. Buffer Part 3	901727	
	Hyb. Buffer Part 4	901728	
CytoSc	an™ Module 4: Post-Lab Stain, Holding Buffer, Beads and Water	904005	
•	Stain Buffer 1	901751	2°C to 8°C
	Stain Buffer 2	901752	
	Array Holding Buffer	901733	
\circ	Purification Beads	901807	
\circ	Water, Nuclease-free	901781	
CytoSc	an™ Module 5: Post-Lab Elution Buffer and Purification Wash Buffer	903000	15°C to 30°C
0	Elution Buffer	901738	
\circ	Purification Wash Buffer	901372	
0	Wash A	901680	15°C to 30°C
0	Wash B	901681	

Array and reagent kit bundles

 Table 5
 Applied Biosystems array and reagent bundles available

Bundle name	Items	Cat. No.
CytoScan™ HD Kit Plus 24	CytoScan HD Arrays+ CytoScan Reagent Kit + CytoScan Amplification Kit, sufficient for 24 reactions	905824
CytoScan HD Kit Plus 96	CytoScan HD Arrays + CytoScan reagent kit + CytoScan Amplification Kit, sufficient for 96 reactions	905896
CytoScan™ 750K Kit Plus 24	CytoScan 750K Arrays+ CytoScan Reagent Kit + CytoScan Amplification Kit, sufficient for 24 reactions	905924
CytoScan 750K Plus 96	CytoScan 750K Arrays+ CytoScan Reagent Kit + CytoScan Amplification Kit, sufficient for 96 reactions	905996
CytoScan™ HD Kit	CytoScan HD Arrays + CytoScan Reagent Kit sufficient for 24 reactions	901835
CytoScan™ 750K Kit	CytoScan 750K Arrays + CytoScan Reagent Kit sufficient for 24 reactions	901859

Other equipment

Table 6 GeneChip[™] Hybridization Oven 645

✓	Item	Specifications
	Input Voltage	100–120 VAC, 5A maximum or 220–240 VAC, 2.5A maximum 50–60 Hz
	Rotisserie Rotation Speed	10-80 RPM, programmable to 1 RPM
	Oven Temperature Set Point Programmable Range	30–70°C, programmable to 0.1°C
	Time to Temperature	30 minutes from ambient to 60°C
	Oven Temperature Accuracy	±1.0°C from 35–60°C
	Communications (optional)	Nine-pin RS-232 port, 9600 Baud rate Monitors and reports oven temperature, rotisserie rotation rate, and oven status

From other suppliers

Reagents required but not provided.

Table 7 Reagents from other suppliers, required

✓	Item
	Bleach, Sodium Hypochlorite prepared from a concentrate solution without additives at final working concentration of 0.615% (v/v)
	PCR gel DNA ladder: 50 to 2000 bp
	Fragmentation gel DNA ladder: 25 to 2600 bp
	Ethanol, absolute
	Loading buffer solution
	TBE buffer 5X

Consumables required but not provided

Table 8 Consumables from other suppliers, required

✓	Item
	Adhesive films, clear, PCR-certified, 96-well plates
	Adhesive label dot, 1/2-inch and 3/8-inch
	Agarose gel, 2%
	Agarose gel, 4%
	Microcentrifuge tubes, nuclease-free, sterile, 1.5 mL polypropylene
	Microcentrifuge tubes, nuclease-free, sterile, 2.0 mL polypropylene
	Microcentrifuge tubes, nuclease-free, sterile, non-stick, 1.5 mL amber polypropylene
	Microcentrifuge tubes, nuclease-free, sterile, non-stick, 1.5 mL blue polypropylene
	Microcentrifuge tubes, nuclease-free, sterile, non-stick, 1.5 mL natural polypropylene
	Microcentrifuge tubes, nuclease-free, sterile, non-stick, 50 mL polypropylene microcentrifuge tubes
	Pipette tips with aerosol barriers, 20 μL, 200 μL, and 1000 μL
	Plate, OD for UV, 96-well (required only if using microplate spectrophotometer)
	Plates, unskirted PCR with a maximum volume of 330 μL
	Reagent reservoir, 25 mL
	Tube strips, nuclease-free, sterile, 8-well, 0.2 mL polypropylene

Pre-PCR Clean Area equipment required but not provided

Table 9 Pre-PCR Clean Room equipment, required

✓	Item		
	If assay is to be performed in one room: • Laminar Flow Cabinet, 6 foot or		
	PCR Cabinet		
	Benchtop Cooler, with the capacity to hold 8 to 32 tubes (1.5 mL) and ability to maintain temperature below –15°C for 2 hours.		
	Centrifuge, plate, multipurpose, 330 µL capacity		
	Cooling chamber, double-block, with the capacity to hold 96 well plates with a maximum volume capacity of 330 µL		
	Freezer, -25 to -15°C; deep freeze; manual defrost; 17 cu ft		
	Microfuge (for tubes and strip tubes)		
	Pipettors: • 12-channel, 2–20 μL • 12-channel, 20–200 μL • single-channel, 100–1000 μL • single-channel, 2–20 μL • single-channel, 20–200 μL		
	Rectangular Ice Tray Large – 9L (16 x 13 in, 41 x 33cm)		
	Storage Racks, Tube, 96-well		
	Thermal cycler: capable of holding 200 μ L volume and 96-well plate; heat block capable of holding temperature of 4–99.9°C; temperature accuracy of ± 0.25 °C (at 35–99.9°C); average heating and cooling rate of 2.6°C per second; thermal uniformity of ± 0.5 °C.		
	Vortexer, 60 Hz, 75 W, 600-3200 RPM		

Post-PCR Room equipment required but not provided

Table 10 Post-PCR Room equipment required

✓	Item		
	Adhesive film applicator (hard plastic)		
	Anti-vibration pad, used with vortexer to prevent movement during operation		
	Cooler, benchtop, with capacity to hold 8 to 32 tubes (1.5 mL) and maintain temperature below –15°C for 2 hours		
	Cooling chamber, double-block, with capacity to hold 96-well plates with a maximum volume capacity of 330 µL		
	Electrophoresis supplies		
	Freezer, -25 to -15°C; deep freeze; manual defrost; 17 cu ft		
	Gel imager		
	Magnetic rack with magnet on the side and capable of holding 8–12 tubes of 1.5–2-mL capacity		
	Microcentrifuge, non-refrigerated with capacity to hold 24 tubes and maximum rotation speed of 16,200 ${\it xg}$		
	Microfuge (for tubes and strip tubes)		
	Microtube foam insert		
	Pipettors: • 12-channel, 100–1200 μL • 12-channel, 2–20 μL • 12-channel, 20–200 μL • single-channel, 100–1000 μL • single-channel, 2–20 μL • single-channel, 2–20 μL		
	Plate centrifuge, refrigerated multipurpose, plate carriers for 4 x 96-well assay plates		
	Platform Head, 6-inch, for microtube foam insert		
	Rectangular ice tray, large - 9L (16 x 13 in; 41 x 33 cm)		
	Refrigerator, 2–8°C, 6 cu ft		
	Spectrophotometer, UV/VIS, single or multichannel		
	Storage racks, tube, 96-well		
	Thermal cycler: capable of holding 200 μ L volume and 96-well plate; heat block capable of holding temperature of 4–99.9°C; temperature accuracy of ± 0.25 °C (at 35–99.9°C); average heating and cooling rate of 2.6°C per second; thermal uniformity of ± 0.5 °C.		
	Vortexer, 60 Hz, 75 W, 600-3200 RPM		

Thermal cycler programs

Before you begin processing samples, enter and save these programs into the appropriate thermal cycler(s).

Use only calibrated thermal cyclers. We recommend that thermal cyclers be serviced at least once per year to ensure that they are operating within the manufacturer's specifications. The thermal cycler programs listed in Table 11 and Table 12 are used in this protocol. Enter and store these programs on the appropriate thermal cycler in the Pre-PCR Clean Area and the Post-PCR Area.

Pre- and Post-PCR programs

Table 11 Pre-PCR clean room

Number of thermal cyclers required	Program name
	CytoScan Digest
1	CytoScan Ligate

Table 12 Post-PCR room

Number of thermal cyclers required	Program name
1	CytoScan PCR Assay
	CytoScan Assay Fragment
·	CytoScan Assay Label
	CytoScan Assay Hyb

Program your thermal cyclers

CytoScan Digest

 Table 13
 CytoScan Digest thermal cycler program

Temperature	Time
37°C	2 hours
65°C	20 minutes
4°C	∞

CytoScan Ligate

 Table 14
 CytoScan Ligate thermal cycler program

Temperature	Time
16°C	3 hours
70°C	20 minutes
4°C	∞

CytoScan PCR

You must use thermal cyclers with silver or gold-plated silver blocks. Do not use thermal cyclers with aluminum blocks.

Ramp speed: MaxVolume: 100 µL

Table 15 CytoScan PCR thermal cycler program

Temperature	Time	Cycles
94°C	3 minutes	1X
94°C	30 seconds)
60°C	45 seconds	> 30X
68°C	15 seconds	•
68°C	7 minutes	1X
4°C	Hold (for up to 24 hrs)	

CytoScan Fragment

 Table 16
 CytoScan Fragment thermal cycler program

Temperature	Time
37°C	35 minutes
95°C	15 minutes
4°C	∞

CytoScan Label

 Table 17
 CytoScan Label thermal cycler program

Temperature	Time
37°C	4 hr
95°C	15 minutes
4°C	∞

CytoScan Hyb

Table 18 CytoScan Hyb thermal cycler program

Temperature	Time
95°C	10 minutes
49°C	∞

Consumables required from other suppliers

Table 19 Consumables required from other suppliers

✓	Item	Vendor	Cat. No.
	MicroAmp Clear Adhesive Film for 96-well plates	Applied Biosystems	4306311
	Pipette tips, 20 μL filter tips	Rainin	GP-L10F
	Pipette tips, 200 μL filter tips	Rainin	GP-L200F
	Pipette tips, 1000 μL filter tips	Rainin	GP-L1000F
	Plates, 96-well unskirted PCR	Bio-Rad	MLP-9601
	Plate, OD for UV spec, 96-well (required only if using microplate spectrophotometer)	E & K Scientific	EK-25801
	Reagent Reservoir, 25 mL	Diversified Biotech	RESE-3000
	TBE Gel, 4%, BMA Reliant precast	Lonza Group LTD	54929
	TBE Gel, 2%, BMA Reliant precast	Lonza Group LTD	54939
	TBE for electrophoresis	Any vendor or house made	
	TrackIt [™] 25 bp DNA Ladder	Life Technologies	10488-022
	Tough-Spots, 1/2"	Diversified Biotech	Spot 2200
	Tough-Spots, 3/8"	USA Scientific	9185-0000
	Tube, Safe-Lock Tube 1.5 mL, Amber	Eppendorf	022363221
	Tube, Safe-Lock Tube 1.5 mL, Blue	Eppendorf	022363247
	Tube, Safe-Lock Tube 1.5 mL, Natural	Eppendorf	022363204
	Tube, centrifuge 50 mL	VWR	93000-036
	Tube, centrifuge 15 mL	VWR	21008-103
	Tube strips, 8-well, 0.2 mL	VWR	20170-004

Symbols

Symbol/ Label	Statement/Meaning
REF	Part/Catalog Number
LOT	Lot Number

Symbol/ Label	Statement/Meaning
\square	Expiration Date YYYY-MM Kit will expire on the last day of the month.
ł	Temperature Limitation

