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OXOID QUALITY ASSURANCE PRODUCT SPECIFICATION		
GIOLITTI-CANTONI BROTH CM0523		

## GIOLITTI-CANTONI BROTH

CM0523

### Typical Formula\*

Tryptone	grams per litre	10.0
'Lab-Lemco' powder		5.0
Yeast extract		5.0
Lithium chloride		5.0
Mannitol		20.0
Sodium chloride		5.0
Glycine		1.2
Sodium pyruvate		3.0

\* adjusted as required to meet performance standards

### Directions

Suspend 54.2g in 1 litre of distilled water. Heat until dissolved completely. Mix well and distribute 19ml volumes into final containers. Sterilize by autoclaving at 121°C for 15 minutes. Cool rapidly and aseptically add 0.2ml of 1% v/v potassium tellurite solution.\*\*

To comply with ISO 6888-3:2003, sorbitan mono-oleate should also be added prior to autoclaving to give a final concentration of 0.1% v/v.

\*\* A 1% v/v potassium tellurite solution may be obtained by adding 1ml Potassium Tellurite 3.5% Solution (SR0030) to 2.5ml of sterile distilled water.

### Physical Characteristics

Straw, free-flowing powder  
 Colour on reconstitution - straw 2-3  
 Moisture level - less than or equal to 7%  
 pH -  $6.9 \pm 0.2$  at 25°C  
 Clarity - clear

### Microbiological Tests Using Optimum Inoculum Dilution

Control Medium: Tryptone Soya Agar and Baird-Parker (ISO) Agar + Egg Yolk Tellurite Emulsion (CM1127 + SR0054)

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#### Reactions after incubation at 37 ± 2°C for 24 ± 2 hours under anaerobic conditions

Tested with the addition of 0.01% v/v potassium tellurite

Medium is challenged with 10-100 colony-forming units/19ml

*Staphylococcus aureus* ATCC® 6358P Black medium and black button at base

A satisfactory result is represented by visible growth and blackening of the medium.


#### Testing performed in accordance with ISO11133:2014

##### Inoculation with mixed cultures

Inoculate 19ml quantities of medium to achieve 10–100 colony-forming units/19ml (cfu/19ml) of *Staphylococcus aureus* and greater than 1E+03 cfu/19ml of *Escherichia coli*. Incubate broths anaerobically at 37 ± 2°C for 48 ± 2 hours. After incubation, subculture onto Baird-Parker (ISO) Agar (CM1127) + Egg Yolk Tellurite Emulsion (SR0054) and incubate plates at 37 ± 2°C for 24 to 48 hours.

#### Reactions after incubation at 37 ± 2°C for 48 ± 2 hours under anaerobic conditions

<i>Staphylococcus aureus</i>	ATCC® 25923	WDCM00034	0.25-3mm black colonies with clear zone
+ <i>Escherichia coli</i>	ATCC® 8739	WDCM00012	No growth
<i>Staphylococcus aureus</i>	ATCC® 25923	WDCM00034	0.25-3mm black colonies with clear zone
+ <i>Escherichia coli</i>	ATCC® 25922	WDCM00013	No growth
<i>Staphylococcus aureus</i>	ATCC® 6538	WDCM00032	0.25-3mm black colonies with clear zone
+ <i>Escherichia coli</i>	ATCC® 8739	WDCM00012	No growth
<i>Staphylococcus aureus</i>	ATCC® 6538	WDCM00032	0.25-3mm black colonies with clear zone
+ <i>Escherichia coli</i>	ATCC® 25922	WDCM00013	No growth

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A satisfactory result is represented by recovery of >10 cfu of *Staphylococcus aureus* on Baird-Parker (ISO) Agar (CM1127) + Egg Yolk Tellurite Emulsion (SR0054).


### Inoculation with pure cultures

Inoculate 19ml quantities of medium to achieve 1E+03 to 1E+04 colony-forming units/19ml (cfu/19ml) of *Escherichia coli*. Incubate broths anaerobically at 37 ± 2°C for 48 ± 2 hours. After incubation, subculture onto Tryptone Soya Agar (CM0131) and Baird-Parker (ISO) Agar (CM1127) + Egg Yolk Tellurite Emulsion (SR0054). Incubate plates at 37 ± 2°C for 24 ± 2 hours.

### Reactions after incubation at 37 ± 2°C for 48 ± 2 hours

<i>Escherichia coli</i>	ATCC® 8739	WDCM00012	No growth (CM1127+SR0054)
<i>Escherichia coli</i>	ATCC® 8739	WDCM00012	No growth or cream colonies + no blackening to slight greying in tube (CM0131)
<i>Escherichia coli</i>	ATCC® 25922	WDCM00013	No growth (CM1127+SR0054)
<i>Escherichia coli</i>	ATCC® 25922	WDCM00013	No growth or cream colonies + no blackening to slight greying in tube (CM0131)

Negative strains are inhibited or produce a negative diagnostic reaction (i.e. no blackening to slight greying in tube).

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### Revision History

Section / Step	Description of Change	Reason for Change	Reference
Creation of ISO11133 section	Update to include testing of ISO11133:2014	Change control	BT-CC-1397
Inoculation with pure cultures	Update to allow for typical product limitation of slight greying in tubes	Change control	BT-CC-2381