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OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION		
EOSIN METHYLENE BLUE AGAR (LEVINE) (CM0069)		

EOSIN METHYLENE BLUE AGAR (LEVINE)

CM0069

Typical Formula*

Peptone	grams per litre	10.0
Lactose		10.0
Di-potassium hydrogen phosphate		2.0
Eosin Y		0.4
Methylene blue		0.06
Agar		15.0

* adjusted as required to meet performance standards

Directions

Suspend 37.5g in 1 litre of distilled water. Bring to the boil to dissolve completely. Sterilize by autoclaving at 121°C for 15 minutes. Cool to 50°C. Mix well to ensure even dispersion of the medium in order to oxidize the methylene blue, which is an essential part of this medium, and pour into sterile Petri dishes.

Physical Characteristics

Purple, grey or pink free-flowing powder
 Colour on reconstitution - dark purple
 Moisture level - less than 7%
 pH 6.8 ± 0.2 at 25°C
 Clarity - clear
 Gel strength - firm, comparable to 15.0g/litre of agar

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Microbiological Tests Using Optimum Inoculum Dilution

Control Media: Tryptone Soya Agar or Sabouraud Dextrose Agar, as appropriate

Reactions after incubation at 37°C for 24 hours

Medium is challenged with 10-100 colony-forming units

<i>Escherichia coli</i>	ATCC® 10536	0.5-2mm purple colonies, with or without metallic green sheen
<i>Escherichia coli</i>	ATCC® 8739	0.5-2mm purple colonies, with or without metallic green sheen
<i>Enterobacter aerogenes</i>	ATCC® 13048	1-2mm purple, mucoid colonies, no sheen
<i>Pseudomonas aeruginosa</i>	ATCC® 9027	0.5-3mm translucent, colourless colonies

A satisfactory result is represented by recovery of positive strains equal to or greater than 70% of the control medium.

Some strains of *Escherichia coli* may produce little or no sheen with single colonies.

Escherichia coli are also tested using high inocula.

Reactions after incubation at 37°C for 24 hours

Medium is challenged with 1E+04 to 1E+06 colony-forming units

Inoculation using diminishing sweep technique

<i>Escherichia coli</i>	ATCC® 10536	0.5-2mm purple, green metallic colonies
<i>Escherichia coli</i>	ATCC® 8739	0.5-2mm purple, green metallic colonies
<i>Escherichia coli</i>	ATCC® 25922	0.5-2mm purple, green metallic colonies

Inoculation using diminishing sweep technique

<i>Staphylococcus aureus</i>	ATCC® 6538	No growth to confluent micro-colonies, no sheen
<i>Staphylococcus aureus</i>	ATCC® 25923	No growth to confluent micro-colonies, no sheen

Negative strains are inhibited or shall produce confluent micro-colonies.

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**Reactions after incubation at 37°C for 48 hours 10% CO₂ atmosphere
(for details, refer to Oxoid Manual - Atmosphere Generation Systems)**

Medium is challenged with 10-100 colony-forming units

Candida albicans ATCC® 10231 0.25-1mm grey, feather-edged colonies

A satisfactory result is represented by recovery of positive strains equal to or greater than 70% of the control medium.

Equivalent results are obtained after incubation at 35°C for 48 hours in 10% CO₂ atmosphere.


Tested in accordance with current CLSI M22 A

Medium is challenged with 10-100 colony-forming units

<i>Escherichia coli</i>	ATCC® 25922	0.5-2mm purple, green metallic colonies
<i>Salmonella typhimurium</i>	ATCC® 14028	0.5-1mm grey, translucent colonies
<i>Enterococcus faecalis</i>	ATCC® 29212	Pinpoint, colourless colonies with no sheen

A satisfactory result is represented by recovery of positive strains equal to or greater than 70% of the control medium.

For *Enterococcus faecalis* ATCC®29212, a satisfactory result is represented by recovery of positive strains equal to or greater than 40% of the control medium.

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Revision History

Section / Step	Description of Change	Reason for Change	Reference
Entire Document	Addition of CLSI specification	Change control	BT-CC-1502