

# pCR<sup>™</sup>8/GW/T0P0<sup>®</sup> TA Cloning<sup>®</sup> Kits

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#### Description

Instructions are provided to TOPO<sup>®</sup> Clone your PCR product into the TOPO<sup>®</sup> vector and transform the reaction into chemically-competent *E. coli* cells. For transformation of electrocompetent *E. coli* cells, a map of the TOPO<sup>®</sup> vector, a diagram of the multiple cloning site, and a manual, refer to **www.lifetechnologies.com** or contact Technical Support.

#### **Produce PCR Products**

Produce PCR products using *Taq* polymerase and your own protocol. End the PCR reaction with a final 7–30 minute extension step.

### TOPO<sup>®</sup> Cloning Reaction

1. Set up the following 6 µL TOPO<sup>®</sup> Cloning reaction:

| Reagent                  | Amount                             |
|--------------------------|------------------------------------|
| Fresh PCR Product        | 0.5–4 μL                           |
| Salt Solution            | 1 μL                               |
| Sterile Water            | add to a total volume of 5 $\mu$ L |
| TOPO <sup>®</sup> Vector | 1 μL                               |
| Final Volume             | 6 μL                               |

- 2. Mix gently and incubate for 5 minutes at room temperature.
- 3. Place tube on ice. Proceed to **Transformation and Analysis**.



## **Transformation and Analysis**

Protocols to transform chemically-competent cells and to analyze positive clones are provided below. Use the **Rapid One Shot**<sup>®</sup> **Chemical Transformation** protocol if you wish to obtain transformants as quickly as possible. To transform electrocompetent cells, refer to the pCR<sup>®</sup>8/GW/TOPO<sup>®</sup> TA Cloning<sup>®</sup> Kit manual for instructions.

### One Shot<sup>®</sup> Chemical Transformation

- 1. Thaw on ice 1 vial of One Shot<sup>®</sup> *E. coli* cells for each transformation.
- 2. Add 2 μL of the TOPO<sup>®</sup> Cloning reaction from step 3 on page 1 to a vial of One Shot<sup>®</sup> *E. coli* and mix gently.
- 3. Incubate the tube on ice for 5–30 minutes.
- 4. Heat-shock the cells for 30 seconds at 42°C without shaking.
- 5. Add 250 µL of room temperature S.O.C. Medium to the cells.
- 6. Cap the tubes and shake at 37°C for 1 hour.
- Spread 10–50 μL from each transformation on prewarmed LB plates containing 100 μg/mL spectinomycin.
- 8. Incubate plates overnight at 37°C.
- An efficient TOPO<sup>®</sup> Cloning reaction should produce several hundred colonies, with cloning efficiency > 95%. Pick ~10 colonies and proceed to Analyze Positive Clones.

## Transformation and Analysis, Continued

## Rapid One Shot<sup>®</sup> Chemical Transformation

- 1. Thaw **on ice** 1 vial of One Shot<sup>®</sup> *E. coli* for each transformation.
- 2. Add 4 μL of the TOPO<sup>®</sup> Cloning reaction from step 3 on page 1 to a vial of One Shot<sup>®</sup> *E. coli* and mix gently.
- 3. Incubate on ice for 5 minutes.
- 4. Spread 50 μL from each transformation on **prewarmed** LB plates containing 100 μg/mL spectinomycin. Incubate plates overnight at 37°C.
- An efficient TOPO<sup>®</sup> Cloning reaction should produce several hundred colonies, with cloning efficiency > 95%. Pick ~10 colonies and proceed to Analyze Positive Clones.

### Analyze Positive Clones

- Culture the 10 colonies overnight in LB medium containing 100 μg/mL spectinomycin.
- Isolate plasmid DNA using your method of choice. To prepare plasmid DNA for sequencing, use the PureLink<sup>®</sup> HQ Mini Plasmid Purification Kit (Cat. no. K2100-01).
- 3. Analyze the plasmid by restriction digest or sequencing to confirm the presence and orientation of the insert. Use the GW1 and GW2 primers included in the kit for sequencing.
- 4. *Optional:* Transfer your gene into a destination vector of choice using Gateway<sup>®</sup> Technology. Refer to the pCR<sup>®</sup>8/GW/ TOPO<sup>®</sup> TA Cloning<sup>®</sup> Kit manual for guidelines.

## **Purchaser Notification**

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